

# Draft Genome Sequence of *Pasteurella multocida* Isolate P1062, Isolated from Bovine Respiratory Disease

## Juan E. Abrahante,<sup>a</sup> Samuel S. Hunter,<sup>b</sup> Samuel K. Maheswaran,<sup>a</sup> Melissa J. Hauglund,<sup>c\*</sup> Fred M. Tatum,<sup>c</sup> Robert E. Briggs<sup>c</sup>

Department of Veterinary and Biomedical Sciences, University of Minnesota, St. Paul, Minnesota, USA<sup>a</sup>; Institute for Bioinformatics and Evolutionary Studies, University of Idaho, Moscow, Idaho, USA<sup>b</sup>; National Animal Disease Center, Agricultural Research Service, U.S. Department of Agriculture, Ames, Iowa, USA<sup>c</sup>

\* Present address: Melissa J. Hauglund, Boehringer Ingelheim Vetmedica, Inc., Fort Dodge, Iowa, USA.

## Here, we report the draft genome of *Pasteurella multocida* isolate P1062 recovered from pneumonic bovine lung in the United States in 1959.

Received 11 September 2015 Accepted 15 September 2015 Published 22 October 2015

Citation Abrahante JE, Hunter SS, Maheswaran SK, Hauglund MJ, Tatum FM, Briggs RE. 2015. Draft genome sequence of *Pasteurella multocida* isolate P1062, isolated from bovine respiratory disease. Genome Announc 3(5):e01254-15. doi:10.1128/genomeA.01254-15.

Copyright © 2015 Abrahante et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Robert E. Briggs, robert.briggs@ars.usda.gov.

asteurella multocida, a Gram-negative bacterium, is a pathogen which affects many species of agricultural importance including cattle, sheep, goats, swine, and poultry. In cattle it is considered an opportunistic pathogen associated with respiratory disease in beef and dairy cattle (1). The bacterium is also found as a commensal in the respiratory tract of apparently healthy cattle. In the United States, P. multocida serotype A:3 is the most common isolate associated with this disease (1). The P1062 strain of P. multocida serotype A:3 presented in this study originated from the pneumonic lung of a Holstein-Friesian calf that died with respiratory disease in 1959 (2). It is pathogenic to cattle and experimental pneumonic pasteurellosis can be reproduced by intra-pulmonic instillation with logarithmic-phase culture of the isolate. Antimicrobial resistance among bacterial bovine respiratory disease pathogens is of growing concern (3, 4), and multidrug-resistant isolates of Pasteurella multocida and Mannheimia haemolytica were recently sequenced (5, 6). The genome sequencing of this isolate was undertaken to facilitate the study of species specificity among known virulent P. multocida and to provide insight into the acquisition of antimicrobial resistance.

The genome sequencing of *P. multocida* was achieved using 3 platforms: Roche 454 GS FLX Titanium resulting in 30-fold coverage; Illumina GA IIx resulting in 13-fold coverage; and PacBio RS resulting in 9-fold coverage. Illumina reads were used to error-correct the PacBio reads using CLC-Genomics Workbench v6.0.2. A hybrid assembly using the CLC software was performed and the resultant contigs were aligned to an optical map (OpGen, MapSolver software, Gaithersburg, Maryland) to confirm the assembly and generate a single scaffold. Reiterative alignments of the 454 and corrected PacBio reads >500 bp against the scaffold, using the CLC software, failed to close the remaining 5 gaps in the single scaffold. The P1062 genome scaffold consists of 2.70 Mb and 5 contigs with a total length of 2.51 Mb, a G+C content of 40.3%,  $N_{50}$  of 140,532 bp, and 100% of contigs >500 bp.

Annotation of the genome was accomplished with the NCBI Prokaryotic Genome Annotation Pipeline revision 2.2. The genome contained a total of 2,456 genes including 2,324 predicted protein-encoding genes, 58 pseudogenes, 19 rRNA, and 53 tRNA genes. Two CRISPR arrays were detected. In contrast to the multiresistant *P. multocida* isolate 36950 (7), an intact tRNA<sup>leu</sup> is present at the site of ICEPmu1 integration. BLAST analysis revealed no significant homology in the P1062 chromosome with ICEPmu1. The determination of genes involved in host specificity of *P. multocida* will require additional analysis.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ASZP00000000. The version described in this paper is the second version, ASZP02000000.

## ACKNOWLEDGMENTS

This work was supported by the Biotechnology Research and Development Corporation, Peoria, Illinois, USA.

The mention of a trade name, proprietary product, or specific equipment does not constitute a guarantee or warranty by the USDA and does not imply approval to the exclusion of other products that might be suitable.

## REFERENCES

- Dabo SM, Taylor JD, Confer AW. 2007. Pasteurella multocida and bovine respiratory disease. Anim Health Res Rev 8:129–150. http://dx.doi.org/ 10.1017/S1466252307001399.
- Heddleston KL, Reisinger RC, Watco LP. 1962. Studies on the transmission and etiology of bovine shipping fever pneumonia. Am J Vet Res 23: 548–553.
- Watts JL, Sweeney MT. 2010. Antimicrobial resistance in bovine respiratory disease pathogens: measures, trends, and impact on efficacy. Vet Clin North Am Food Anim Pract 26:79–88. http://dx.doi.org/10.1016/ j.cvfa.2009.10.009.
- Katsuda K, Kohmoto M, Mikami O. 2013. Relationship between serotype and the antimicrobial susceptibility of *Mannheimia haemolytica* isolates collected between 1991 and 2010. Res Vet Sci 94:205–208. http:// dx.doi.org/10.1016/j.rvsc.2012.09.015.
- 5. Eidam C, Poehlein A, Brenner Michael G, Kadlec K, Liesegang H,

Brzuszkiewicz E, Daniel R, Sweeney MT, Murray RW, Watts JL, Schwarz S. 2013. Complete genome sequence of *Mannheimia haemolytica* strain 42548 from a case of bovine respiratory disease. Genome Announc 1(3): e00318-13. http://dx.doi.org/10.1128/genomeA.00318-13.

6. Brenner Michael G, Kadlec K, Sweeney MT, Brzuszkiewicz E, Liesegang H, Daniel R, Murray RW, Watts JL, Schwarz S. 2012. ICEPmu1, an integrative conjugative element (ICE) of *Pasteurella multocida*: analysis of

the regions that comprise 12 antimicrobial resistance genes. J Antimicrob Chemother 67:84–90. http://dx.doi.org/10.1093/jac/dkr406.

 Brenner Michael G, Kadlec K, Sweeney MT, Brzuszkiewicz E, Liesegang H, Daniel R, Murray RW, Watts JL, Schwarz S. 2012. *ICEPmu1*, an integrative conjugative element (ICE) of *Pasteurella multocida*: structure and transfer. J Antimicrob Chemother 67:91–100. http://dx.doi.org/ 10.1093/jac/dkr411.