## Research Article

# Children Living near a Sanitary Landfill Have Increased Breath Methane and *Methanobrevibacter smithii* in Their Intestinal Microbiota

Humberto Bezerra de Araujo Filho,<sup>1,2</sup> Mirian Silva Carmo-Rodrigues,<sup>2</sup> Carolina Santos Mello,<sup>1</sup> Lígia Cristina Fonseca Lahoz Melli,<sup>1,2</sup> Soraia Tahan,<sup>1</sup> Antonio Carlos Campos Pignatari,<sup>3</sup> and Mauro Batista de Morais<sup>1</sup>

<sup>1</sup> Division of Pediatric Gastroenterology, Universidade Federal de São Paulo, 598 Botucatu Street, Vila Clementino, 04023-062 São Paulo, SP, Brazil

<sup>2</sup> Centro Universitário FIEO, 300 Franz Voegeli Avenida, Vila Yara, 06020-190 Osasco, SP, Brazil

<sup>3</sup> Division of Medicine, Universidade Federal de São Paulo, 188 Leandro Dupret Street, Vila Clementino, 04025-010 São Paulo, SP, Brazil

Correspondence should be addressed to Mauro Batista de Morais; maurobmorais@gmail.com

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This study evaluated the breath  $CH_4$  excretion and concentration of *M. smithii* in intestinal microbiota of schoolchildren from 2 slums. One hundred and eleven children from a slum near a sanitary landfill, 35 children of a slum located away from the sanitary landfill, and 32 children from a high socioeconomic level school were included in the study. Real-time PCR was performed to quantify the *M. smithii nifH* gene and it was present in the microbiota of all the participating children, with higher (*P* < 0.05) concentrations in those who lived in the slum near the landfill ( $3.16 \times 10^7$  CFU/g of feces), comparing with the children from the slum away from the landfill ( $2.05 \times 10^6$  CFU/g of feces) and those from the high socioeconomic level group ( $3.93 \times 10^5$  CFU/g of feces). The prevalence of children who present breath methane was 53% in the slum near the landfill, 31% in the slum further away from the landfill and, 22% in the high socioeconomic level group. To live near a landfill is associated with higher concentrations of *M. smithii* in intestinal microbiota, comparing with those who live away from the landfill, regardless of their socioeconomics conditions.

## 1. Introduction

The human intestinal microbiota consists of a diverse group of microorganisms that play an important role in controlling the colonization of the gastrointestinal tract and the maturation and proliferation of intestinal cells as well as regulating the immunologic system, nutrition adsorption, and metabolism [1]. Methanogenic archaea are among the anaerobic microorganisms present in the human microbiota [2]. These archaeal species produce methane (CH<sub>4</sub>) by metabolizing hydrogen (H<sub>2</sub>) and carbon dioxide (CO<sub>2</sub>) gases, acetate, formate, and methanol [3, 4]. In humans, the predominant methanogenic archaea are *Methanobrevibacter smithii*, which can comprise up to 10% of all of the anaerobic organisms in the intestinal microbiota [4, 5]. To detect methanogenic archaea in humans, stool samples are cultured [5], examined using molecular biology techniques such as real-time PCR [6, 7] or indirectly by breath methane excretion assessed by gas chromatography [8].

Some studies have associated breath methane excretion with colorectal cancer [9], irritable bowel syndrome [10], diverticulosis [11], and chronic constipation with retentive fecal incontinence [12, 13]. However, the exact role of *M. smithii* in the development or outcome of these illnesses has not yet been established [14]. In the pediatric population, the presence of *M. smithii* in gut microbiota has not been carefully examined. There is little information about  $CH_4$  production, which is found almost exclusively in children with fecal retentive incontinence secondary to chronic constipation [12, 13, 15].

A previous study [16] reported that a large proportion of children living in a slum near a sanitary landfill were breath  $CH_4$  producers. In this slum, the environmental concentration of  $CH_4$  was higher than in locations away from this landfill. The proportion of methane-producing children in this slum was higher than that found in areas with good environmental conditions and was not associated with chronic constipation [16].

Based on these results, the purpose of this study was to evaluate the relationship between living near a sanitary landfill, the socioeconomic and environmental conditions, and the presence of *M. smithii* in the microbiota of children. We evaluated the socioeconomic and environmental conditions, the breath  $CH_4$  excretion, and the concentration of *M. smithii* in the intestinal microbiota of children living in a slum near a sanitary landfill, in a different slum away from the landfill and students from a high socioeconomic level school.

#### 2. Material and Methods

2.1. Study Design. This was a community-based, crosssectional study of children aged 6 to 11 years living in three different socioenvironmental conditions. The study included 111 children living in a slum approximately 50 meters distant from Osasco's sanitary landfill, São Paulo, Brazil, representing approximately 9% of the children in this slum within the age range of the study. The control groups were composed of 35 children from a slum approximately 7.5 kilometers distant from the landfill, representing approximately 11% of the children in this slum within the age range of this study and 32 children from a high socioeconomic level school in the same city, which corresponds to approximately 5% of total students in the age range of this study.

The inclusion criteria considered were age between 6 and 11 years, absence of diarrhea for at least 30 days, nonuse of antibiotics during the 15 days prior to the breath test, and absence of clinical evidence that would characterize serious illnesses such as cardiopathy, nephropathy, type 1 diabetes, or neuropathy. The parents or guardians of children enrolled in the study signed a term of free and informed consent.

2.2. Socioeconomic Questionnaire. The socioeconomic characteristics were evaluated through interviews with the parents or guardians using a socioeconomic questionnaire. The variables analyzed were family income, economic class, mother's schooling, demand for health services (public and private), family density, living conditions, and basic sanitation. The division of the families into social classes was performed using the Brazil Economic Classification Criteria.

2.3. Breath  $CH_4$  Dosage. Breath samples were collected from patients after an overnight fast. Moreover, a mouthwash

followed by teeth brushing was performed before breath collection. End-expiratory breath samples were collected for testing using a GaSampler system (QuinTron Instrument, Milwaukee, Wisconsin, USA). This apparatus consists of a mouthpiece attached to two bags linked by a T-valve. The first 250 mL of expired air (dead space) enters into a polyvinyl bag, and the valve then automatically shunts, directing the subsequent expired air into the other gas-impermeable bag. The alveolar air sample thus obtained was transferred to a 20 mL plastic syringe with a stopcock. The samples were analyzed immediately with a gas chromatograph MicroLyzer model 12i (QuinTron Instrument, Wisconsin, USA), and the results were expressed in parts per million (ppm). The chromatograph was calibrated using a standard gas mixture containing 92 ppm of hydrogen and 54 ppm of methane (White Martins, Sao Paulo, Brazil).

A child was considered a methane producer if his or her breath methane concentration was greater than or equal to 3 ppm in relation to the methane in the environment [8, 13, 17]. Therefore, air samples were collected in the environment where the breath tests were performed for all three groups.

2.4. Stool Collection and DNA Extraction. The stool collection was performed by each child's parents using a clean container and following established guidelines, with the objective of securing bacterial DNA of sufficient quality and quantity. Approximately 1g of each stool sample was transferred to a microtube containing ASL buffer from a DNA extraction QIAamp Mini Stool Kit (Qiagen, Hilden, Germany) and then frozen at  $-20^{\circ}$ C until the DNA was to be extracted.

The bacterial genomic DNA was extracted according to the protocol recommended by the extraction kit manufacturer (Qiagen, Hilden, Germany). The purified DNA was diluted to a final volume of  $200 \,\mu$ L. The DNA concentration was quantified using a NanoDrop 1000 spectrophotometer (Thermo Scientific, Waltham, MA, USA). All of the DNA samples were diluted to a final concentration of  $20 \text{ ng}/\mu$ L and stored at  $-20^{\circ}$ C.

2.5. Real-Time PCR. In the real-time PCR reactions, a fragment of 151 base pairs (bp) of the M. smithii-specific gene nifH was used as the target [18]. All of the reactions were performed in duplicate with a final volume of  $10 \,\mu L$ containing 5  $\mu$ L of de Rotor-gene SYBR Green PCR Master Mix (Qiagen, Hilden, Germany), 0.2 µL each of the primers Mnif 202F and Mnif 353R (10 pmol/ $\mu$ L), 0.5  $\mu$ L of the DNA sample, and  $4.1 \,\mu\text{L}$  of DEPC-treated water (Qiagen, Hilden, Germany). The thermocycling was performed using a Rotorgene Q thermocycler (Qiagen, Hilden, Germany) with the following conditions: 95°C for 5 minutes followed by 40 cycles of 95°C for 10s and 60°C for 15s, a dissociation cycle for the melting curve of 95°C for 1 minute, and a melting curve program of 70-95°C with a gradual temperature increase of 1°C/s. As a negative control, a reaction containing all of the reagents except the DNA sample was included and its specificity was confirmed by sequencing and alignment using the BLAST system.

#### Archaea

	Slum near landfill group		Slum away from the landfill group		High socioeconomic group		$P^1$
	п	%	п	%	п	%	Г
Brick house	69 <sup>a</sup>	63.4%	11 <sup>b</sup>	31.4%	32 <sup>c</sup>	100.0%	< 0.001
Sewage network	$10^{a}$	9.2%	$4^{a}$	11.4%	32 <sup>b</sup>	100.0%	< 0.001
Piped water	60 <sup>a</sup>	55.5%	23 <sup>a</sup>	65.7%	32 <sup>b</sup>	100.0%	< 0.001
Regularized electricity	70 <sup>a</sup>	64.9%	$1^{b}$	2.9%	32 <sup>c</sup>	100.0%	< 0.001
Paved street	14 <sup>a</sup>	13.0%	$0^{\mathrm{a}}$	0.0%	32 <sup>b</sup>	100.0%	< 0.001
Backyard of land	39 <sup>a</sup>	36.1%	15 <sup>a</sup>	42.9%	$0^{\mathrm{b}}$	0.0%	< 0.001
Pets	52 <sup>a</sup>	48.1%	$8^{\mathrm{b}}$	22.9%	18 <sup>a</sup>	46.1%	0.019
Income per capita >1/2 MW	22 <sup>a</sup>	20.4%	2 <sup>a</sup>	5.7%	32 <sup>b</sup>	100.0%	< 0.001
Social class							
А	$0^{a}$	0.0%	$0^{\mathrm{a}}$	0.0%	9 <sup>b</sup>	26.5%	
В	$4^{a}$	3.7%	$0^{a}$	0.0%	20 <sup>b</sup>	58.8%	
С	67 <sup>a</sup>	62.0%	$17^{a}$	48.6%	5 <sup>b</sup>	14.7%	< 0.001
D	31 <sup>a</sup>	28.7%	$18^{a}$	51.4%	$0^{\mathrm{b}}$	0.0%	
E	6 <sup>a</sup>	5.6%	$0^{\mathrm{a}}$	0.0%	$0^{\mathrm{b}}$	0.0%	

TABLE 1: Socioeconomic and environmental conditions of children living in the three distinct areas.

<sup>1</sup>Chi-squared test: different letters on the line represent statistically significant differences (P < 0.05); MW = minimum wage.

TABLE 2: Prevalence of breath  $CH_4$  producers and children breath  $CH_4$  concentration (ppm) in the three distinct groups.

	Slum near the landfill	Slum away from the landfill	High socioeconomic group	Р
Breath CH <sub>4</sub> producer prevalence	53.1% (59/111) <sup>a</sup>	31.4% (11/35) <sup>b</sup>	21.9% (7/32) <sup>b</sup>	0.001 <sup>2</sup>
CH <sub>4</sub> in breath CH <sub>4</sub> producers	24 (18.0-35.0) <sup>a</sup>	17.0 (10.0–31.0) <sup>b</sup>	17 (11.0–19.0) <sup>b</sup>	$0.007^{1}$
Total breath CH <sub>4</sub>	$14 (0.0-25)^{a}$	2.0 (0.0–9.5) <sup>b</sup>	$0.0 (0.0-2.5)^{\rm b}$	$< 0.001^{1}$
	2 cl .			(7)

<sup>1</sup>Mann-Whitney test: the median and percentiles 25 and 75; <sup>2</sup>Chi-squared test; different letters on the lines represent statistically significant differences (P < 0.05).

The standard curve for all of the analyses was created by amplifying a TopoTA plasmid (Invitrogen) carrying a fragment of the reference gene previously amplified by conventional PCR. With the molecular mass of the plasmid and insert known, it is possible to calculate the copy number as follows: mass in Daltons (g/mol)= (size of double-stranded [ds] product in base pairs [bp])  $(330 \text{ Da} \times 2 \text{ nucleotides})$ [nt]/bp) [19]. Hence, the g/mol value divided by Avogadro's number equals the g/molecule value, which equals the copy number [19]. Knowing the copy number and concentration of plasmid DNA, the precise number of molecules added to subsequent real-time PCR runs can be calculated, thus providing a standard for specific copy number of genes quantification. The real-time PCR results were expressed as colony forming units/g of feces (CFU/g of feces), once M. smithii possesses 1 copy of the nifH gene per cell [20].

2.6. Statistical Analyses. For the statistical analysis of the numerical variables, either an ANOVA or the Kruskal-Wallis test complemented by the Dunn or Mann-Whitney test, when appropriate, was used. The frequencies and proportions were compared using Pearson's chi-square test ( $\chi^2$ ) and its partition or Fischer's exact test. The correlations were evaluated using the Spearman coefficient. The alpha error was established as 5%.

#### 3. Results

The socioeconomic and environmental data of the studied groups are shown in Table 1. The median age of the participating children was 8.0 years in the slum near the landfill, 8.4 years in the slum away from the landfill, and 8.1 in the high socioeconomic group, with no statistical differences between the groups (P = 0.317). A statistically significant difference was found among the 3 groups with respect to the type of housing and electrical energy supply. The frequency of a brick house and regularized electricity was higher in the slum near the landfill than in the slum away from the landfill. The presence of a sewage network, water supply, paved street, and earthen backyard as well as the income per capita and distribution of social classes was similar between the two slum groups, whereas the control high socioeconomic group was significantly better off to all of the evaluated aspects than either of the 2 slum groups.

The environmental air samples collected in the slum near the sanitary landfill, the slum away from the landfill, and the high socioeconomic school environment contained 9 ppm, 1 ppm, and 0 ppm of  $CH_4$ , respectively. The cut-off point from which a child was considered as  $CH_4$  producers in the slum near the landfill is 12 ppm of breath  $CH_4$ , in the slum away from landfill more than 4 ppm and above 3 ppm in the high socioeconomic group. Table 2 shows the prevalence of

$CH_4$ Producer	Slum near the landfill		Slum away from landfill		High socioeconomic group		D
	n	Concentration	п	Concentration	п	Concentration	1
Yes 5	59	$4.26 \times 10^7$	11	$1.42 \times 10^7$	7	$1.26 \times 10^{7}$	0.094 <sup>1</sup>
	57	$(1.41 \times 10^7 - 1.46 \times 10^8)$	11	$(7.97 \times 10^{6} - 7.39 \times 10^{8})$		$(5.32 \times 10^{5} - 3.94 \times 10^{7})$	
No 52	52	$1.64 \times 10^{7a}$	24	$5.63 \times 10^{5b}$	25	$2.06 \times 10^{5b}$	$< 0.001^{1}$
	52	$(2.83 \times 10^{6} - 5.42 \times 10^{7})$	24	$(6.13 \times 10^4 - 2.97 \times 10^6)$	25	$(3.11 \times 10^4 - 2.59 \times 10^6)$	
Total	111	$3.16 \times 10^{7a}$	35	$2.05 \times 10^{6b}$	32	$3.93 \times 10^{5b}$	$< 0.001^{1}$
		$(6.36 \times 10^{6} - 8.63 \times 10^{7})$	55	$(1.55 \times 10^{5} - 1.66 \times 10^{7})$		$(5.05 \times 10^4 - 5.54 \times 10^6)$	

TABLE 3: Comparison of *M. smithii* concentrations in CFU/g of feces, in accordance with breath CH<sub>4</sub> production in the three distinct groups.

<sup>1</sup>One-way ANOVA complemented by Dunn's Test, median and percentiles 25 and 75; Different letters in the same line represent statistically significant differences: P < 0.05.

methane producer's children and the median concentration of breath  $CH_4$  in all 3 groups. The median concentration of breath methane was higher in children living near the landfill than in the other two groups.

Using real-time PCR, *M. smithii* was detected in all of the stool specimens from the three groups. Table 3 presents the quantitative results of *M. smithii* concentrations in the fecal microbiota of  $CH_4$ -producers and non- $CH_4$  producers in the 3 studied groups. The total concentration of *M. smithii* in the feces of the children living in the slum near the sanitary landfill was higher than that found in the other two groups.

The correlation coefficient between the breath CH<sub>4</sub> concentration and the fecal concentration of *M. smithii* was +0.556 (P < 0.001) for the children from the slum near the sanitary landfill, +0.754 (P < 0.001) for the children living in the slum away from the sanitary landfill, and +0.464 (P = 0.007) for the children in the high socioeconomic group.

#### 4. Discussion

The real-time PCR results demonstrated the presence of M. *smithii* in the stools of all of the children who participated in this study. As we expected, the group of children living in the slum near the sanitary landfill had a higher concentration of M. *smithii* in their stools and greater prevalence of breath CH<sub>4</sub> producers that agrees with our previous study [16]. This result is also in agreement with methane concentration in the environment air; however the fecal M. *smithii* concentrations of the children living in a slum away from the landfill were similar to those observed in children living in better socioeconomic conditions.

*M. smithii* is difficult to grow *in vitro*; therefore, molecular methods that test for the presence of 16S rRNA genes and other *M. smithii*-specific genes, such as *nifH*, have become popular [6, 7, 21]. The successful use of the *nifH* gene in the detection of *M. smithii* in contaminated water has demonstrated the high specificity and sensibility of such an approach, making it a good target gene for identification and quantification [18, 21]. With the use of real-time PCR, another study quantified the presence of *M. smithii* in the microbiota of obese, normal, and anorexic adult patients, demonstrating the presence of *M. smithii* in approximately 80% of the samples with average concentrations varying from

 $9.78 \times 10^7$  to  $1.68 \times 10^8$  copies/g of feces [22]. Dridi et al. [6] tested for the presence of *M. smithii* in 700 stool samples from children and adults using real-time PCR and found that 95.7% of the samples were positive, with concentrations varying between  $1.09 \times 10^1$  and  $1.45 \times 10^{11}$  copies/g of feces. Stewart et al. [7] found concentrations of M. smithii ranging from  $7.45 \times 10^5$  to  $4.91 \times 10^7$  CFU/g of feces from 12 adults and 40 children's samples using real-time PCR assays. Our study found concentrations of *M. smithii* ranging from  $10^4$  to  $10^8$  CFU/g of stool, which is consistent with the data in the studies described above. On the other hand, Weaver et al. [11] found concentrations of M. smithii ranging from  $1.0 \times 10^7$  to  $3.0 \times 10^{10}$  CFU/g of feces from 130 adults before sigmoidoscopy, including individuals with normal colon, diverticulosis, inflammatory bowel disease, colon polyps, and colon cancer using culture methods.

*M. smithii* was evaluated on fecal microbiota through molecular methods only in France [6, 22] and New Zealand [7]; this is the first study in Brazil evaluating the presence of *M. smithii* on pediatric populations with different socioeconomic conditions. The studies in Brazil have used only the dosage of breath  $CH_4$  as an indirect marker for the presence of methanogenic archaea in children with severe chronic constipation in a specialized outpatient clinic of pediatric gastroenterology [12, 13] and children living in different environmental conditions [16].

The concentration of breath  $CH_4$  is used to classify a population into  $CH_4$  producers and  $CH_4$  nonproducers [8]. Considering only this criterion, the prevalence of children considered to be  $CH_4$  producers reported in the literature varies between 6% and 40% [23, 24], whereas among adults, the reported prevalence of producers is higher, varying from 33% to 70% [8, 17, 24]. In the present study, the prevalence of  $CH_4$  producers was 53.1% among the children residing in the slum near the sanitary landfill; this percentage was higher than that found in the children living in the slum away from the landfill and children in the higher socioeconomic group (31.4% and 21.9%, resp.).

The prevalence of  $CH_4$ -producing children encountered in the two control groups who live away from the landfill was similar to the range of 14.3 to 18.2% reported in a study in Israel that included children aged 7 to 14 [23]. In another study conducted in a rural population in Nigeria, the prevalence of breath  $CH_4$  producers between the ages of 2 and 6 was 40% [24]. Taken together, the findings of these studies suggest that there are differences in the proportions of breath  $CH_4$  producers in groups from different socioeconomic classes who live in different environmental conditions.

The present study showed the presence of *M. smithii* in all fecal samples analyzed, even in those children without breath CH<sub>4</sub> excretion. It should be emphasized that it was not possible to establish a cut-off point between the microbiota concentration of *M. smithii* and the minimum breath CH<sub>4</sub> concentration detected. The correlation between breath  $CH_4$ excretion and the microbiota concentration of M. smithii varied among the studied groups. For the children residing in the slum away from the sanitary landfill, a stronger correlation was found between breath CH<sub>4</sub> excretion and the M. smithii being better than that observed for the children in the high socioeconomic level group. Approximately 20% of all of the CH<sub>4</sub> produced in the gastrointestinal tract is expelled through the lungs [8]; this can explain the differences found in the correlation between the breath  $CH_4$ and the concentration of M. smithii in the microbiota of the children.

The children living near the sanitary landfill show higher concentrations of M. smithii in microbiota and breath CH<sub>4</sub> compared with the other groups. Among all the children considered breath CH<sub>4</sub> producers the concentration of M. smithii in the microbiota was similar in the three groups. The children living near the sanitary landfill present higher counts of M. smithii in the microbiota, even when they do not produce breath  $CH_4$ . Despite the socioeconomic differences, the children living in the slum away from the landfill and those from the high socioeconomic group had similar fecal concentrations of M. smithii between the non-CH<sub>4</sub> producers. The children living near the sanitary landfill were classified into CH<sub>4</sub> with breath excretion above 12 ppm of CH<sub>4</sub> because of the high concentration of CH<sub>4</sub> in the environment, while in the other groups the cut-off point was 4 ppm and 3 ppm, which can explain the higher counts of *M*. smithii among the nonproducers CH<sub>4</sub> from the slum near the sanitary landfill.

Residing close to sanitary landfills can increase exposure to microorganisms and toxic gases emitted by these types of installations [25]. In Finland, studies have been conducted on two different sanitary landfills, evaluating the concentration of microorganisms and gases in the surrounding air [26]. The researchers found that the concentrations of bacteria and viable fungi dispersed in the air into those sanitary landfills were approximately 5 to 20 times higher than those found in the exterior environment [26].

The development of methanogens is not directly related to the introduction of particular foods and the main factors that influence the occurrence of methanogenics archaea are the environment factors [27]. By analyzing the socioeconomic and environment conditions found among the groups, we can deduce that living near a sanitary landfill was a major factor contributing to the differences observed in the microbiota of the children. The children living in the slum away from the sanitary landfill and those who live near the sanitary landfill present the same socioeconomics conditions, including income per capita and basic sanitation, while the high socioeconomic group presents better conditions for both characteristics.

Increased concentration of breath  $CH_4$  and *M. smithii* in the intestinal microbiota has been associated with diverticulosis [11], constipation-predominant irritable bowel syndrome [10], and chronic constipation with retentive fecal incontinence in children [13]. Pimentel et al. have also shown that  $CH_4$  slows intestinal transit and augments small intestinal contractile activity and these contractions are isolated, segmental, and nonpropagating [28]. *M. smithii* can interact with other bacteria of the gut microbiota, enhance the activities and growth of polysaccharide consumers like *Bacteroidetes* and *Firmicutes* by removing  $H_2$ , and promote caloric intake [29].

The role *M. smithii* might have in pathological conditions is still unclear, but through syntrophic interactions methanogens might support the growth of fermenting bacteria, which themselves could be either true pathogens or at least opportunistic pathogens which influence our health in other indirect ways [30, 31]. The children living near the sanitary landfill possess significant increase of *M. smithii* in the microbiota, which can cause microbiota's alterations that might influence their health in the future; their pattern of microbiota may help to clarify which bacteria are more sensible to variations of *M. smithii* concentrations through further investigations.

## 5. Conclusion

The present study is the first report showing the distribution of *M. smithii* in children living in different socioeconomics conditions in Brazil and worldwide. To live near a sanitary landfill is associated with higher concentrations of *M. smithii* in intestinal microbiota, comparing with those who live away from the sanitary landfill, regardless of their socioeconomics conditions. The effects of these changes cannot be seen in the health of the children; however investigating the alterations on microbiota of peoples living in those conditions could help to understand the relations between *M. smithii* and other microorganisms.

## **Conflict of Interests**

The authors declare that there is no conflict of interests regarding the publication of this paper.

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#### References

 P. B. Eckburg, E. M. Bik, C. N. Bernstein et al., "Diversity of the human intestinal microbial flora," *Science*, vol. 308, no. 5728, pp. 1635–1638, 2005.

- [2] W. E. Balch, G. E. Fox, L. J. Magrum, C. R. Woese, and R. S. Wolfe, "Methanogens: reevaluation of a unique biological group," *Microbiological Reviews*, vol. 43, no. 2, pp. 260–296, 1979.
- [3] M. J. Wolin and T. L. Miller, "Bacterial strains from human feces that reduce CO<sub>2</sub> to acetic acid," *Applied and Environmental Microbiology*, vol. 59, no. 11, pp. 3551–3556, 1993.
- [4] T. L. Miller and M. J. Wolin, "Oxidation of hydrogen and reduction of methanol to methane is the sole energy source for a methanogen isolated from human feces," *Journal of Bacteriology*, vol. 153, no. 2, pp. 1051–1055, 1983.
- [5] T. L. Miller, M. J. Wolin, E. C. de Macario, and A. J. L. Macario, "Isolation of Methanobrevibacter smithii from human feces," *Applied and Environmental Microbiology*, vol. 43, no. 1, pp. 227– 232, 1982.
- [6] B. Dridi, M. Henry, A. El Khéchine, D. Raoult, and M. Drancourt, "High prevalence of *Methanobrevibacter smithii* and *Methanosphaera stadtmanae* detected in the human gut using an improved DNA detection protocol," *PLoS ONE*, vol. 4, no. 9, Article ID e7063, 2009.
- [7] J. A. Stewart, V. S. Chadwick, and A. Murray, "Carriage, quantification, and predominance of methanogens and sulfatereducing bacteria in faecal samples," *Letters in Applied Microbiology*, vol. 43, no. 1, pp. 58–63, 2006.
- [8] L. F. McKay, M. A. Eastwood, and W. G. Brydon, "Methane excretion in man—a study of breath, flatus, and faeces," *Gut*, vol. 26, no. 1, pp. 69–74, 1985.
- [9] A. Haines, J. Dilawari, G. Metz, L. Blendis, and H. Wiggins, "Breath methane in patients with cancer of the large bowel," *The Lancet*, vol. 2, no. 8036, pp. 481–483, 1977.
- [10] M. Pimentel, A. G. Mayer, S. Park, E. J. Chow, A. Hasan, and Y. Kong, "Methane production during lactulose breath test is associated with gastrointestinal disease presentation," *Digestive Diseases and Sciences*, vol. 48, no. 1, pp. 86–92, 2003.
- [11] G. A. Weaver, J. A. Krause, T. L. Miller, and M. J. Wolin, "Incidence of methanogenic bacteria in a sigmoidoscopy population: an association of methanogenic bacteria and diverticulosis," *Gut*, vol. 27, no. 6, pp. 698–704, 1986.
- [12] A. C. F. Soares, S. Tahan, U. Fagundes-Neto, and M. B. de Morais, "Breath methane in children with chronic constipation," *Arquivos de Gastroenterologia*, vol. 39, no. 1, pp. 66–72, 2002.
- [13] A. C. F. Soares, H. M. Lederman, U. Fagundes-Neto, and M. B. De Morais, "Breath methane associated with slow colonic transit time in children with chronic constipation," *Journal of Clinical Gastroenterology*, vol. 39, no. 6, pp. 512–515, 2005.
- [14] E. C. de Macario and A. J. L. Macario, "Methanogenic archaea in health and disease: a novel paradigm of microbial pathogenesis," *International Journal of Medical Microbiology*, vol. 299, no. 2, pp. 99–108, 2009.
- [15] A. Leiby, D. Mehta, V. Gopalareddy, S. Jackson-Walker, and K. Horvath, "Bacterial overgrowth and methane production in children with encopresis," *Journal of Pediatrics*, vol. 156, no. 5, pp. 766–770, 2010.
- [16] C. S. Mello, S. Tahan, L. C. F. L. Melli et al., "Methane production and small intestinal bacterial overgrowth in children living in a slum," *World Journal of Gastroenterology*, vol. 18, no. 41, pp. 5932–5939, 2012.
- [17] J. H. Bond Jr., R. R. Engel, and M. D. Levitt, "Factors influencing pulmonary methane excretion in man. An indirect method of studying the in situ metabolism of the methane-producing colonic bacteria.," *Journal of Experimental Medicine*, vol. 133, no. 3, pp. 572–588, 1971.

- [18] J. A. Ufnar, S. Y. Wang, J. M. Christiansen, H. Yampara-Iquise, C. A. Carson, and R. D. Ellender, "Detection of the nifH gene of *Methanobrevibacter smithii*: a potential tool to identify sewage pollution in recreational waters," *Journal of Applied Microbiology*, vol. 101, no. 1, pp. 44–52, 2006.
- [19] R. Verma, A. K. Verma, V. Ahuja, and J. Paul, "Real-time analysis of mucosal flora in patients with inflammatory bowel disease in India," *Journal of Clinical Microbiology*, vol. 48, no. 11, pp. 4279– 4282, 2010.
- [20] B. S. Samuel, E. E. Hansen, J. K. Manchester et al., "Genomic and metabolic adaptations of Methanobrevibacter smithii to the human gut," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 25, pp. 10643–10648, 2007.
- [21] C. Johnston, J. A. Ufnar, J. F. Griffith, J. A. Gooch, and J. R. Stewart, "A real-time qPCR assay for the detection of the nifH gene of *Methanobrevibacter smithii*, a potential indicator of sewage pollution," *Journal of Applied Microbiology*, vol. 109, no. 6, pp. 1946–1956, 2010.
- [22] M. Million, M. Maraninchi, M. Henry et al., "Obesity-associated gut microbiota is enriched in *Lactobacillus reuteri* and depleted in *Bifidobacterium animalis* and *Methanobrevibacter smithii*," *International Journal of Obesity*, vol. 36, no. 6, pp. 817–825, 2012.
- [23] Y. Peled, T. Gilat, E. Liberman, and Y. Bujanover, "The development of methane production in childhood and adolescence," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 4, no. 4, pp. 575–579, 1985.
- [24] M. J. Hudson, A. M. Tomkins, H. S. Wiggins, and B. S. Drasar, "Breath methane excretion and intestinal methanogenesis in children and adults in rural Nigeria," *Scandinavian Journal of Gastroenterology*, vol. 28, no. 11, pp. 993–998, 1993.
- [25] E. R. Fielding, D. B. Archer, E. Conway de Macario, and A. J. Macario, "Isolation and characterization of methanogenic bacteria from landfills.," *Applied and environmental microbiology*, vol. 54, no. 3, pp. 835–836, 1988.
- [26] P. Rahkonen, M. Ettala, M. Laukkanen, and M. Salkinoja-Salonen, "Airborne microbes and endotoxins in the work environment of two sanitary landfills in Finland," *Aerosol Science and Technology*, vol. 13, no. 4, pp. 505–513, 1990.
- [27] Y. Liu and W. B. Whitman, "Metabolic, phylogenetic, and ecological diversity of the methanogenic archaea," *Annals of the New York Academy of Sciences*, vol. 1125, pp. 171–189, 2008.
- [28] M. Pimentel, H. C. Lin, P. Enayati et al., "Methane, a gas produced by enteric bacteria, slows intestinal transit and augments small intestinal contractile activity," *American Journal of Physiology—Gastrointestinal and Liver Physiology*, vol. 290, no. 6, pp. G1089–G1095, 2006.
- [29] B. S. Samuel and J. I. Gordon, "A humanized gnotobiotic mouse model of host-archaeal-bacterial mutualism," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 26, pp. 10011–10016, 2006.
- [30] H.-P. Horz and G. Conrads, "The discussion goes on: what is the role of Euryarchaeota in humans?" *Archaea*, vol. 2010, Article ID 967271, 8 pages, 2010.
- [31] E. Conway de Macario and A. J. L. Macario, "Methanogenic archaea in health and disease: a novel paradigm of microbial pathogenesis," *International Journal of Medical Microbiology*, vol. 299, no. 2, pp. 99–108, 2009.