nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
\boxtimes	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on statistics for highgrists contains articles on many of the points above

Software and code

Policy information about availability of computer code

Data collection

NA, all data were already collected and de-identified and provided for researchers to use.

Data analysis

All data analysis methods utilize publicly available and published software packages. The SNP-based heritability was estimated using SumHer (Idak5.2.linux). The lambda intercept was estimated using LD Score Regression v1.0.1. Conditional analyses were conducted using Genomewide Complex Trait Analysis v1.93.0. Functional annotations of enrichment tests of GWAS results were completed using Functional Mapping and Annotation Web tool (accessed August 2023). Predicted gene expression as completed using S-PrediXcan/MetaXcan v0.7.1. Gene expression colocalization was conducted using coloc R library v5.2.2. OpenTargets Genetics was used to map variants to genes. Pathway analysis of colocalized genes was completed with Ingenuity Pathway Analysis (accessed March 2024).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Genetic association summary statistics for each meta-analysis have been deposited in the GWAS Catalog under accession codes GCST90461957 [https://www.ebi.ac.uk/gwas/studies/GCST90461957] (multi-ancestry), GCST90461958 [https://www.ebi.ac.uk/gwas/studies/GCST90461958] (European ancestry), GCST90461959 [https://www.ebi.ac.uk/gwas/studies/GCST90461959] (East Asian/Central South Asian ancestry), GCST90461960 [https://www.ebi.ac.uk/gwas/studies/GCST90461960] (African ancestry). Study-specific summary statistics for BioBank Japan [https://www.pheweb.jp/pheno/UF] and FinnGen [https://r7.finngen.fi/pheno/CD2_BENIGN_LEIOMYOMA_UTERI] are available at their respective web portals. UKBB data access can be requested [https://www.ukbiobank.ac.uk/enable-your-research/apply-for-access]. BioVU [https://ictr.vumc.org/how-to-use-biovu/] and eMERGE [https://emerge-network.org/collaborate/] data also require approved access, which can be requested at their respective links. The All of Us [https://www.researchallofus.org/] data is accessible on the Researcher Workbench with registered access. The data from CARDIA have been deposited at dbGaP accession phs000285.v3.p2 [https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000285.v3.p2] and the BWHS data at dbGaP accession phs001409.v2 [https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs001409].The predicted expression models [https://predictdb.org/post/2021/07/21/gtex-v8-models-on-eqtl-and-sqtl/] used are publicly available. The other data generated in this study are provided in the Supplementary Data files.

Research involving human participants, their data, or biological material

Policy information about studies with <u>human participants or human data</u>. See also policy information about <u>sex, gender (identity/presentation)</u>, <u>and sexual orientation</u> and <u>race, ethnicity and racism</u>.

Reporting on sex and gender

The data sources used in this varied on inclusion as self-reported female sex or woman gender. Fibroids are a disease unique to individuals with a uterus.

Reporting on race, ethnicity, or other socially relevant groupings

Our data stratification included use of EHR and self-reported race and ethnicity. We recognize that race and ethnicity are not ideal substitutes for genetic ancestry. We delineate in supplemental table 1 whether each data source uses race and ethnicity or genetic ancestry to determine the meta-analysis strata grouping. Non-Hispanic Black individuals were part of the African ancestry analyses, Non-Hispanic White and Finnish individuals were part of the European ancestry analyses, and Japanese individuals were part of the East Asian/Central South Asian analyses.

Population characteristics

Case and control definitions for each data source varied, with a range of use of ICD billing codes, CPT codes, survey response, imaging-confirmed, and self-report. These are described in the methods portion of the paper. Mean age and other population characteristics are also shown in Supplemental Table 1.

Recruitment

Each data source used had its own recruitment method, which have all been previously published. There are both hospital and community recruitment methods across the data sources. There is some ascertainment bias due to these different recruitment methods, but heterogeneity measures revealed sufficiently homogeneous meta-data

Ethics oversight

This research was approved by the Vanderbilt University Medical Center Institutional Review Board.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one belo	w that is the best fit for your research	. If you are not sure, read the appropriate sections before making your selection.
X Life sciences	Behavioural & social sciences	Ecological, evolutionary & environmental sciences

 $For a \ reference \ copy \ of the \ document \ with \ all \ sections, see \ \underline{nature.com/documents/nr-reporting-summary-flat.pdf}$

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

We report the complete sample size for each data source and combined meta-analyses. No sample size calculation was performed due to use of all available data.

Data exclusions

Genetic quality control excluded low quality samples and was performed separately by each data source used. Samples were also excluded if consent had been revoked, sample was duplicated, or sex concordance checks failed.

Replication

Our study was able to replicate other previously reported fibroid-associated variants and genes. Each analysis was performed independently as the cohorts do not have overlapping individuals, and each meta-analysis only includes independent data sets.

Randomization

This is an observational study of associations between genetic variants and uterine fibroids, and randomization is not required as cases and

Randomization	control definitions are based on presence or absence of fibroids.
Blinding	Blinding is not relevant to genome-wide association studies because the case control statuses are already determined, and the subsequent

Reporting for specific materials, systems and methods

statistical analyses performed in this study.

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Ma	terials & experimental systems	Methods
n/a	Involved in the study	n/a Involved in the study
\boxtimes	Antibodies	ChIP-seq
\boxtimes	Eukaryotic cell lines	Flow cytometry
\times	Palaeontology and archaeology	MRI-based neuroimaging
\boxtimes	Animals and other organisms	
\boxtimes	Clinical data	
\times	Dual use research of concern	
\boxtimes	Plants	
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Plants

Seed stocks

Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.

Novel plant genotypes

Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor was applied. Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to

Authentication

assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosiacism, off-target gene editing) were examined.