iScience

CelPress

Article

Highly sensitive and simultaneous detection of ascorbic acid, dopamine, and uric acid using Pt@g-C₃N₄/N-CNTs nanocomposites



Lin Zhang, Liu Yu, Junyang Peng, Xiaoying Hou, Hongzhi Du

3162@hbtcm.edu.cn (L.Z.) jhhxy2021@jhun.edu.cn (X.H.) 3163dhz@hbtcm.edu.cn (H.D.)

Highlights

Pt@g-C₃N₄ were synthesized and combined with N-doped carbon nanotubes

A simple, quick, and cheap electrochemical sensor was established

Simultaneous detection of AA, DA, and UA was

The proposed sensor was well applied in practical measurement in human

Zhang et al., iScience 27, 109241 March 15, 2024 © 2024 The Author(s). https://doi.org/10.1016/ j.isci.2024.109241

iScience

Article



Highly sensitive and simultaneous detection of ascorbic acid, dopamine, and uric acid using $Pt@q-C_3N_4/N-CNTs$ nanocomposites

Lin Zhang,^{1,2,4,5,*} Liu Yu,^{1,4} Junyang Peng,¹ Xiaoying Hou,^{1,3,*} and Hongzhi Du^{1,2,*}

SUMMARY

The detection of ascorbic acid (AA), dopamine (DA), and uric acid (UA) is crucial for understanding and managing various illnesses. In this research, $Pt@g-C_3N_4$ nanoparticles were synthesized via hydrothermal method and combined with N-doped carbon nanotubes (N-CNTs). The Pt@g-C₃N₄/N-CNTs-modified glassy carbon (GC) electrode was fabricated as an electrochemical sensor for the determination of AA, DA, and UA. The linear response range of AA, DA, and UA in the optimal condition was 100–3,000 μ M, 1–100 μ M, and 2–215 μ M boasting a low detection limit (S/N = 3) of 29.44 μ M (AA), 0.21 μ M (UA), and 2.99 μ M (DA), respectively. Additionally, the recoveries of AA, DA, and UA in serum sample were 100.4%–106.7%. These results corroborate the feasibility of the proposed method for the simultaneous, sensitive, and reliable detection of AA, DA, and UA. Our Pt@g-C₃N₄/N-CNTs/GC electrode can provide a potential strategy for disease diagnosis and health monitoring in clinical settings.

INTRODUCTION

The detection of ascorbic acid (AA), dopamine (DA), and uric acid (UA) has garnered significant attention in recent years. These three active components play crucial roles in human metabolic physiology and have a profound impact on human health.¹⁻³ AA is essential for organism growth and promoting antibody synthesis. It is a vital nutrient that supports immune system and helps fight off infections.⁴ DA is a small-molecule substance found in human central nervous system that directly influences mental activities.⁵ It is responsible for regulating mood, motivation, and cognition. When purine metabolism is disrupted, excess UA may be produced, leading to a condition called hyperuricemia. This accumulation of UA in the body can alter the pH of body fluids, creating an acidic internal environment. This acidic environment can have detrimental effect on cellular function and contribute to the development of various illnesses.⁶ Mental illness,⁷ Parkinson's disease,² hyperuricemia, and leukemia^{8,9} can arise from abnormal levels of AA, DA, and UA. Detecting and monitoring these substances in the body are crucial for early diagnosis and effective treatment of these diseases.

AA, DA, and UA coexist in biological fluids, such as in human plasma and urine. While the basal concentration of DA is very low (0.01–1 µM), AA and UA are present at much higher concentrations (0.1–0.5 mM and 0.12–0.45 mM).¹⁰ This large concentration difference makes it difficult to accurately detect and quantify these three substances simultaneously. Additionally, the presence of interfering substances further complicates the simultaneous detection of AA, DA, and UA. Traditional methods for detecting these substances are relatively time-consuming and expensive and require skilled labor, such as enzyme-linked immunosorbent assay, ^{11,12} high-performance liquid chromatography,¹³⁻¹⁵ spectrophotometry,¹⁶ capillary electrophoresis,¹⁷ and electrochemiluminescence analysis.¹⁸⁻²⁰ However, there is a growing interest in electrochemical techniques due to their rapid response, low cost, in situ monitoring capability, high sensitivity, good stability, and selectivity.²¹⁻²³ Electrochemical analysis offers a promising solution for the simultaneous detection of AA, DA, and UA.

One of the main challenges in electrochemical analysis is the overlapping oxidation waves of interfering AA and DA analytes, which often disrupts the accurate determination of UA levels. To address this issue, researchers have been exploring innovative approaches to enhance the selectivity and sensitivity of electrochemical detection methods. For example, various modifications have been utilized on conventional electrodes, including carbon-based modifiers,^{22,24-26} metals/metal oxides-based electrode materials,^{21,27-29} conducting polymers,^{30,31} and so on. However, the current modifications still cannot meet clinical requirements.

One promising avenue for the development of electrochemical sensors is the utilization of carbon nanotubes (CNTs). These nanotubes are allotropes of carbon composed of a rolled sheet of graphene, with each end capped by half a fullerene-like molecule. They have attracted considerable attention due to their potential in sensor development. Multi-walled CNTs are concentric and enclosed tubules, with two or

²Hubei Shizhen Laboratory, Wuhan 430065, China ³Cancer Institute, School of Medicine, Jianghan University, Wuhan, China

⁵Lead contact

¹School of Pharmacy, Hubei University of Chinese Medicine, Wuhan 430065, China

⁴These authors contributed equally

^{*}Correspondence: 3162@hbtcm.edu.cn (L.Z.), jhhxy2021@jhun.edu.cn (X.H.), 3163dhz@hbtcm.edu.cn (H.D.) https://doi.org/10.1016/j.isci.2024.109241



multiple layers of graphene separated by approximately 0.34 nm. Nitrogen can be easily incorporated into the structure of CNTs and forming N-doped carbon nanotube (N-CNT).³²⁻³⁴ It has been reported that pyridine and the tetravalent N atoms of N-CNTs can provide high localized electron densities and surface energy differences, which are favorable for electrocatalysis.

By utilizing N-CNTs, the electrochemical sensors achieve high sensitivity in the detection of AA, DA, and UA. Moreover, it exhibits excellent stability, ensuring reliable and consistent measurements. Tsierkezos et al.²⁸ fabricated a powerful analytical tool consisting of N-CNTs decorated with rhodium, palladium, iridium, platinum, and gold nanoparticles and investigated N-CNTs/AuNPs enabling simultaneous analysis of AA, DA, and UA. Wang and co-workers²² found a three-dimensional g-C₃N₄/MWCNTs/GO hybrid modified electrode, which was constructed for AA, DA, and UA simultaneous detection. Fernandes et al.³⁵ reported an Fe₃O₄@N-CNT modified glass carbon electrode and demonstrate Fe₃O₄@N-CNT composite materials are an efficient electrocatalyst. In this work, Pt@g-C₃N₄ was applied for modification of N-CNTs-based electrodes to facilitate the electron transfer rate and the catalytic reaction. The local electronic structure of g-C₃N₄ is expected to be modified by adding metallic elements, resulting in enhanced catalytic performance.³⁶ Firstly, due to the presence of active sites and high surface area of g-C₃N₄, Pt nanoparticles absorption on carbon nitride surface is enhanced. Secondly, porous structure of g-C₃N₄ makes it easy for reactants to reach active sites or edges and form products quickly. Therefore, decoration of Pt@g-C₃N₄ on N-CNTs is quite evident to exhibit better electrocatalytic activity.

In this work, we have developed $Pt@g-C_3N_4/N-CNTs$ nanomaterials to fabricate an electrochemical sensor for simultaneous determination of AA, DA, and UA. A large specific surface area and plenty of electrochemically active sites are attributed to the superior features of the prepared nanohybrids. This sensor exhibited high sensitivity and good stability, making it a promising platform for practical applications. Notably, the sensor has been successfully employed to analyze these three substances in human serum samples.

RESULTS AND DISCUSSION

Characterizations

To characterize the structure, morphology, and assembly mode of the hybrid nanomaterials, scanning electron microscope (SEM) images was obtained. Figures 1A–1C shows the SEM images of N-CNTs, Pt@g-C₃N₄, and Pt@g-C₃N₄/N-CNTs. The N-CNTs display distinct tubular _structures (Figure 1A), while the g-C₃N₄ nanosheets exhibit a unique two-dimensional block structure with folds (Figure 1B). In Figure 1C, the Pt@g-C₃N₄ nanoparticles are uniformly distributed and remarkably small in size, effectively attached to the surface of N-CNTs. The elemental mapping analysis (Figure 1D) and energy dispersive spectroscopy (EDS) (Figure 1E) reveal the presence of C, N, O, and Pt elements of Pt@g-C₃N₄/N-CNTs, confirming the successful attachment of Pt@g-C₃N₄ nanoparticles. And it is beneficial to enhance the effective surface area and boost the electrocatalytic performance of the sensor.

The structures of Pt@g-C₃N₄, N-CNTs, and Pt@g-C₃N₄/N-CNTs were analyzed by X-ray diffraction (XRD) (Figure 1F). The diffraction peak at $2\vartheta = 27.8^{\circ}$ corresponds to (002) peak of g-C₃N₄, and the XRD of Pt@g-C₃N₄ reveals the presence of metallic Pt, which conforms to *JCPDS card* no. 04-0802. The average crystallite particle size of Pt@g-C₃N₄/N-CNTs is about 7.1 nm according to Scherrer equation.

X-ray photoelectron spectroscopy (XPS) was used to further characterize the elemental composition and valence states of N-CNTs, Pt@g-C₃N₄, and Pt@g-C₃N₄/N-CNTs. From the full XPS survey spectrum in Figure 2, the characteristic peaks of C, N, O, and Pt are located at 284, 399, 531, and 73/76 eV, respectively.^{22,37–39} As can be seen from the C 1s high-resolution XPS spectra, the hydrothermally treated Pt@g-C₃N₄ composites exhibited smaller C-C (284.3 eV) and C-O (285.9 eV) peaks, and higher C=O (288.1 eV) peak, compared to g-C₃N₄.⁴⁰ In the C 1s high-resolution XPS spectra of Pt@g-C₃N₄/N-CNTs, there is an increase in the ratio of C-C and C-N bonds, corresponding to the carbon skeleton of N-CNTs. Another peak observed at 291.2 eV can be attributed to the binding energy of CH₂-CF₂ of Nafion. The N 1s spectra of the N-CNTs indicate the presence of three types of nitrogen-doped sites, including pyridinic N (398.6 eV), graphitic N (400.5 eV), and quaternary N (402 eV).^{41,42} Compared with N-CNTs, the pyridinic N of sample Pt@g-C₃N₄ shifted negatively. N-C=N and N(-C)₃ in Pt@g-C₃N₄/N-CNTs are the main components of g-C₃N₄ and N-CNTs. Pyridine and the tetravalent N atoms of N-doped carbon materials can provide high localized electron densities and are responsible for the electrocatalytic activity in the oxidation of UA.⁴³ The spectrum of O 1s shows -COOH (532.1 eV) and C-OH (535.3 eV) peaks of sample Pt@g-C₃N₄/N-CNTs, indicating the presence of defects in the composite nanomaterials and their enhanced electrocatalytic activity.⁴⁴ Because of Pt 4f high-resolution spectrum, two peaks at 75.7 and 73.1 eV were identified as Pt 4f_{5/2} and Pt 4f_{3/2}, which indicate the existence of zero-valence Pt in Pt@g-C₃N₄ and Pt@g-C₃N₄/N-CNTs. The result showed that Pt nanoparticles were homogeneously distributed on the surface of N-CNTs sample.⁴³

Electrochemical characterizations of the modified electrodes

Before AA, DA, and UA detection, the electrochemical features of modified electrodes were characterized by the $[Fe(CN)_6]^{3-/4-}$ redox probe. Cyclic voltammetry (CV) and electrochemical impedance spectroscopy (EIS) tests were performed. As can be seen in Figure 3A, a pair of reversible redox peaks could be observed at the bare GC electrode. After the modification of N-CNTs nanoparticles on the electrode surface, the gap between redox peak potential became smaller and the peak current density decreased slightly. When Pt@g-C₃N₄ nanoparticles are attached and dispersed on the surface of N-CNTs, the redox peak current density increased significantly. It indicates that Pt@g-C₃N₄/N-CNTs hybrid nanomaterials facilitate electron transport and the electroactive surface area of Pt@g-C₃N₄/N-CNTs was 0.27 cm² (calculated on the basis of the Randles-Ševčík equation), which is about 2.2 times higher than that of bare GC electrode.

Nyquist plots were in Figure 3B. The Randles circuit was used to interpret the electron transfer characteristics. Mass transport is characterized at low frequencies, and Warburg impedance relates diffusion of electroactive species. In this work, ZView software was used to fit the





Figure 1. Microstructural characterizations of N-CNTs, Pt@g-C₃N₄, and Pt@g-C₃N₄/N-CNTs samples

(A) SEM image of N-CNTs.

(B) SEM image of $Pt@g-C_3N_4$.

(C) SEM image of $Pt@g-C_3N_4/N-CNTs$.

(D) EDS mapping images of C, N, O, and Pt element.

(E) EDS line image of the synthesized $Pt@g-C_{3}N_{4}/N\text{-}CNTs$ nanomaterials.

(F) XRD patterns of N-CNTs, Pt@g-C₃N₄, and Pt@g-C₃N₄/N-CNTs.

semicircle diameter in the EIS Nyquist plot. The electron transfer resistance (Rct) value of Pt@g-C₃N₄/N-CNTs/GC electrode is 6.8 Ω , which is much lower than that of GC electrode (53 Ω), meaning electron transfer ability of Pt@g-C₃N₄/N-CNTs/GC electrode is faster.

Electrochemical behaviors of AA, DA, and UA

The electrochemical response of the three different electrodes to AA, DA, and UA in 0.1 M PBS (pH 7.0) was performed. Figure 4 shows the CV and differential pulse voltammetry (DPV) performances of 0.5 mM AA, 0.1 mM DA, and 0.1 mM UA simultaneously at bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes. CV signals (Figure 4A) show that, compared to the bare GC and N-CNTs/GC electrode, at Pt@g-C₃N₄/N-CNTs/GC electrode three distinct oxidation peaks of AA, DA, and UA occur at a lower overpotential, accompanied by a slightly higher peak current. The presence of N-CNTs electrode, resulting in the separation of the overlapping oxidation waves of AA and DA. The electrooxidation of DA shows a tendency toward reversibility on both N-CNTs/GC and Pt@g-C₃N₄/N-CNTs/GC electrodes. The peak potential (Δ Ep) value for oxidation of DA obtained onto Pt@g-C₃N₄/N-CNTs/GC electrode is about 0.023 V, which is much smaller compared to that measured under the same conditions onto N-CNTs/GC electrode (0.072 V). The difference in Δ Ep is related to the rate of heterogeneous electron transfer. These findings indicate that the oxidation current of UA is higher on Pt@g-C₃N₄/N-CNTs/GC electrode than on either GC or N-CNTs/GC electrode.







Figure 2. Chemical analysis and electrical properties of N-CNTs, Pt@g-C₃N₄, and Pt@g-C₃N₄/N-CNTs samples

(A) XPS survey spectra of N-CNTs, $Pt@g-C_3N_4$, and $Pt@g-C_3N_4/N-CNTs$.

(B) High-resolution XPS spectra of C1s for $Pt@g-C_3N_4$ (top) and $Pt@g-C_3N_4/N-CNTs$ (bottom) samples.

(C) High-resolution XPS spectra of N1s for $Pt@g-C_3N_4$ (top) and $Pt@g-C_3N_4/N-CNTs$ (bottom) samples.

(D) High-resolution XPS spectra of O1s for $Pt@g-C_3N_4$ (top) and $Pt@g-C_3N_4/N-CNTs$ (bottom) samples.

(E) High-resolution XPS spectra of Pt4f for $Pt@g-C_3N_4$ (top) and $Pt@g-C_3N_4/N-CNTs$ (bottom) samples.

These results have been validated by DPV, a powerful electrochemical technique in analytical measurements. The electrochemical behaviors of AA, DA, and UA exhibit distinct differences, with three oxidation peaks observed at -0.028, 0.172, and 0.292 V, respectively (Figure 4B). Notably, the oxidation of AA on GC electrode lies at about 0.400 V vs. saturated calomel electrode (SCE),⁴⁵ whereas the oxidation overpotential of AA decreases on N-CNTs and Pt@g-C₃N₄/N-CNTs-modified electrodes. Furthermore, the anodic peak potentials for AA, DA, and UA show a shift toward a less positive potential at Pt@g-C₃N₄/N-CNTs/GC electrode, as compared to the GC or N-CNTs/GC electrode. The response signals of Pt@g-C₃N₄/N-CNTs/GC electrode to the mixture of the three substances are notably higher than those of N-CNTs/GCE. This enhancement in electrocatalytic activity is attributed to the increased electrode surface area and active sites, as well as the high electron transfer performance of Pt@g-C₃N₄/N-CNTs composite. The profound results demonstrate the good electrocatalytic activity of Pt@g-C₃N₄/N-CNTs/GC electrode results about different modified electrodes are provided in the supplemental material.

Optimization of the experimental conditions

To achieve the best performance, the pH of detection solution and materials proportion were optimized.

The electrolyte solution's pH has an impact on the peak current of AA, DA, and UA; 0.1 M PBS buffer solutions with varying pH levels ranging from 6 to 8 were measured. The results, illustrated in Figure 5A, revealed a notable trend: as the pH of the medium increased from 6 to 8, the anodic peak potentials for AA, DA, and UA shifted toward less positive potential. This shift can be attributed to the involvement of protons in the electrooxidation reactions of AA, DA, and UA.^{22,46}

Plotting Ep against pH within the working pH range (Figure S3) further elucidated the relationship between the compounds and the buffer pH. The plotted data exhibited a linear correlation, with the Ep of all compounds demonstrating a consistent linear relationship with the buffer pH. Notably, the observed slopes of 0.046, 0.048, and 0.064 V/pH for AA, DA, and UA, respectively, closely approximated the expected



Figure 3. Electrochemical characterizations of bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes in 5 mM [Fe(CN)₆]^{3-/4-} solution containing 0.5 M KCl

(A) CV curves of bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes. (B) EIS spectra of bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes.

Nernstian value of 0.0585 V/pH for a two-electron, two-proton transfer in the electrochemical reaction. The possible redox reaction mechanism can be expressed as Scheme 1.^{21,22}

The peak currents of AA, DA, and UA with the increase of solution pH are shown in Figure 5B. The response signal of AA was observed to remain nearly stable, while the maximum current signals of DA and UA were obtained at pH 7. This finding is indicative of the fact that PBS at pH 7 gave the best response.

The optimized proportional of hybrid Pt@g-C3N4/N-CNTs nanomaterials was crucial in determining the electrochemical performance. Figure 5C shows the electrochemical performance of Pt@g-C₃N₄/N-CNTs (1:1, 2:1, 10:1, 20:1) in a 0.1 M PBS at pH 7 solution containing 0.5 mM AA + 0.1 mM DA + 0.1 mM UA. DPV signals varied with the three substances on different modifier GC electrode. As shown in Figure 5D, anodic peak currents of AA, DA, and UA on Pt@g-C₃N₄/N-CNTs (2:1) modified GC electrode are higher than those of GC electrode modified by other substances. The resulting data indicate that the hybrid nanomaterials exhibit strong synergy and high electron transfer properties, particularly at the 2:1 proportion of Pt@g-C₃N₄/N-CNTs.

Feasibility study

The Pt@g-C₃N₄/N-CNTs GC electrode detection of AA, DA, and UA was evaluated using DPV. Through an analysis of the relationship between the peak current and the concentration of the three substances, the electrocatalytic oxidation of AA, DA, and UA was explored. In Figure 6A, it is found that the DPV profiles of the Pt@g-C₃N₄/N-CNTs GC electrode exhibited a linear relationship with the concentration of AA, with a regression equation of $l_{(AA)}(\mu A) = 0.0259 C_{(AA)}(\mu M) + 30.667 (R² = 0.998) within the concentration range of 2–2,000 <math>\mu$ M. For DA, the linear regression equation between peak current and concentration is $l_{(DA)}(\mu A) = 0.709 C_{(DA)}(\mu M) + 30.282 (R² = 0.996) within the concentration range$ $of 5–100 <math>\mu$ M. As the concentration of DA continues to increase, the linear regression equation will be different. This is because the diffusion rate of oxidation products may differ. The linear regression equation between peak current and UA concentration is $l_{(UA)}(\mu A) = 0.417$ $C_{(UA)}(\mu M) + 11.074 (R² = 0.996)$ within the concentrations range of 1–110 μ M. The detection limits of Pt@g-C₃N₄/N-CNTs GC electrode for determination of AA, DA, and UA are 29.44, 0.21, and 2.99 μ M (S/N = 3), respectively.

The concentrations of AA, DA, and UA in the ternary mixture were simultaneously determined on the $Pt@g-C_3N_4/N-CNTs$ GC electrode. The analytical performance was examined by the relationship between peak current and AA, DA, and UA in the ternary mixtures. The DPV curves of the concentrations changes of three analytes are shown in Figure 7A. The concentration of AA increased from 100 to 3,000 μ M,



Figure 4. Electrochemical detection of 0.5 mM AA, 0.1 mM DA, and 0.1 mM UA in 0.1 M PBS
(A) CV curves of bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes.
(B) DPV curves of bare GC, N-CNTs/GC, and Pt@g-C₃N₄/N-CNTs/GC electrodes.







Figure 5. Optimization of experimental conditions

(A) DPV curves of Pt@g-C₃N₄/N-CNTs/GC electrode in different pH solutions.

(B) Histograms showing the peak currents of AA, DA, and UA in different pH solutions. Error bars represent the standard errors (n = 3).

(C) DPV curves of modified electrodes with different proportions of hybrid nanomaterials.

(D) Histograms showing the peak currents of AA, DA, and UA at modified electrodes with different proportions of hybrid nanomaterials. Error bars represent the standard errors (n = 3).

and the concentrations ranges of DA and UA are 1–100 μ M and 2–215 μ M, respectively. The results show that peak currents of all three analytes increased linearly with increasing concentrations of AA, DA, and UA. The fitted linear regression equations are as follow: $l_{(AA)}(\mu A) = 0.0372 \text{ C}_{(AA)}(\mu M) + 641.4 \text{ (R}^2 = 0.990) \text{ for AA}, l_{(DA)}(\mu A) = 38.174\text{ C}_{(DA)}(\mu M) + 566.821 \text{ (R}^2 = 0.990) \text{ and } l_{(DA)}(\mu A) = 7.408\text{ C}_{(DA)}(\mu M) + 715.135 \text{ (R}^2 = 0.993) \text{ for DA}, and l_{(UA)}(\mu A) = 6.009\text{ C}_{(UA)}(\mu M) + 450.914 \text{ (R}^2 = 0.988) \text{ and } l_{(UA)}(\mu A) = 0.518\text{ C}_{(UA)}(\mu M) + 486.106 \text{ (R}^2 = 0.990) \text{ for UA}.$



Scheme 1. Oxidation process of AA, DA, and UA







Figure 6. Individual detection of AA, DA, and UA

(A) DPV curves of Pt@q-C₃N₄/N-CNTs GC electrode in different concentrations of AA (2–2,000 μ M).

(B) Linear fitting between the concentrations of AA and peak currents. Error bars represent the standard errors (n = 3).

- (C) DPV curves of Pt@g-C₃N₄/N-CNTs GC electrode in different concentrations of DA (5–100 μ M).
- (D) Linear fitting between the concentrations of DA and peak currents. Error bars represent the standard errors (n = 3).
- (E) DPV curves of $Pt@g-C_3N_4/N-CNTs$ GC electrode in different concentrations of UA (1–110 μ M).

(F) Linear fitting between the concentrations of UA and peak currents. Error bars represent the standard errors (n = 3).

Table 1 provides a comparison of the performance of the Pt@g-C₃N₄/N-CNTs GC electrode with several other electrodes that have been previously studied. The results clearly demonstrate that the Pt@g-C₃N₄/N-CNTs GC electrode is capable of detecting AA, DA, and UA simultaneously, with a high level of sensitivity, low detection limits, and wide linear ranges. This good biosensing performance is likely due to the advantageous properties of the N-CNTs-based composite for electrochemical analysis of AA, DA, and UA. The presence of nitrogen atoms in g-C₃N₄, along with the unique sp² hybridization of C and N structures, provides a large number of active surface sites. Additionally, the incorporation of metal elements can adjust the electronic and catalytic properties of g-C₃N₄. Pt nanoparticles appear to improve better the electrocatalytic activity and sensitivity.

Repeatability, stability, and interferences effect

Repeatability, stability, and selectivity are important factors for electrochemical sensors in practical analysis. Pt@g-C₃N₄/N-CNTs GC electrode sensor was employed to detect the peak current of the ternary mixture containing 0.5 mM AA, 0.1 mM DA, and 0.1 mM UA. 10 times repeated determination of the peak current of AA, DA, and UA via DPV experiments was carried out. The relative standard deviations of peak currents are 0.73% for AA, 0.22% for DA, and 0.75% for UA. The results indicate that Pt@g-C₃N₄/N-CNTs GC electrode has good reproducibility for simultaneous determination of AA, DA, and UA.







Figure 7. Simultaneous detection of AA, DA, and UA

(A) DPV curves of Pt@g-C₃N₄/N-CNTs GC electrode in the ternary mixture with different concentrations of AA (2–2,000 μ M), DA (5–100 μ M), and UA (1–110 μ M). (B) Linear fitting between the concentrations of AA and peak currents. Error bars represent the standard errors (n = 3). (C) Linear fitting between the concentrations of DA and peak currents. Error bars represent the standard errors (n = 3). (D) Linear fitting between the concentrations of UA and peak currents. Error bars represent the standard errors (n = 3).

Figure 8A shows the stability test. $Pt@g-C_3N_4/N-CNTs$ GC electrode was stored at room temperature. Three paralleled tests were performed to detect the peak current of the ternary mixture containing 0.5 mM AA, 0.1 mM DA, and 0.1 mM UA. The relative standard deviation of peak current are 1.65% for AA, 0.67% for DA, and 0.65% for UA, suggesting that $Pt@g-C_3N_4/N-CNTs$ GC electrode has good stability for simultaneous determination of AA, DA, and UA.

Figures 8B, 8C, and 8D show several potential interferences evaluated by amperometric method. The applying potentials are 0.1 V for AA, 0.35 V for DA, and 0.5 V for UA, respectively. The physiological interfering molecules such as glucose, glycine, and citric acid (1 mM) and

Table 1. Comparison of Pt@g-C₃N₄/N-CNTs GC electrode sensor with those of reported electrode materials										
	Sensitivity/($\mu A \ \mu M^{-1}$ cm ⁻²)			Linear range/(µM)			LOD/(µM)			
Electrode	AA	DA	UA	AA	DA	UA	AA	DA	UA	Reference
g-C ₃ N ₄ /MWNTs/GO/GCE	0.0236	5.666	2.682	200–7500	2–100	4–200	96	0.22	1.36	Wang et al. ²²
PdNPsª/rGO/GCE	0.079	10.893	0.481	500–3500	3–42	300–1400	100	1	16.67	Wei et al. 47
Fe ₃ O ₄ /rGO	0.0335	38.8	4.5	160–7200	0.4–3.5	4–20	20	0.08	0.5	Teymourian et al. ⁴⁸
Pt@NP-AuSn ^b /CFP ^c	0.0004	0.0017	0.0003	200–1200	0.5–10	25–500	13.4	0.31	0.76	Yang et al. ⁴⁹
CB ^d /GCE	0.214	1.570	0.680	1.91–37.8	0.599–11.8	1.01–14	0.03	0.0524	0.0407	Vicentini et al. ⁵⁰
Pd ₃ Pt ₁ °/PDDA ^f -rGO/GCE	0.359	0.639	0.498	40–1200	4–200	4–400	0.61	0.04	0.10	Yan et al. ⁵¹
SnO ₂ /chitosan/GCE	0.127	2.773	2.391	20–220	0.1–18	1–100	6.45	0.77	0.89	Selvarajan et al. ⁵²
Pt@g-C ₃ N ₄ /N-CNTs/GCE	0.258	0.709	0.417	2–2000	5–100	1–110	29.44	0.21	2.99	This work

^apalladium nanoparticles;

^bPt nanoparticle-modified nanoporous AuSn;

^cNi-buffered flexible carbon fiber paper;

^dnanostructured carbon black;

^ePd-Pt bimetallic nanoparticles;

^fpoly(diallyldimethylammonium chloride).







Figure 8. Stability and selectivity test

Interfering substance: 5 mM Na⁺, 5 mM K⁺, 5 mM Mg²⁺, 1 mM glucose, 1 mM glycine, and 1 mM citric acid.

(A) Histograms of $Pt@g-C_3N_4/N-CNTs$ GC electrode for stability test. Error bars represent the standard errors (n = 3).

(B) Amperometric response to continuous addition of 0.5 mM AA and interfering substances.

(C) Amperometric response to continuous addition of 0.1 mM DA and interfering substances.

(D) Amperometric response to continuous addition of 0.1 mM UA and interfering substances.

common ions such as Na⁺, K⁺, and Mg²⁺ (5 mM) were added into 0.1 M PBS solution containing 0.5 mM AA, 0.1 mM DA, and 0.1 mM UA, respectively. It was found that the response current remained unchanged when interfering substances were added, despite the fact that the concentrations of interfering substances were almost 10 or 50 times higher than the concentrations of AA, DA, and UA. The results show Pt@g-C₃N₄/N-CNTs GC electrode has good stability for simultaneous determination of AA, DA, and UA in the presence of the aforementioned interfering substances.

Real sample analysis

In order to verify the reliability of the practical application of the proposed Pt@g-C₃N₄/N-CNTs GC electrode sensor, the feasibility was performed by determining the recovery rates of AA, DA, and UA in the human serum samples provided by local hospital. The serum sample was injected into the electrolyte and diluted 40 times. Without any pretreatment process, DPV records Pt@g-C₃N₄/N-CNTs GC electrode. As shown in Figure 9 and Table 2, UA has been detected in real serum sample with the value about 73.14 μ M. Standard addition method was applied for the quantitation of AA, DA, and UA. Based on the obtained recovery values 106.7%, 100.4%, and 113.2%, respectively, the presence of the possibly interfering species in human serum sample has minor impact on the detection of AA, DA, and UA. The Pt@g-C₃N₄/N-CNTs GC electrode is a suitable electrochemical sensor in simultaneous determination of AA, DA, and UA.

Limitations of the study

A new electrochemical sensor has been developed using $Pt@g-C_3N_4/N$ -CNTs composite. Incorporating heteroatoms such as N into the carbon supports as anchors to disperse and stabilize the supported metal nanoparticles, which shows striking effects to the improved catalytic performance. However, the synthesis of Pt nanoparticles with controllable shape and size and customized catalytic properties still faces challenges.

Moreover, the advanced sensor demonstrates high sensitivity and selectivity, allowing for the simultaneous detection of AA, DA, and UA. The whole process is still at laboratory scale. A detailed analysis of electrocatalytic process in modified electrodes is required for an in-depth explanation. Further endeavors are imperative in achieving the desired electrochemical catalysis.







Figure 9. Real sample analysis by standard addition method

DPV responses of human blood serum sample and 150 μ M AA, 25 μ M DA, and 15 μ M UA at Pt@g-C₃N₄/N-CNTs GC electrode.

STAR***METHODS**

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- RESOURCE AVAILABILITY
 - O Lead contact
 - Materials availability
 - Data and code availability
- METHOD DETAILS
 - Materials
 - Synthesis of Pt@g-C₃N₄
 - Synthesis of Pt@g-C₃N₄/N-CNTs/GC electrode
 - Materials characterization
 - O Electrochemical measurements and calculations
- QUANTIFICATION AND STATISTICAL ANALYSIS

SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.isci.2024.109241.

ACKNOWLEDGMENTS

This work was financially supported by the National Natural Science Foundation of China (82204416), the Department of Education of Hubei Province (no. B2022119), and the Young Elite Scientists Sponsorship Program by CAST (2021-QNRC2-B16).

AUTHOR CONTRIBUTIONS

L.Z.: conceptualization, methodology, investigation, writing – original draft, writing – review & editing; L.Y.: validation, investigation; J.P.: validation; X.H.: writing – review & editing; H.D.: supervision, funding acquisition, writing – review & editing. All authors reviewed, revised, and approved of this manuscript.

Table 2. Recovery data for the determination of AA, DA, and UA in human blood serum using the calibration curve method						
Sample	Species	Detected (µM)	Added (µM)	Found (µM)	Recovery (%)	
Human blood serum	AA	-	150	160.05	106.7	
	DA	-	25	26.14	100.4	
	UA	73.14	15	89.12	106.5	

iScience Article

DECLARATION OF INTERESTS

The authors declare no competing interests.

Received: November 2, 2023 Revised: December 21, 2023 Accepted: February 12, 2024 Published: February 15, 2024

REFERENCES

- Ballaz, S.J., and Rebec, G.V. (2019). Neurobiology of vitamin C: Expanding the focus from antioxidant to endogenous neuromodulator. Pharmacol. Res. 146, 104321. https://doi.org/10.1016/j.phrs.2019. 104321.
- Ciobanu, C.A., Ionita, I., Mihailescu, A.I., and Ciobanu, A.M. (2023). Cyclic voltammetry as a measuring tool in Parkinson's disease and associated psychiatric commorbidities. Eur. Psychiatr. 66, S757–S758. https://doi.org/10. 1192/j.eurpsy.2023.1597.
- Aoki, T., Nakai, S., Uchino, J., Shibata, K., Shinzato, T., Masakane, I., Toma, S., Fukui, H., Matsuoka, T., Kato, S., et al. (2022). Comparison of Body Fluid Volumes Determined by Kinetic Modeling and by Bioimpedance Spectroscopy. Blood Purif. 51, 866–874. https://doi.org/10.1159/ 000521563.
- 4. Tzounakas, V.L., Anastasiadi, A.T., Arvaniti, V.-Z., Lelli, V., Fanelli, G., Paronis, E.C., Apostolidou, A.C., Balafas, E.G., Kostomitsopoulos, N.G., Papageorgiou, E.G., et al. (2022). Supplementation with uric and ascorbic acid protects stored red blood cells through enhancement of non-enzymatic antioxidant activity and metabolic rewiring. Redox Biol. 57, 102477. https://doi.org/10. 1016/j.redox.2022.102477.
- Costa, K.M., and Schoenbaum, G. (2022). Dopamine. Curr. Biol. 32, R817–R824. https:// doi.org/10.1016/j.cub.2022.06.060.
- Lin, Z., Jayachandran, M., Haskic, Z., Kumar, S., and Lieske, J.C. (2022). Differences of Uric Acid Transporters Carrying Extracellular Vesicles in the Urine from Uric Acid and Calcium Stone Formers and Non-Stone Formers. Int. J. Mol. Sci. 23, 10010. https:// doi.org/10.3390/ijms231710010.
- Trossbach, S.V., Bader, V., Hecher, L., Pum, M.E., Masoud, S.T., Prikulis, I., Schäble, S., de Souza Silva, M.A., Su, P., Boulat, B., et al. (2016). Misassembly of full-length Disrupted-in-Schizophrenia 1 protein is linked to altered dopamine homeostasis and behavioral deficits. Mol. Psychiatry 21, 1561–1572. https://doi.org/10.1038/mp. 2015.194.
- Ahmed, A.S., Mona, M.M., Elsisy, R.A., and Hantash, E.M. (2023). Potential of Ascorbic Acid Supplements to Ameliorate the Deleterious Effects of Hyperuricemia on Albino Wistar Rats' Hippocampus (Structural and Functional Study). Neuroendocrinology 113, 615–624. https://doi.org/10.1159/ 000529061.
- Feng, Y., Fu, M., Guan, X., Wang, C., Meng, H., Zhou, Y., He, M., and Guo, H. (2022). Associations of exposure to perfluoroalkyl substances with serum uric acid change and hyperuricemia among Chinese women: Results from a longitudinal study. Chemosphere 308,

136438. https://doi.org/10.1016/j. chemosphere.2022.136438.

- Alba, A.F., Totoricaguena-Gorriño, J., Sánchez-Ilárduya, M.B., Ruiz-Rubio, L., Vilas-Vilela, J.L., Lanceros-Méndez, S., and del Campo, F.J. (2021). Laser-activated screenprinted carbon electrodes for enhanced dopamine determination in the presence of ascorbic and uric acid. Electrochim. Acta 399, 139374. https://doi.org/10.1016/j.electacta. 2021.139374.
- Zhao, D., Li, J., Peng, C., Zhu, S., Sun, J., and Yang, X. (2019). Fluorescence Immunoassay Based on the Alkaline Phosphatase Triggered in Situ Fluorogenic Reaction of o-Phenylenediamine and Ascorbic Acid. Anal. Chem. *91*, 2978–2984. https://doi.org/10. 1021/acs.analchem.8b05203.
- Kim, J., Jeon, M., Paeng, K.-J., and Paeng, I.R. (2008). Competitive enzyme-linked immunosorbent assay for the determination of catecholamine, dopamine in serum. Anal. Chim. Acta 619, 87–93. https://doi.org/10. 1016/j.aca.2008.02.042.
- Darczuk, D., Krzyściak, W., Bystrowska, B., Kęsek, B., Kościelniak, D., Chomyszyn-Gajewska, M., and Kaczmarzyk, T. (2019). The Relationship between the Concentration of Salivary Tyrosine and Antioxidants in Patients with Oral Lichen Planus. Oxid. Med. Cell. Longev. 2019, 5801570. https://doi.org/10. 1155/2019/5801570.
- van Gorkom, G., Gijsbers, B., Ververs, E.-J., El Molla, A., Sarodnik, C., Riess, C., Wodzig, W., Bos, G., and Van Elssen, C. (2022). Easy-to-Use HPLC Method to Measure Intracellular Ascorbic Acid Levels in Human Peripheral Blood Mononuclear Cells and in Plasma. Antioxidants 11, 134. https://doi.org/10. 3390/antiox11010134.
- Qiao, F., Wang, X., Han, Y., Kang, Y., and Yan, H. (2023). Preparation of poly (methacrylic acid)/graphene oxide aerogel as solid-phase extraction adsorbent for extraction and determination of dopamine and tyrosine in urine of patients with depression. Anal. Chim. Acta 1269, 341404. https://doi.org/10.1016/j. aca.2023.341404.
- Chaiendoo, K., Ittisanronnachai, S., Promarak, V., and Ngeontae, W. (2019). Polydopamine-coated carbon nanodots are a highly selective turn-on fluorescent probe for dopamine. Carbon 146, 728–735. https://doi. org/10.1016/j.carbon.2019.02.030.
- Zinellu, A., Sotgia, S., Deiana, L., and Carru, C. (2011). Field-amplified sample injection combined with pressure-assisted capillary electrophoresis UV detection for the simultaneous analysis of allantoin, uric acid, and malondialdehyde in human plasma. Anal. Bioanal. Chem. 399, 2855–2861. https:// doi.org/10.1007/s00216-010-4648-x.
- Liang, Y., Lai, W., Su, Y., and Zhang, C. (2022). A novel cloth-based multiway closed bipolar electrochemiluminescence biosensor for

accurate detection of uric acid. Microchem. J. 179, 107657. https://doi.org/10.1016/j. microc.2022.107657.

- Tao, X.-L., Pan, M.-C., Yang, X., Yuan, R., and Zhuo, Y. (2022). CDs assembled metalorganic framework: Exogenous coreactantfree biosensing platform with pore confinement-enhanced electrochemiluminescence. Chin. Chem. Lett. 33, 4803–4807. https://doi.org/10.1016/ i.cclet.2022.01.010.
- Wang, Z., Wang, X., Zhu, X., Lv, J., Zhang, J., Zhu, Q., and Dai, Z. (2019). Fabrication of nondestructive and enhanced electrochemiluminescence interface for reusable detection of cell-released dopamine. Sensor. Actuator. B Chem. 285, 438–444. https://doi.org/10.1016/j.snb.2019. 01.087.
- Pan, Y., Zuo, J., Hou, Z., Huang, Y., and Huang, C. (2020). Preparation of Electrochemical Sensor Based on Zinc Oxide Nanoparticles for Simultaneous Determination of AA, DA, and UA. Front. Chem. 8, 592538. https://doi.org/10.3389/ fchem.2020.592538.
- Wang, H., Xie, A., Li, S., Wang, J., Chen, K., Su, Z., Song, N., and Luo, S. (2022). Threedimensional g-C(3)N(4)/MWNTs/GO hybrid electrode as electrochemical sensor for simultaneous determination of ascorbic acid, dopamine and uric acid. Anal. Chim. Acta 1211, 339907. https://doi.org/10.1016/j.aca. 2022.339907.
- Hatefi-Mehrjardi, A., Karimi, M.A., Soleymanzadeh, M., and Barani, A. (2020). Highly sensitive detection of dopamine, ascorbic and uric acid with a nanostructure of dianix yellow/multi-walled carbon nanotubes modified electrode. Measurement 163, 107893. https://doi.org/10.1016/j. measurement.2020.107893.
- Lin, K.-C., Tsai, T.-H., and Chen, S.-M. (2010). Performing enzyme-free H2O2 biosensor and simultaneous determination for AA, DA, and UA by MWCNT-PEDOT film. Biosens. Bioelectron. 26, 608–614. https://doi.org/10. 1016/j.bios.2010.07.019.
- Noroozifar, M., Khorasani-Motlagh, M., Akbari, R., and Bemanadi Parizi, M. (2011). Simultaneous and sensitive determination of a quaternary mixture of AA, DA, UA and Trp using a modified GCE by iron ion-doped natrolite zeolite-multiwall carbon nanotube. Biosens. Bioelectron. 28, 56–63. https://doi.org/10.1016/j.bios.2011. 06.042.
- Savk, A., Özdil, B., Demirkan, B., Nas, M.S., Calimli, M.H., Alma, M.H., Inamuddin Asiri, A.M., Asiri, A.M., and Şen, F. (2019). Multiwalled carbon nanotube-based nanosensor for ultrasensitive detection of uric acid, dopamine, and ascorbic acid. Mater. Sci. Eng. C Mater. Biol. Appl. 99,





CellPress

OPEN ACCESS

- Xue, Y., Zheng, Y., Wang, E., Yang, T., Wang, H., and Hou, X. (2022). Ti3C2Tx (MXene)/Pt nanoparticle electrode for the accurate detection of DA coexisting with AA and UA. Dalton Trans. 51, 4549–4559. https://doi.org/ 10.1039/D2DT00110A.
- Tsierkezos, N.G., Othman, S.H., Ritter, U., Hafermann, L., Knauer, A., Köhler, J.M., Downing, C., and McCarthy, E.K. (2016). Electrochemical analysis of ascorbic acid, dopamine, and uric acid on nobel metal modified nitrogen-doped carbon nanotubes. Sensor. Actuator. B Chem. 231, 218–229. https://doi.org/10.1016/j.snb.2016. 03.032.
- Xu, Y., Meng, Z., Meng, Y., Li, X., and Xiao, D. (2021). Lithium cobalt phosphate electrode for the simultaneous determination of ascorbic acid, dopamine, and serum uric acid by differential pulse voltammetry. Mikrochim. Acta 188, 190. https://doi.org/10.1007/ s00604-021-04839-5.
- Văduva, M., Baibarac, M., and Cramariuc, O. (2022). Functionalization of Graphene Derivatives with Conducting Polymers and Their Applications in Uric Acid Detection. Molecules 28, 135. https://doi.org/10.3390/ molecules28010135.
- Wu, Y., Deng, P., Tian, Y., Feng, J., Xiao, J., Li, J., Liu, J., Li, G., and He, Q. (2020). Simultaneous and sensitive determination of ascorbic acid, dopamine and uric acid via an electrochemical sensor based on PVPgraphene composite. J. Nanobiotechnol. 18, 112. https://doi.org/10.1186/s12951-020-00672-9.
- Lee, W.J., Hwang, T.H., Hwang, J.O., Kim, H.W., Lim, J., Jeong, H.Y., Shim, J., Han, T.H., Kim, J.Y., Choi, J.W., and Kim, S.O. (2014). N-doped graphitic selfencapsulation for high performance silicon anodes in lithium-ion batteries. Energy Environ. Sci. 7, 621–626. https://doi.org/10. 1039/C3EE43322F.
- Zhao, Z., Kang, B., Xu, J., Zhu, C., Zhang, X., Zhang, X., and Chen, Y. (2023). N-doped carbon hollow spheres supported N-doped carbon nanotubes for efficient electromagnetic wave absorption. Carbon 209, 117995. https://doi.org/10.1016/j. carbon.2023.04.001.
- 34. Li, S., Liu, Y., Gong, H., Wu, K.-H., Ba, H., Duong-Viet, C., Jiang, C., Pham-Huu, C., and Su, D. (2019). N-Doped 3D Mesoporous Carbon/Carbon Nanotubes Monolithic Catalyst for H2S Selective Oxidation. ACS Appl. Nano Mater. 2, 3780–3792. https://doi. org/10.1021/acsanm.9b00654.
- Fernandes, D.M., Costa, M., Pereira, C., Bachiller-Baeza, B., Rodríguez-Ramos, I., Guerrero-Ruiz, A., and Freire, C. (2014). Novel electrochemical sensor based on

N-doped carbon nanotubes and Fe3O4 nanoparticles: simultaneous voltammetric determination of ascorbic acid, dopamine and uric acid. J. Colloid Interface Sci. 432, 207–213. https://doi.org/10.1016/j.jcis.2014. 06.050.

- Mao, S., Wang, C., and Wang, Y. (2019). The chemical nature of N doping on N doped carbon supported noble metal catalysts. J. Catal. 375, 456–465. https://doi.org/10. 1016/j.jcat.2019.06.039.
- Xie, L., Ni, J., Tang, B., He, G., and Chen, H. (2018). A self-assembled 2D/2Dtype protonated carbon nitridemodified graphene oxide nanocomposite with improved photocatalytic activity. Appl. Surf. Sci. 434, 456–463. https://doi.org/10.1016/j.apsusc. 2017.10.193.
- Zhou, Y., Zhang, Y., Xue, J., Wang, R., Yin, Z., Ding, L., and Wang, H. (2021). Graphene oxide-modified g-C3N4 nanosheet membranes for efficient hydrogen purification. Chem. Eng. J. 420, 129574. https://doi.org/10.1016/j.cej.2021. 129574.
- Nazir, R., Fageria, P., Basu, M., Gangopadhyay, S., and Pande, S. (2017). Decoration of Pd and Pt nanoparticles on a carbon nitride (C3N4) surface for nitro-compounds reduction and hydrogen evolution reaction. New J. Chem. 41, 9658–9667. https://doi.org/10.1039/ c7nj01221g.
- Choi, Y.J., Kim, H.-K., Lee, S.-W., Kim, Y.H., Youn, H.-C., Roh, K.C., and Kim, K.-B. (2017). Surfactant-free synthesis of a nanoperforated graphene/nitrogen-doped carbon nanotube composite for supercapacitors. J. Mater. Chem. A Mater. 5, 22607–22617. https://doi. org/10.1039/c7ta06742a.
- Duan, C., Tian, Y., Wang, X., Wu, J., Liu, B., Fu, D., Zhang, Y., Lv, W., Hu, L., Wang, F., et al. (2023). Anchoring Mo single atoms on N-CNTs synchronizes hydrogenation/ dehydrogenation property of Mg/MgH2. Nano Energy 113, 108536. https://doi.org/10. 1016/j.nanoen.2023.108536.
- Skudin, V., Andreeva, T., Myachina, M., and Gavrilova, N. (2022). CVD-Synthesis of N-CNT Using Propane and Ammonia. Materials 15, 2241. https://doi.org/10.3390/ma15062241.
- Kočí, K., Dang Van, H., Edelmannová, M., Reli, M., and Wu, J.C. (2020). Photocatalytic reduction of CO2 using Pt/C3N4 photocatalyts. Appl. Surf. Sci. 503, 144426. https://doi.org/10.1016/j.apsusc.2019. 144426.
- Liu, J., Li, W., Duan, L., Li, X., Ji, L., Geng, Z., Huang, K., Lu, L., Zhou, L., Liu, Z., et al. (2015). A Graphene-like Oxygenated Carbon Nitride Material for Improved Cycle-Life Lithium/ Sulfur Batteries. Nano Lett. 15, 5137–5142.

https://doi.org/10.1021/acs.nanolett. 5b01919.

- 45. Karim-Nezhad, G., Hasanzadeh, M., Saghatforoush, L., Shadjou, N., Khalilzadeh, B., and Ershad, S. (2009). Electro-oxidation of ascorbic acid catalyzed on cobalthydroxidemodified glassy carbon electrode. J. Serb. Chem. Soc. 74, 581–593.
- Shastan, Z.O., Ganesh, H.V.S., Noroozifar, M., and Kerman, K. (2018). Carbon ceramic microelectrodes modified with buckyballs for simultaneous determination of redox-active biomolecules. RSC Adv. 8, 5960–5966. https://doi.org/10.1039/ C7RA09603H.
- Wei, Y., Liu, Y., Xu, Z., Wang, S., Chen, B., Zhang, D., and Fang, Y. (2020). Simultaneous Detection of Ascorbic Acid, Dopamine, and Uric Acid Using a Novel Electrochemical Sensor Based on Palladium Nanoparticles/ Reduced Graphene Oxide Nanocomposite. Int. J. Anal. Chem. 2020, 8812443. https://doi. org/10.1155/2020/8812443.
- Teymourian, H., Salimi, A., and Khezrian, S. (2013). Fe3O4 magnetic nanoparticles/ reduced graphene oxide nanosheets as a novel electrochemical and bioeletrochemical sensing platform. Biosens. Bioelectron. 49, 1–8. https://doi.org/10.1016/j.bios.2013. 04.034.
- Yang, H., Zhao, J., Qiu, M., Sun, P., Han, D., Niu, L., and Cui, G. (2019). Hierarchical bicontinuous Pt decorated nanoporous Au-Sn alloy on carbon fiber paper for ascorbic acid, dopamine and uric acid simultaneous sensing. Biosens. Bioelectron. 124–125, 191–198. https://doi.org/10.1016/j.bios.2018. 10.012.
- Vicentini, F.C., Raymundo-Pereira, P.A., Janegitz, B.C., Machado, S.A., and Fatibello-Filho, O. (2016). Nanostructured carbon black for simultaneous sensing in biological fluids. Sensor. Actuator. B Chem. 227, 610–618. https://doi.org/10.1016/j.snb. 2015.12.094.
- 51. Yan, J., Liu, S., Zhang, Z., He, G., Zhou, P., Liang, H., Tian, L., Zhou, X., and Jiang, H. (2013). Simultaneous electrochemical detection of ascorbic acid, dopamine and uric acid based on graphene anchored with Pd–Pt nanoparticles. Colloids Surf. B Biointerfaces 111, 392–397. https://doi.org/10.1016/j.colsurfb. 2013.06.030.
- 52. Selvarajan, S., Suganthi, A., and Rajarajan, M. (2017). A facile approach to synthesis of mesoporous SnO2/ chitosan nanocomposite modified electrode for simultaneous determination of ascorbic acid, dopamine and uric acid. Surface. Interfac. 7, 146–156. https://doi.org/10.1016/j. surfin.2017.03.008.





STAR*METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Chemicals, peptides, and recombinant proteins		
Uric acid	Sinopharm	CAS: 69-93-2
Ascorbic acid	Sinopharm	CAS: 50-81-7
Dopamine	Macklin	CAS: 51-61-6
Chloroplatinic acid hexahydrate	Sinopharm	CAS: 18497-13-7
Nitrogen-doped multi-walled carbon	XFNANO	CAS: 1333-86-4
nanotubes		
graphitic carbon nitride	XFNANO	CAS: 7440-44-0
Critical commercial assays		
electrochemical workstation	CHI 660E	http://www.chinstr.com/
Software and algorithms		
Origin 8.5	OriginLab Corporation	https://www.originlab.com/

RESOURCE AVAILABILITY

Lead contact

Further information and requests for materials should be directed to and will be fulfilled by the lead contact, Lin Zhang (3162@hbtcm.edu.cn).

Materials availability

This study did not generate new unique reagents.

Data and code availability

- Data reported in this paper will be shared by the lead contact upon request.
- This paper does not report original code.
- Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

METHOD DETAILS

Materials

All chemical reagents were of analytical grade and used without further purification. All solutions were freshly prepared with ultrapure water.

Synthesis of Pt@g-C₃N₄

The noble platinum metal was loaded onto the surface of $g-C_3N_4$ via hydrothermal methods. Briefly, 10 mg $g-C_3N_4$ and 5 mg NaOH were dispersed in 10 mL deionized water by magnetic stirring, and 226 μ L H₂PtCl₆ solution (1%) was slowly dropped into the suspension solution. Then, the mixture was sealed in an autoclave for hydrothermally treated at 180°C for 10 h. The target product was collected by centrifugation, followed by drying.

Synthesis of Pt@g-C₃N₄/N-CNTs/GC electrode

Different mass of Pt@g-C₃N₄ and N-CNTs were dispersed and ultrasonicated for 20 min 1 mg mL⁻¹ Pt@g-C₃N₄/N-CNTs solution was obtained by redispersing Pt@g-C₃N₄/N-CNTs (1:2) in a mixture of water and 5% Nafion (v/v = 1:0.008) for 20 min under ultrasonic treatment. 10 μ L of the suspension solution was dropped onto the surface of GC electrode and after drying Pt@g-C₃N₄/N-CNTs/GC electrode was obtained.

Materials characterization

X-ray diffraction (XRD) was performed using an X-ray diffractometer (D/Max 2500 PC, Rigaku Corporation, Japan). Scanning electron microscope (SEM) images were obtained by scanning electron microscope (Sigma 500, Zeiss, Germany), while X-ray photoelectron spectroscopy (XPS) were collected by X-ray photoelectron spectrometer (Escalab 250 XI, Thermo Scientific, USA).





Electrochemical measurements and calculations

The electrochemical test was carried out using CHI 660E electrochemical workstation (Shanghai CH Instrument Co., China). A three-electrode system was used with a bare or modified glassy carbon electrode (GC electrode, 3 mm in diameter) as the working electrode, a platinum plate (5×5 mm) as the auxiliary electrode, and KCl saturated Ag/AgCl as the reference electrode.

The Randles-Ševčík equation was used to calculate the electroactive surface area:

$I_P = (2.69 \times 10^5) A D^{1/2} n^{3/2} v^{1/2} C$

where I_P relates to the peak current (A), A is the effective surface area (cm²), D is the diffusion coefficient of $[Fe(CN)_6]^{4-/3-}$ (6.7 × 10⁻⁶ cm² s⁻¹), n denotes the electron transfer number (n = 1), and v is the scanning rate (V s⁻¹).

QUANTIFICATION AND STATISTICAL ANALYSIS

Origin software was used to compile and analyze data.