



Genome Sequence of Lymphocystis Disease Virus 2 LCDV-JP_Oita_2018, Isolated from a Diseased Japanese Flounder (*Paralichthys olivaceus*) in Japan

Satoshi Kawato,^a Reiko Nozaki,^a ^(D)Ikuo Hirono,^a ^(D)Hidehiro Kondo^a

^aLaboratory of Genome Science, Tokyo University of Marine Science and Technology, Tokyo, Japan

ABSTRACT Here, we present the genome sequence of lymphocystis disease virus 2 LCDV-JP_Oita_2018 (genus *Lymphocystivirus*, family *Iridoviridae*), which was isolated from a diseased Japanese flounder (*Paralichthys olivaceus*) in Japan. The LCDV-JP_Oita_2018 genome was assembled into a circular contig of 186,627 bp, with 140 predicted protein-coding genes and a GC content of 27%.

ymphocystis disease (LCD), which is caused by LCD virus (LCDV) (genus *Lymphocystivirus*, family *Iridoviridae*), is a common viral disease in fish (1–6). Here, we sequenced the genome of LCDV 2 LCDV-JP_Oita_2018, which was isolated from a Japanese flounder (*Paralichthys olivaceus*).

A dead *P. olivaceus* fish exhibiting tumor-like lesions on the body surface (a typical clinical sign of LCD) was provided by a commercial farm in Oita Prefecture, Japan, in 2018. Skin lesions were excised, homogenized in phosphate-buffered saline, and stored at -30° C. Two milliliters of thawed lysate was diluted with 6 ml of TNES-urea buffer (6 M urea, 10 mM Tris-HCI [pH 7.5], 125 mM NaCl, 10 mM EDTA, 1% SDS) (7) and treated with proteinase K (final concentration, 500 ng/ml) for 2 h at 60°C. Five milliliters of 5 M NaCl was added, and the lysate was cleared by centrifugation. Genomic DNA was extracted from the supernatant using the cetyltrimethylammonium bromide (CTAB) method (8). The crude DNA was purified using a NucleoBond AXG 100 column (Macherey-Nagel). No shearing or size selection of the extracted DNA was performed before library preparation. Default parameters were used for all software unless otherwise noted.

A paired-end library was prepared with the Nextera XT library preparation kit (Illumina) and was sequenced with the MiSeq reagent kit v2 (2×150 bp) (Table 1). The raw Illumina reads were quality trimmed using Fastp v0.20.0 (9) and were *de novo* assembled by SPAdes v3.13.0 (10), to generate a draft LCDV-JP_Oita_2018 genome.

A long-read library was prepared with a ligation sequencing kit (SQK-LSK109; Oxford Nanopore Technologies) and was sequenced using an R9.4.1 flow cell on a GridlON platform (Table 1). The fast5 files were base called using Guppy v3.2.8+bd67289 with the fast mode.

The Nanopore reads were mapped to the draft LCDV-JP_Oita_2018 genome using minimap2 v2.17-r963-dirty (11). The mapped Nanopore reads were extracted using SAMtools v1.10 (12) and filtered by length (>50 kb) using SeqKit v0.10.0 (13). The filtered Nanopore reads (Table 1) were *de novo* assembled using Flye v2.6 (14), yielding a single contig. The filtered Nanopore reads and trimmed Illumina reads were mapped back using minimap2 to prepare input for polishing with HyPo v1.0.2 (15). Protein-coding genes were predicted using Prodigal v2.6.3 (16). The proteins were manually annotated based on BLASTP search results with the NCBI nonredundant protein database (accessed February 2020).

The LCDV-JP_Oita_2018 genome was assembled into a 186,627-bp circular sequence with a GC content of 27% and 140 predicted protein-coding genes. BLASTN pairwise alignment (task megablast v2.11.0+) with a Chinese isolate (LCDV-C, 186,250 bp [GenBank accession number AY380826]) revealed 99.91% nucleotide identity with 99% coverage. The small

from a diseased Japanese flounder (*Paralichthys olivaceus*) in Japan. Microbiol Resour Announc 10:e00547-21. https://doi.org/ 10.1128/MRA.00547-21. Editor John J. Dennehy, Queens College CUNY Copyright © 2021 Kawato et al. This is an

open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Citation Kawato S, Nozaki R, Hirono I, Kondo H.

2021. Genome sequence of lymphocystis disease virus 2 LCDV-JP_Oita_2018, isolated

Address correspondence to Hidehiro Kondo, h-kondo@kaiyodai.ac.jp.

Received 26 May 2021 **Accepted** 27 July 2021 **Published** 19 August 2021

TABLE 1 LCDV-JP_Oita_2018 sequencing statistics

Parameter	Data for:			
			Nanopore reads (accession no. DRR213900)	
	Raw	Filtered	Raw	Filtered
No. of reads	3,682,911	3,659,510	97,124	1,065
Total length (bp)	867,436,261	863,771,560	630,612,147	71,205,129
Avg read length (bp)	118	118	6,493	66,859
N ₅₀ (bp)			15,501	64,930
No. of mapped reads	2,570,459	2,530,934	66,333	1,065
Proportion mapped (%)	69.79	69.16	68.30	100.00
Mean coverage ^{a} (×)	3,137	3,161	2,480	372

^a Calculated using SAMtools coverage v1.10 (12) from alignments generated by minimap2 v2.17 (11), with the settings ax sr for Illumina reads and ax map-ont for Nanopore reads.

amount of divergence between the two LCDV isolates supports the view that LCDV strains affecting *P. olivaceus* in East Asia are virologically identical, as suggested by the high nucleotide identities (\geq 99.6%) of major capsid protein gene sequences (5).

Data availability. The LCDV-JP_Oita_2018 genome is available in DDBJ/EMBL/GenBank with the accession number LC534415. The raw read data are available with the accession numbers DRR213899 and DRR213900.

ACKNOWLEDGMENTS

This research was supported by Grants-in-Aid for Scientific Research from the Japan Society for Promotion of Science (JSPS) and by SATREPS from the Japan Science and Technology Agency (JST).

S.K. and H.K. conceived the study, and S.K., I.H., and H.K. designed the experiments. S.K. and R.N. prepared and sequenced the libraries. S.K. analyzed the data and wrote the manuscript.

REFERENCES

- Tidona CA, Darai G. 1997. Molecular anatomy of lymphocystis disease virus. Arch Virol Suppl 13:49–56. https://doi.org/10.1007/978-3-7091-6534-8_5.
- López-Bueno A, Mavian C, Labella AM, Castro D, Borrego JJ, Alcami A, Alejo A. 2016. Concurrence of iridovirus, polyomavirus, and a unique member of a new group of fish papillomaviruses in lymphocystis disease-affected gilthead sea bream. J Virol 90:8768–8779. https://doi.org/10.1128/JVI.01369-16.
- Iwakiri S, Song J-Y, Nakayama K, Oh M-J, Ishida M, Kitamura S-I. 2014. Host responses of Japanese flounder *Paralichthys olivaceus* with lymphocystis cell formation. Fish Shellfish Immunol 38:406–411. https://doi.org/10.1016/j .fsi.2014.03.028.
- Zhang Q-Y, Xiao F, Xie J, Li Z-Q, Gui J-F. 2004. Complete genome sequence of lymphocystis disease virus isolated from China. J Virol 78: 6982–6994. https://doi.org/10.1128/JVI.78.13.6982-6994.2004.
- Kitamura S-I, Jung S-J, Kim W-S, Nishizawa T, Yoshimizu M, Oh M-J. 2006. A new genotype of lymphocystivirus, LCDV-RF, from lymphocystis diseased rockfish. Arch Virol 151:607–615. https://doi.org/10.1007/s00705-005-0661-3.
- Chinchar VG, Hick P, Ince IA, Jancovich JK, Marschang R, Qin Q, Subramaniam K, Waltzek TB, Whittington R, Williams T, Zhang Q-Y, ICTV Report Consortium. 2017. ICTV virus taxonomy profile: *Iridoviridae*. J Gen Virol 98:890–891. https://doi.org/10.1099/jgv.0.000818.
- Asahida T, Kobayashi T, Saitoh K, Nakayama I. 1996. Tissue preservation and total DNA extraction from fish stored at ambient temperature using buffers containing high concentration of urea. Fish Sci 62:727–730. https://doi .org/10.2331/fishsci.62.727.
- Murray MG, Thompson WF. 1980. Rapid isolation of high molecular weight plant DNA. Nucleic Acids Res 8:4321–4326. https://doi.org/10.1093/nar/8.19 .4321.

- Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/bioinformatics/ bty560.
- Nurk S, Bankevich A, Antipov D, Gurevich A, Korobeynikov A, Lapidus A, Prjibelsky A, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, McLean J, Lasken R, Clingenpeel SR, Woyke T, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling genomes and mini-metagenomes from highly chimeric reads, p 158–170. *In* Deng M, Jiang R, Sun F, Zhang X (ed), Research in computational molecular biology. Springer, Berlin, Germany.
- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. Bioinformatics 34:3094–3100. https://doi.org/10.1093/bioinformatics/bty191.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–2079. https://doi .org/10.1093/bioinformatics/btp352.
- Shen W, Le S, Li Y, Hu F. 2016. SeqKit: a cross-platform and ultrafast toolkit for FASTA/Q file manipulation. PLoS One 11:e0163962. https://doi.org/10 .1371/journal.pone.0163962.
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, errorprone reads using repeat graphs. Nat Biotechnol 37:540–546. https://doi .org/10.1038/s41587-019-0072-8.
- Kundu R, Casey J, Sung W-K. 2019. HyPo: super fast & accurate polisher for long read genome assemblies. bioRxiv 2019.12.19.882506. https://doi .org/10.1101/2019.12.19.882506.
- Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. https://doi.org/10.1186/1471-2105-11-119.