

Naureen Mushtaq<sup>41</sup>, Andrew Walter<sup>42</sup>, Nada Jabado<sup>43</sup>, Aysa Alsahlawi<sup>43</sup>, Jean-Pierre Farmer<sup>43</sup>, Christina Coleman Abadi<sup>44</sup>, Sabine Mueller<sup>44</sup>, Claire Mazewski<sup>45</sup>, Dolly Aguilera<sup>45</sup>, Nathan Robison<sup>46</sup>, Katrina O'Halloran<sup>46</sup>, Samuel Abbou<sup>47</sup>, Pablo Berlanga<sup>47</sup>, Birgit Georger<sup>47</sup>, Ingrid Øra<sup>48,49</sup>, Christopher L. Moertel<sup>50</sup>, Evangelia D. Razis<sup>51</sup>, Anastasia Vernadou<sup>51</sup>, François Doz<sup>3,52</sup>, Theodore W. Laetsch<sup>2</sup>, Sébastien Perreault<sup>1</sup>; <sup>1</sup>CHU Sainte-Justine, Montréal, Canada. <sup>2</sup>Children's Hospital of Philadelphia, Philadelphia, USA. <sup>3</sup>Institut Curie, Paris, France. <sup>4</sup>Hopp Children's Cancer Center Heidelberg (KITZ); German Cancer Research Center (DKFZ) and German Cancer Consortium (DKTK); Heidelberg University Hospital, Heidelberg, Germany. <sup>5</sup>University Medical Center Göttingen, Göttingen, Germany. <sup>6</sup>Ghent University Hospital, Ghent, Belgium. <sup>7</sup>Department of Pediatric Hematology/Oncology and BMT, Wrocław Medical University, Wrocław, Poland. <sup>8</sup>St. Anna Children's Hospital, Department of Pediatrics, Medical University of Vienna, and St. Anna Children's Cancer Research Institute (CCRI), Vienna, Austria. <sup>9</sup>Cincinnati Children's Hospital Medical Center, Cincinnati, USA. <sup>10</sup>Memorial Sloan Kettering Cancer Center, New York City, USA. <sup>11</sup>Department of Paediatric Haematology and Oncology, Second Faculty of Medicine, Charles University and University Hospital Motol, Prague, Czech Republic. <sup>12</sup>Hospices Civils de Lyon, Lyon, France. <sup>13</sup>Institut d'Hématologie et d'Oncologie Pédiatrique and Pluridisciplinaire Research in pediatric Oncology for Perspectives in Evaluation Care and Therapy (PROSPECT), Centre Leon Berard, Lyon, France. <sup>14</sup>Hospital for Sick Children, Toronto, Canada. <sup>15</sup>Hematology department, faculty of Medicine, King Abdulaziz University, Jeddah, Saudi Arabia. <sup>16</sup>Children's Cancer Centre, Royal Children's Hospital; Murdoch Children's Research Institute; Department of Pediatrics, University of Melbourne, Melbourne, Australia. <sup>17</sup>IWK Health Centre, Halifax, Canada. <sup>18</sup>University of Iowa Stead Family Children's Hospital, Iowa City, USA. <sup>19</sup>Perth Children's Hospital; Brain Tumour Research Programme, Telethon Kids Institute; Paediatrics, School of Medicine, University of Western Australia, Perth, Australia. <sup>20</sup>Rigshospitalet, Copenhagen, Denmark. <sup>21</sup>Kanagawa Children's Medical Center, Yokohama, Japan. <sup>22</sup>Sydney Children's Hospital, Sydney, Australia. <sup>23</sup>Texas Children's Hospital, Houston, USA. <sup>24</sup>Taipei Medical University Hospital, Taipei, Taiwan. <sup>25</sup>University of Michigan, Michigan, USA. <sup>26</sup>Children's National Hospital, Washington, D.C., USA. <sup>27</sup>Arnold Palmer Hospital, Orlando, USA. <sup>28</sup>B.C. Children's Hospital, Vancouver, Canada. <sup>29</sup>Children's Hospital, London Health Sciences Centre, London, Canada. <sup>30</sup>Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy. <sup>31</sup>Monash Children's Hospital, Clayton, Australia. <sup>32</sup>Monash University, Clayton, Australia. The Hudson Institute of Medical Research, Clayton, Australia. <sup>33</sup>Phoenix Children's Hospital, Phoenix, USA. <sup>34</sup>IOP-GRAACC/UNIFESP, Sao Paulo, Brazil. <sup>35</sup>Hospital Infantil Universitario Niño Jesús, Madrid, Spain. <sup>36</sup>Cliniques universitaires Saint-Luc, Bruxelles, Belgium. <sup>37</sup>Vall d'Hebron Children's Hospital, Barcelona, Spain. <sup>38</sup>Department of Oncology, University Children's Hospital, Zurich, Switzerland. <sup>39</sup>Institut Bergonié, Bordeaux, France. <sup>40</sup>Children's Minnesota, Minneapolis, USA. <sup>41</sup>American University of Beirut Medical Center, Beirut, Lebanon. <sup>42</sup>Aga Khan University Hospital, Karachi, Pakistan. <sup>43</sup>Nemour Alfred I duPont Hospital for Children, Wilmington, USA. <sup>44</sup>Montreal Children's Hospital, Montréal, Canada. <sup>45</sup>University of California, San Francisco, USA. <sup>46</sup>Children's Health Care of Atlanta, Emory University School of Medicine, Atlanta, USA. <sup>47</sup>Children's Hospital Los Angeles, Los Angeles, USA. <sup>48</sup>Gustave Roussy Cancer Center, Université Paris-Saclay, Villejuif, France. <sup>49</sup>Lund University, Lund, Sweden. <sup>50</sup>Karolinska University Hospital, Stockholm, Sweden. <sup>51</sup>University of Minnesota Masonic Children's Hospital, Minneapolis, USA. <sup>52</sup>Hygeia Hospital, Athènes, Greece. <sup>53</sup>University of Paris, Paris, France

**BACKGROUND:** TRK fusions are detected in less than 3% of CNS tumors. Given their rarity, there are limited data on the clinical course of these patients. **METHODS:** We contacted 166 oncology centers worldwide to retrieve data on patients with TRK fusion-driven CNS tumors. Data extracted included demographics, histopathology, NTRK gene fusion, treatment modalities and outcomes. Patients less than 18 years of age at diagnosis were included in this analysis. **RESULTS:** Seventy-three pediatric patients with TRK fusion-driven primary CNS tumors were identified. Median age at diagnosis was 2.4 years (range 0.0–17.8) and 60.2% were male. NTRK2 gene fusions were found in 37 patients (50.7%), NTRK1 and NTRK3 aberrations were detected in 19 (26.0%) and 17 (23.3%), respectively. Tumor types included 38 high-grade gliomas (HGG; 52.1%), 20 low-grade gliomas (LGG; 27.4%), 4 embryonal tumors (5.5%) and 11 others (15.1%). Median follow-up was 46.5 months (range 3–226). During the course of their disease, a total of 62 (84.9%) patients underwent surgery with a treatment intent, 50 (68.5%) patients received chemotherapy, 35 (47.9%) patients received radiation therapy, while 34 (46.6%) patients received NTRK inhibitors (3 as first line treatment). Twenty-four (32.9%) had no progression including 9 LGG (45%) and 9 HGG (23.6%). At last follow-up, only one (5.6%–18 evaluable) patient with LGG died compared to 11 with HGG (35.5%–31 evaluable). For LGG the median progression-free survival (PFS) after the first line of treatment was 17 months (95% CI: 0.0–35.5)

and median overall survival (OS) was not reached. For patients with HGG the median PFS was 30 months (95% CI: 11.9–48.1) and median OS was 182 months (95% CI 20.2–343.8). **CONCLUSIONS:** We report the largest cohort of pediatric patients with TRK fusion-driven primary CNS tumors. These results will help us to better understand clinical evolution and compare outcomes with ongoing clinical trials.

#### HGG-12. RAPID PTEFB-DEPENDENT TRANSCRIPTIONAL REORGANIZATION UNDERPINS THE GLIOMA ADAPTIVE RESPONSE TO RADIOTHERAPY

Faye Walker<sup>1</sup>, Lays Martin Sobral<sup>1</sup>, Etienne Danis<sup>1</sup>, Bridget Sanford<sup>1</sup>, Ilango Balakrishnan<sup>1</sup>, Dong Wang<sup>1</sup>, Angela Pierce<sup>1</sup>, Sana Karam<sup>1</sup>, Natalie Serkova<sup>1</sup>, Nicholas Foreman<sup>1</sup>, Sujatha Venkataraman<sup>1</sup>, Robin Dowell<sup>2</sup>, Rajeev Vibhakar<sup>1</sup>, Nathan Dahl<sup>1</sup>; <sup>1</sup>University of Colorado, Aurora, CO, USA. <sup>2</sup>University of Colorado, Boulder, CO, USA

**BACKGROUND:** Dynamic regulation of gene expression is fundamental for cellular adaptation to exogenous stressors. PTEFb-mediated promoter proximal pause-release of Pol II is a conserved regulatory mechanism for synchronous transcriptional induction best described in response to heat shock, but this pro-survival role has not been examined in the applied context of cancer therapy. **DESIGN/METHOD:** In order to examine the dynamics of chromatin reorganization following radiotherapy, we performed a combination of ChIP-, ATAC-, and RNA-seq in model systems of diffuse intrinsic pontine glioma (DIPG) and other pediatric high-grade gliomas (pHGG) following IR exposure. We interrogated IR-induced gene expression in the presence or absence of PTEFb blockade, including both mechanistic and functional consequences of concurrent inhibition or genetic depletion. We utilized culture models with live cell imaging to assess the therapeutic synergy of PTEFb inhibition with IR, as well as the therapeutic index of this intervention relative to normal controls. Finally, we employed orthotopic models of pHGG treated with conformal radiotherapy and CNS-penetrant PTEFb inhibitors in order to assess tolerability and anti-tumor effect in vivo. **RESULTS:** Rapid genome-wide redistribution of active chromatin features and PTEFb facilitates Pol II pause-release to drive nascent transcriptional induction within hours of exposure to therapeutic ionizing radiation. Concurrent inhibition of PTEFb imparts a transcription elongation defect, abrogating canonical adaptive programs such as DNA damage repair and cell cycle regulation. This combination demonstrates a potent, synergistic therapeutic potential agnostic of glioma subtype, leading to a marked induction of tumor cell apoptosis and prolongation of xenograft survival. **CONCLUSION:** These studies reveal a central role for PTEFb underpinning the early adaptive response to radiotherapy, opening new avenues for combinatorial treatment in these lethal malignancies.

#### HGG-13. COMBINED CDK INHIBITION AND ARGININE-DEPRIVATION AS TARGETED THERAPY FOR ARGININE-AUXOTROPHIC GLIOBLASTOMA MULTIFORME CELLS

Christin Riess<sup>1,2</sup>, Katharina del Moral<sup>3</sup>, Adina Fiebig<sup>4</sup>, Philipp Kaps<sup>3</sup>, Charlotte Linke<sup>3</sup>, Burkhard Hinze<sup>5</sup>, Anne Rupprecht<sup>1</sup>, Markus Frank<sup>6</sup>, Tomas Fiedler<sup>4</sup>, Dirk Koczan<sup>7</sup>, Sascha Troschke-Meurer<sup>8</sup>, Holger N. Lode<sup>8</sup>, Nadja Engel<sup>9</sup>, Carl Friedrich Classen<sup>3</sup>, Claudia Maletzki<sup>10</sup>; <sup>1</sup>Univ.-Children's Hospital, Rostock, MV, Germany. <sup>2</sup>University Medicine Clinic III, Rostock, MV, Germany. <sup>3</sup>Univ.-Children's Hospital, Rostock, mv, Germany. <sup>4</sup>Institute for Microbiology, Rostock, mv, Germany. <sup>5</sup>Institute for Pharmacology, Rostock, mv, Germany. <sup>6</sup>Medical Biology and Electron Microscopy Center, Rostock, mv, Germany. <sup>7</sup>Inst.f. Immunology, Rostock, mv, Germany. <sup>8</sup>Ped. Hematology and Oncology, Greifswald, mv, Germany. <sup>9</sup>Oral Surgery, Rostock, mv, Germany. <sup>10</sup>University Medicine Clinic III, Rostock, mv, Germany

**INTRODUCTION/BACKGROUND:** Glioblastoma multiforme show constitutive activation of cyclin-dependent kinases (CDKs) or arginine auxotrophy. This renders tumor cells vulnerable towards arginine-depleting substances, such as arginine deiminase from *Streptococcus pyogenes* (SpyADI). Previously, we confirmed the susceptibility of patient-derived GBM cells towards administration of SpyADI as well as CDK inhibitors (CDKis). To improve effects, we applied a sequential (SEQ) CDKi/SpyADI approach to examine mechanistic insights and drug susceptibility. **MATERIALS AND METHODS:** Three arginine-auxotrophic patient-derived GBM lines with different molecular characteristics were cultured in 2D and 3D (spheres and glioma stem-like cells (GSC)) and effects of this combined CDKi/SpyADI approach were analyzed. This included viability staining via Calcein AM in 2D and 3D-Glo in 3D culture and cell death analysis via flow cytometry. Therapy-induced morphological changes were identified with transmission electron microscopy (TEM). Besides, 3D-invasiveness, cellular stress, and DNA damage responses were measured. **RESULTS:** All SEQ-CDKi/SpyADI combinations yielded synergistic antitumor effects, characterized by impaired cell proliferation, invasiveness, and viability. Notably, this SEQ-CDKi/SpyADI approach was most effective in 3D models. Mitochondrial impairment was demonstrated by increasing mitochondrial