

# Effects of Formic Acid-Treated Shrimp Meal on Growth Performance and Nutrient Digestibility in Broilers

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This study was conducted to know the effect of formic acid-treated shrimp meal as a protein source on growth performance, digestibilities, and nitrogen (N) retention for broilers. Shrimp meal (SM) was treated with 3% formic acid (w/v) at room temperature for 20 minutes, sun-dried, ground through a 1.0 mm mesh screen, and then ready to use as the treated SM (TSM). Forty-two male broiler chicks (8 d old, Ross 308) were randomly divided into 7 dietary groups (6 birds each), namely control diet, diets containing 5, 10, and 15% of SM, and diets containing 5, 10, and 15% of TSM and offered diets till 35 d old. Final body weight, body weight gain and feed intake decreased significantly with increasing levels of SM in diets. Feed conversion ratio also decreased with increasing levels of the SM (*P***<** 0.05). Similar trend was observed in the TSM group, but the adverse effects of the TSM were milder in comparison to the SM group ( $P$  $\leq$ **0.05). Dry matter digestibility tended to decrease (** $P$  $\leq$ **0.05) with increasing levels of the SM but** unchanged with increasing level of the TSM. Availability of ash decreased with increasing levels of the SM and TSM in diets (*P***<**0.05). Although N retention decreased (*P***<**0.05) with increasing level of the SM and TSM in diets but the decreasing trend was milder in the TSM groups than the SM groups. Moreover, chitin digestibility was significantly greater in the TSM groups than the SM groups. In conclusion, broilers received diets containing the TSM showed better growth performance along with improved nutrient digestibility and N retention which suggests that formic acid-treated SM can be used as a potential protein source in broiler diets.

**Key words**: broiler, digestibilities, formic acid, growth performance, shrimp meal

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## **Introduction**

In general, the nutritional quality of shrimp meal (SM), as a protein source for chicken diets, is poor, although it depends on the species of shrimp and the body parts used (Meyers, 1986; Ngoan *et al*., 2000; Rahman and Koh, 2014). The maximum feed inclusion levels for shells and heads of black tiger shrimp (*Penaeus monodon*), and heads and white leg shrimp (*Litopenaeus vannamei*) were 4, 5, and 10%, respectively (Khempaka *et al*., 2006a; Rahman and Koh, 2016a). Some researchers have reported that these limited inclusion levels could be explained, in part, by the presence of chitin, which can decrease digestibility in broilers (Khempaka *et al*., 2006a) and rats (Oduguwa *et al*., 1998). In this regard, our previous *in vitro* study (Rahman and Koh, 2016b) revealed that formic acid could successfully reduce

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the chitin level in SM, and *in vitro* digestibility was greater for formic acid-treated SM (TSM) than for untreated SM. Therefore, TSM is a promising protein source for broiler diets.

The purpose of our present study was to measure growth performance, digestibilities, and nitrogen (N) retention in broilers that received diets containing TSM, and to discuss the suitability of this shrimp meal as a protein source for broilers.

## **Materials and Methods**

This research was conducted in accordance with guidelines for regulation of animal experimentation of Shinshu University, Japan.

# *Preparation of Treated SM*

The sun-dried SM, composed of heads and hulls of black tiger shrimp (*Penaeus monodon*), was treated with formic acid. In brief, approximately 100 g of SM was suspended with 300 mL of 3% formic acid at room temperature for 20 minutes. The SM was then sun-dried and ground through a 1.0 mm mesh screen, and was then ready to use as the TSM.

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Proximate components, calcium (Ca), phosphorus, and chitin content of the SM and TSM were analysed according to AOAC (1990) and Ghanem *et al*. (2003) methods, respectively (Table 1).

### *Birds, Diets and Sampling*

Forty-two male broiler chicks (8 d old, Ross 308) were distributed into seven dietary groups based on similar body weight (BW). A control diet, diets containing 5, 10, and 15% of SM, and diets containing 5, 10, and 15% of TSM were prepared. In the SM and TSM diets, SM was included mainly as a substitute for soybean meal. Corn and corn oil were also used to adjust the nutrient requirements. Diets (approximately 3180 kcal/kg of energy and approximately

Table 1. **Chemical composition of untreated and treated shrimp meal and soybean meal (air dry matter basis)**

	$TSM^{\dagger}$ $SM*$		Soybean meal <sup>1</sup>	
Components		g/kg		
Crude protein	454	533	450	
Crude fibre	159	145	53	
Ether extract	36	42	19	
Ash	285	163	64	
Chitin	173	153		
Calcium	89	68	37	
Phosphorus	19	11	7.2	
ME, kcal/kg	1230 <sup>1</sup>	1230 <sup>1</sup>	2400	

<sup>1</sup> Standard Tables of Feed Composition in Japan (NARO, 2009).

\* SM**=**untreated shrimp meal; **†**TSM**=**treated shrimp meal.

235 g/kg of CP) were formulated to meet or exceed the nutrient requirements for broilers (Japanese feeding standard for poultry, 2011) (Table 2). Diets and water were provided *ad libitum* for the 28 d experimental period (from 8 to 35 d old). BW and feed intake (FI) were recorded weekly and daily, respectively. Feed conversion ratio (FCR) was also calculated. Excreta were collected from 32 to 36 d of age and stored in a freezer (**−**20**℃**) until analysis.

# *Chemical Analysis*

Dry matter (DM) and ash in diets and excreta were measured to estimate their digestibilities according to standard methods (AOAC, 1990). N in diets and excreta was measured using a CHNS/O analyser (PerkinElmer 2400 Series II), and chitin in excreta was analysed according to the method of Ghanem *et al*. (2003) to estimate their retention and digestibility, respectively.

# *Statistical Analysis*

Data were initially analysed with ANOVA using JMP version 10.0 (SAS Institute, 2012) and significant differences among the dietary groups were evaluated with Tukey's multiple comparison tests. Statements of statistical significance are based on *P***<**0.05. Further, regression analyses were performed to determine the relationships between dietary chitin levels, and digestibilities and N retention.

## **Results and Discussion**

#### *Nutrient Composition of SM and TSM*

The results of the chemical analysis of the SM and TSM, (Table 1), revealed that there was higher CP and lower crude ash levels in the TSM than in the SM. This may be the result

Items		$SM^*$ (%)			$TSM^{\dagger}$ $(^{0}/_{0})$			
	Control	5	10	15	5	10	15	
<b>Ingredients</b>								
Commercial diet <sup>1</sup>	550	550	550	550	550	550	550	
Soybean meal	185	135	88	42	130	78	25	
Corn	239	225	210	193	234	222	214	
Shrimp meal	$\theta$	50	100	150	50	100	150	
Corn oil	10.5	24.5	36.5	49.5	20.5	34.5	45.5	
$Premix^2$	15.5	15.5	15.5	15.5	15.5	15.5	15.5	
Calculated composition (as fed basis)								
ME, kcal/kg	3180	3180	3174	3173	3182	3180	3175	
Calcium	10.8	9.5	13.7	17.9	8.5	11.5	14.7	
Available phosphorus	4.8	5.3	6.3	6.7	4.8	5.1	5.5	
Analysed composition (as fed basis)								
Crude protein	236	235	234	234	235	236	235	
Crude fibre	39.7	44.7	45.9	55.1	43.5	47.4	51.4	
Ash	49.5	60.9	72.4	83.9	54.8	60.2	65.6	
Chitin	$\theta$	9.2	17.9	26.6	8.1	15.7	23.6	

Table 2. **Ingredients and chemical composition of experimental diets (g/kg)**

\* SM=untreated shrimp meal;  $^{\dagger}$  TSM=treated shrimp meal.<br><sup>1</sup> Broiler starter diet (CP ≥ 23.5%, ME ≥ 3050 kcal/kg, Nippon Formula Feed Mfg. Kanagawa, Japan).<br><sup>2</sup> Premix (units/ kg): vitamin A, 5,00,000 IU; vitamin D<sub>3</sub> min B<sub>1</sub>, 800 mg; vitamin B<sub>2</sub>, 600 mg; vitamin B<sub>6</sub>, 600 mg; vitamin B<sub>12</sub>, 5.4 mg; pantothenic acid, 800 mg; nicotinic acid, 800; choline chloride, 20,000 mg; foliate, 104 mg; phosphorus, 106 g; iron, 2 mg; copper, 362 mg; zinc, 3368 mg; manganese, 2,560 mg; iodine, 45 mg.

		$SM^*$ (%)			$TSM^{\dagger}$ (%)		
Parameters	Control		10	15			
Final BW, g		$2135.8 \pm 14.1^a$ $2094.2 \pm 14.4^{ab}$ $1923.3 \pm 33.8^{cd}$ $1795.0 \pm 38.7^d$ $2188.7 \pm 14.7^a$ $2120.3 \pm 46.4^{ab}$ $1995.9 \pm 21.9^{bc}$					
BWG, g		$1944.6 \pm 12.4^a$ $1912.0 \pm 14.9^{ab}$ $1738.6 \pm 32.8^c$ $1601.3 \pm 37.9^d$ $2001.3 \pm 15.0^a$ $1915.5 \pm 51.1^{ab}$ $1803.6 \pm 20.8^{bc}$					
Feed intake, g/b/d $111.5\pm0.8^a$ $111.2\pm0.7^a$ $105.9\pm1.7^{bc}$ $104.4\pm1.4^c$ $112.9\pm0.6^a$ $110.7\pm1.6^{ab}$ $108.5\pm0.8^{abc}$							
FCR, g feed/g BW $1.61 \pm 0.01^{ad}$ $1.63 \pm 0.01^{ad}$ $1.71 \pm 0.01^{b}$ $1.83 \pm 0.03^{c}$ $1.58 \pm 0.02^{ad}$ $1.62 \pm 0.02^{ad}$ $1.67 \pm 0.05^{bd}$							

Table 3. **The effects of dietary untreated and treated shrimp meal on growth performance in broilers<sup>1</sup>**

<sup>1</sup> Values for each parameter represent mean  $\pm$  SE ( $n=6$ ).<br>\* SM=untreated shrimp meal; <sup>†</sup> TSM=treated shrimp meal.

 $^{\mathrm{a-d}}$  Means in a row with different superscripts are significantly different (*P*<0.05).

Table 4. **The effects of dietary untreated and treated shrimp meal on nutrient digestibilities and N retention in broilers<sup>1</sup>**

		$SM^*(\%)$			(9/0) TSM <sup>'</sup>		
Parameters	Control		10			10	
DM digestibility, $\%$	$78.5 \pm 0.53^{\circ}$	77.2 $\pm$ 0.48 <sup>ab</sup>	74.3 $\pm$ 0.75 <sup>bc</sup>	$73.2 \pm 0.98$ <sup>c</sup>	$77.7 + 0.76^a$	76.8 $\pm$ 0.82 <sup>ab</sup>	75 $8+0$ $84^{\text{abc}}$
Ash digestibility, %	42.5 $\pm$ 0.53 <sup>a</sup>	41.4 $\pm$ 0.39 <sup>ab</sup>	$35.7 \pm 0.55$ <sup>c</sup>	30.3 $\pm$ 0.37 <sup>d</sup>	42.1 $\pm$ 0.43 <sup>ab</sup>	40.4 ± 0.40 <sup>b</sup>	37 $4+0.26^{\circ}$
Chitin digestibility, %	$\left($	$29.3 \pm 0.38^a$	$25.5 \pm 0.66^{\circ}$	$19.3 \pm 0.55^{\circ}$	33.6 $\pm$ 0.77 <sup>b</sup>	$28.5 \pm 0.51^{\circ}$	$25.1 \pm 0.61$ <sup>c</sup>
N retention, $\%$	$68.1 \pm 0.23^{\text{a}}$	65.4 $\pm$ 0.31 <sup>b</sup>	58.1 $\pm$ 0.17 <sup>c</sup>	53.7 $\pm$ 0.39 <sup>d</sup>	68.2 $\pm$ 0.29 <sup>a</sup>	66.8 ± 0.49 <sup>ab</sup>	66.0 $\pm$ 0.42 <sup>b</sup>

<sup>1</sup> Values for each parameter represent mean  $\pm$  SE (*n*=6).

\* SM**=**untreated shrimp meal; **†**TSM**=**treated shrimp meal. <sup>a</sup>**-**<sup>d</sup> Means in a row with different superscripts are significantly different (*P***<**0.05).

of the formic acid leaching the minerals from the shrimp exoskeleton, and accordingly, the CP content increasing in the TSM, similar to previous studies (Fox *et al*., 1994; Oduguwa *et al*., 1998; Rahman and Koh, 2016b). Consequently, levels of crude fibre and chitin were lower in the TSM than in the SM, suggesting that chitin, the main source of crude fibre and the cause of the decrease in digestibility (Khempaka *et al*., 2006b), was leached from the SM by the formic acid (Rahman and Koh, 2016b). Overall, the TSM used in this study, meets the nutrient requirements for broilers, defined by the Japanese feeding standard for poultry (2011).

#### *Growth Performance*

In the control group, final BW, body weight gain (BWG), FI, and FCR were similar to those noted in the broiler performance objectives (Aviagen, 2007), but these values deteriorated, dose-responsively, with increasing levels of the SM (Table 3). Rahman and Koh (2016a) reported similar findings, noting decreased growth performance for broilers that received diets containing more than 5% SM. These results suggest that decreased growth performance in the SM group may be, in part, due to decreased FI, and that SM contains one or more anorectic factors. Regarding the FCR, generally, this value improves when FI decreases (Rosenfeld *et al*., 1997; Gernat, 2001; El-Ghousein and Al-Beitawi, 2009), but our data showed the opposite trend, which may be explained by the decreased DM digestibility (Table 4). On the other hand, in the TSM group, although final BW and BWG decreased with increasing levels of SM, this trend was more pronounced in the SM group. In addition, FI and FCR

were better in the TSM group than in the SM group. In this connection, decreased DM digestibility in SM group was restored in TSM group (Table 4). Based on these results, it appears that the formic acid treatment improves the growth performance of broilers by reducing the effects of the anorectic and anti-digestive factors contained in the SM.

In the present and our previous studies (Rahman and Koh, 2016b), we confirmed that chitin and Ca levels were reduced by formic acid treatment, but both of these constituents may not be the anorectic factor, because of the following reasons: decreased FI was not found in broilers given a diet containing purified chitin at the same levels of as chitin in the SM diets (Khempaka *et al*., 2006b); and increasing the dietary level of Ca up to 2.12% (Shafey and McDonald, 1990) and 3.0% (Smith and Kabaiji, 1985) did not cause any detrimental effects on FI or the growth performance of broilers. *Digestibilities and N Retention*

In the control group, DM and ash digestibilities, and N retention were 78.5, 42.5, and 68.1% (Table 4), respectively, which are reasonable values for 35 d old broilers (Apata, 2008; Khempaka *et al*., 2011). Similar to the previous results (Fanimo *et al*., 2004; Khempaka *et al*., 2006b), these values in the SM group decreased with increasing levels of SM. Although a similar trend was observed in the TSM group, the trend was less prominent, except for ash digestibility, which was similar between the SM and TSM groups. These results were supported by our previous *in vitro* study (Rahman and Koh, 2016b), which revealed higher DM and CP digestibilities in the TSM than in the SM. As previously discussed, the higher digestibilities, and N

Parameters		Slope	Intercept		
	$SM*$	$TSM^{\dagger}$	<b>SM</b>	<b>TSM</b>	
DM digestibility, %	$-2.27 \pm 0.44^a$	$-1.22 \pm 0.21^{\rm b}$	$78.9 \pm 0.85$	$78.7 \pm 0.35$	
Ash digestibility, %	$-7,27+0,42$	$-5.95\pm0.63$	47.1 $\pm$ 0.79	49.2 $\pm$ 1.07	
Chitin digestibility, %	$-5.73 \pm 0.47$	$-5.52 \pm 0.59$	$34.9 \pm 0.91$	$37.7 \pm 1.00$	
N retention, $\%$	$-6.25 \pm 0.29$ <sup>a</sup>	$-4$ 81+0 31 <sup>b</sup>	$69.9 \pm 0.56$	$72.3 \pm 0.53$	

Table 5. **Results of the regressions of digestibilities and N retention on chitin levels in untreated and treated shrimp meal diets<sup>1</sup>**

<sup>1</sup> Values for each parameter represent mean $\pm$ SE (*n*=6).<br>\* SM=untreated shrimp meal; <sup>†</sup> TSM=treated shrimp meal.

 $x$ <sup>-b</sup> Means in a row with different superscripts are significantly different (*P*<0.05).

retention in the TSM group may be the reason for the better growth performance in this group.

Chitin digestibility in the SM group ranged from 19.3%  $(15\% \text{ group})$  to  $29.3\%$  (5% group), and decreased with increasing levels of SM (Table 4). Similar findings are reported by Rahman and Koh (2016a). This trend was also observed in the TSM group, but was less prominent. As previously mentioned, there was lower amount of chitin, the factor responsible for decreased digestibility (Fox *et al*., 1994; Rahman and Koh, 2016b), in the TSM than in the SM, and thus it may be interesting to examine whether the improved digestibilities in the TSM group can be explained by the decreased chitin level. Therefore, we conducted regression analyses to determine the relationships between dietary chitin levels, and digestibilities and N retention in the SM and the TSM groups (Table 5). The results showed that the slopes for DM digestibility and N retention were gentler in the TSM group than in the SM group  $(P \le 0.05)$ , which not only chitin, but also some other unknown factor(s) may be involved in the improved digestibility and N retention in the TSM group. The decreased chitin level in the TSM suggests a partially degraded chitin-protein complex in the shrimp shell, which would lead to an increased level of free protein in the shell (i.e. a more digestible from of protein).

In order to generate the TSM for the industry, some potential disadvantages of formic acid handling need to be considered. The hazards of formic acid treatment depend on its concentration, with higher concentrations (**>**10%) considered to be corrosive to skin and eyes, and a risk to unprotected workers (EFSA, 2014). Formic acid is currently listed in the European Union registered feed additives as a technological additive (functional group: preservative) and as a sensory additive (functional group: flavouring compounds) for use in feed for all animal species (EFSA, 2014). It is allowed for the processing of by-products of fish origin (Regulation (EC) No 93/2005), and its use in animal nutrition is safe for the environment (EFSA, 2014). Moreover, formic acid treatment of chicken feed could have important benefits for public health (Humphrey and Lanning, 1988). Therefore, from all perspectives, the use of formic acid at a 3% level is considered safe (EFSA, 2014).

In conclusion, the beneficiary effects of the TSM (up to the level of 10%) on growth performance, along with improved nutrient digestibilities and N retention, suggest that formic acid-treated SM can be used as a potential protein source in broiler diets.

#### **References**

- Apata DF. Growth performance, nutrient digestibility and immune response of broiler chicks fed diets supplemented with a culture of *Lactobacillus bulgaricus*. Journal of the Science of Food and Agriculture, 88: 1253**-**1258. 2008.
- Association of Official Analytical Chemists. Official Method of Analysis. 15<sup>th</sup> edition. Association of Analytical Chemists, Washington, DC. 1990.
- Aviagen. Ross 308 Broiler: performance objectives, June 2007. Ross Breeders Limited, Newbridge, Midlothian, EH28 8SZ, Scotland, UK. 2007.
- EFSA. Scientific opinion on the safety and efficiency of formic acid when used as a technological additive for all animal species. EFSA Journal, 12: 1**-**16. 2014.
- El-Ghousein SS and Al-Beitawi NA. The effect of feeding of crushed thyme (*Thymus valgaris* L) on growth, blood constituents, gastrointestinal tract and carcass characteristics of broiler chickens. Journal of Poultry Science, 46: 100**-**104. 2009.
- Fanimo AO, Mudama E, Umukoro TO and Oduguwa OO. Substitution of shrimp waste meal for fish meal in broiler chicken rations. Tropical Agriculture (Trinidad), 73: 201**-**205. 1996.
- Fanimo AO, Oduguwa BO, Oduguwa OO, Ajasa OY and Jegede O. Feeding value of shrimp meal for growing pigs. Archivos de Zootecnia, 53: 77**-**85. 2004.
- Fox CJ, Blow P, Brown JH and Watson I. The effect of various processing methods on the physical and biochemical properties of shrimp head meals and their utilization by juvenile *Penaeus monodon* Fab. Aquaculture, 122: 209**-**226. 1994.
- Gernat AG. The effect of using different levels of shrimp meal in laying hen diets. Poultry Science, 80: 633**-**636. 2001.
- Ghanem A, Ghaly AE and Chaulk M. Effect of shrimp processing procedures on the quality and quantity of extracted chitin from the shells of northern shrimp *Pandalus borealis*. Journal of Aquatic Food Product Technology, 12: 63**-**79. 2003.
- Humphrey TJ and Lanning DG. The vertical transmission of salmonellas and formic acid treatment of chicken feed. Epidemiology and Infection, 100: 43**-**49. 1988.
- Japanese Feeding Standard for Poultry. National Agriculture and Food Research Organization. Japan Livestock Industry Association, Japan. 2011.
- Khempaka S, Koh K and Karasawa Y. Effect of shrimp meal on

growth performance and digestibility in growing broilers. Journal of Poultry Science, 43: 250**-**254. 2006a.

- Khempaka S, Mochizuki M, Koh K and Karasawa Y. Effect of chitin in shrimp meal on growth performance and digestibility in growing broilers. Journal of Poultry Science, 43: 339**-**343. 2006b.
- Khempaka S, Chitsatchapong C and Molee W. Effect of chitin and protein constituents in shrimp head meal on growth performance, nutrient digestibility, intestinal microbial populations, volatile fatty acids, and ammonia production in broilers. Journal of Applied Poultry Research. 20: 1**-**11. 2011.
- Meyers SP. Utilisation of shrimp processing wastes. Infofish Marketing Digest, 4: 18**-**19. 1986.
- National Agricultural Research Council Organization, NARO. Standard Tables of Feed Composition in Japan. Japan Livestock Industry Association, Japan. 2009.
- Ngoan LD, An LV, Ogle B and Lindberg JE. Ensiling techniques for shrimp by-products and their nutritive value for pigs. Asian-Australasian Journal of Animal Science, 13: 1278**-**1284. 2000.
- Oduguwa OO, Fanimo AO, Iyayi EA, Kalejaiye OO and Oyekola OA. Preliminary studies on the effects of different processing methods on the nutritive value of shrimp waste meal. Nigerian Journal of Animal Production, 25: 139**-**144. 1998.
- Oduguwa OO, Fanimo AO, Olayemi VO and Oteri N. The feeding value of sun-dried shrimp waste-meal based diets for starter

and finisher broilers. Archivos de Zootecnia, 53: 87**-**90. 2004.

- Rahman M and Koh K. Nutritional quality and *in vitro* digestibility of shrimp meal made of heads and hulls of black tiger (*Penaeus monodon*), white leg (*Litopenaeus vannamei*) and argentine red (*Pleoticus muelleri*) Shrimps. Journal of Poultry Science, 51: 411**-**415. 2014.
- Rahman M and Koh K. Effect of shrimp meal made of heads of black tiger (*Penaeus monodon*) and white leg (*Litopenaeus vannamei*) shrimps on growth performance in broilers. Journal of Poultry Science, 53: 149**-**152. 2016a.
- Rahman M and Koh K. Improvement in nutritional quality of shrimp meal with autoclave and chemical treatments: an *in vitro* study. Journal of Poultry Science, 53: 124**-**127. 2016b.
- Rosenfeld DJ, Gernat AG, Marcano JD, Murillo JG, Lopez GH and Flores JA. The effect of using different levels of shrimp meal in broiler diets. Poultry Science, 76: 581**-**587. 1997.
- SAS Institute. SAS Institute Inc (SAS). JMP, the Statistical Discovery Software. Version 10. Cary, NC. 2012.
- Shafey TM and McDonald MW. Effects of dietary calcium: available phosphorus on calcium tolerance of broiler chickens. Australian Journal of Experimental Agriculture, 30: 483**-**490. 1990.
- Smith OB and Kabaija E. Effect of high dietary calcium and wide calcium-phosphorus ratios in broiler diets. Poultry Science, 64: 1713**-**1720. 1985.