#### RESEARCH



# Ultra-Sensitive Fluorimetric Method for the First Estimation of Vonoprazan in Real Human Plasma and Content Uniformity Test

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## Abstract

Vonoprazan (VON) has been approved recently via US-FDA in 2015 as the first in class of potassium competitive acid blocker group. VON is used for management of GIT ulcer, reflux esophagitis and for eradication of Helicobacter pylori. So, the first spectrofluorimetric method was developed for estimation of VON in real human plasma and content uniformity test. The fluorimetric methodology based on reaction of secondary amine group in VON with benzofurazan (0.05% w/v NBD-Cl) reagent as nucleophilic substitution reaction in alkaline medium (0.1 M borate buffer pH 8.2) to produce highly fluorescent product measure at 530 nm after excitation at 465 nm. The linear calibration range was found 15 to 200 ng mL<sup>-1</sup> with lower limit of quantitation (LOQ) equal to  $8.57 \text{ ng mL}^{-1}$ . The method was successfully applied for estimation of VON in pharmacokinetic (PK) and content uniformity studies. The maximum plasma concentration was found to be ( $C_{max}$ ) 71.03 ng mL<sup>-1</sup> after maximum time ( $t_{max}$ ) equal to  $1.5 \pm 0.15$  h. The presented strategy also applied to ensure concentration of drug in each tablet using content uniformity test with high percent of recovery  $100.05 \pm 0.66$ . The proposed method was established for clinical laboratories and therapeutic drug monitoring studies.

Keywords Vonoprazan · Content uniformity · Pharmacokinetics · NBD-Cl

#### Abbreviations

NBD-Cl7-Chloro-4-nitrobenz-2-oxa-1,3-diazoleVONVonoprazanPKPharmacokineticAUCArea under curve

# Introduction

VON (Fig. 1), IUPAC name: 1-(5-(2-fluorophenyl)-1-(pyridin-3- ylsulfonyl)-1H-pyrrol – 3-yl)-Nmethylmethanamine fumarate. VON is the first drug as potassium competitive acid blocker group used for management GIT ulcer and esophageal reflux [1]. In last few years, the fluorimetric approach is the most applicable approach due to high sensitivity, simplicity, economical, easy handling in clinical laboratories and its selectivity for estimation of varying pharmaceutical products [2–4]. Few analytical methods have been reported for estimation of VON as high performance liquid chromatography (HPLC) [5, 6] and electrochemical method [7]. Some drawbacks were reported in these methods as using expensive equipment's, reagents, time consuming, and lack to sensitivity. The presented work is the first spectrofluorimetric method for estimation of VON in human plasma and content uniformity test with high sensitivity. 7-chloro-4-nitrobenz-2-oxa-1,3-diazole is neucliphic substitution reagent used for fluorimetric derivatization reactions with amine groups (primary and secondary) to produce highly fluorogenic product in alkaline medium [2, 8–11].

The aim of this study is to develop ultra-sensitive, selective, and economical fluorimetric method. The method was successfully applied to a pharmacokinetic study in healthy human volunteers and content uniformity test in tablet dosage form. The pharmacokinetic parameters were studied as maximum plasma concentration, half-life and maximum time for absorption which make the presented method easily used for therapeutic drug monitoring system.

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Ν

s′=0

Fluorescent product (530nm)

Fig. 1 The proposed reaction mechanism between VON and NBD-Cl

Ν

# **Experimental Section**

## Instruments

The spectrofluorimetric measurements were done using Scinco (FS-2 fluorimeter, Scinco, Korea), with 1 cm quartz cell, excitation and emission slit width set at 5 nm and photo multiplied tube (PMT) voltage 400 V). Water bath (WB-1R1H-3 thermostatic) China, Benchtop pH meter (China), high speed centrifuge (China), MS-TS analytical digital balance (USA). MATLAP software for PK analysis.

## **Chemical and Solutions**

Vonoprazan (VON 99.89%) and vonaspire® tablets (20 mg/ tablet) was obtained from obtained from Alrowad pharmaceutics, Egypt. 4-Chloro-7-nitrobenzofurazan (NBD-Cl) was obtained from Sigma Aldrich (USA). Methanol, ethanol, acetonitrile, boric acid, acetone, borax, sodium hydroxide and hydrochloric acid were obtained from El-Gomhoria Co, Assiut, Egypt. Human plasma samples were achieved from 6 healthy human volunteers following ethical standards of the responsible committee and kept at -10 degree.

# **Preparation of Solutions**

Stock solution of VON was prepared using 5 mg of authentic powder dissolved in ultra-pure water 50 mL into 100 mLvolumetric flask. The solution was sonicated for 3 mint to ensure the complete solubility of VON drug then completed to the mark with the same solvent. Further dilutions with distilled water were performed to prepare working solution in a concentration range from  $(150 - 2000 \text{ ng mL}^{-1})$ .

Borate buffer (0.1 M) was prepared [9, 10] and adjusted to pH range using 0.1 M NaOH and 0.1 M HCl.

NBD-Cl (0.05% w/v) reagent prepared by 5 mg powder was dissolved using 50 mL methanol into a 100 mL volumetric flask, then completed with methanol to the mark.

## **Preparation of Samples**

## **Biological Samples**

Plasma samples were collected and carried out according to (ethical standards of the responsible committee on human experimentation) (institutional and national) and with Helsinki Declaration of 1975, as revised in 2008.

## **Spiked Plasma Samples**

Five milliliters of the blood samples were collected from 6 healthy human volunteers (males and females aged from 30

to 40 years) into 10 mL screw-capped heparinized tube. The tubes were centrifuged at 5000 rpm for 25 min to separate the plasma. The plasma samples (300  $\mu$ L) were spiked with different concentrations of working solution of VON. Two milliliters of methanol were added as protein precipitating agent. The resultant solution was completed to 10 mL with ultra-pure water and then centrifuged at 4000 rpm for 20 min. One milliliter from the supernatant was used at general procedures.

#### **Pharmacokinetic Study**

*The blood samples* were collected from 6 healthy human (males and females aged from 30–40 years) volunteers after administration vonaspire® tablets (40 mg/tablet VON) as a single oral dose. After that, serial venous blood samples (5 mL) were collected in heparinized tubes for 24 h after dosing (0 h) and after 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6, 10, 15, 20 and 24 h). The tubes were centrifuged at 5000 rpm for 25 min to separate the plasma. The plasma samples frozen at -20 until analysis. Then the samples treated as spiked method without addition of the drug.

#### Pharmaceutical Dosage Form

Ten vonaspire (1) tablets (40 mg VON/tablet) were accurately weighed, finely powdered and mixed. An accurately weighed amount of the powdered tablets equivalent to 10 mg of VON was transferred into 100-mL volumetric flask followed by addition of 60 mL of ultra-pure water. The solution was sonicated for 5 min then followed by filtration and dilution with ultra-pure water to provide final concentration of 100  $\mu$ g mL<sup>-1</sup>. A series of working standard solutions with concentration range between 150 ng mL<sup>-1</sup>- 2000 ng mL<sup>-1</sup> were prepared in volumetric flasks.

*The content uniformity* testing was assessed and performed through analysis of ten tablets individually according to official USP guidelines [12]. Each tablet was individually weighed, finely powdered, and analyzed using the fluorescence procedure.

## **Fluorescence Procedure**

One milliliter from VON working solution (150 – 2000 ng mL<sup>-1</sup>) was transferred into a series of 10-mL stoppered autoclaved test tubes, 1.0 mL of 0.1 M borate buffer (pH 8.2) was added followed by 0.5 mL of NBD-Cl (0.05% w/v). The tubes were heated in water bath at 85 °C for 15 min. The tubes were rapidly cooled in ice water bath and 0.75 mL from 2 M HCl solution was added. The solution quantitatively transferred to 10-mL calibrated flask and completed to the mark with methanol in order to provide final concentration range (15 – 200 ng mL<sup>-1</sup>). The fluorescent product was measured at 530 nm ( $\lambda$ ex at 465 nm). The blank experiment was performed similarly omitting the drug.

## **Stoichiometry Mechanism**

The Stoichiometry of the reaction was studied using equimolar concentration  $(1 \times 10^{-3} \text{ M})$  from VON and NBD-Cl (0.05% w/v) by application continuous variation method [2, 9, 13]. A series of solutions where portions of mixture of master solutions of VON and NBD-Cl were made up as (0.1: 0.9, 02: 0.8, 0.3: 0.7, 0.4: 0.6, 0.5: 0.5, 0.6: 0.4, 0.7: 0.3, 0.8: 0.2 and 0.9: 0.1, respectively). The fluorescence procedure was followed.

# **Results and Discussion Part**

NBD-Cl is nucleophilic substitution reagent reacts with primary and secondry amine groups producing highly fluorescent product in alkaline medium [9, 10, 13]. However, some drawbacks were reported in published method as long heating time (55 min) which consider time consuming [13, 14], high concentration of the reagent [11, 13, 15]. While, the presented study provides sensitive, time saving and economical method for analysis of VON in human plasma which applied in pharmacokinetic study and content uniformity test. In the developed work, the reaction of secondary amine moiety in VON structure with NBD-Cl (0.05%w/v) was achieved using borate buffer at pH  $8.2 \pm 0.2$  and heating at 85 °C for only 15 min as in Fig. 1 with stoichiometry of the reaction 1:1. The fluorogenic product measure at 530 nm after excitation at 465 nm (Fig. 2).

# **Optimization of the Methodology**

The fluorescence reaction between VON and NBD-Cl is based on different factors as; pH, temperature, heating time, NBD-Cl concentrations, and diluting solvents.

**Fig. 2** Excitation and emission spectra for reaction of VON (100 ng mL.<sup>-1</sup>) with NBD-Cl



Fig. 3 Effect of pH range (0.1 M borate buffer) on the reaction of VON (100 ng mL. $^{-1}$ ) with NBD-Cl (0.05% w/v)

# Effect of Buffer System and pH

The nucleophilic substitution reaction between VON and NBD-Cl reagent for formation highly fluorescent product is pH dependent. Hence, different type of buffers was studied as (Teorell, borate, phosphate and BR) as shown in Fig. S1 (supplementary Materials). The maximum fluorescent intensity was obtained using 0.1 M borate buffer (pH  $8.2\pm0.3$ ) (Fig. 3).

Using borate buffer at pH more than pH 8.6 led to decrease in the fluorescence intensity due to the increase in hydroxide ions concentration which holds back the reaction between VON and NBD-Cl and this agrees with previously reported studies [2, 9, 10, 15]. Moreover, the effect of buffer volume on the fluorescence intensity was studied, where different volumes from 0.25 to 3 mL were investigated. It was found that maximum RFI obtained using  $1.0 \pm 0.25$  mL of borate buffer pH 8.2 (Fig. S2).

# Effect of NBD-Cl Concertation

Varying volumes of NBD-Cl reagent (0.1 to 0.9 mL) were utilized to study the effect of the reagent. As shown in Fig. 4







the maximum RFI was achieved via applying 0.5 mL of (0.05% w/v) NBD-Cl, after trying varying volumes of NBD-Cl reagent. Increasing the volume of NBD-Cl than 0.7 mL led to decrease RFI and increasing of the blank intensity. So, 0.5 mL of (0.05% w/v) NBD-Cl was used as the optimum condition.

# Effect of Temperature and Heating Time Effect

The nucleophilic substitution reaction between secondary amine moiety in VON and Cl<sup>-</sup> in NBD-Cl is based on temperature and heating time. As seen in Fig. 5, varying temperature degrees were studied under time intervals, the maximum RFI was obtained using  $85 \pm 5$  °C for 15 min.

# **Effect of HCI Concentration**

Acidification of the reaction media between VON and NBD-Cl is required before the measuring of results because formation of hydroxy-7-nitrobenzo-2-oxa-1,3-diazole bi- product (NBD-OH) [2, 9, 10]. The NBD-OH was quenched using  $0.75 \pm 0.25$  mL from 2 M HCl without any effect on the reaction results (Fig. S3).

# **Effect of Diluting Solvents**

The influence of diluting solvents as distilled water, methanol, ethanol, acetonitrile, acetone and DMF was studied. Methanol was observed to be the best diluting solvent as it gave the highest fluorescence with good reproducibility and low blank readings.

# Validation of the Methodology

## **Linearity Range and Sensitivity**

The calibration range of the reaction was achieved by plotting concentration of VON versus RFI. The linear equation was found y = 1.5401x + 3.103 with standard



Fig. 5 Effect of temperature and heating time for reaction of VON (100 ng mL. $^{-1}$ ) with NBD-Cl (0.05% w/v)

deviation (SD) equal to 1.31. the linear range was found to be  $15-200 \text{ ng mL}^{-1}$  with LOQ equal to 8.5 ng mL<sup>-1</sup> which indicated to the ultra-sensitivity of the fluorimetric method as in Table 1. Lower limit of quantitation and detection were calculated using International Council for Harmonization guidelines [16] using following equations: LOQ = 10  $\sigma$ / slope and LOD = 3.3  $\sigma$ / slope. ( $\sigma$ ) is the standard deviation.

## **Accuracy and Precession**

Five concentrations within the calibration range were used for studying the accuracy of the proposed method (20,30, 50, 100, 150 ng mL<sup>-1</sup>) as seen in Table 2. The results refer to high accuracy of the proposed method with high percent of recovery ranged from 99.53 to 101.62%.

Besides, 3 concentrations were applied for intraday and inter-day precession study. The results refer to excellent precision of the proposed method with high percent of recovery  $101.43 \pm 0.34$ .

In addition, bio-analytical validation of the reaction between VON and NBD-Cl was studied using six concentrations according to US-FDA recommendations [17]. According to the results in Table 3, the results refer to high accuracy and precision of the proposed method in human plasma.

#### Robustness

The robustness of this study was checked by applying minor changes on parameters of analytical procedure. The results in table (Table S1) there was no significant effect for small change in method variables.

 Table 1 Quantitative analytical parameters for estimation of VON using the proposed method

Parameters	VON
λex (nm)	465
λem(nm)	530
Concentration range (ng mL <sup>-1</sup> )	15 - 200
Correlation coefficient (r)	0.9997
Determination coefficient (r <sup>2</sup> )	0.9998
Slope	1.54
Intercept	3.10
SD the intercept (Sa)	1.31
*LOD ( ng mL <sup><math>-1</math></sup> )	2.80
**LOQ (ng mL <sup>-1</sup> )	8.50

\*LOD Limit of detection

\*\*LOQ Limit of quantitation

Table 2 Accuracy of the proposed method for estimation of VON using NBD-Cl reagent

Sample	VON				
number	Taken (ng mL <sup>-1</sup> )	Found (ng mL <sup>-1</sup> )	% Recovery*	% RSD	
1	20	19.94	99.70	0.61	
2	30	29.86	99.53	0.94	
3	50	50.20	100.04	1.09	
4	100	100.16	100.16	0.98	
5	150	152.44	101.62	0.81	

\*Average of five determinations

# Incurred Sample Reanalysis (ISR)

Further incurred sample reanalysis (IRS) was carried out to evaluate accuracy and precision of incurred sample using FDA guidelines. The difference values between initial samples and incurred samples do not exceed than 1.82 to 4.03% (Table S2).

# Selectivity of the Proposed Method

The method selectivity was checked to observe any interference from excipient. VON was studied in the presence of varying different excipient as talc 10 mg, starch 100 mg, mannitol 10 mg, magnesium stearate 10 mg, lactose 10 mg, and sodium chloride 10 mg. The results refer to there was no interference from the excipients which refer to the high selectivity of the proposed methods (Table S3).

# **Stability of VON in Human Plasma**

Varying parameters were applied to check the stability of VON in plasma samples as three Freeze–thaw cycle stability (-24 °C), long-term stability (1 month at -24 °C), short-term stability (12 h at -24 °C), post-preparative stability (6 h at

 Table 3
 Accuracy and precision of the fluorimetric reaction in human plasma

Intra-day assay(n=6)		Inter-day assay(n = 18)		
Conc. (ng mL <sup>-1</sup> )	Accuracy (%)	Precision (% RSD)	Accuracy (%)	Precision (% RSD)
15	97.33	1.18	97.00	0.98
20	97.15	1.55	96.94	1.08
60	97.00	1.30	96.94	1.90
100	98.09	1.15	97.60	1.11
150	96.21	1.76	96.08	1.46
200	97.38	1.32	96.88	1.80

room temperature 25 °C), post-preparative stability (12 h at room temperature 25 °C) using three concentrations, low quality control (LQC 20 ng mL<sup>-1</sup>), medium quality (MQC 100 ng mL<sup>-1</sup>) and high quality control (HQC 150 ng mL<sup>-1</sup>) levels. The percentage of recoveries compared with freshly prepared samples between  $95.11 \pm 1.79$  and  $98.02 \pm 1.60$  (Table S4).

## **Applications of the Proposed Method**

The ultra-sensitivity of the fluorimetric method was utilized for estimation of VON in spiked human plasma. The percent of recovery was found to be in the range  $96.16 \pm 1.21$  to  $98.80 \pm 0.92\%$  for the investigated method at 6 different concentration levels (Table 4).

In addition to, VON was determined in real human plasma (pharmacokinetic study) using 6 healthy human volunteers, VON concentration was calculated in the plasma samples after time intervals (0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6, 10, 15, 20 and 24 h).

The peak plasma level (C max) of cited drug was found to be  $71.03 \pm 1.12$  ng mL<sup>-1</sup> after administrated single oral dose 40 mg/tablet. The maximum time for absorption was found (t<sub>max</sub>) 1.5 h. with AUC equal to  $429.45 \pm 55.45$  ng h. mL<sup>-1</sup> (Fig. S4).

The results of the proposed method agrees with previously reported method [18] which refer to high sensitivity and good reliability of the proposed method.

Besides, the proposed method was successfully applied for determination of VON in pharmaceutical dosage form, and the obtained results were found to be satisfactory with a good recovery ( $102.40 \pm 0.70$ ). Using *student*'s *t*-test (1.40) and *F*-test (2.65) with respect to accuracy and precision and showed no significant difference between the calculated and theoretical values of both the proposed and the reported method [5] at 95% confidence level.

To ensure the consistency of dosage units, each unit in a batch should have a drug substance content within a narrow range around the label claim. For VON, due to the high sensitivity of the proposed method and its ability to rapidly measure the fluorescence intensity of a single

Table 4 Application of the proposed method in spiked human plasma

Added conc. (ng mL <sup>-1</sup> )	% Recovery <sup>*</sup> ± RSD
15	$96.02 \pm 1.20$
50	$97.10 \pm 1.40$
100	$98.00 \pm 1.52$
130	$97.41 \pm 1.71$
150	$96.45 \pm 1.60$
200	$97.30 \pm 1.08$

\*Average of six determinations

 Table 5
 Content uniformity for determination of VON using the proposed method

	(vonaspire® tablets 40 mg)
1	99.81
2	99.95
3	100.19
4	99.77
5	100.62
6	98.90
7	101.01
8	100.60
9	98.63
10	101.02
Mean	100.05
SD	0.66
RSD	0.65
Acceptance value (AV)*	1.58
Max. allowed AV (L1)*	15

\*Acceptance value =  $2.4 \times SD$ 

tablet extract with sufficient accuracy, the method was ideally suited for content uniformity testing which is a timesaving process when using other conventional assay techniques. The steps of the test were adopted according to the USP procedure [3, 12]. The content of individual dosage form was analyzed. The acceptance value (AV) was calculated, and it was found to be smaller than the maximum allowed acceptance value (L1). The results demonstrated an excellent drug uniformity as shown in Table 5.

# Conclusion

Eventually, as far as we know, this is the most sensitive (LOQ 8.50 ng mL<sup>-1</sup>) and selective method for rapid and economic estimation of VON in real human plasm and in formulations using NBD-Cl reagent in alkaline medium (pH 8.2). The method was applied in pharmacokinetic study with maximum plasma concentration equal to  $71.03 \pm 1.12$  ng mL<sup>-1</sup>. The presented study easily applied in clinical laboratories and therapeutic drug monitoring system.

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**Availability of Data and Material/Data Availability** All data generated or analyzed during this study are included in this published article (and its supplementary information files).

# Declarations

**Ethics Approval** Plasma samples were collected and carried out according to (ethical standards of the responsible committee on human experimentation) (institutional and national) and with Helsinki Declaration of 1975, as revised in 2008.

**Consent to Participate** The human experimental carried out at human healthy volunteers according to Helsinki Declaration of 1975, as revised in 2008.

**Conflicts of Interest** The authors declare they have no competing interests.

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# References

- Garnock-Jones KP (2015) Vonoprazan: first global approval. Drugs 75:439–443. https://doi.org/10.1007/s40265-015-0368-z
- Hussein SA, Salman BI, Ali MF, Marzouq MA (2019) Development of sensitive benzofurazan-based spectrometric methods for analysis of spectinomycin in vials and human biological samples. Luminescence 34:895–902. https://doi.org/10.1002/bio.3688
- Hammad MA, Omar MA, Salman BI, Derayea SM (2016) Highly sensitive spectrofluorimetric method for determination of doxazosin through derivatization with fluorescamine; Application to content uniformity testing. Spectrochim Acta A 157:55–60. https:// doi.org/10.1016/j.saa.2015.12.012
- Marzouq MA, Salman BI, Hussein SA, Ali MF (2019) Hantzsch reaction approach for determination of teicoplanin and vancomycin in real human plasma: Application to pharmaceutical preparations and to synthetic mixture with rifampicin for drug-resistant strain of Staphylococcus aureus. Microchem J 147:25–29. https:// doi.org/10.1016/j.microc.2019.03.002
- 5. Yoneyama T, Teshima K, Jinno F, Kondo T, Asahi S (2016) A validated simultaneous quantification method for vonoprazan

(TAK-438F) and its 4 metabolites in human plasma by the liquid chromatography-tandem mass spectrometry. J Chromatog B 1015:42–49. https://doi.org/10.1016/j.jchromb.2016.01.051

- Qiao Y, Huang J, Xu Y, Zhao J, Wang Q (2018) Determination of vonoprazan pyroglutamate and vonoprazan fumarate by HPLC. China Pharma 12:535–538
- Al-Qahtani SD, Al-nami SY (2021) Copper oxide nanopowder modified carbon paste electrode for the voltammetric assay of vonoprazan. Arab J Chem 14:103254. https://doi.org/10.1016/j. arabjc.2021.103254
- Salman BI, Hussein SA, Ali MF, Marzouq MA (2019) Innovative ultra-sensitive spectrofluorimetric method for nanogram detection of doripenem monohydrate in human plasma, urine and pharmaceutical formulation. Microchem J 145:959–965. https://doi.org/ 10.1016/j.microc.2018.12.018
- Salman BI, Ali MF, Marzouq MA, Hussein AS (2019) Utility of the fluorogenic characters of benzofurazan for analysis of tigecycline using spectrometric technique; application to pharmacokinetic study, urine and pharmaceutical formulations. Luminescence 34:175–182. https://doi.org/10.1002/bio.3590
- Omar MA, Hammad MA, Awad M (2017) Utility of 4-chloro-7nitrobenzo-2-oxa-1, 3-diazole for development of a highly sensitive stability indicating spectrofluorimetric method for determination of salmeterol xinafoate; application to human plasma. RSC Adv 7:44773–44779. https://doi.org/10.1039/C7RA08106E
- Darwish IA, Amer SM, Abdine HH, Al-Rayes LI (2009) Spectrofluorimetric determination of fluvoxamine in dosage forms and plasma via derivatization with 4-chloro-7-nitrobenzo-2-oxa-1,3-diazole. J Fluoresc 19:463–471. https://doi.org/10.1007/ s10895-008-0433-z
- U.S. Pharmacopoeial Convention (2011) (905) Uniformity of Dosage Units. Stage 6 Harmonization 3:4–6
- Walsh M, Belal F, El-Enany N, Elmansi H (2011) Spectrofluorimetric determination of paroxetine HCl in pharmaceuticals via derivatization with 4-chloro-7-nitrobenzo-2-oxa-1, 3-diazole (NBD-Cl). J Fluoresc 21:105–112. https://doi.org/10.1007/ s10895-010-0693-2
- Aktas ES, Ersoy L, Sagırlı O (2003) A new spectrofluorimetric method for the determination of lisinopril in tablets. Il Farmaco 58:165–168. https://doi.org/10.1016/S0014-827X(02)00013-7
- Kepekci SE, Önal E (2011) Spectrofluorimetric methods for the determination of gemifloxacin in tablets and spiked plasma samples. J Fluoresc 21:1001–1007. https://doi.org/10.1007/ s10895-010-0759-1
- 16. International Conference on Harmonization (2005) Topic Q2 (R1) Validation of analytical procedures: text and methodology
- Zimmer D (2014) New US-FDA draft guidance on bio-analytical method validation versus current FDA and EMA guidelines: chromatographic methods and ISR. Bioanalysis 6:13–19. https://doi. org/10.4155/bio.13.298
- Sakurai Y, Nishimura A, Kennedy G, Hibberd M, Jenkins R, Okamoto H, Yoneyama T, Jenkins H, Ashida K, Irie S, Taubel J (2015) Safety, tolerability, pharmacokinetics, and pharmacodynamics of single rising TAK-438 (vonoprazan) doses in healthy male Japanese/non-Japanese subjects. Clin Transl Gastroenterol 6:e94. https://doi.org/10.1038/ctg.2015.18

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