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ORIGINAL RESEARCH

# MicroRNA 885-5p Inhibits Hepatocellular Carcinoma Metastasis by Repressing AEGI

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**Background:** Hepatocellular carcinoma (HCC) is the third major cause of cancer-related death. Mounting evidence shows that microRNAs play critical roles in the initiation and progression of HCC and may potentially serve as diagnostic markers for HCC.

**Methods and Results:** In the present study, we explored the biological effects of miR-885-5p on HCC progression. We performed flow cytometry analyses of miR-885-5p in HCC cell lines and identified miR-885-5p as a recurrence-related microRNA. Overexpression of miR-885-5p significantly inhibited cell migration, invasion, proliferation, angiogenesis and EMT. Then, the correlation of miR-885-5p and AEG1 were confirmed by using luciferase assays, quantitative real-time PCR analysis and Western blotting. It was subsequently confirmed that Astrocyte Elevated Gene1 (AEG1) was a direct target gene of miR-885-5p.

**Conclusion:** miR-885-5p likely acts as a tumor suppressor by regulating AEG1, suggesting that miR-885-5p may be a potential biomarker and can be targeted in therapeutic strategies against HCC in the future.

Keywords: miR-885-5p, AEG1, biomarker, EMT, hepatocellular carcinoma

#### Introduction

Hepatocellular carcinoma (HCC) is one of the most common malignancies in the world, and a highly invasive tumor with frequent distant metastasis in China.<sup>1,2</sup> The underlying pathogenesis of HCC may be very complicated, though most HCC cases may be caused by hepatitis B or hepatitis C virus infection and cirrhosis.<sup>3,4</sup> Unfortunately, despite recent advances in HCC early diagnosis and developed treatment approaches, HCC patients still suffer from poor prognosis and high recurrence rate.<sup>5</sup> Consequently, it is of crucial importance to identify molecular pathways regulating HCC metastasis in order to develop novel treatment strategies.

MicroRNAs, small non-coding endogenous RNAs, act as post-transcriptional regulators by binding to 3'-untranslated region (3'-UTR) of the target mRNAs and have been demonstrated to be associated with a wide range of pathological processes, including cell differentiation, proliferation, apoptosis, invasion and migration.<sup>6,7</sup> In the past few years, there has been growing evidence focused on the function of miR-885-5p in tumorigenesis. MiR-885-5p has been identified as a tumor suppressor in glioblastomas, neuroblastomas and pancreatic cancer.<sup>8–10</sup> Although Zhang et al reported that miR-885-5p contributes to suppression of HCC metastasis,<sup>11</sup> whether miR-885-5p may also be dysregulated and function as miR-885-5p-sponging epigenetic regulators in HCC is not clear.

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# **Methods**

### Cell Culture and Transfection

Human cancer cell lines HepG2 and Hela cells purchased from Type Culture Collection of Chinese Academy of Sciences (Shanghai, China) were maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (GIBCO, Carlsbad, CA, USA), 100 U/mL penicillin G and 100  $\mu$ g/mL streptomycin at 37 °C in a humidified incubator containing 5% CO<sub>2</sub>. HepG2 cells were transfected with either miR-885-5p mimics (5'-UCC AUU ACA CUA CCC UGC CUC U-3') or negative control (5'-UUC UCC GAA CGU GUC ACG UTT-3') (RiboBio, Guangzhou, China) using LipofectamineTM RNA iMax transfection reagent according to the manufacturer's protocols.

# Cell Viability and Flow Cytometry

The cell viability of HepG2 was assessed at 72 hrs after transfection with either miR-885-5p mimics or negative control, using a Cell Counting Kit-8 (Dojino, Japan), following the manufacturer's instructions. Absorbance was then measured at 450 nm using Elx800 Reader (Bio-Tek, USA).

For flow cytometry studies, HepG2 cells were transfected with either miR-885-5p mimics or negative control for 48 h, respectively. Then, the cells were double-stained with Annexin V or propidium iodide, according to the manufacturer's instructions. Cells were analyzed using FACS Calibur flow cytometer (BD Biosciences, USA), followed by flowjo7.6 software analysis. Experiments were conducted using three independent biological samples.

# Transwell Migration and Tube Formation Assay

Transwells with 8-µm pore size filters covered with (BD Biosciences, USA) were inserted into 24-well plates for invasion or migration assays, respectively. Medium (600 µL) containing 10% FBS was added to the lower chamber, and 100 µL of a serum free cell suspension ( $1 \times 10^5$  cells) was placed onto the upper chamber. The cells that did not migrate or invade after 24 h of incubation were removed from the upper face of the filters by scrubbing with a cotton swab. Membranes were then

fixed with 4% formaldehyde for 30 min at room temperature and stained with DAPI. In each sample, the number of migrating or invading cells was counted at  $20 \times$  magnification from five different fields for each filter and analyzed to determine statistically significance. For invasion assays, cells were seeded onto a matrigel-coated chamber and were incubated at 37 °C.

HUVEC purchased from Type Culture Collection of Chinese Academy of Sciences (Shanghai, China) was seeded onto a 96-well plate coated with Matrigel (BD Biosciences, USA) followed by transfection with miR-885-5p. Peak tube formation was observed at 8 h post-treatment. The number of tubes for each treatment was quantified.

### Immunofluorescence

Cells seeded on glass coverslips in 6-well plates were fixed in 4% of formaldehyde solution and permeabilized with 1% of Triton/PBS. Cells were blocked with 1% of BSA/PBS for 1 h at room temperature and incubated with E-cadherin or Vimentin antibodies (Proteintech, USA) for 2 h at room temperature, followed by the incubation with fluorescent-dye conjugated secondary antibody (Cell Signaling Technology, USA) for 1 h. Samples were stained with DAPI and images were taken under an inverted fluorescence microscope.

# Western Blotting and Real-Time PCR

The expression of cellular proteins was analyzed by Western blotting. The following antibodies were used in our study: AEG1, PD-L1 and EGFR (Abacm, USA),  $\beta$ -actin (Proteintech, USA) and the secondary antibodies (Cell Signaling Technology, USA).

Total RNA was extracted using RNA Trizol (Invitrogen, USA) according to the manufacturer's instructions. PrimeScript RT reagent kit (Takara, Japan) was used to synthesize cDNA according to the manufacturer's instructions. Real-time PCR (qRT-PCR) and Stem-loop qRT-PCR were performed using SYBR Mixture (Vazyme Biotech, China) in 96 well optical plates at 95 °C for 5 mins, followed by 40 cycles of 95 °C for 10 seconds and 60 °C for 30 min. After the reactions, the cycle threshold (Ct) data were determined using the default threshold settings. The relative gene expression levels were normalized to GAPDH. The primers for PCR were listed (5'-3'): AEG1 forward: CCT GGC CTT GCT GAA GAA TC; AEG1 reverse: GGC TGC TTT GCT GTT ACA CT; PD-L1 forward: GGC ATT TGC TGA ACG CAT; PD-L1 reverse: CAA TTA GTG CAG CCA GGT; EGFR forward: TCT ACA ACC CCA CCA CGT AC; EGFR reverse: TTC CGT TAC ACA CTT TGC GG; GAPDH forward: CAT GAG AAG TAT GAC AAC AGC CT; GAPDH reverse: AGT CCT TCC ACG ATA CCA AAG T. The relative miR-885-5p expression was normalized to U6 snRNA. The primers for PCR were listed (5'-3'): miR-885-5p forward: GTC CAT TAC ACT ACC CTG CCT C; miR-885-5p reverse: CGC GAG CAC AGA ATT AAT ACG; U6 snRNA forward: CTC GCT TCG GCA GCA CA; U6 snRNA reverse: AAC GCT TCA CGA ATT TGC GT.

#### Dual Luciferase Reporter Assay

The 3'-UTR region of AEG1 with the potential binding sites for intact or mutant sites of miR-885-5p were generated, followed by the fusion into pGL3 promoter vector (Promega, USA) named as AEG-1 3'-UTR (AEG1-WT) or a mutated 3'-UTR of AEG-1 (AEG1-MU). Sequences were amplified according to the following primers: AEG1-WT forward: 5'- CTA GTC TAG AAT TAC CTC CCC CAC CCC CAA GAA CAC TT -3', and reverse: 5'- CCG

GAA TTC GGA ACA TTA TTA GCT GCT ACA CTA AGC -3'; AEG1-MU forward: 5'- TCT TCT CAA ATC GAA TTA CCT CAC CTC TAA CAG TGT CTG TC -3', and reverse: 5'- AGG TAA TTC GAT TTG AGA AGA TCC TAA CAG AAT CTC AAA AT-3'. Luciferase reporter experiments were conducted in 96 well plates by using Dual Luciferase Reporter Assay kit (Promega, USA), and SpectraMax M5 instrument software (Molecular Devices, USA) was used to analyze the results. AEG1-WT or AEG1-MU was co-transfected with either miR-885-5p mimics or the negative control (miR-NC) using 50 nM Lipofectamine 2000 (Life Technologies, USA). After 48 h, cells were analyzed using a Dual-Luciferase Reporter Assay kit in accordance with the manufacturer's guidance. Data were normalized using the relative luciferase activity was normalized to Renilla luciferase activity.

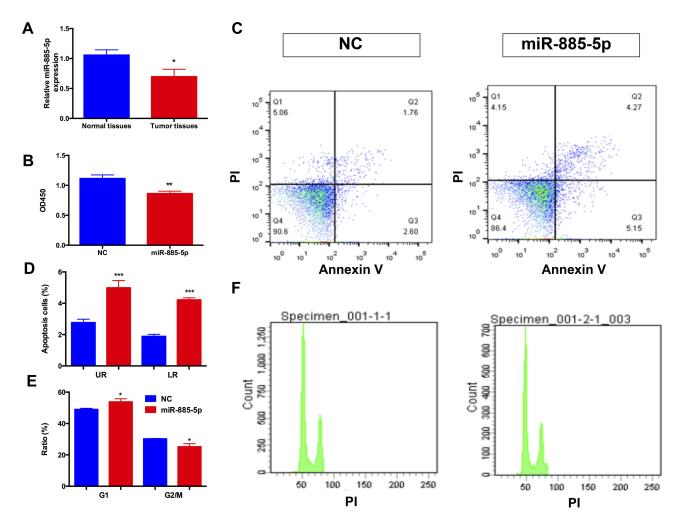


Figure I Overexpression miR-885-5p inhibits the metastasis of HCC cells.

**Notes:** (**A**) Relative miR-885-5p expressions measured by qRT-PCR in liver tumor tissues and paired normal tissues (\*p < 0.05). (**B**) CCK8 assays were applied to determine the effect of miR-885-5p on HCC cells proliferation ability (\*p < 0.01). (**C**) (**D**) Flow cytometry assays were employed to exam the function of miR-885-5p on the apoptosis capacity of HCC cells (\*\*p < 0.001). (**E**, **F**) Flow cytometry assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK8 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05). (**B**) **C**CK9 assays were employed to exam the function of miR-885-5p on the cell cycle capacity of HCC cells (\*p < 0.05).

### Statistical Analysis

All experiments were performed in triplicate. All the data are expressed as the mean  $\pm$  SEM and were analyzed using GraphPad Prism 6 software (GraphPad Software, Inc, USA). Differences between individual groups were analyzed by Student's *t*-tests. *p* < 0.05 was considered statistically significant.

## Results

# Overexpression miR-885-5p Inhibits Proliferation as Well as Cell Cycle Progression and Promotes Cell Apoptosis in HCC Cells

The relative expression levels of miR-885-5p in HCC samples and adjacent normal tissue samples were determined by qRT-PCR. As presented in Figure 1A, the expression of miR-885-5p in tumor tissue samples were reduced compared with adjacent normal tissues,

respectively. Next, to explore the function of miR-885-5p on metastasis of HCC cells, we used HepG2 cells were transfected with miR-885-5p mimics, respectively by using miR-NC as a negative control (NC). We performed CCK-8 assay to determine the effect of miR-885-5p on the proliferative capacity of HepG2 cells. As demonstrated in Figure 1B, miR-885-5p significantly decreased the proliferation of HepG2 cells at 3 days post-transfection. Then we evaluated the effect of miR-885-5p on cell apoptosis and cell cycle progression, as shown in Figure 1C–F, flow cytometry analysis indicated that miR-885-5p significantly promoted apoptosis and caused cell cycle arrest at G1 phase in HepG2 cells.

# Overexpression of miR-885-5p Inhibits Tumor Angiogenesis and EMT

To examine whether miR-885-5p functionally contributed to migratory capacities of HCC cells, transwell assay was

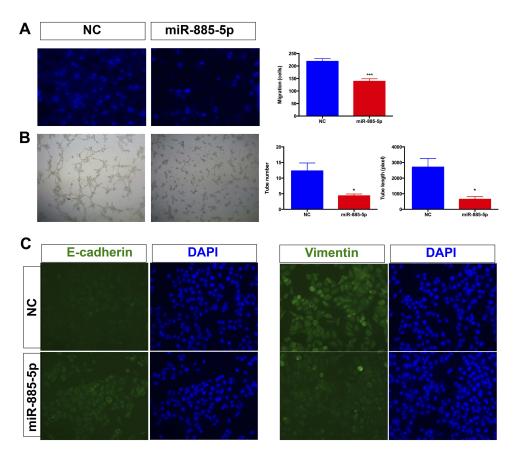


Figure 2 Overexpression of miR-885-5p inhibits tumor angiogenesis and EMT.

**Notes:** (A) Transwell assays were employed to exam the function of miR-885-5p on the migration capacity of HCC cells (\*\*\*p < 0.001). (B) Tube number and tube length in HUVEC cells were performed to investigate the function of miR-885-5p on angiogenesis (\*p < 0.05). (C) Immunofluorescence was performed to investigate the function of miR-885-5p on EMT formation of HCC cells.

Abbreviations: HCC, hepatocellular carcinoma; EMT, Epithelial-Mesenchymal Transition; NC, negative control.

performed in HCC cells after transfection with miR-885-5p mimics or miR-NC. The results demonstrated that overexpression of miR-885-5p potentially suppresses the migratory ability of HCC cells (Figure 2A). To evaluate the effects of miR-885-5p on angiogenesis, we conducted angiogenesis assay by assessing the tubes formation of HUVEC cells in vitro for angiogenesis assays. Our results demonstrated that miR-885-5p reduces the angiogenic tubes formation of HUVEC cells. Furthermore, miR-885-5p also inhibited the formation of new blood vessels in the matrix gel (Figure 2B). It is established that EMT process can promote the migratory capacities of cancer cells. Next, examined the EMT markers in HCC cells. we Immunofluorescence analysis was employed to examine the levels of epithelial marker (E-cadherin) and mesenchymal marker (Vimentin) in cells transfected with miR-885-5p mimics or miR-NC. As shown in Figure 2C, the result of immunofluorescence hinted that the overexpression of miR-885-5p in HepG2 cells increased level of epithelial marker (E-cadherin) and decreased level of mesenchymal (Vimentin) marker. These data indicated that miR-885-5p reduces migratory ability of HCC cells through inhibiting the EMT pathway.

#### AEG1 Is Identified as a Target of miR-885-5p

Overexpression of AEG1 has been documented to be involved in many tumors' biological processes, including tumor proliferation, metastasis, EMT.<sup>12-15</sup> To evaluate the possibility that AEG1 is important for HCC, we examined AEG1 expression in corresponding noncancerous tissues and HCC using Gene Expression Profiling Interactive Analysis (GEPIA) website based on TCGA database.<sup>16</sup> Notably, AEG1 expression in HCC samples was significantly higher compared to normal samples (Figure 3A). To confirm whether AEG1 mediated the function of miR-885-5p, we used two bioinformatic analysis tools (TargetScan and RNA22), and along with luciferase reporter assays were utilized to confirm the binding sites and regulation relationship between miR-885-5p and AEG1. As presented in Figure 3B-D, miR-885-5p could bind to the 3'-UTR of AEG1, and miR-885-5p mimics reduced the luciferase activity of wild-type (WT) AEG1 reporter vector but not that of mutant reporter vector (MU). What's more, we found that miR-885-5p overexpression decreased the AEG1 mRNA and protein levels decrease at 72h-post transfection (Figure 3E and F). Our findings indicated that miR-885-5p directly targets the AEG1 3'UTR, thus reducing AEG1 expression in HCC cells.

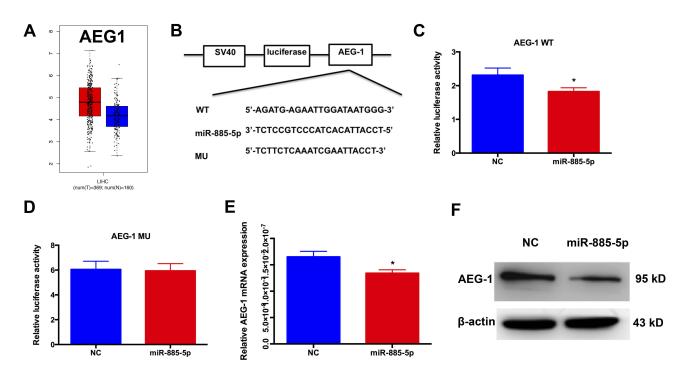


Figure 3 AEG1 is identified as a target of miR-885-5p.

**Notes:** (**A**) AEG1 messenger RNA expression was higher in HCC tissues than in normal tissues. (**B**–**D**) the binding sites between miR-885-5p and AEG1; miR-885-5p mimics decreased the luciferase reporter analysis of AEG1 (\*p < 0.05). (**E**) Real-time PCR results for endogenous AEG1 infected with miR-885-5p or NC (\*p < 0.05). (**F**) Western blotting for endogenous AEG1 infected with miR-885-5p or NC.

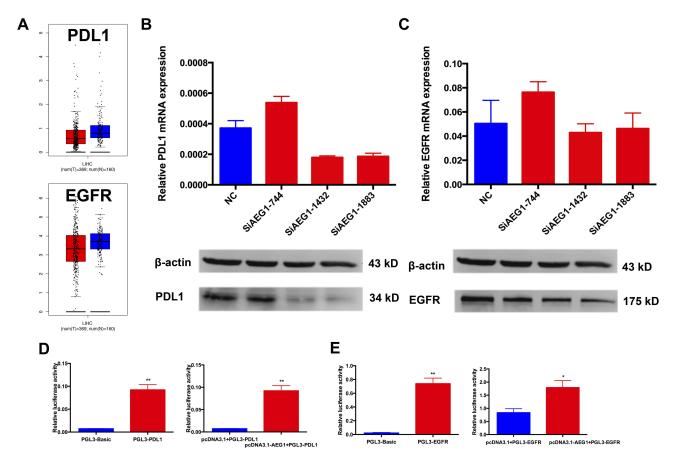
Abbreviations: HCC, hepatocellular carcinoma; MU, mutant; WT, wild-type; NC, negative control.

# AEGI Silencing Inhibits the PD-LI and EGFR Expression in HCC

To investigate the functions of AEG1 in HCC metastasis, we transfected HepG2 cells with SiAEG1 into HepG2 cells and measured the PD-L1 and EGFR mRNA and protein levels using qRT-PCR and Western blotting. We found that when AEG-1 was down-regulated, the levels of both PD-L1 and EGFR mRNA and protein both decreased (Figure 4B and C). Next, we cloned the PD-L1 and EGFR gene promoter regions into a luciferase reporter construct (pGL3-basic). The luciferase activity of PD-L1 and EGFR increased when AEG1 was over-expressed compared with controls (Figure 4D and E). These findings indicated that AEG1 transcriptionally activates PD-L1 and EGFR.

# Discussion

Recently, more and more evidences have showed that microRNAs play crucial roles in tumorigenesis, Searching a microRNA signature can potentially be effective clinical tools for diagnosis, prognosis, and therapy in HCC.<sup>17</sup> MiR-885-5p was studied in the context of cancer for several years; however, its effect on HCC still needs to be investigated. Afanasyeva et al reported that miR-885-5p was downregulated upon deletion of 3p25.3 region in neuroblastoma. They showed that miR-885-5p has a tumor-suppressive role in neuroblastoma via interfering with cell cycle progression and cell survival;<sup>8</sup> Yan et al demonstrated that miR-885-5p reduces the levels of MMP-9 expression and inhibits cellular invasion in glioma cells;9 Cybula et al illustrated that miR-885-5p may act as a tumor suppressor genes, and it is an indicator for the classification of laryngeal cancer tissue and normal mucosa.<sup>18</sup> Such contradictory effects of miR-885-5p prompted us to investigate its role in HCC. Currently, the clinical significance of miR-885-5p was disclosed in HCC. We found that miR-885-5p inhibited proliferation, migration and reduced EMT in HCC cells. Therefore, miR-885-5p



#### Figure 4 AEG1 silencing inhibits the PD-L1 and EGFR expression in HCC.

**Notes:** (**A**) There is no difference of *PD-L1* and *EGFR* messenger RNA expression between HCC tissues and normal tissues. (**B**) qRT-PCR assay and Western blot assay to measure PD-L1 mRNA and protein levels when AEG1 was down-regulated, and that the siRNA efficiency of AEG1 was most marked at the 744 and 1883 sites in HCC cells. (**C**) qRT-PCR assay and Western blot assay to measure EGFR mRNA and protein levels when AEG1 was down-regulated, and that the siRNA efficiency of AEG1 was most marked at the 744 and 1883 sites in HCC cells. (**D**) Luciferase assay showing activity for co-transfection of AEG1 and PDL1 promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HCC cells (\*\*p < 0.01). (**E**) Luciferase assay showing activity for co-transfection of AEG1 and EGFR promoter vector in HC

Abbreviations: HCC, hepatocellular carcinoma; MU, mutant; WT, wild-type; qRT-PCR, quantitative real-time PCR; NC, negative control

potentially serves as a promising biomarker for the prognosis of patients with HCC.

AEG1 has a crucial role in tumor progression, including transformation, evasion of apoptosis, invasion, metastasis and chemoresistance.<sup>19</sup> Notably, the present study also identified that AEG1 silencing inhibits the expression of cyclin D1, c-Myc and MMP9. Previous studies suggested that cyclin D1 and c-Myc are associated with cell proliferation, and are involved in tumorigenesis and deterioration.<sup>20,21</sup> In addition, AEG1 activates various signaling factors involved in mediating EMT, including sonic hedgehog protein, transforming growth factor- $\beta$  and protein Wnt signaling.<sup>22–24</sup> To explore whether AEG1 was involved in the miR-885-5pmediated regulation in HCC cells, we first utilized the bioinformatics and dual-luciferase report assays to confirm the potential binding sites in miR-885-5p and AEG1 in HCC cells. Next, we measured the expression level of AEG1 in HCC cells. Furthermore, qRT-PCR and Western blotting were employed to examine the function of miR-885-5p on the expression levels of AEG1. We found that up-regulated miR-885-5p could remarkably suppress the level of AEG1 in both mRNA and protein levels. Furthermore, luciferase assays were performed to confirm the regulatory interaction between miR-885-5p and AEG1.

In summary, this study revealed that overexpression of miR-885-5p significantly inhibits the metastatic ability and reverses the EMT to MET in HCC cells. Our data suggested that the function of miR-885-5p in HCC cells is partially exerted through targeting AEG1, and the signal pathway including miR-885-5p/AEG1 might be a potential target for treating HCC.

#### **Author Contributions**

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

#### Disclosure

The authors report no conflicts of interest in this work.

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