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Impact of gut microbiota on metabolic dysfunction-associated steatohepatitis and hepatocellular carcinoma: pathways, diagnostic opportunities and therapeutic advances

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Abstract

Metabolic dysfunction-associated steatohepatitis (MASH) and progression to hepatocellular carcinoma (HCC) exhibits distinct molecular and immune characteristics. These traits are influenced by multiple factors, including the gut microbiome, which interacts with the liver through the "gut-liver axis". This bidirectional relationship between the gut and its microbiota and the liver plays a key role in driving various liver diseases, with microbial metabolites and immune responses being central to these processes. Our review consolidates the latest research on how gut microbiota contributes to MASH development and its progression to HCC, emphasizing new diagnostic and therapeutic possibilities. We performed a comprehensive literature review across PubMed/MedLine, Scopus, and Web of Science from January 2000 to August 2024, focusing on both preclinical and clinical studies that investigate the gut microbiota's roles in MASH and HCC. This includes research on pathogenesis, as well as diagnostic and therapeutic advancements related to the gut microbiota. This evidence emphasizes the critical role of the gut microbiome in the pathogenesis of MASH and HCC, highlighting the need for further clinical studies and trials. This is to refine diagnostic techniques and develop targeted therapies that exploit the microbiome's capabilities, aiming to enhance patient care in liver diseases.

Keywords Gut microbiota, Metabolic dysfunction-associated steatohepatitis/non-alcoholic steatohepatitis, Hepatocellular carcinoma, Immune modulation

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Introduction

The human gut consists of an abundant and diverse microbial population and the bacterial density in the colon has been estimated to be 10^{11} – 10^{12} per ml [1]. The molecular techniques involving metabolomic, lipidomic, metatranscriptomic and metagenomic deciphered the impact of gut microbial populations in different organs [2]. The alterations in the microbial composition in the gut lead to the development of various diseases including inflammatory bowel disease (IBD), irritable bowel syndrome (IBS), type-2 diabetes, atopy, autoimmune diseases (ulcerative colitis, lupus, psoriasis, multiple sclerosis, and Crohn's disease), hepatic steatosis and several types of carcinomas (oral cancer, gastric cancer, colorectal cancer, lung cancer, cervical cancer, gall bladder cancer, hepatocellular carcinoma, etc.) [3–8]. Gut microbiota consists of several microorganisms belonging to the category of bacteria, viruses and yeast. The most prominent bacterial phyla in the composition of gut microbiota are Firmicutes and Bacteroidetes [9, 10]. Other microbes including Actinobacteria, Fusobacteria, Proteobacteria, and Verrucomicrobia are also present. The Firmicutes phylum mainly possesses 200 different genera, for example, Lactobacillus, Clostridium, Bacillus, Ruminococcus and Enterococcus. Clostridium genera are most prominent in (around 95%) the Firmicutes phyla. Bacteroidetes are composed of prime genera such as *Bacteroides* and Prevotella [11, 12]. The clinical study revealed that the presence of bacteria named Ruminococcus obeum and Alistipes was reduced while Dorea, Lactobacillus, and Megasphaera were enriched in NAFLD patients compared to healthy individuals [1, 13, 14]. Compared to fatty liver patients, NASH patients possess higher levels of Firmicutes but lower levels of Bacteroidetes at the phylum level [15]. Studies demonstrated that the patients with cirrhosis showed higher levels of Enterobacteriaceae and Streptococcus and low levels in Akkermansia. In HCC patients, the presence of *Bacteroides* and *Ruminococ*caceae was elevated, while Bifidobacterium was reduced. Bacteria such as Akkermansia and Bifidobacterium were inversely correlated with calprotectin concentration (cellular inflammatory markers). The fecal microbial diversity is evident in cirrhosis to early HCC, and the presence of phylum Actinobacteria was high in the early stage of HCC. Similarly, Gemmiger and Parabacteroides were higher in early HCC than in cirrhosis. On the other hand, butyrate-producing genera declined and lipopolysaccharide-producing genera were enriched in early HCC than healthy individuals [16, 17]. The abnormal increase of Bacteroidetes/Firmicutes ratio in NASH patients may increase the occurrence of HCC [18]. Evidence suggests that the changes in the composition and diversity of gut microbiome may lead to the development and progression of different liver diseases. Metabolic dysfunction-associated steatotic liver disease (MASLD), previously known as non-alcoholic fatty liver disease (NAFLD), refers to a range of liver conditions characterized by the accumulation of excess fat in the liver ($\geq 5\%$ hepatic steatosis). This spectrum of disease begins with steatotic liver disease (SLD), which represents the early stage of liver fat accumulation [19]. As the condition progresses, it may develop into metabolic MASH, formerly known as non-alcoholic steatohepatitis (NASH), which involves liver inflammation and damage, potentially with or without fibrosis. In more advanced stages, MASLD can lead to cirrhosis, liver failure, and even liver cancer [20]. Epidemiological evidence shows that NAFLD, along with its more severe form, non-alcoholic steatohepatitis (NASH), is increasingly recognized as a major contributor to hepatocellular carcinoma (HCC). Dysbiosis in the gut microbiome can disrupt homeostasis, exacerbating liver cell injury by triggering various immune-mediated responses. Several genomic factors interrupt the gut-liver axis and increase microbial exposure to the liver [21]. Evidences indicate that microbial metabolites such as secondary bile acids, trimethylamine, short-chain fatty acids, etc., are responsible for the onset and progression of liver diseases [22]. Improper microbial production and the entry of microbial products into the liver via the portal vein can cause hepatic inflammation and lead to the development of NAFLD to NASH progression [23]. The gut microbiota influences NASH to HCC progression, via modulating different factors, such as gut epithelial permeability, hepatic Toll-like receptor (TLR) endogenous alcohol production, choline metabolism, bile acid metabolism, and release of inflammatory cytokines [24– 30]. The present review elaborates on the importance of gut microbiota-mediated influences in MASH and HCC. Also, it highlights the diagnostic and therapeutic importance of gut microbiota in MASH and HCC (Fig. 1) (Table 1).

Methodology

This review was conducted by searching electronic databases to identify studies that report on the role of gut microbiota in the development of metabolic dysfunction-associated steatohepatitis /non-alcoholic steatohepatitis (MASH/NASH) and its progression to hepatocellular carcinoma (HCC). The databases included PubMed/MedLine, Scopus, and Web of Science. The search was conducted from January 2000 to August 2024 to encompass recent developments in the field (Fig. 2). The search strategy employed both Medical Subject Headings (MeSH) and free-text terms to ensure comprehensive coverage of the literature. The MeSH terms used were: "Gut Microbiota" [MeSH];

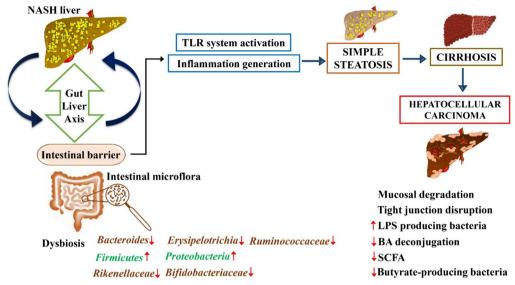


Fig. 1 Progression of liver disease from MASH to hepatocellular carcinoma via gut–liver axis dysregulation. The figure illustrates the pathological progression from MASH to HCC through disruptions in the gut–liver axis. The MASH liver exhibits alterations in the intestinal barrier and gut microbiota, leading to dysbiosis characterized by fluctuations in key bacterial populations. The dysregulated gut microbiota affects the intestinal barrier's integrity, fostering mucosal degradation and tight junction disruption. This breakdown facilitates the systemic infiltration of lipopolysaccharides (LPS) and other bacterial metabolites into the liver through the portal circulation. Increased TLR (Toll-like receptor) activation in the liver induces inflammation, progressing from simple steatosis to cirrhosis and ultimately culminating in hepatocellular carcinoma. Key changes in microbial populations include increased Firmicutes and Proteobacteria, with a decrease in Bacteroides, Erysipelotrichia, Ruminococcaceae, Rikenellaceae, and Bifidobacteriaceae. The figure also notes a decrease in bile acid (BA) deconjugation, short-chain fatty acids (SCFA), and butyrate-producing bacteria, which are critical to maintaining hepatic and intestinal health. Symbols: ↑increase, ↓decrease

Table 1 Expression of different microbiome in MASH-induced HCC

SI. No	Gut microbiome	Status in MASH	Status in HCC
1	Bacteroides	Bacteroides ↑ [104]	Bacteroides ↑ [105]
2	Enterococcus	Enterococcus ↑ [104]	Enterococcus ↑ [105]
3	Ruminococcaceae	Ruminococcaceae ↑ [104]	Ruminococcaceae ↑ [105]
4	Bifidobacterium	Bifidobacterium ↓ [104]	Bifidobacterium ↓ [105]
5	Oscillospira	Oscillospira ↓ [104]	Oscillospira ↑ [105]
6	Lachnospiraceae	Lachnospiraceae 1 [104]	Lachnospiraceae↑ [105]

"Hepatocellular Carcinoma" [MeSH]; " Metabolic dysfunction-associated fatty liver disease/Non-alcoholic Fatty Liver Disease" [MeSH]; " Metabolic dysfunction-associated steatohepatitis/Non-alcoholic Steatohepatitis" [MeSH]; "Microbiome" [MeSH]; These terms were combined with additional keywords and phrases relevant to the study topic, such as "intestinal microbiome", "liver cancer", "MASH/NASH", "immune modulation", and "therapeutic implications". Boolean operators (AND, OR) were used to combine these terms effectively.

Inclusion criteria:

- Studies published in English.
- Studies that directly investigated the impact of gut microbiota on the pathogenesis, progression, or treatment of MASH/NASH and HCC.
- Both preclinical and clinical studies.
- Reviews, meta-analyses, randomized controlled trials, cohort studies, and case—control studies.

Exclusion criteria:

- Studies published before the year 2000.
- Studies not in English.

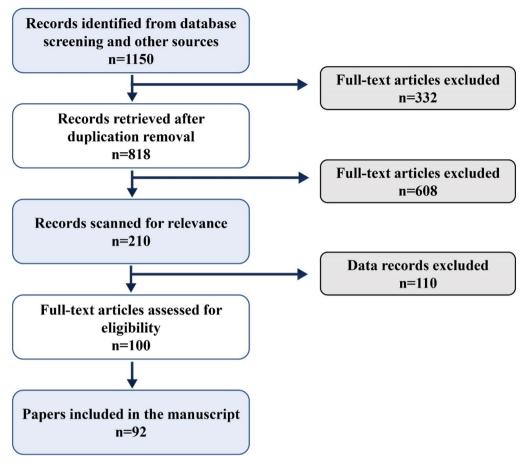


Fig. 2 Flow diagram of the study selection

- Studies focusing on alcoholic liver disease or other forms of liver disease not directly related to MASH/ NASH or HCC.
- Commentaries, editorials, and expert opinions without original data or systematic analysis.
- Studies with incomplete data or unclear methodologies.

The most representative data are summarized in tables and figures.

Immune modulatory role of gut microbiota in MAFLD/MASH to HCC progression

Numerous preclinical and clinical studies have established that abnormal expression of gut microbiota and its metabolites are closely associated with liver diseases such as MAFLD, MASH (formerly known as NAFLD and NASH), Cirrhosis and HCC [31, 32]. The alternations in the gut microflora can facilitate the synthesis of free fatty acids (FFA) in the intestine and increase the permeability of FFA across the intestinal area which may lead to the

development of NAFLD [33, 34]. The studies indicated that the high-fat diet could increase the number of alcohol-producing bacteria such as Escherichia genus members of the Proteobacteria phylum in the gut, which may produce acetate and acetaldehyde via the oxidation of ethanol and facilitate the synthesis of fatty acids and contributes to the development of NAFLD [35]. Recent studies suggest that the gut microflora is altered due to genetic predisposition and improper diet which may affect the lipid and hepatic carbohydrate metabolism and also influence the activities of anti-inflammatory and pro-inflammatory agents in the liver and may lead to the development of NAFLD and its progression to NASH [32]. The consumption of obesogenic foods such as a high-fat diet may disrupt the Gram-negative bacteria that are present in the intestinal tract and increase the level of lipopolysaccharides (LPS), which can act as a key regulator for producing inflammatory responses in the liver tissue and produce liver injury via activating TLR4 signaling and cause the development of NAFLD and its progression [34]. Moreover, the high-fat diet can also modulate

the enzyme produced by the gut microbiome, it may act as a catalyst for the conversion of choline into toxic metabolites, known as dimethylamine and trimethylamine. These metabolites can be converted into trimethylamine oxide (TMAO) in the liver, which produces inflammation in the hepatocytes and the progression of NAFLD into NASH [36]. The clinical studies indicated that the overgrowth of bacteria in the small intestine due to high-fat diet and genetic factors in NAFLD patients may increase the risk for the development of NASH [37-39]. Small intestinal bacterial overgrowth (SIBO) prominently affects the progression of NASH in NAFLD patients. Wigg et al. [41] carried out a comparative study to evaluate the presence of SIBO in NASH patients as well as in healthy controls [40]. The report indicated that 50% of NASH patients were observed with SIBO whereas SIBO was limited up to 22% in healthy controls. They evaluated the mean levels of tumor necrosis factor TNF-α in NASH patients and healthy people and it was found to be 14.2 and 7.5 pg/ml, respectively. The amount of intestinal bacteria was quantified via glucose hydrogen breath test and quantitative jejunal aspirate culture (the removal and culture of a sample of intestinal fluid) and outcome indicated that low-grade around≥103 CFU/ml in NASH patients compared with that of controls. These data established that patients with NASH have the highest prevalence of SIBO [41]. Shanab et al. [43] reported that an increased level of SIBO in NASH patients increases the hepatic release of interleukin-8 (IL-8) and increases the expression of Toll-like receptors-4 (TLR-4) which facilitates the development and progression of NASH [42]. The imbalance in toll-like receptor (TLR) also contributes to the progression of NAFLD to NASH. Abnormal bacterial DNA, LPS and other endogenous substances can activate the innate immune system via TLR 4 and TLR 9 which will facilitate the production of kuffer cells and interleukin- 1β (IL- 1β). IL-1β can uphold the accumulation of lipids and also increase the cell death of hepatocytes followed by inflammation and steatosis [43]. Moreover, microbial pathogenassociated molecular patterns can activate different inflammasome such as NLRP1 (NALP1), NLRP3 (NALP3, cryporin), NLRC4 (IPAF), AIM2 and NLRP6 can facilitate the progression of NAFLD and contribute to the initiation of steatosis [44]. The evidence suggests that TLR 4 and microbacteria-derived LPS are the key factors that lead to the progression of cirrhosis [45]. Gut dysbiosis may cause systemic inflammation and immunodeficiency by impairing the functions of immune cells such as T cells, B cells, macrophages, etc., and cause the development of cirrhosis-associated immune dysfunction (CAID). CAID can promote the translocation of bacterial products into the bloodstream and facilitate the

intensity of inflammation in the body [46]. Moreover, gut dysbiosis can disrupt the bacterial flora and facilitate the LPS/TLR4-mediated signaling. It can also initiate the cirrhotic to cancer progression via increased secretion of chemokines from HSCs and chemotaxis of Kupffer cells which stimulate the profibrogenic cytokine TGF-b [47]. The cirrhosis to HCC progression is driven by different inflammatory pathways, initiated via the crosstalk between the intestinal bacteria, immune system, and liver. The inflammatory process mainly encompasses the interplay between the macrophages, Kupffer cells, and PAMPs in the liver cells. These Kupffer cells, macrophages, and PAMPs can elicit the NF-κB pathway through binding with nucleotide-binding oligomerization domain-like receptors (NOD-like receptors) and TLRs, particularly TLR-4 and TLR-9 [48]. The gut microbiota-mediated TLR-4 signaling pathway participates in the pro-inflammatory response in the liver and promotes HCC development [49]. The inflammatory chain reactions may elicit excess cytokine release and inflammation in the liver in turn causing dysbiosis of the microbiota. This increases Kupffer cells mediated secretion of inflammatory cytokines, such as tumor necrosis factor-α (TNFα), interleukin-8 (IL-8), and IL-1β. Excessive cytokines may elicit lipid accumulation and apoptosis in hepatocytes and can cause steatosis and inflammation. Increased levels of pro-inflammatory cytokines due to the abnormal regulation of gut microbiota were observed in almost all NAFLD and NASH patients, which promote the development and progression of NASH through TLR-triggered pathways [50, 51]. Thus, gut microbiotamediated cytokines play a key role in the initiation and progression of NAFLD to NASH to HCC [52–54]. Dysbiosis induced by cirrhosis, coupled with increased intestipermeability, may trigger the release pathogen-associated molecular patterns (PAMPs) and metabolites mediated by the gut microbiome, leading to enhanced inflammation, damage, and fat production in the liver [55]. Research indicates that PAMPs initiate the release of cytokines and chemokines such as IL8, IL-17, and IL1B through TLR activation, intensifying immune cell presence in the liver [56, 57]. Continuous production of these cytokines can lead to DNA damage and oxidative stress, thereby initiating and advancing hepatocellular carcinoma (HCC) [57, 58]. Furthermore, metabolites produced by the microbiota, including bile acids, shortchain fatty acids, PAMPs, lipoteichoic acid (LTA), and branched-chain amino acids, are known to activate hepatic stellate cells (HSCs) via the senescence-associated secretory phenotype (SASP), promoting hepatocyte proliferation and increasing susceptibility to HCC [59]. Clinical and preclinical studies demonstrated that bile acid (BA) metabolism also plays a significant role in

NASH to HCC progression. [60, 61]. Increased levels of bile acids in the liver can prompt inflammation, hepatocyte DNA damage, and apoptosis; hence tumorigenesis will occur in the liver [62]. Furthermore, dysbiosis in NASH will increase the abundance of Gram-positive microorganisms in the microflora, thus stimulating HCC through an increase in the synthesis of secondary BA including deoxycholic acid (DCA), which restricts the activation of liver sinusoidal endothelial cell (LSEC) and facilitates the suppression of chemokine ligand 6 (CXCL6), natural killer T cell recruitment (NK), and produce tumorigenesis [63, 64]. Additionally, secondary bile acids directly initiate the development of HCC from NASH via stimulating mTOR signaling [65]. Guerra Ruiz et al. [64] demonstrated that the serum levels of lipopolysaccharides (LBP) were significantly increased in NASH patients compared with healthy patients with simple steatosis. The augmented serum LBPlevel was connected with abnormal expression of tumor necrosis factor (TNF- α) in the liver tissue. The increased level of TNF- α plays a key role in the development of HCC [66, 67]. Obesity is another risk factor that influences the changes in the composition of the microbiota and its metabolites such as LPS or PAMPs [68]. The injured hepatocytes can produce damage-associated molecular patterns (DAMPs) which prompt the inflammatory molecules via TLR and activation of target immune cells and arouse the transition from NAFLD-NASH-HCC [69]. Current evidence indicates that gut microbiome can also have an impact on antitumor responses, which may provide a novel perspective on refining the effectiveness of cancer immunotherapy [70]. Emerging evidence established that the abnormal characterization of gut microbiota can elicit immunosuppression by inducing M2 (pro-tumor)-like tumor-associated macrophage (TAM). Dysbacteriosis associated with IL-25-persuaded activation of M2 macrophages can augment HCC progression by secreting C-X-C motif chemokine ligand 10 (CXCL10) and augment epithelial–mesenchymal transition pathway (EMT) [71]. Studies revealed that gut microbiota can produce oncogenesis and cancer progression in myeloid-derived suppressor cells (MDSC) dependent manner [72]. Abnormal gut microbiome-mediated dysbiosis can disturb homeostasis, consequently prompting immune-mediated hepatocyte injury which further prompts the HCC progression. Metabolomics and metagenomic studies related to gut** microbiota discovered that the gut microbiota causes T cell-mediated immune suppression via increasing the level of regulatory T cells (T reg) and reducing the level of CD8+T cells including cytotoxic T cells [73]. Moreover, Kang, Y et al., 2021 demonstrated that the gut microbiota can increase the generation of prostaglandin E2 (PGE2) and cyclooxygenase-2 (COX-2) enzymes

which inhibits the antitumor immune responses through Prostaglandin E2 receptor 4 (EP4 receptor), hence facilitate HCC progression [29]. The expression of different proteins such as CD68 (Cluster of Differentiation 68) is considered a marker of macrophages and TLR (TLR-2, TLR-4, TLR-5 and TLR-9) plays a negative role in the activation of the innate immune system. Studies demonstrated that CD68 is a tumor-associated macrophage and it leads to the development of NAFLD and NASH-HCC progression [31]. It also indicates that a leaky gut can result in the overproduction of gut microbiota-derived metabolites, potentially impacting the hepatic immune system and increasing the risk of HCC [74] (Fig. 3).

Table 2 summarizes the roles of microbial and immune factors in the progression from MASH to HCC.

Immunotherapeutic significance of modulating gut microbiota in MASH and HCC

NASH-induced dysbiosis in the gut microbiota leads to an increase in intestinal permeability, thereby increasing exposure to bacterial metabolites in the liver and causing severe inflammation, contributing to HCC [76]. Modulating gut microbiota-mediated bile acids (BAs) metabolism, toll-like receptor (TLR) activity, regulating farnesoid X receptor (FXR)/Takeda G protein-coupled receptor 5 (TGR5) activation, choline metabolism, and targeting inflammatory cytokines are considered novel therapeutic options against NASH and NASHassociated HCC [18]. Therapeutic targeting of the gut microbiota against NASH and HCC is highly attractive; meanwhile, these treatment modalities show a low risk of adverse effects and a high safety profile, including fecal microbiota transplantation (FMT), probiotics, prebiotics, synbiotics (combination of prebiotics and probiotics), antibiotics, and immunotherapies [29]. The predominant mechanisms behind the gut microbiota-targeted therapies are as follows: controlling the T helper 17 (Th17) cell proliferation which may raise the secretion of interleukin-17 (IL-17); decreasing the level of metastasis through reduction of the overexpression of vascular endothelial growth factor (VEGF) limiting the angiogenesis lymphangiogenesis, and inflammations [77]. Additionally, alterations in regulating gut microbiota may stimulate the synthesis of short-chain fatty acids (SCFAs) and eventually restrict the progression of NASH to HCC. Modulations in the composition of gut microbiota may upsurge the production of propionate that may help the patients to recover from HCC through cyclic adenosine 3',5'-monophosphate (cAMP) leveldependent pathway and the stimulation of G proteincoupled receptors 43 (GPR43) [78]. Moreover, regulation of gut microbiota may accomplish an anti-HCC effect by increasing the level of hepatic CXCR6+NKT cells and

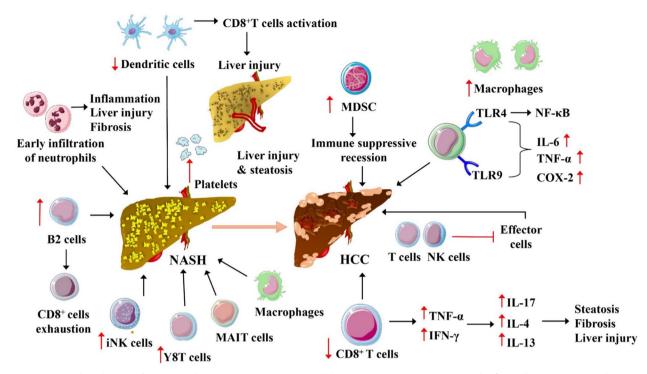


Fig. 3 Potential mechanisms for gut microbiota-associated immune modulation in MASH–HCC progression. This figure illustrates the complex immune interactions and cellular transformations involved in the progression from MASH to HCC. Key features include the early infiltration of neutrophils leading to inflammation, liver injury, and fibrosis, and the role of B2 cells and CD8+T cells in modulating the immune response. Activation of CD8+T cells contributes to further liver injury and steatosis, while the presence of myeloid-derived suppressor cells (MDSC) indicates an immune suppressive state facilitating cancer progression. The diagram also highlights the activation of macrophages through toll-like receptors (TLR4 and TLR9) leading to an increase in inflammatory cytokines (IL-6, TNF-α) and COX-2, which are important in the development of HCC. Additionally, the impact of various cytokines such as TNF-α, IFN-γ, IL-17, IL-4, and IL-13 on the hepatic environment, promoting steatosis, fibrosis, and liver injury, is depicted. Abbreviations and symbols: B2 cells: a type of B cell involved in immune response; CD8+T cells: cytotoxic T cells which are a part of the immune system that kills cancer cells, virus-infected cells, and other damaged cells; COX-2: cyclooxygenase-2, an enzyme that plays a crucial role in inflammation; HCC: hepatocellular carcinoma; IFN-γ: interferon gamma, a cytokine critical for innate and adaptive immunity; IL-4: interleukin 4, a cytokine involved in the regulation of immune responses; IL-17: interleukin 17, a pro-inflammatory cytokine; iNK cells: invariant natural killer T cells, a component of the immune system that recognizes lipid antigens; MAIT cells: mucosal-associated invariant T cells, involved in the mucosal immunity; MDSC: myeloid-derived suppressor cells, regulate immune responses in cancer; NASH: non-alcoholic steatohepatitis

elevating the level of interferon-gamma (IFN- γ). Simultaneously, CXCR6+NKT cell accumulation was controlled through the expression of CXCL16 in the liver sinusoidal endothelial cells, which was connected with microbiometriggered primary-to-secondary bile acid conversion [79].

Probiotics are primarily utilized to correct microbial imbalances [65]. Clinical applications of probiotic bacteria have demonstrated effectiveness in decelerating the progression of NASH and reducing the spread of HCC cells by diminishing the activation of inflammation mediated by toll-like receptors (TLRs). Pathogen-associated molecular patterns (PAMPs) contribute to the development of NASH and HCC by activating inflammatory responses through TLRs. Using microbial agents, particularly probiotic bacteria, has proven to mitigate liver metastasis by curtailing the excessive inflammatory

responses triggered by TLRs [80]. In experiments, rats with liver cirrhosis treated with Lactiplantibacillus plantarum exhibited reduced TLR4 expression and minimal liver damage. Additionally, sterilizing the gut and deactivating the TLR4 receptor significantly slowed HCC progression by 80-90%, suggesting their potential as a preventative measure against HCC [81]. Research by Li et al. in 2016, confirmed that a combination of probiotics could limit NASH and its progression to HCC by lowering the levels of pro-inflammatory cytokines like IL-17 in mouse models. It also indicated that probiotics could reduce liver fat and aspartate aminotransferase (AST) levels in NASH patients [82], [NCT00870012, NCT01791959]. Probiotics triggered the growth of gut microbiota composition towards specific beneficial bacteria including Prevotella and Oscillibacter. Prevotella

Table 2 Impact of gut microbiota and immune factors on MAFLD/MASH to HCC progression

Microbial and immune factors	Impact on MAFLD/MASH	Impact on HCC progression	Evidence	References
Small intestinal bacterial overgrowth (SIBO)	Associated with 50% of NASH patients Linked to higher levels of TNF-α	↑ Hepatic inflammation and cytokine production ↑ IL-8, ↑ TLR-4	Clinical study	[41, 42] [43]
TLR-4-mediated pathway	Augments hepatic inflammation by enhancing secretion of pro- inflammatory cytokines (†) † level of LPS Increase the expression of CD68 (tumor-associated macrophage)	Directly linked to the development of HCC via sustained inflammatory responses (†) NAFLD and NASH-HCC progression	Preclinical study Germ-free animal model	[31, 48, 49]
Pathogen-associated molecular patterns (PAMPs)	†lintestinal permeability allow microbial metabolites to pro- mote liver inflammation (†)	↑ IL-8, ↑ IL-17, ↑ IL-1 β ↑ chemokines release, exacerbating liver injury and initiating HCC (↑)	Germ-free animal model Clinical study	[55, 56] [56]
Bile acid metabolism	Dysregulated metabolism contributes to liver inflammation (†)	Altered metabolism leads to increased secondary bile acids, promoting HCC via mTOR signaling and inflammation (†)	Clinical study	[58–60]
Prostaglandin E2 (PGE2) and COX-2 enzymes	Not directly mentioned in NAFLD/ NASH context	Suppress antitumor immune responses, facilitating HCC progression (1)	Preclinical study	[30]
DAMPs and cytokine production	Activates inflammatory pathways through TLR stimulation (†)	↑ Chronic cytokine release ↑DNA damage ↑Oxidative stress ↑HCC progression	Clinical study	[67]
Microbial dysbiosis	This leads to SIBO and imbalances in key bacterial populations, aggravating liver conditions (↑) Promote steatohepatitis via modulating Toll-like receptor 4 (TLR4) and TLR9	Increases the risk of HCC by altering liver immune responses and metabolic functions (†) HCC was developed due to hepatic inflammation	Clinical study germ-free animal model	[75]

1: indicates an increase in activity; ↓: indicates a decrease in activity. COX-2: cyclooxygenase-2; DAMPs: damage-associated molecular patterns; HCC: hepatocellular carcinoma; IL-1β: interleukin-1 beta; IL-8: interleukin-8; IL-17: interleukin-17; mTOR: mechanistic target of rapamycin; NAFLD: non-alcoholic fatty liver disease; NASH: non-alcoholic steatohepatitis; PAMPs: pathogen-associated molecular patterns; PGE2: prostaglandin E2; SIBO: small intestinal bacterial overgrowth; TLR-4: toll-like receptor 4; TNF-α: tumor necrosis factor-alpha

and Oscillibacter elicit anti-inflammatory metabolites, which afterward diminished the T helper 17 cells (Th17) polarization and increase the differentiation of regulatory T cells ((Treg)/Type 1 regulatory T (Tr1)) cells in the gut and produce anti-inflammatory responses in the cancer cells. Besides, probiotics can control the abnormal growth of segmented filamentous bacteria (SFB), which are the foremost bacteria to increase the level of Th17 in the body. Thus, the administration of probiotics intensely decreases the level of SFB which leads to reduce the production of pro-inflammatory cytokines such as IL-17. The IL-17A formed from Th17 could favor angiogenesis, thus reduction of Th17 and IL-17 level may reduce HCC progression. Different clinical and preclinical studies have shown that probiotics are effective against NASH and HCC [83]. Furthermore, Helicobacter sp. was found in the surroundings of NASH cells and its translocation might be possible to elicit HCC. To this end, the intestinal microbial profile might be prominently exhibit the beneficial rates in HCC patients experiencing treatment with immunotherapy such as immune checkpoint inhibitors (ICIs), indicating that the gut micro flora targeted immunotherapy could be beneficial for liver cancer [84]. A double-blind, randomized, placebo-controlled trial of probiotics in patients with Child–Pugh A-B cirrhosis was conducted to evaluate the predictive role and risk of the microbiome in HCC development. In this particular study, they evaluate the role of probiotics towards the presence of endotoxins (LPS) and different cytokines (IL-6 and TNF- α) in the tumor microenvironment (TME) and also evaluate the expression of TLR4 in mononuclear cells (NCT03853928) [85].

Prebiotics are non-absorbent oligosaccharide substances that accelerate the growth of bacteria. It can also reduce the growth of harmful bacteria and maintain the balance of gut microbiota. They mainly initiate the production of SCFAs and regulate the immune responses in the liver cells. Hence, prebiotics alter the gut microbiota to reduce the progression of NASH as well as NASH-associated HCC [86]. Dietary polyphenols are significant prebiotics used in this modern era due to their great therapeutic value. They mainly include flavonoids including lignins, and phenolic acids found in tea, vegetables, nuts, fruits and wine. One of the significant prebiotic

polyphenols is ellagic acid which is an antioxidant having anti-cancer properties. The ellagic acid is metabolized by micro-flora present in the colon producing urolithins that are abundantly present in certain nuts and berries [87]. Urolithins can suppress the COX-2-associated inflammation in liver cells [88]. Another polyphenol such as resveratrol which is naturally found in grapes can also reduce or prevent NASH [89, 90] and HCC progression by destroying the metastatic invasion and tumor cell migration in liver cancer [91, 92]. Resveratrol acts as an immunomodulatory agent by either stimulating the immune cells situated in the tumor microenvironment (TME) or by sensitizing tumor cells toward the cytotoxic signaling of immune cells [93]. Quercetin is another dietary flavonoid that works as a prebiotic through the suppression of activated nuclear factor kappa B (NF-kB) in hepatocytes [94, 95]. A prospective cohort study demonstrated that increasing the consumption of tree nuts such as almonds, hazelnuts, pistachios, macadamias, cashews and pecans was related to a reduced risk of NASH and HCC [96-98]. The combinatorial effects of pectins and fructo-oligosaccharide (FOS) with raspberry polyphenols on microbial fermentation and modulation of inflammation and lipid metabolism in the liver was evaluated and thus suggests, FOS and pectins improved the action of the raspberry polyphenolic extract against NASH and HCC [99, 100]. Moreover, a study on hepatocytes demonstrated that polyphenols extracted from raspberries also control immunometabolic signals connected with the development of obesity [90, 101]. Supplementation with prebiotics will also help the activation of AMPK [102, 103]. Astragalus polysaccharides, grifolan, lentinan, and krestin (PSK) display anti-cancer properties by regulating the activity of the immune system and eliciting direct actions against cancer cells [104]. Clinical studies revealed that Omega 3 fatty acid and EPA (eicosapentaenoic acid) are active against HCC [NCT04682665]. Some of the clinical trials also established the effectiveness of synbiotic and prebiotics against NASH [NCT02530138, NCT01791959, NCT03184376 and NCT03897218].

Antibiotics can be also used to reduce or remove the altered gut microbial content; this can help restrict the inflammatory signals from leaky guts. Different preclinical evidences suggest that different antibiotics such as vancomycin, metronidazole, ampicillin, and neomycin significantly decrease HCC proliferation [105]. Antibiotic cocktails (ABX, including vancomycin, primaxin, and neomycin) produce anti-HCC effect. These antibiotics can increase the hepatic CXCR6+NKT cells and also enhance the level of INF- γ and inhibit cancer cell growth [105]. A phase 2, interventional study was also evaluate the safety and efficacy of solithromycin against NASH without cirrhosis [NCT02510599]. A randomized

interventional clinical trial established the effect of rifaximin on the lipopolysaccharides (LPS) and related cytokine levels in NAFLD and NASH [NCT02009592]. The prolonged antibiotics belonging to β -lactams, tetracyclines, fluoroquinolones, sulfonamides, and aminoglycosides impacts human gut flora. They can modify the diversity of bacterial flora and composition leading to the occurrence of various metabolic alterations in the body that contribute to the onset and progression of NAFLD [106]. Deregulated metabolism in the body especially in the metabolism of SCFA may lead to obesity, metabolic syndrome, and diabetes. Moreover, studies indicated that the continuous use of antibiotics can cause the depletion in gut bacterial diversity and may increase the susceptibility to infections [107]. The continuous use of antibiotics may increase the level of the antibiotic-resistant gene in the microbiome. These pools of resistant genes can initiate antibiotic resistance [108]. In this scenario, the major challenge is to facilitate the growth of beneficial microorganisms, meanwhile reducing the proportion of microorganisms that are responsible for dysbiosis to promote the patient's health. Thus, the development of novel antibiotics can be personalized for a patient based on intestinal and biochemical individuality. The use of selective antibiotics will minimize the negative impact of antibiotics on human health due to changes in the gut microbiome [109]. Fecal microbiota transplantation (FMT) is a medical procedure for the transfer of a small sample of stool (faces) from a healthy person to a diseased person [110]. The healthy stool sample comprises trillions of beneficial microbiomes that can ameliorate the health of the diseased person. Studies suggest that FMT can restore the healthy bacteria in the lower intestine, which will also help to terminate the growth of Clostridium difficile from the intestinal area [111, 112]. As mentioned earlier a healthy intestinal tract possesses a large number of healthy bacteria, but in certain conditions, the use of antibiotics may restrict the growth of good bacteria, and it may promote the development of unhealthy bacteria in the colon. FMT is usually preferred to treat Clostridioides difficile infection (CDI) and also in patients who suffer from IBD [114, 115]. Based on clinical trial of FMT is extended to irritable bowel syndrome, hepatic encephalopathy, diabetes mellitus, refractory diarrhea, fatty liver disease, metabolic syndrome, neurological disease (parkinsonism), and neuropsychiatric disease (autism spectrum disorder) [115, 116]. FMT procedure can be achieved via using different techniques such as colonoscopy, enema, nasogastric (NG) tube and oral capsules (VOWST, SER-109) [117, 118]. Currently, the U.S. Food and Drug Administration (FDA) approves FMT only for the treatment of recurrent CDI that is not responsive to standard antibiotic therapy. Two different FMT therapies have been approved by the FDA REBY-OTA (fecal microbiota, live—JSLM) and VOWST [119]. Studies revealed that FMT elicits around 80–90% in preventing CDI from recurring after antibiotics. Nevertheless, there are several short-term and long-term adverse effects are also associated with FMT [120]. Thus, rigorous donor screening and testing should be mandated to minimize the risk of FMT.

Table 3 outlines the impact of gut microbiota modulation on immune responses in MASH and HCC, detailing therapeutic strategies and their outcomes.

Limitations and challenges

Despite the comprehensive analysis of the role of gut microbiota in the development of non-alcoholic steatohepatitis (NASH) and its progression to hepatocellular carcinoma (HCC) presented in this manuscript, several limitations and challenges remain that need to be addressed:

- i. The majority of studies discussed are preclinical, involving animal models or in vitro systems. These studies provide valuable insights, but may not fully replicate the complex interactions and environmental factors influencing human gut microbiota and liver disease progression. Thus, translating these findings into clinical practice requires much attention, as human studies are more variable and complex.
- ii. The gut microbiome is extraordinarily complex, with a vast number of microbial species that have not been fully characterized. This complexity makes it challenging to determine causal relationships between specific microbial changes and disease states. The functional roles of many species within the microbiome and their interactions with host metabolism and immunity are still poorly understood.
- iii. There is significant variability in microbiota composition among different populations due to factors such as diet, genetics, lifestyle, and antibiotic

Table 3 Immunotherapeutic effect of gut microbiota modulation in MASH and HCC

Category	Agent	Mechanism of Action	Outcome and Benefits	References
Prebiotics	Kappaphycus striatum	Carrageenan polysaccharide with different molecular weights	Immunostimulating activities and antitumor effect via increasing NK cell activity	[72]
	Ganoderma lucidum	Elicits immune regulation, decreases blood sugar and lipid levels	Upsurge anti-inflammatory and anti- hypoxia effects; scavenges free radicals	[121]
	Antrodia cinnamomea	Stimulates immune modulatory action via TLR5 and NLRP3	†Immune response, potentially beneficial against cancer	[122]
	Hirsutella sinensis	↑Cytotoxic T cells ↓Regulatory T cell production in the TME	†Anti-cancer effects by stimulating T cell activity and inhibiting immune inhibitors	[123]
	Polyphenols	fimmune cells in the TME	↑ immune cells to potentially counteract cancerous growth	[124]
	Lactobacillus acidophilus, Lactobacillus reuteri + Inulin	↑Secretion of Th1 mediated T cells	Augments cytokine secretion (e.g., IFN-y), Jangiogenesis, 1cytotoxicity and antigen presentation	[125]
Probiotics	Lactobacillus rhamnosus	Modulates gut microbiota; ↓endotoxemia	$\downarrow TNF-\alpha$ expression, \downarrow inflammation and liver damage	[126]
	Streptococcus thermophilus	↑Gut homeostasis ↓Intestinal and hepatic inflammation	Helps restrict progression of cirrhosis to HCC	[127]
	Lactiplantibacillus plantarum	↓TLR4 expression, gut sterilization and TLR4 receptor inactivation	Abridges the progression of HCC by 80% to 90%	[56]
Antibiotics	Vancomycin	Inhibits fermentable fiber-induced liver cancer by downregulating TLR	Potentially reduces liver cancer progression	[29, 128]
	Metronidazole	↓Butyrate-producing bacteria	↓Occurrence of HCC by impacting bacterial profiles linked to cancer progression	[129]
	Ampicillin	Interferes with TLR protein	↓HCC progression	[130]
	Neomycin	↓IL-6, ↓TNF-α,↓Ki67	↓Inflammatory and proliferative markers associated with HCC	[130]
Fecal micro- biota transplan- tation	REBYOTA and VOWST	The spore suspension is produced by treating fecal matter with ethanol to kill live organisms that are not spores	Helps to destroy <i>Clostridioides difficile</i> from the digestive tract	[113]

HCC: hepatocellular carcinoma; IL-6: interleukin 6; Ki67: a proliferation marker; NASH: non-alcoholic steatohepatitis; NK cells: natural killer cells; NLRP3: NLR family pyrin domain containing 3; TME: tumor microenvironment; TLR: toll-like receptor; TNF-α: tumor necrosis factor alpha; VEGF: vascular endothelial growth factor. Symbols: ↓decrease; ↑increase

- use. This variability can affect the reproducibility and applicability of findings across different demographic and geographic groups.
- iv. Current methodologies for analyzing the gut microbiome, such as 16S rRNA sequencing and metagenomic sequencing, have limitations in resolution, and accuracy and may not capture the full spectrum of microbial diversity or the functional potential of the microbiome. Additionally, these methods are susceptible to contamination and other technical issues that can affect data quality and interpretation.
- v. Modulating the gut microbiota presents a promising therapeutic avenue, but developing effective microbiota-based therapies is challenging. Issues include ensuring the stability and survival of probiotic strains, the unpredictability of prebiotic effects on the existing gut flora, and the potential for adverse effects from broad-spectrum antibiotics.
- vi. The regulatory pathway for microbiota-targeted therapies is not fully established, which may pose challenges in clinical trial design, approval, and market access. Safety concerns also remain, particularly regarding the long-term impacts of altering the gut microbiome on immune function and susceptibility to other diseases.
- vii. The interactions between gut microbiota-modulating therapies and existing treatments for NASH and HCC are not well understood. These interactions could affect the efficacy and safety profiles of treatments.
- viii. The cost of developing microbiota-targeted therapies and the technological demands of such treatments may limit their accessibility, especially in low-resource settings where NASH and HCC are increasingly prevalent.

These limitations underscore the need for further research to better understand the gut microbiome's role in liver diseases and to develop safe, effective, and accessible therapies. Future studies should aim to incorporate larger, more diverse human cohorts, utilize advanced technologies for microbiome analysis, and explore the mechanistic pathways connecting the gut microbiome to liver disease outcomes.

Conclusions

Unhealthy gut microbiota and its metabolites lead to the generation of improper immune signaling in the liver leading to the initiation and progression of different kinds of liver diseases such as MAFLD, MASH, and especially HCC progression. Probiotics, prebiotics, and synbiotics may exemplify advanced, safe, and affordable treatment strategies against these diseases. However, preclinical and well-designed human trials prove that the modifications in the gut microbiota elicit immune modulations in the TME along with anti-tumor response. Understanding the pivotal role of the gut microbiota in the cancer progression may empower the discovery of more effective diagnostic and prevention modalities against HCC. Thus, the treatment of MASH and HCC via targeting the gut microbiota will be an effective research direction in the future for treating MASH-induced HCC. Gene sequencing and machine learning-based data analysis help to identify a key biomarker for the detection of liver illnesses, particularly in MASH-associated HCC. In these circumstances, more numbers of laboratory-based mechanistic evaluations and detailed clinical trials are needed to estimate the composition of gut microflora and this will help to select appropriate useful bacterial strains for the treatment of cancer. Hence, more evidence is needed to translate the existing knowledge relating to the functional aspects of the gut microbiome into diagnostic, prognostic, and therapeutic strategies in patients suffering with HCC. However, evidence suggests that modulation of gut microbiota paves way to a promising therapeutic strategy for the treatment and prevention of MASH and MASH-associated HCC.

Abbreviation lists

ABX Antibiotic cocktails
AMPK AMP-activated protein kinase
AST Aspartate aminotransferase
BA Bile acid

BAS Bile acids
COX-2 Cyclooxygenase-2

cAMP Cyclic adenosine monophosphate CXCL16 Chemokine (C-X-C motif) ligand 16 CXCR6 C-X-C chemokine receptor type 6

DCA Deoxycholic acid

DAMPs Damage-associated molecular patterns

EPA Eicosapentaenoic acid
FMT Fecal microbiota transplantation
FXR Farnesoid X receptor
GPR43 G protein-coupled receptor 43
HCC Hepatocellular carcinoma
HSCs Hepatic stellate cells
ICIs Immune checkpoint inhibitors
IFN-v Interferon gamma

IFN-γ Interferon gamma
 IL-1β Interleukin-1 beta
 IL-6 Interleukin 6
 IL-8 Interleukin 8
 IL-17 Interleukin 17
 LPS Lipopolysaccharides

LSEC Liver sinusoidal endothelial cell

MAFLD Metabolic dysfunction-associated fatty liver disease
MASLD Metabolic dysfunction-associated steatotic liver disease
MASH Metabolic dysfunction-associated steatohepatitis

MDSC Myeloid-derived suppressor cells
NAFLD Non-alcoholic fatty liver disease
NASH Non-alcoholic steatohepatitis
NF-κB Nuclear factor kappa B

NK Natural killer NKT Natural killer T

NLRP3 NLR family pyrin domain containing 3

PAMPs Pathogen-associated molecular patterns

PGE2 Prostaglandin E2
PSK Polysaccharide krestin
RCTs Randomized controlled trials

SASP Senescence-associated secretory phenotype

SCFAs Short-chain fatty acids

SFB Segmented filamentous bacteria
SIBO Small intestinal bacterial overgrowth
TGR5 Takeda G protein-coupled receptor 5

Th17 T helper 17
TLR Toll-like receptor
TLR4 Toll-like receptor 4
TME Tumor microenvironment
TNF-a Tumor necrosis factor-alpha
Treg Regulatory T cells
Tr1 Type 1 regulatory T

VEGF Vascular endothelial growth factor

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References

Rinninella E, Raoul P, Cintoni M, Franceschi F, Miggiano GA, Gasbarrini A, Mele MC. What is the healthy gut microbiota composition?
 A changing ecosystem across age, environment, diet, and diseases. Microorganisms. 2019;7(1):14.

- de Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: mechanistic insights. Gut. 2022;71(5):1020–32.
- 3. Khan AA, Sirsat AT, Singh H, Cash P. Microbiota and cancer: current understanding and mechanistic implications. Clin Transl Oncol. 2022;24(2):193–202.
- Zhang Y, Shen J, Shi X, Du Y, Niu Y, Jin G, Wang Z, Lyu J. Gut microbiome analysis as a predictive marker for the gastric cancer patients. Appl Microbiol Biotechnol. 2021;105:803–14.
- 5. Zhao Y, Liu Y, Li S, Peng Z, Liu X, Chen J, Zheng X. Role of lung and gut microbiota on lung cancer pathogenesis. J Cancer Res Clin Oncol. 2021;147(8):2177–86.
- Sánchez-Alcoholado L, Ramos-Molina B, Otero A, Laborda-Illanes A, Ordóñez R, Medina JA, Gómez-Millán J, Queipo-Ortuño MI. The role of the gut microbiome in colorectal cancer development and therapy response. Cancers. 2020;12(6):1406.
- Sharma J, Huda F, Naithani M, Singh SK, Kumar N, Basu S. Role of gut microbiome and enteric bacteria in gallbladder cancer. In immunology of the GI tract-recent advances 2022. London: IntechOpen; 2022.
- 8. Park SY, Hwang BO, Lim M, Ok SH, Lee SK, Chun KS, Park KK, Hu Y, Chung WY, Song NY. Oral–gut microbiome axis in gastrointestinal disease and cancer. Cancers. 2021;13(9):2124.
- Chen YH, Wu WK, Wu MS. Microbiota-associated therapy for nonalcoholic steatohepatitis-induced liver cancer: a review. Int J Mol Sci. 2020;21(17):5999.
- Yang C, Xu J, Xu X, Xu W, Tong B, Wang S, Ji R, Tan Y, Zhu Y. Characteristics of gut microbiota in patients with metabolic associated fatty liver disease. Sci Rep. 2023;13(1):9988.
- Huang F, Lyu B, Xie F, Li F, Xing Y, Han Z, Lai J, Ma J, Zou Y, Zeng H, Xu Z. From gut to liver: unveiling the differences of intestinal microbiota in NAFL and NASH patients. Front Microbiol. 2024;4(15):1366744.
- Ren Z, Li A, Jiang J, Zhou L, Yu Z, Lu H, Xie H, Chen X, Shao L, Zhang R, Xu S. Gut microbiome analysis as a tool towards targeted non-invasive biomarkers for early hepatocellular carcinoma. Gut. 2019;68(6):1014–23.
- Caligiuri A, Gentilini A, Marra F. Molecular pathogenesis of NASH. Int J Mol Sci. 2016;17(9):1575.
- Thursby E, Juge N. Introduction to the human gut microbiota. Biochem J. 2017;474(11):1823–36.
- Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M. Enterotypes of the human gut microbiome. Nature. 2011;473(7346):174–80.
- Ponziani FR, Nicoletti A, Gasbarrini A, Pompili M. Diagnostic and therapeutic potential of the gut microbiota in patients with early hepatocellular carcinoma. Therapeut Adv Med Oncol. 2019;11:1758835919848184.
- 17. Cholankeril G, Patel R, Khurana S, Satapathy SK. Hepatocellular carcinoma in non-alcoholic steatohepatitis: Current knowledge and implications for management. World J Hepatol. 2017;9(11):533.
- Sobhonslidsuk A, Chanprasertyothin S, Pongrujikorn T, Kaewduang P, Promson K, Petraksa S, Ongphiphadhanakul B. The association of gut microbiota with nonalcoholic steatohepatitis in Thais. Biomed Res Int. 2018;16:2018.
- Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, Harrison SA, Brunt EM, Sanyal AJ. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. Hepatology. 2018;67(1):328–57.
- Nair B, Kamath AJ, Tergaonkar V, Sethi G, Nath LR. Mast cells and the gut-liver Axis: implications for liver disease progression and therapy. Life Sci. 2024;10: 122818.
- 21. Yu J, Marsh S, Hu J, Feng W, Wu C. The pathogenesis of nonalcoholic fatty liver disease: interplay between diet, gut microbiota, and genetic background. Gastroenterol Res Pract. 2016;2016(1):2862173.
- Albillos A, De Gottardi A, Rescigno M. The gut-liver axis in liver disease: pathophysiological basis for therapy. J Hepatol. 2020;72(3):558–77.
- Gupta H, Youn GS, Shin MJ, Suk KT. Role of gut microbiota in hepatocarcinogenesis. Microorganisms. 2019;7(5):121.
- Zhang C, Yang M, Ericsson AC. The potential gut microbiota-mediated treatment options for liver cancer. Front Oncol. 2020;14(10): 524205.

- Ohtani N, Hara E. Gut-liver axis-mediated mechanism of liver cancer: a special focus on the role of gut microbiota. Cancer Sci. 2021;112(11):4433–43.
- 26. Chu H, Williams B, Schnabl B. Gut microbiota, fatty liver disease, and hepatocellular carcinoma. Liver Res. 2018;2(1):43–51.
- 27. Young VB. The role of the microbiome in human health and disease: an introduction for clinicians. BMJ. 2017;15:356.
- 28. Olszak T, An D, Zeissig S, Vera MP, Richter J, Franke A, Glickman JN, Siebert R, Baron RM, Kasper DL, Blumberg RS. Microbial exposure during early life has persistent effects on natural killer T cell function. Science. 2012;336(6080):489–93.
- Loo TM, Kamachi F, Watanabe Y, Yoshimoto S, Kanda H, Arai Y, Nakajima-Takagi Y, Iwama A, Koga T, Sugimoto Y, Ozawa T. Gut microbiota promotes obesity-associated liver cancer through PGE2-mediated suppression of antitumor immunitygut microbiota promotes obesitylinked HCC via immune escape. Cancer Discov. 2017;7(5):522–38.
- Kang Y, Cai Y, Yang Y. The gut microbiome and hepatocellular carcinoma: Implications for early diagnostic biomarkers and novel therapies. Liver Cancer. 2021;11:113.
- 31. Le Roy T, Llopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, Martin P, Philippe C, Walker F, Bado A, Perlemuter G. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. Gut. 2013;62(12):1787–94.
- 32. Wang X, Zhang L, Dong B. Molecular mechanisms in MASLD/MASH related HCC. Hepatology. 2024;13:10–97.
- Kolodziejczyk AA, Zheng D, Shibolet O, Elinav E. The role of the microbiome in NAFLD and NASH. EMBO Mol Med. 2019;11(2): e9302.
- Abdul-Hai A, Abdallah A, Malnick SD. Influence of gut bacteria on development and progression of non-alcoholic fatty liver disease. World J Hepatol. 2015;7(12):1679.
- 35. Chakraborti CK. New-found link between microbiota and obesity. World J Gastrointest Pathophysiol. 2015;6(4):110.
- Arslan N. Obesity, fatty liver disease and intestinal microbiota. World J Gastroenterol WJG. 2014;20(44):16452.
- 37. Mazzotti A, Caletti MT, Sasdelli AS, Brodosi L, Marchesini G. Pathophysiology of nonalcoholic fatty liver disease: lifestyle-gut-gene interaction. Dig Dis. 2016;34(Suppl. 1):3–10.
- 38. Augustyn M, Grys I, Kukla M. Small intestinal bacterial overgrowth and nonalcoholic fatty liver disease. Clin Exp Hepatol. 2019;5(1):1.
- Wijarnpreecha K, Lou S, Watthanasuntorn K, Kroner PT, Cheungpasitporn W, Lukens FJ, Pungpapong S, Keaveny AP, Ungprasert P. Small intestinal bacterial overgrowth and nonalcoholic fatty liver disease: a systematic review and meta-analysis. Eur J Gastroenterol Hepatol. 2020;32(5):601–8.
- Gudan A, Jamioł-Milc D, Hawryłkowicz V, Skonieczna-Żydecka K, Stachowska E. The prevalence of small intestinal bacterial overgrowth in patients with non-alcoholic liver diseases: NAFLD, NASH, fibrosis, cirrhosis—a systematic review, meta-analysis and meta-regression. Nutrients. 2022;14(24):5261.
- 41. Wigg AJ, Roberts-Thomson IC, Dymock RB, McCarthy PJ, Grose RH, Cummins AG. The role of small intestinal bacterial overgrowth, intestinal permeability, endotoxaemia, and tumour necrosis factor α in the pathogenesis of non-alcoholic steatohepatitis. Gut. 2001;48(2):206–11.
- 42. Ghoshal UC, Baba CS, Ghoshal U, Alexander G, Misra A, Saraswat VA, Choudhuri G. Low-grade small intestinal bacterial overgrowth is common in patients with non-alcoholic steatohepatitis on quantitative jejunal aspirate culture. Indian J Gastroenterol. 2017;36:390–9.
- Shanab AA, Scully P, Crosbie O, Buckley M, O'Mahony L, Shanahan F, Gazareen S, Murphy E, Quigley EM. Small intestinal bacterial overgrowth in nonalcoholic steatohepatitis: association with toll-like receptor 4 expression and plasma levels of interleukin 8. Dig Dis Sci. 2011;56:1524–34.
- Thuy S, Ladurner R, Volynets V, Wagner S, Strahl S, Königsrainer A, Maier KP, Bischoff SC, Bergheim I. Nonalcoholic fatty liver disease in humans is associated with increased plasma endotoxin and plasminogen activator inhibitor 1 concentrations and with fructose intake. J Nutr. 2008;138(8):1452–5.
- Tranah TH, Edwards LA, Schnabl B, Shawcross DL. Targeting the gut– liver–immune axis to treat cirrhosis. Gut. 2021;70(5):982–94.
- 46. Odena G, Chen J, Lozano JJ, Altamirano J, Rodrigo-Torres D, Affo S, Morales-Ibanez O, Matsushita H, Zou J, Dumitru R, Caballeria J. LPS-TLR4

- pathway mediates ductular cell expansion in alcoholic hepatitis. Sci Rep. 2016;6(1):35610.
- 47. Soares JB, Pimentel-Nunes P, Roncon-Albuquerque R, Leite-Moreira A. The role of lipopolysaccharide/toll-like receptor 4 signaling in chronic liver diseases. Hep Intl. 2010;4:659–72.
- 48. Chassaing B, Gewirtz AT. Gut microbiota, low-grade inflammation, and metabolic syndrome. Toxicol Pathol. 2014;42(1):49–53.
- Liu J, Zhuang ZJ, Bian DX, Ma XJ, Xun YH, Yang WJ, Luo Y, Liu YL, Jia L, Wang Y, Zhu ML. Toll-like receptor-4 signalling in the progression of non-alcoholic fatty liver disease induced by high-fat and high-fructose diet in mice. Clin Exp Pharmacol Physiol. 2014;41(7):482–8.
- Nobili V, Carpino G, Alisi A, Franchitto A, Alpini G, De Vito R, Onori P, Alvaro D, Gaudio E. Hepatic progenitor cells activation, fibrosis, and adipokines production in pediatric nonalcoholic fatty liver disease. Hepatology. 2012;56(6):2142–53.
- 51. Yoon HJ, Cha BS. Pathogenesis and therapeutic approaches for nonalcoholic fatty liver disease. World J Hepatol. 2014;6(11):800.
- Brandi G, De Lorenzo S, Candela M, Pantaleo MA, Bellentani S, Tovoli F, Saccoccio G, Biasco G. Microbiota, NASH, HCC and the potential role of probiotics. Carcinogenesis. 2017;38(3):231–40.
- Miura K, Kodama Y, İnokuchi S, Schnabl B, Aoyama T, Ohnishi H, Olefsky JM, Brenner DA, Seki E. Toll-like receptor 9 promotes steatohepatitis by induction of interleukin-1β in mice. Gastroenterology. 2010;139(1):323–34.
- 54. Rivera CA, Adegboyega P, van Rooijen N, Tagalicud A, Allman M, Wallace M. Toll-like receptor-4 signaling and Kupffer cells play pivotal roles in the pathogenesis of non-alcoholic steatohepatitis. J Hepatol. 2007;47(4):571–9.
- Del Chierico F, Nobili V, Vernocchi P, Russo A, De Stefanis C, Gnani D, Furlanello C, Zandonà A, Paci P, Capuani G, Dallapiccola B. Gut microbiota profiling of pediatric nonalcoholic fatty liver disease and obese patients unveiled by an integrated meta-omics-based approach. Hepatology. 2017;65(2):451–64.
- Dapito DH, Mencin A, Gwak GY, Pradere JP, Jang MK, Mederacke I, Caviglia JM, Khiabanian H, Adeyemi A, Bataller R, Lefkowitch JH. Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. Cancer Cell. 2012;21(4):504–16.
- 57. Yuan D, Huang S, Berger E, Liu L, Gross N, Heinzmann F, Ringelhan M, Connor TO, Stadler M, Meister M, Weber J. Kupffer cell-derived Tnf triggers cholangiocellular tumorigenesis through JNK due to chronic mitochondrial dysfunction and ROS. Cancer Cell. 2017;31(6):771–89.
- 58. Wang X, Fu X, Van Ness C, Meng Z, Ma X, Huang W. Bile acid receptors and liver cancer. Curr Pathobiol Reports. 2013;1:29–35.
- Chen T, Xie G, Wang X, Fan J, Qiu Y, Zheng X, Qi X, Cao Y, Su M, Wang X, Xu LX. Serum and urine metabolite profiling reveals potential biomarkers of human hepatocellular carcinoma. Mol Cell Proteom. 2011. https://doi.org/10.1074/mcp.M110.004945.
- 60. Jansen PL. Endogenous bile acids as carcinogens. J Hepatol. 2007;47(3):434–5.
- Ma C, Han M, Heinrich B, Fu Q, Zhang Q, Sandhu M, Agdashian D, Terabe M, Berzofsky JA, Fako V, Ritz T. Gut microbiome—mediated bile acid metabolism regulates liver cancer via NKT cells. Science. 2018;360(6391):eaan5931.
- 62. Jia B. Commentary: gut microbiome–mediated bile acid metabolism regulates liver cancer via NKT cells. Front Immunol. 2019;20(10):282.
- Yamada S, Takashina Y, Watanabe M, Nagamine R, Saito Y, Kamada N, Saito H. Bile acid metabolism regulated by the gut microbiota promotes non-alcoholic steatohepatitis-associated hepatocellular carcinoma in mice. Oncotarget. 2018;9(11):9925.
- 64. Guerra Ruiz A, Casafont F, Crespo J, Cayón A, Mayorga M, Estebanez A, Fernadez-Escalante JC, Pons-Romero F. Lipopolysaccharide-binding protein plasma levels and liver TNF-alpha gene expression in obese patients: evidence for the potential role of endotoxin in the pathogenesis of non-alcoholic steatohepatitis. Obes Surg. 2007;17:1374–80.
- Shao G, Liu Y, Lu L, Zhang G, Zhou W, Wu T, Wang L, Xu H, Ji G. The pathogenesis of HCC driven by NASH and the preventive and therapeutic effects of natural products. Front Pharmacol. 2022. https://doi. org/10.3389/fphar.2022.944088.
- Duseja A, Chawla YK. Obesity and NAFLD: the role of bacteria and microbiota. Clin Liver Dis. 2014;18(1):59–71.

- Arrese M, Cabrera D, Kalergis AM, Feldstein AE. Innate immunity and inflammation in NAFLD/NASH. Dig Dis Sci. 2016;61:1294–303.
- Liu X, Chen Y, Zhang S, Dong L. Gut microbiota-mediated immunomodulation in tumor. J Exp Clin Cancer Res. 2021;40(1):1–20.
- Li Q, Ma L, Shen S, Guo Y, Cao Q, Cai X, Feng J, Yan Y, Hu T, Luo S, Zhou L. Intestinal dysbacteriosis-induced IL-25 promotes development of HCC via alternative activation of macrophages in tumor microenvironment. J Exp Clin Cancer Res. 2019;38:1–3.
- Zhang Q, Ma C, Duan Y, Heinrich B, Rosato U, Diggs LP, Ma L, Roy S, Fu Q, Brown ZJ, Wabitsch S. Gut Microbiome directs hepatocytes to recruit MDSCS and promote cholangiocarcinoma the gut microbiome controls hepatic MDSCs. Cancer Discov. 2021;11(5):1248–67.
- Behary J, Amorim N, Jiang XT, Raposo A, Gong L, McGovern E, Ibrahim R, Chu F, Stephens C, Jebeili H, Fragomeli V. Gut microbiota impact on the peripheral immune response in non-alcoholic fatty liver disease related hepatocellular carcinoma. Nat Commun. 2021;12(1):187.
- 72. Zhou A, Tang L, Zeng S, Lei Y, Yang S, Tang B. Gut microbiota: a new piece in understanding hepatocarcinogenesis. Cancer Lett. 2020;1(474):15–22.
- 73 Zhou J, Tripathi M, Sinha RA, Singh BK, Yen PM. Gut microbiota and their metabolites in the progression of non-alcoholic fatty liver disease. Hepatoma Res. 2021. https://doi.org/10.20517/2394-5079.2020.134.
- Scarpellini E, Scarlata GG, Santori V, Scarcella M, Kobyliak N, Abenavoli L. Gut microbiota, deranged immunity, and hepatocellular carcinoma. Biomedicines. 2024;12(8):1797.
- Schneider KM, Mohs A, Gui W, Galvez EJ, Candels LS, Hoenicke L, Muthukumarasamy U, Holland CH, Elfers C, Kilic K. Imbalanced gut microbiota fuels hepatocellular carcinoma development by shaping the hepatic inflammatory microenvironment. Nat Commun. 2022. https://doi.org/10.1038/s41467-022-31312-5.
- Dai X, Hou H, Zhang W, Liu T, Li Y, Wang S, Wang B, Cao H. Microbial metabolites: critical regulators in NAFLD. Front Microbiol. 2020;7(11): 567654
- Liu G, Abas O, Strickland AB, Chen Y, Shi M. CXCR6+ CD4+ T cells promote mortality during trypanosoma Brucei infection. PLoS Pathog. 2021;17(10): e1009968.
- Culligan EP, Hill C, Sleator RD. Probiotics and gastrointestinal disease: successes, problems and future prospects. Gut pathogens. 2009;1:1–2.
- 79 Elshaer AM, El-Kharashi OA, Hamam GG, Nabih ES, Magdy YM, Abd El Samad AA. Involvement of TLR4/CXCL9/PREX-2 pathway in the development of hepatocellular carcinoma (HCC) and the promising role of early administration of lactobacillus plantarum in Wistar rats. Tissue Cell. 2019;60:38–47.
- Wong VW, Wong GL, Chim AM, Chu WC, Yeung DK, Li KC, Chan HL. Treatment of nonalcoholic steatohepatitis with probiotics. A proof-of-concept study. Ann Hepatol. 2013;12(2):256–62.
- 81. Murugaiyan G, Saha B. Protumor vs antitumor functions of IL-17. J Immunol. 2009;183(7):4169–75.
- Russo E, Nannini G, Dinu M, Pagliai G, Sofi F, Amedei A. Exploring the food–gut axis in immunotherapy response of cancer patients. World J Gastroenterol. 2020;26(33):4919.
- Fotiadis CI, Stoidis CN, Spyropoulos BG, Zografos ED. Role of probiotics, prebiotics and synbiotics in chemoprevention for colorectal cancer. World J Gastroenterol WJG. 2008;14(42):6453.
- Larrosa M, González-Sarrías A, García-Conesa MT, Tomás-Barberán FA, Espín JC. Urolithins, ellagic acid-derived metabolites produced by human colonic microflora, exhibit estrogenic and antiestrogenic activities. J Agric Food Chem. 2006;54(5):1611–20.
- Rodríguez-Lara A, Rueda-Robles A, Sáez-Lara MJ, Plaza-Diaz J, Álvarez-Mercado Al. From non-alcoholic fatty liver disease to liver cancer: microbiota and inflammation as key players. Pathogens. 2023;12(7):940.
- González-Sarrías A, Larrosa M, Tomás-Barberán FA, Dolara P, Espín JC. NF-kB-dependent anti-inflammatory activity of urolithins, gut microbiota ellagic acid-derived metabolites, in human colonic fibroblasts. Br J Nutr. 2010;104(4):503–12.
- 87. Heebøll S, El-Houri RB, Hellberg YE, Haldrup D, Pedersen SB, Jessen N, Christensen LP, Grønbæk H. Effect of resveratrol on experimental non-alcoholic fatty liver disease depends on severity of pathology and timing of treatment. J Gastroenterol Hepatol. 2016;31(3):668–75.
- 88. Carpi RZ, Barbalho SM, Sloan KP, Laurindo LF, Gonzaga HF, Grippa PC, Zutin TL, Girio RJ, Repetti CS, Detregiachi CR, Bueno PC. The effects of

- probiotics, prebiotics and synbiotics in non-alcoholic fat liver disease (NAFLD) and non-alcoholic steatohepatitis (NASH): a systematic review. Int J Mol Sci. 2022;23(15):8805.
- Yeh CB, Hsieh MJ, Lin CW, Chiou HL, Lin PY, Chen TY, Yang SF. The antimetastatic effects of resveratrol on hepatocellular carcinoma through the downregulation of a metastasis-associated protease by SP-1 modulation. PLoS ONE. 2013;8(2): e56661.
- Athar M, Back JH, Kopelovich L, Bickers DR, Kim AL. Multiple molecular targets of resveratrol: anti-carcinogenic mechanisms. Arch Biochem Biophys. 2009;486(2):95–102.
- 91. Malaguarnera L. Influence of resveratrol on the immune response. Nutrients. 2019;11(5):946.
- Martínez-Flórez S, Gutiérrez-Fernández B, Sánchez-Campos S, González-Gallego J, Tuñón MJ. Quercetin attenuates nuclear factor-κB activation and nitric oxide production in interleukin-1β–activated rat hepatocytes. J Nutr. 2005;135(6):1359–65.
- Marcolin E, Forgiarini LF, Rodrigues G, Tieppo J, Borghetti GS, Bassani VL, Picada JN, Marroni NP. Retracted: quercetin decreases liver damage in mice with non-alcoholic steatohepatitis. Basic Clin Pharmacol Toxicol. 2013;112(6):385–91.
- 94. Acharya S, Adamová D, Adhya SP, Adler A, Adolfsson J, Aggarwal MM, Rinella GA, Agnello M, Agrawal N, Ahammed Z, Ahmad S. Investigations of anisotropic flow using multiparticle azimuthal correlations in p p, p—Pb, Xe–Xe, and Pb–Pb collisions at the LHC. Phys Rev Lett. 2019;123(14): 142301.
- 95. Lamuel-Raventos RM, Onge MP. Prebiotic nut compounds and human microbiota. Crit Rev Food Sci Nutr. 2017;57(14):3154–63.
- 96. Plaz Torres MC, Bodini G, Furnari M, Marabotto E, Zentilin P, Giannini EG. Nuts and non-alcoholic fatty liver disease: are nuts safe for patients with fatty liver disease? Nutrients. 2020;12(11):3363.
- 97. Fotschki B, Juśkiewicz J, Jurgoński A, Sójka M. Fructo-oligosaccharides and pectins enhance beneficial effects of raspberry polyphenols in rats with nonalcoholic fatty liver. Nutrients. 2021;13(3):833.
- Russo E, Fiorindi C, Giudici F, Amedei A. Immunomodulation by probiotics and prebiotics in hepatocellular carcinoma. World J Hepatol. 2022;14(2):372.
- Fotschki B, Laparra JM, Sójka M. Raspberry polyphenolic extract regulates obesogenic signals in hepatocytes. Molecules. 2018;23(9):2103.
- Wongkrasant P, Pongkorpsakol P, Ariyadamrongkwan J, Meesomboon R, Satitsri S, Pichyangkura R, Barrett KE, Muanprasat C. A prebiotic fructo-oligosaccharide promotes tight junction assembly in intestinal epithelial cells via an AMPK-dependent pathway. Biomed Pharmacother. 2020;1(129): 110415.
- 101. Wang J, Tian S, Yu H, Wang J, Zhu W. Response of colonic mucosaassociated microbiota composition, mucosal immune homeostasis, and barrier function to early life galactooligosaccharides intervention in suckling piglets. J Agric Food Chem. 2018;67(2):578–88.
- 102 Fatima N, Akhtar T, Sheikh N. Prebiotics: a novel approach to treat hepatocellular carcinoma. Canadian J Gastroenterol Hepatol. 2017. https:// doi.org/10.1155/2017/6238106.
- 103. Yoshimoto S, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, Iwakura Y, Oshima K, Morita H, Hattori M, Honda K. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. Nature. 2013;499(7456):97–101.
- Xiang H, Sun D, Liu X, She ZG, Chen Y. The role of the intestinal microbiota in nonalcoholic steatohepatitis. Front Endocrinol. 2022;8(13):15.
- Li K, Liu J, Qin X. Research progress of gut microbiota in hepatocellular carcinoma. J Clin Lab Anal. 2022;36(7): e24512.
- Tarantino G, Citro V. Could adverse effects of antibiotics due to their use/misuse be linked to some mechanisms related to nonalcoholic fatty liver disease? Int J Mol Sci. 2024;25(4):1993.
- Francino M. Antibiotics and the human gut microbiome: dysbioses and accumulation of resistances. Front Microbiol. 2016;12(6): 164577.
- Ramirez J, Guarner F, Bustos Fernandez L, Maruy A, Sdepanian VL, Cohen H. Antibiotics as major disruptors of gut microbiota. Front Cell Infect Microbiol. 2020;24(10): 572912.
- Ribeiro CF, Silveira GG, Candido ED, Cardoso MH, Espinola Carvalho CM, Franco OL. Effects of antibiotic treatment on gut microbiota and how to overcome its negative impacts on human health. ACS Infect Dis. 2020;6(10):2544–59.

- Bassetti M, Villa G, Pecori D, Arzese A, Wilcox M. Epidemiology, diagnosis and treatment of clostridium difficile infection. Expert Rev Anti Infect Ther. 2012;10(12):1405–23.
- 111. Sinh P, Barrett TA, Yun L. Clostridium difficile infection and inflammatory bowel disease: a review. Gastroenterol Res Pract. 2011;2011(1): 136064.
- Cojocariu C, Stanciu C, Stoica O, Singeap AM, Sfarti C, Girleanu I, Trifan A. Clostridium difficile infection and inflammatory bowel disease. Turk J Gastroenterol. 2014;25(6):603–10.
- 113. Choi HH, Cho YS. Fecal microbiota transplantation: current applications, effectiveness, and future perspectives. Clin Endosc. 2016;49(3):257–65.
- 114. Craven L, Rahman A, Parvathy SN, Beaton M, Silverman J, Qumosani K, Hramiak I, Hegele R, Joy T, Meddings J, Urquhart B. Allogenic fecal microbiota transplantation in patients with nonalcoholic fatty liver disease improves abnormal small intestinal permeability: a randomized control trial. Off J Am Coll Gastroenterol ACG. 2020;115(7):1055–65.
- Gulati M, Singh SK, Corrie L, Kaur IP, Chandwani L. Delivery routes for faecal microbiota transplants: available, anticipated and aspired. Pharmacol Res. 2020;1(159): 104954.
- Blair HA. SER-109 (VOWST[™]): a review in the prevention of recurrent clostridioides difficile infection. Drugs. 2024;84(3):329–36.
- Wang Y, Hunt A, Danziger L, Drwiega EN. A comparison of currently available and investigational fecal microbiota transplant products for recurrent clostridioides difficile infection. Antibiotics. 2024;13(5):436.
- 118. Quraishi MN, Widlak M, Bhala NA, Moore D, Price M, Sharma N, Iqbal TH. Systematic review with meta-analysis: the efficacy of faecal microbiota transplantation for the treatment of recurrent and refractory Clostridium difficile infection. Aliment Pharmacol Ther. 2017;46(5):479–93.
- 119. Vrieze A, Out C, Fuentes S, Jonker L, Reuling I, Kootte RS, Van Nood E, Holleman F, Knaapen M, Romijn JA, Soeters MR. Impact of oral vancomycin on gut microbiota, bile acid metabolism, and insulin sensitivity. J Hepatol. 2014;60(4):824–31.
- Panda S, El Khader I, Casellas F, Lopez Vivancos J, Garcia Cors M, Santiago A, Cuenca S, Guarner F, Manichanh C. Short-term effect of antibiotics on human gut microbiota. PLoS ONE. 2014;9(4): e95476.
- 121. Zhu J, Ding J, Li S, Jin J. Ganoderic acid A ameliorates non-alcoholic steatohepatitis (NASH) induced by high-fat high-cholesterol diet in mice. Exp Ther Med. 2022;23(4):1–9.
- 122. Yen IC, Lin JC, Chen Y, Tu QW, Lee SY. Antrodia cinnamomea attenuates non-alcoholic steatohepatitis by suppressing NLRP3 inflammasome activation in vitro and in vivo. Am J Chin Med. 2020;48(08):1859–74.
- Chen L, Zhang L, Wang W, Qiu W, Liu L, Ning A, Cao J, Huang M, Zhong M. Polysaccharides isolated from Cordyceps Sinensis contribute to the progression of NASH by modifying the gut microbiota in mice fed a high-fat diet. PLoS ONE. 2020;15(6): e0232972.
- 124 Li S, Tan HY, Wang N, Cheung F, Hong M, Feng Y. The potential and action mechanism of polyphenols in the treatment of liver diseases. Oxidat Med Cell Longevity. 2018. https://doi.org/10.1155/2018/83948 18.
- Jeong JJ, Park HJ, Cha MG, Park E, Won SM, Ganesan R, Gupta H, Gebru YA, Sharma SP, Lee SB, Kwon GH. The lactobacillus as a probiotic: focusing on liver diseases. Microorganisms. 2022;10(2):288.
- YU J, LIU Q. Advances in lactobacillus Rhamnosus GG and pediatric nonalcoholic fatty liver disease. Chin J Appl Clin Pediatr. 2022:389–92.
- 127. Zhang HL, Yu LX, Yang W, Tang L, Lin Y, Wu H, Zhai B, Tan YX, Shan L, Liu Q, Chen HY. Profound impact of gut homeostasis on chemicallyinduced pro-tumorigenic inflammation and hepatocarcinogenesis in rats. J Hepatol. 2012;57(4):803–12.
- Singh V, Yeoh BS, Abokor AA, Golonka RM, Tian Y, Patterson AD, Joe B, Heikenwalder M, Vijay-Kumar M. Vancomycin prevents fermentable fiber-induced liver cancer in mice with dysbiotic gut microbiota. Gut Microbes. 2020;11(4):1077–91.
- 129. Eguchi A, Mizukami S, Nakamura M, Masuda S, Murayama H, Kawashima M, Inohana M, Nagahara R, Kobayashi M, Yamashita R, Uomoto S. Metronidazole enhances steatosis-related early-stage hepatocarcinogenesis in high fat diet-fed rats through DNA doublestrand breaks and modulation of autophagy. Environ Sci Pollut Res. 2022:29:779–89
- 130. Yamagishi R, Kamachi F, Nakamura M, Yamazaki S, Kamiya T, Takasugi M, Cheng Y, Nonaka Y, Yukawa-Muto Y, Thuy LT, Harada Y. Gasdermin D-mediated release of IL-33 from senescent hepatic stellate cells

promotes obesity-associated hepatocellular carcinoma. Sci Immunol. 2022;7(72):eabll7209.

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