



REVIEW

REVISED **Current trends in the treatment of pneumonia due to multidrug-resistant Gram-negative bacteria [version 2; referees: 2 approved]**

Richard R. Watkins^{1,2}, David Van Duin³

¹Division of Infectious Diseases, Cleveland Clinic Akron General, Akron, OH, 44302, USA

²Department of Medicine, Northeast Ohio Medical University, Rootstown, OH, 44272, USA

³Department of Medicine, University of North Carolina, Chapel Hill, NC, 27514, USA

v2 **First published:** 30 Jan 2019, 8(F1000 Faculty Rev):121 (<https://doi.org/10.12688/f1000research.16517.1>)
Latest published: 06 Feb 2019, 8(F1000 Faculty Rev):121 (<https://doi.org/10.12688/f1000research.16517.2>)

Abstract

Pneumonia is one of the most common infections worldwide. Morbidity, mortality, and healthcare costs increase substantially when pneumonia is caused by multidrug-resistant Gram-negative bacteria (MDR-GNB). The ongoing spread of antimicrobial resistance has made treating MDR-GNB pneumonia increasingly difficult. Fortunately, there have been some recent additions to our antibiotic armamentarium in the US and Europe for MDR-GNB, along with several agents that are in advanced stages of development. In this article, we review the risk factors for and current management of MDR-GNB pneumonia as well as novel agents with activity against these important and challenging pathogens.

Keywords

Enterobacteriaceae, Pseudomonas aeruginosa, Acinetobacter baumannii, antibiotics

Open Peer Review

Referee Status:

	Invited Referees	
	1	2
REVISED version 2 published 06 Feb 2019		
version 1 published 30 Jan 2019		

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty. In order to make these reviews as comprehensive and accessible as possible, peer review takes place before publication; the referees are listed below, but their reports are not formally published.

- 1 **Frank Schweizer**, University of Manitoba, Canada
- 2 **Dean Winslow**, Stanford University School of Medicine, USA

Discuss this article

Comments (0)

Corresponding author: Richard R. Watkins (WatkinR2@ccf.org)

Author roles: **Watkins RR:** Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Van Duin D:** Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: RRW serves on an advisory board and speakers' bureau and has received research support from Allergan. DvD serves on advisory boards for Allergan, Achaogen, Shionogi, Tetraphase, Sanofi Pasteur, MedImmune, and Astellas and has received research funding from Steris Inc. and Scynexis.

Grant information: The author(s) declared that no grants were involved in supporting this work.

Copyright: © 2019 Watkins RR and Van Duin D. This is an open access article distributed under the terms of the [Creative Commons Attribution Licence](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Watkins RR and Van Duin D. **Current trends in the treatment of pneumonia due to multidrug-resistant Gram-negative bacteria [version 2; referees: 2 approved]** *F1000Research* 2019, 8(F1000 Faculty Rev):121 (<https://doi.org/10.12688/f1000research.16517.2>)

First published: 30 Jan 2019, 8(F1000 Faculty Rev):121 (<https://doi.org/10.12688/f1000research.16517.1>)

REVISED Amendments from Version 1

This version corrects an error in version 1 that stated there is an oral formulation of eravacycline, which there is not.

See referee reports

Introduction

Infections due to multidrug-resistant Gram-negative bacteria (MDR-GNB) pose a serious and increasing threat to human health. This is particularly true in the pathogenesis of pneumonia, where escalating rates of antimicrobial resistance (AMR) have been associated with excessive morbidity, mortality, and health-care costs¹. It is well recognized that timely and effective therapy is vital for improving outcomes, especially for pneumonia that is hospital acquired². The choice of initial antibiotic therapy for pneumonia is based on several factors, including recommendations from guidelines, national and local antimicrobial susceptibility data, and patient characteristics such as allergies and renal function. For many years, the backbone of treatment for pneumonia has been the β -lactam class of antibiotics, including the third- and fourth-generation cephalosporins and β -lactam/ β -lactamase inhibitor combinations like piperacillin/tazobactam³. Unfortunately, the ongoing spread of extended-spectrum β -lactamases (ESBLs) and carbapenemases such as *Klebsiella pneumoniae* carbapenemase (KPC) has begun to limit the clinical effectiveness of β -lactam agents over the last decade^{4,5}.

The diagnosis of pneumonia can be challenging, especially in cases of hospital-acquired pneumonia (HAP) and ventilator-associated pneumonia (VAP). Indeed, pulmonary infiltrates on imaging in critically ill patients are common and can be due to non-infectious etiologies, including atelectasis, acute respiratory distress syndrome (ARDS), congestive heart failure (CHF), pulmonary hemorrhages, and pulmonary infarction. Moreover, upper airways and endotracheal tubes of hospitalized patients are often colonized by MDR-GNB and their presence does not necessarily mean that they are the cause of the pulmonary abnormalities seen on imaging studies. A careful clinical assessment is therefore imperative when evaluating for pneumonia, especially in patients who have had a prolonged hospitalization. The current HAP/VAP guidelines from the Infectious Diseases Society of America are an excellent reference for help with diagnosing these cases².

The initial approach to pneumonia is most often empirical because results of antimicrobial susceptibility testing typically take 48 to 72 hours. Rapid diagnostic tests (RDTs), including molecular methods that identify specific resistance genes or automated microscopy that can quickly determine antibiotic susceptibility, have great potential for guiding empiric antibiotic therapy. But current RDTs have limitations and most have not been validated for respiratory secretions⁶. Deciding on an appropriate empirical regimen can be difficult because clinicians must consider the benefits of starting therapy early versus the harms of unnecessary coverage. Indeed, inappropriate antimicrobial treatment or delays in starting appropriate treatment in VAP are associated with increased morbidity and mortality⁷. Once susceptibility testing

results are available, empiric antibiotic therapy should be de-escalated. Most cases of MDR-GNB pneumonia can be successfully treated with 7 days of therapy².

Several risk factors for MDR-GNB pneumonia have been identified. These include prior infection or colonization with MDR-GNB, antibiotic therapy in the past 90 days, poor functional status performance, hospitalization for more than 2 days in the past 90 days, occurrence 5 or more days after admission to an acute hospital, receiving hemodialysis, and immunosuppression^{8,9}. Moreover, prior receipt of carbapenems, broad-spectrum cephalosporins, and fluorquinolones has been associated specifically with MDR *Pseudomonas aeruginosa*¹⁰.

Recently, the high mortality and mortality associated with MDR-GNB pneumonia along with limited treatment options have led to a resurgence in the use of the nephrotoxic drug colistin¹¹. Fortunately, several new antibiotic agents with activity against MDR-GNB, including plazomicin, ceftazidime/avibactam, and meropenem/vaborbactam, have become available. This review discusses new antibiotic options for MDR-GNB and those in late stages of clinical development and provides guidance for their use in treating MDR-GNB pneumonia.

Multidrug-resistant Enterobacteriaceae

Similar to other MDR-GNB, MDR Enterobacteriaceae are usually encountered as the cause of healthcare-associated pneumonia and are less commonly seen in community-acquired pneumonia (CAP). In a recent large intensive care unit (ICU) study of 75 US centers, the most common Enterobacteriaceae isolated from patients with pneumonia were *Klebsiella pneumoniae*, *Enterobacter* spp., and *Escherichia coli*, which accounted for 12%, 8%, and 7% of all bacterial isolates included in the study, respectively¹². Important MDR Enterobacteriaceae that cause pneumonia include those bacteria that produce AmpC enzymes, ESBL, or carbapenemases or a combination of these. AmpC and ESBL producers are usually resistant to most, if not all, cephalosporins. ESBL but not AmpC producers are variably inhibited by β -lactamase inhibitors. Also, AmpC enzymes are frequently found in *Enterobacter* spp. and may be induced by antibiotic treatment, leading to treatment-emergent resistance¹³. As AmpC enzymes do not effectively hydrolyze cefepime, AmpC-producing Enterobacteriaceae often retain *in vitro* susceptibility to cefepime¹³. The management of pneumonia caused by carbapenem-resistant Enterobacteriaceae (CRE) is the most challenging. In a longitudinal cohort study of patients with CRE, pneumonia and bloodstream infections (BSIs) were found to be associated with the highest mortality rates¹⁴. When compared with comparable patients colonized with CRE, CRE pneumonia had an excess hospital mortality of 27% and adjusted hazard ratio of 3.44 (95% confidence interval [CI] 1.80–6.48, $p < 0.001$) for time to death¹⁴. Risk factors for MDR Enterobacteriaceae as a cause of pneumonia overlap with those of other MDR organisms and include prior exposure to antibiotics, healthcare exposure, and use of medical devices such as urinary catheters^{15,16}.

An excellent comprehensive review on the therapy of MDR Enterobacteriaceae was recently published by Rodríguez-Baño *et al.* and the reader is referred to this review for additional

background¹⁷. It is important to note that the MERINO trial (Meropenem versus piperacillin-tazobactam for definitive treatment of bloodstream infections due to ceftriaxone non-susceptible *Escherichia coli* and *Klebsiella* spp.) had not yet been published at the time of that review. In the MERINO trial, patients with BSI caused by ceftriaxone-resistant Enterobacteriaceae were randomly assigned to receive either piperacillin/tazobactam or meropenem in an open-label non-inferiority design¹⁸. The mechanism of resistance in these isolates was an ESBL in about 85% and AmpC in about 10%. In contrast to some observational studies, the mortality in piperacillin/tazobactam-treated patients was significantly higher as compared with those treated with meropenem (12% versus 4%)^{18,19}. A total of 12 out of 379 patients had pneumonia as a source of BSI in this study. Similarly, in a randomized controlled trial comparing cefepime versus imipenem in the treatment of pneumonia, a decreased efficacy of cefepime was noted in patients infected with ESBL-producing Enterobacteriaceae²⁰. Based on these data, carbapenems should be considered first-line treatment for pneumonia caused by ESBL-producing Enterobacteriaceae. In an observational study of AmpC-producing Enterobacteriaceae BSI, patients who received definitive therapy with piperacillin/tazobactam were compared with a group of patients who received either cefepime or piperacillin/tazobactam. Only 14% and 20% of patients had pneumonia as the source of BSI²¹. Although no statistically significant differences were found, it was notable that the 30-day mortality in the piperacillin/tazobactam-treated patients was 15% versus 7% in the meropenem- or cefepime-treated patients²¹. Based on these clinical data and the known hydrolysis of piperacillin by AmpC, piperacillin/tazobactam cannot be considered first-line treatment for AmpC-producing Enterobacteriaceae. As mechanisms of resistance are usually not available to treating clinicians, practically speaking, piperacillin/tazobactam should not be used for pneumonia caused by *Enterobacter* spp., *Serratia* spp., or *Citrobacter* spp.

Whether cefepime can be successfully used in the treatment of pneumonia caused by AmpC-producing Enterobacteriaceae remains unclear. In a propensity-adjusted analysis of observational data on patients with infections caused by AmpC-producing Enterobacteriaceae, treatment with cefepime (n = 32) was compared with meropenem (n = 32)²². Pneumonia was present in 53% and 41% of cefepime- and meropenem-treated patients, respectively. Overall, no difference in 30-day mortality was seen (odds ratio [OR] 0.63, 95% CI 0.23–211), but it should be noted that this OR had a wide CI that included both a fourfold decreased odds of 30-day mortality in cefepime-treated patients and a twofold increased odds²². Given the absence of activity of AmpC against cefepime and these limited clinical results, cefepime is a reasonable carbapenem-sparing option for pneumonia caused by AmpC-positive Enterobacteriaceae.

Carbapenem resistance in Enterobacteriaceae can be mediated through carbapenemases, which are usually carried on mobile genetic elements such as plasmids (carbapenemase-producing Enterobacteriaceae, or CPE), or through a variety of other mechanisms such as porin mutations (non-carbapenemase-producing CRE). Important classes of carbapenemases include KPC, New Delhi metallo- β -lactamases (NDMs), and OXA-48-like

carbapenemases²³. Prior to the availability of newer antibiotics, treatment of invasive CRE infections included the use of polymyxins, tigecycline, and aminoglycosides, often given in combination regimens²⁴. More recently, meropenem/vaborbactam, ceftazidime/avibactam, eravacycline, and plazomicin have become available^{25–27}. These agents have specific *in vitro* anti-CRE activity. Plazomicin has broad activity regardless of carbapenemase but is inactivated by 16S rRNA ribosomal methyltransferases that are present in some NDM-producing CPE²⁸. Meropenem/vaborbactam is active against KPC-producing CPE, and avibactam inhibits KPC and OXA-48-like carbapenemases^{25,29}. Eravacycline is a fluorocycline antibiotic similar in structure to tigecycline with activity against CRE³⁰. In addition, there are several other anti-CRE agents in the pipeline, including cefiderocol, imipenem/relebactam, and meropenem/nacubactam.

With the availability of these new agents, there are some (but no definitive) clinical data available on the best treatment of CRE infections, including CRE pneumonia. In an observational study, patients with CRE infections—caused primarily by KPC producers—who were started on colistin (n = 99) were compared with patients who were started on ceftazidime/avibactam (n = 38)³¹. In an inverse probability of treatment-weighted analysis, the 30-day mortality in patients on colistin was 32% versus 9% (absolute difference 23%, 95% CI 9%–35%; $p = 0.001$) in those on ceftazidime/avibactam. Pneumonia was present in 24% and 32% of patients on colistin and ceftazidime/avibactam, respectively³¹. In a randomized controlled trial, plazomicin was compared with colistin—both given in combination with either tigecycline or meropenem—in CRE BSI (n = 29) or pneumonia (n = 8)³². Plazomicin versus colistin therapy was associated with 2/17 (12%) versus 8/20 (40%) all-cause mortality at day 28³². Similarly, meropenem/vaborbactam (n = 28) was compared with best available therapy (n = 15) for CRE infections, 5 of which were pneumonia²⁶. Rates of clinical cure at test of cure visit were 57% and 27% in the meropenem-vaborbactam and best available therapy arms, respectively²⁶. Based on these data, it is clear that polymyxin-based therapy is inferior to treatment with one of these novel agents. Although only limited numbers of patients with CRE pneumonia were studied, the use of either meropenem-vaborbactam or ceftazidime-avibactam in CRE pneumonia is a reasonable approach while awaiting more data. Ceftazidime/avibactam was shown to be non-inferior to meropenem in a recent large randomized controlled trial of non-CRE pneumonia³³. Meropenem itself has been used as a comparator agent in many pneumonia studies, and vaborbactam achieves high epithelial lining fluid concentrations³⁴. In contrast, in the absence of confirmatory data, plazomicin should not be considered a first-line choice for monotherapy of CRE pneumonia.

Pseudomonas aeruginosa

The acquisition of MDR *P. aeruginosa*, a significant and increasing cause of HAP/VAP in North America and Europe, is related to both patient factors (for example, older age, previous colonization, recent broad-spectrum antibiotic use, malignancy, and presence of shock) and nosocomial factors (for example, admission to a ward with a high incidence of MDR strains)³⁵. Indeed, recent receipt of an anti-pseudomonal antibiotic, especially quinolones and carbapenems, appears to be an important driver of MDR

P. aeruginosa acquisition¹⁰. Compared with less-resistant strains, pneumonia due to MDR *P. aeruginosa* is associated with longer stays in the ICU, prolonged mechanical ventilation, and greater mortality³⁶. Thus, improving the management of MDR *P. aeruginosa* pneumonia must be a priority in order to improve outcomes from both clinical and financial standpoints.

At present, there is no high-grade evidence to guide management decisions for MDR *P. aeruginosa* pneumonia. Current guidelines recommend combination empiric therapy when AMR is a concern and suggest that aminoglycosides and colistin be avoided if alternative agents are available (low-quality evidence)². An anti-pseudomonal cephalosporin, carbapenem, fluoroquinolone, or β -lactam/ β -lactamase inhibitor is a potential option for initial therapy. Once susceptibility results are available, combination therapy can be de-escalated to monotherapy in most cases. However, combination therapy should be continued for patients in septic shock or at a high risk for death². Patients with VAP due to MDR *P. aeruginosa* that is susceptible to only aminoglycosides or polymyxins should receive both inhaled and systemic antibiotics rather than systemic antibiotics alone². In addition to adequate antibiotic coverage, other factors such as adequate drug dosing, appropriate intervals of drug administration, and duration of therapy (usually 7 days) are important for achieving optimal clinical outcomes. For example, a single-center retrospective study found that mortality was significantly lower in patients with *P. aeruginosa* pneumonia who received extended-infusion cefepime versus standard dosing (20% versus 3%, respectively; $p = 0.03$), along with significantly lower length of ICU stay (18.5 versus 8 days, respectively; $p = 0.04$)³⁷.

Several new antibiotics with activity against MDR *P. aeruginosa* have become available recently or are in late stages of development. Ceftolozane/tazobactam is a combination of a novel cephalosporin with a modified side chain and a β -lactamase inhibitor. The potent anti-pseudomonal activity of ceftolozane/tazobactam is attributed to its ability to evade the resistance mechanisms of *P. aeruginosa*, including efflux pumps, reduced uptake through porins, and modification of penicillin-binding proteins (PBPs)³⁸. A higher dose (3 g intravenously [IV] every 8 hours) has been recommended for treating pneumonia compared with the currently approved dose (1.5 g IV every 8 hours) based on pharmacokinetic estimates for achieving a more than 90% probability of target attainment for the 1-log kill target against pathogens with a minimum inhibitory concentration (MIC) of not more than 8 mg/L in epithelial lining fluid³⁹. Of concern is a recent retrospective study from a single center in Germany that reported that 55% of *P. aeruginosa* isolates were resistant to ceftolozane/tazobactam, which the authors suggested may be due to carriage of the VIM-2 carbapenemase⁴⁰. Notably, ceftolozane/tazobactam lacks activity against Ambler class B (metallo-)carbapenemases, such as VIM and NDM.

Ceftazidime/avibactam, a combination of a third-generation cephalosporin and a novel synthetic non- β -lactam, β -lactamase inhibitor, is also ineffective against metallo- β -lactamases. In pooled data from five randomized controlled trials including one on HAP/VAP (REPROVE), 95 cases of MDR *P. aeruginosa* were identified⁴¹. The favorable microbiological response rates at

test of cure for MDR *P. aeruginosa* were 57.1% for ceftazidime/avibactam and 53.8% for comparators, primarily carbapenems⁴¹. Thus, ceftazidime/avibactam likely has a role as a carbapenem-sparing agent for treating MDR *P. aeruginosa* pneumonia. Recent data suggest that ceftazidime/avibactam is a viable option against strains of MDR *P. aeruginosa* that are resistant to ceftolozane/tazobactam⁴².

Cefiderocol is a novel siderophore cephalosporin that inhibits cell wall synthesis through the formation of an iron ion complex that penetrates bacteria via a ferric iron transporter system⁴³. Cefiderocol has demonstrated potent activity against β -lactamase-producing *P. aeruginosa*, including those expressing ESBLs, Ambler class A β -lactamases, and metallo- β -lactamases^{44,45}. Currently, there is a randomized clinical trial under way for the treatment of HAP comparing the combination of cefiderocol and linezolid with linezolid and meropenem (ClinicalTrials.gov Identifier: NCT03032380)⁴⁶. The novel β -lactamase inhibitor relebactam inhibits Ambler class A and C β -lactamases and is in development in combination with imipenem. According to data from a global surveillance program that included Africa, Asia, Europe, Latin America, the Middle East, US/Canada, and the South Pacific, imipenem/relebactam inhibited 90.8% of all *P. aeruginosa* isolates and 70.7% of MDR *P. aeruginosa* isolates ($n = 3,708$)⁴⁷. A study on patients with HAP/VAP is ongoing. Finally, though still in early development, murepavadin is the first member of a novel class of outer membrane protein-targeting antibiotics specifically designed to target *P. aeruginosa*⁴⁸. In a study that included 785 isolates of extremely drug-resistant *P. aeruginosa*, Sader *et al.* reported that murepavadin showed potent activity against isolates that were non-susceptible to colistin, ceftolozane/tazobactam, or tobramycin or a combination of these⁴⁹. These promising findings raise hopes for the further development of murepavadin.

Acinetobacter baumannii

Most cases of *Acinetobacter baumannii* pneumonia occur in hospitalized patients, although it is occasionally seen in CAP⁵⁰. Therefore, the recent report that, after increasing for many years, the rate of AMR in *A. baumannii* hospital-acquired infections may be leveling off is grounds for cautious optimism⁵¹. AMR in *A. baumannii* is the primary reason that clinicians prescribe ineffective empirical antibiotic therapy, often leading to poor outcomes⁵². For example, VAP due to MDR *A. baumannii* results in significantly lower rates of successful ventilator weaning compared with susceptible strains⁵³. A retrospective cohort study that included 175 hospitals found that having pneumonia or sepsis from MDR *A. baumannii* was significantly associated with receiving inappropriate antibiotic therapy and higher hospital mortality⁵⁴. Thus, it is crucial that risk factors for MDR *A. baumannii* be recognized early so that appropriate empiric therapy can be rapidly initiated.

When pneumonia due to MDR *A. baumannii* is suspected (that is, during an *A. baumannii* outbreak, in an endemic setting, or in a previously colonized patient), combination therapy including a polymyxin should be empirically prescribed until susceptibilities are known⁵⁵. If clinical suspicion for resistance is low, then a carbapenem (except ertapenem, which lacks activity against

A. baumannii) should be first-line therapy. Many combination therapies for MDR *A. baumannii* have been investigated and were recently discussed by Vazquez Guillamet and Kollef⁵. Polymyxins remain the backbone of combination regimens. A retrospective cohort study that included patients with pneumonia caused by strains of *A. baumannii* susceptible only to colistin and tigecycline compared three combination regimens: colistin and high-dose sulbactam (n = 93), colistin and tigecycline (n = 43), and colistin and high-dose prolonged infusion of a carbapenem (n = 30)⁵⁶. The 28-day survival rate and mean length of hospital stay were not statistically different between regimens, whereas an elevated Acute Physiologic Assessment and Chronic Health Evaluation (APACHE) score, delay in receipt of an active regimen, underlying malignancy, and chronic kidney disease were all significantly associated with increased mortality⁵⁶. Using a loading dose of IV colistin for MDR *A. baumannii* VAP appears to have no significant effect on clinical cure rates or bacteriologic clearance but does increase the risk for nephrotoxicity⁵⁷. The addition of inhaled colistin to systemic therapies has generally showed favorable results, including better microbiological response. A prospective observational study that compared an IV β -lactam plus IV aminoglycoside, monotherapy

with inhaled colistin, and aerosolized colistin plus IV aminoglycoside found no difference in cure rates⁵⁸. Another study in which IV colistin was compared with IV colistin plus inhaled colistin and inhaled colistin alone also found no differences in mortality or clinical cure, and microbiological cure was better in the aerosolized group⁵⁹. Once susceptibility results of *A. baumannii* isolates become available, combination therapy may be de-escalated to monotherapy. However, tigecycline alone should be avoided, as resistance in *A. baumannii* can develop rapidly⁶⁰.

Although there have been several recent approvals of antibiotics with activity against Gram-negative pathogens (for example, delafloxacin, ceftazidime-avibactam, ceftolozane-tazobactam, and meropenem-vaborbactam), the pipeline is limited for agents effective against MDR *A. baumannii* (Table 1). Cefiderocol has been shown to have potent *in vitro* activity against *A. baumannii* and exhibits high stability against carbapenemase hydrolysis⁶¹. A 52-country collection of clinical isolates obtained between 2014 and 2016 found that 330/368 (89%) of MDR *A. baumannii* strains had cefiderocol MICs of not more than 4 $\mu\text{g}/\text{mL}$ ⁶². Another study evaluated 107 carbapenem-resistant *A. baumannii* isolates from 18 Greek hospitals and determined that the MIC₉₀ of

Table 1. New antibiotics for multidrug-resistant Gram-negative bacteria.

Drug	Class	Development stage	Activity	FDA indication
Aztreonam/avibactam	Monobactam/ β -lactamase inhibitor	Phase II	ESBL, KPC, class C β -lactamase, MBL	Not applicable
Cefiderocol	Siderophore cephalosporin	Phase III	ESBL, CRE (class A, B, and D enzymes), carbapenem-resistant <i>Pseudomonas aeruginosa</i> , <i>Stenotrophomonas maltophilia</i> , and <i>Acinetobacter baumannii</i>	Not applicable
Ceftazidime/avibactam	Cephalosporin/ β -lactamase inhibitor	FDA-approved	ESBL, KPC, AmpC, some class D serine β -lactamases	HABP/VABP, cIAI, cUTI
Ceftolozane/tazobactam	Cephalosporin/ β -lactamase inhibitor	FDA-approved	ESBL, MDR <i>P. aeruginosa</i>	cUTI, cIAI
Delafloxacin	Fluoroquinolone	FDA-approved	<i>Klebsiella pneumoniae</i> , including AmpC and class A ESBL-producers, ciprofloxacin-resistant <i>Escherichia coli</i> and <i>A. baumannii</i>	ABSSSI
Eravacycline	Fluorocycline tetracycline	FDA-approved	ESBL, CRE, MDR <i>A. baumannii</i>	cIAI
Imipenem+cilastatin/relebactam	Carbapenem/ β -lactamase inhibitor	Phase III	KPC, MDR <i>P. aeruginosa</i>	Not applicable
Meropenem/vaborbactam	Carbapenem/boronic acid inhibitor	FDA-approved	CRE (class A and C enzymes)	cUTI
Murepavadin	Cyclic peptide that targets outer membrane	Phase III	MDR <i>P. aeruginosa</i>	Not applicable
Omadacycline	Aminomethylcycline	FDA-approved	ESBL, <i>A. baumannii</i>	ABSSSI, CABP
Plazomicin	Aminoglycoside	FDA-approved	ESBL, CRE excluding NDM producers, <i>A. baumannii</i> , <i>P. aeruginosa</i>	cUTI

ABSSSI, acute bacterial skin and skin structure infection; CABP, community-acquired bacterial pneumonia; cIAI, complicated intra-abdominal infection; CRE, carbapenem-resistant Enterobacteriaceae; cUTI, complicated urinary tract infection; ESBL, extended-spectrum β -lactamase; FDA, US Food and Drug Administration; HABP, hospital-acquired bacterial pneumonia; KPC, *Klebsiella pneumoniae* carbapenemase; MBL, metallo- β -lactamase; MDR, multidrug-resistant; NDM, New Delhi metallo- β -lactamase; VABP, ventilator-acquired bacterial pneumonia.

cefiderocol was 0.5 mg/L, which was more active than either tigecycline or colistin⁶³. Two phase III clinical trials for cefiderocol—APEKS-NP and CREDIBLE-CR—that include VAP and HAP due to GNB are under way⁶⁴.

Eravacycline was recently approved for the treatment of intra-abdominal infections and complicated urinary tract infections (cUTIs). One study showed that against carbapenem- and tigecycline-resistant *Acinetobacter* isolates, eravacycline MICs were about twofold lower versus comparator agents⁶⁵. However, clinical experience with eravacycline for *A. baumannii* pneumonia is limited and its role is unclear, especially given evidence of increased adverse events and mortality with tigecycline when prescribed for pneumonia⁶⁶.

Plazomicin is a novel aminoglycoside derived from sisomicin that was modified to resist aminoglycoside-modifying enzymes and is currently approved for use in cUTIs. Significantly improved activity has been observed in OXA-producing *A. baumannii* compared with other aminoglycosides, and plazomicin MICs are 16 to 32 times lower⁶⁷. A recent clinical trial found favorable results with plazomicin in CRE VAP as well as a favorable safety profile and a low incidence of drug-related adverse events, including serum creatinine elevations⁶⁸. Thus, plazomicin appears promising as part of combination regimens and data on its effectiveness in HAP/VAP due to MDR *A. baumannii* are eagerly awaited.

Zidebactam and WCK 5153 are two novel β -lactam antibiotics under development that display high affinity for PBP2 of GNB and overcome multiple β -lactam resistance mechanisms.

When combined with cefepime or sulbactam, zidebactam and WCK 5153 demonstrated enhanced killing and full bacterial eradication after 24 hours against strains of MDR *A. baumannii*⁶⁹. Arylomycins are a new class of lipopeptide antibiotics whose lead compound, G0775, was demonstrated to have potent activity against 16 strains of MDR *A. baumannii*⁷⁰. The antibiotic adjuvant SPR741 sensitized *A. baumannii* to a panel of antibiotics and permitted strong potentiation of rifampin against multiple MDR *A. baumannii* isolates⁷¹. Several non-antibiotic therapies against *A. baumannii* are in various stages of development (for example, bacteriophage, vaccines, and monoclonal antibodies) and have recently been reviewed⁷². Finally, it may be possible in the future to use CRISPR-Cas systems to target plasmids that spread AMR in GNB⁷³.

Conclusions

Pneumonia due to MDR-GNB represents a serious threat to hospitalized patients. Clinicians must be knowledgeable about local resistance patterns and a patient's risk factors for MDR-GNB to ensure appropriate empiric antimicrobial therapy. Fortunately, several new drugs that target MDR-GNB have been approved or are in late stages of development. Further pragmatic studies are needed to elucidate their place in therapy and their impact on real-world outcomes such as length of stay and mortality, especially for ICU patients with HAP/VAP.

Grant information

The author(s) declared that no grants were involved in supporting this work.

References

- Thorpe KE, Joski P, Johnston KJ: **Antibiotic-Resistant Infection Treatment Costs Have Doubled Since 2002, Now Exceeding \$2 Billion Annually.** *Health Aff (Millwood)*. 2018; **37**(4): 662–9. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Kalil AC, Metersky ML, Klompas M, *et al.*: **Management of Adults With Hospital-acquired and Ventilator-associated Pneumonia: 2016 Clinical Practice Guidelines by the Infectious Diseases Society of America and the American Thoracic Society.** *Clin Infect Dis*. 2016; **63**(5): e61–e111. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Bassetti M, Welte T, Wunderink RG: **Treatment of Gram-negative pneumonia in the critical care setting: is the beta-lactam antibiotic backbone broken beyond repair?** *Crit Care*. 2016; **20**: 19. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Watkins RR, Deresinski S: **Using β -lactam/ β -lactamase inhibitors for infections due to extended-spectrum β -lactamase-producing Enterobacteriaceae to slow the emergence of carbapenem-resistant Enterobacteriaceae.** *Expert Rev Anti Infect Ther*. 2017; **15**(10): 893–5. [PubMed Abstract](#) | [Publisher Full Text](#)
- Grundmann H, Glasner C, Albigler B, *et al.*: **Occurrence of carbapenemase-producing *Klebsiella pneumoniae* and *Escherichia coli* in the European survey of carbapenemase-producing Enterobacteriaceae (EuSCAPE): a prospective, multinational study.** *Lancet Infect Dis*. 2017; **17**(2): 153–63. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Vazquez Guillamet C, Kollef MH: **Acinetobacter Pneumonia: Improving Outcomes With Early Identification and Appropriate Therapy.** *Clin Infect Dis*. 2018; **67**(9): 1455–62. [PubMed Abstract](#) | [Publisher Full Text](#)
- Davey PG, Marwick C: **Appropriate vs. inappropriate antimicrobial therapy.** *Clin Microbiol Infect*. 2008; **14**(Suppl 3): 15–21. [PubMed Abstract](#) | [Publisher Full Text](#)
- Maruyama T, Fujisawa T, Ishida T, *et al.*: **A Therapeutic Strategy for All Pneumonia Patients: A 3-Year Prospective Multicenter Cohort Study Using Risk Factors for Multidrug Resistant Pathogens To Select Initial Empiric Therapy.** *Clin Infect Dis*. 2018. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Lat I, Daley MJ, Shewale A, *et al.*: **A Multicenter, Prospective, Observational Study to Determine Predictive Factors for Multidrug-Resistant Pneumonia in Critically Ill Adults: The DEFINE Study.** *Pharmacotherapy*. 2018. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Montero M, Sala M, Riu M, *et al.*: **Risk factors for multidrug-resistant *Pseudomonas aeruginosa* acquisition. Impact of antibiotic use in a double case-control study.** *Eur J Clin Microbiol Infect Dis*. 2010; **29**(3): 335–9. [PubMed Abstract](#) | [Publisher Full Text](#)
- Gu WJ, Wang F, Tang L, *et al.*: **Colistin for the treatment of ventilator-associated pneumonia caused by multidrug-resistant Gram-negative bacteria: a systematic review and meta-analysis.** *Int J Antimicrob Agents*. 2014; **44**(6): 477–85. [PubMed Abstract](#) | [Publisher Full Text](#)
- Sader HS, Castanheira M, Mendes RE, *et al.*: **Frequency and antimicrobial susceptibility of Gram-negative bacteria isolated from patients with pneumonia hospitalized in ICUs of US medical centres (2015–17).** *J Antimicrob Chemother*. 2018; **73**(11): 3053–9. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Jacoby GA: **AmpC beta-lactamases.** *Clin Microbiol Rev*. 2009; **22**(1): 161–82. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)



14. Hauck C, Cober E, Richter SS, *et al.*: Spectrum of excess mortality due to carbapenem-resistant *Klebsiella pneumoniae* infections. *Clin Microbiol Infect.* 2016; **22**(6): 513–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Safdar N, Maki DG: The commonality of risk factors for nosocomial colonization and infection with antimicrobial-resistant *Staphylococcus aureus*, *enterococcus*, gram-negative bacilli, *Clostridium difficile*, and *Candida*. *Ann Intern Med.* 2002; **136**(11): 834–44.
[PubMed Abstract](#) | [Publisher Full Text](#)
16. Bassetti M, Camelutti A, Peghin M: Patient specific risk stratification for antimicrobial resistance and possible treatment strategies in gram-negative bacterial infections. *Expert Rev Anti Infect Ther.* 2017; **15**(1): 55–65.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
17. Rodríguez-Baño J, Gutiérrez-Gutiérrez B, Machuca I, *et al.*: Treatment of Infections Caused by Extended-Spectrum-Beta-Lactamase-, AmpC-, and Carbapenemase-Producing *Enterobacteriaceae*. *Clin Microbiol Rev.* 2018; **31**(2): pii: e00079-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
18. Harris PNA, Tambyah PA, Lye DC, *et al.*: Effect of Piperacillin-Tazobactam vs Meropenem on 30-Day Mortality for Patients With *E coli* or *Klebsiella pneumoniae* Bloodstream Infection and Ceftriaxone Resistance: A Randomized Clinical Trial *JAMA.* 2018; **320**(10): 984–994.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
19. Tamma PD, Villegas MV: Use of β -Lactam/ β -Lactamase Inhibitors for Extended-Spectrum- β -Lactamase Infections: Defining the Right Patient Population. *Antimicrob Agents Chemother.* 2017; **61**(8): pii: e01094-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
20. Zanetti G, Bally F, Greub G, *et al.*: Cefepime versus imipenem-cilastatin for treatment of nosocomial pneumonia in intensive care unit patients: a multicenter, evaluator-blind, prospective, randomized study. *Antimicrob Agents Chemother.* 2003; **47**(11): 3442–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Cheng L, Nelson BC, Mehta M, *et al.*: Piperacillin-Tazobactam versus Other Antibacterial Agents for Treatment of Bloodstream Infections Due to AmpC β -Lactamase-Producing *Enterobacteriaceae*. *Antimicrob Agents Chemother.* 2017; **61**(1): pii: e00276-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
22. Tamma PD, Girdwood SC, Gopaul R, *et al.*: The use of cefepime for treating AmpC β -lactamase-producing *Enterobacteriaceae*. *Clin Infect Dis.* 2013; **57**(6): 781–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
23. van Duin D, Doi Y: The global epidemiology of carbapenemase-producing *Enterobacteriaceae*. *Virulence.* 2017; **8**(4): 460–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
24. van Duin D, Kaye KS, Neuner EA, *et al.*: Carbapenem-resistant *Enterobacteriaceae*: a review of treatment and outcomes. *Diagn Microbiol Infect Dis.* 2013; **75**(2): 115–20.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. van Duin D, Bonomo RA: Ceftazidime/Avibactam and Ceftolozane/Tazobactam: Second-generation β -Lactam/ β -Lactamase Inhibitor Combinations. *Clin Infect Dis.* 2016; **63**(2): 234–41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
26. Kaye KS, Vazquez J, Mathers A, *et al.*: Clinical Outcomes of Serious Infections due to Carbapenem-Resistant *Enterobacteriaceae* (CRE) in TANGO II, a Phase 3, Randomized, Multi-National, Open-Label Trial of Meropenem-Vaborbactam (M-V) Vs. Best Available Therapy (BAT). *Open Forum Infect Dis.* 2017; **4**(suppl_1): S534–S535.
[Publisher Full Text](#)
27. Karaïskos I, Souli M, Giamarellou H: Plazomicin: an investigational therapy for the treatment of urinary tract infections. *Expert Opin Investig Drugs.* 2015; **24**(11): 1501–11.
[PubMed Abstract](#) | [Publisher Full Text](#)
28. Zhang Y, Kashikar A, Bush K: *In vitro* activity of plazomicin against β -lactamase-producing carbapenem-resistant *Enterobacteriaceae* (CRE). *J Antimicrob Chemother.* 2017; **72**(10): 2792–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
29. Zhanell GG, Lawrence CK, Adam H, *et al.*: Imipenem-Relebactam and Meropenem-Vaborbactam: Two Novel Carbapenem- β -Lactamase Inhibitor Combinations. *Drugs.* 2018; **78**(1): 65–98.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
30. Thakare R, Dasgupta A, Chopra S: Eravacycline for the treatment of patients with bacterial infections. *Drugs Today (Barc).* 2018; **54**(4): 245–54.
[PubMed Abstract](#) | [Publisher Full Text](#)
31. van Duin D, Lok JJ, Earley M, *et al.*: Colistin Versus Ceftazidime-Avibactam in the Treatment of Infections Due to Carbapenem-Resistant *Enterobacteriaceae*. *Clin Infect Dis.* 2018; **66**(2): 163–71.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
32. Achaogen: Achaogen Announces Positive Results in Phase 3 cUTI and CRE Clinical Trials of Plazomicin. Press release. 2016.
[Reference Source](#)
33. Torres A, Zhong N, Pacht J, *et al.*: Ceftazidime-avibactam versus meropenem in nosocomial pneumonia, including ventilator-associated pneumonia (REPROVE): a randomised, double-blind, phase 3 non-inferiority trial. *Lancet Infect Dis.* 2018; **18**(3): 285–95.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
34. Mehta M, Uhlemann AC: Beware of broad-spectrum generalizations: ceftazidime-avibactam compared to meropenem for the treatment of gram-negative pneumonia. *J Emerg Crit Care Med.* 2018; **7**: pii: 45.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
35. Bassetti M, Vena A, Croxatto A, *et al.*: How to manage *Pseudomonas aeruginosa* infections. *Drugs Context.* 2018; **7**: 212527.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
36. Micek ST, Wunderink RG, Kollef MH, *et al.*: An international multicenter retrospective study of *Pseudomonas aeruginosa* nosocomial pneumonia: impact of multidrug resistance. *Crit Care.* 2015; **19**(1): 219.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
37. Bauer KA, West JE, O'Brien JM, *et al.*: Extended-infusion cefepime reduces mortality in patients with *Pseudomonas aeruginosa* infections. *Antimicrob Agents Chemother.* 2013; **57**(7): 2907–12.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
38. Zhanell GG, Chung P, Adam H, *et al.*: Ceftolozane/tazobactam: a novel cephalosporin/ β -lactamase inhibitor combination with activity against multidrug-resistant gram-negative bacilli. *Drugs.* 2014; **74**(1): 31–51.
[PubMed Abstract](#) | [Publisher Full Text](#)
39. Xiao AJ, Miller BW, Huntington JA, *et al.*: Ceftolozane/tazobactam pharmacokinetic/pharmacodynamic-derived dose justification for phase 3 studies in patients with nosocomial pneumonia. *J Clin Pharmacol.* 2016; **56**(1): 56–66.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
40. Katchanov J, Asar L, Klupp EM, *et al.*: Carbapenem-resistant Gram-negative pathogens in a German university medical center: Prevalence, clinical implications and the role of novel β -lactam/ β -lactamase inhibitor combinations. *PLoS One.* 2018; **13**(4): e0195757.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
41. Stone GG, Newell P, Gasink LB, *et al.*: Clinical activity of ceftazidime/avibactam against MDR *Enterobacteriaceae* and *Pseudomonas aeruginosa*: pooled data from the ceftazidime/avibactam Phase III clinical trial programme. *J Antimicrob Chemother.* 2018; **73**(9): 2519–23.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
42. Rodríguez-Núñez O, Ripa M, Morata L, *et al.*: Evaluation of ceftazidime/avibactam for serious infections due to multidrug-resistant and extensively drug-resistant *Pseudomonas aeruginosa*. *J Glob Antimicrob Resist.* 2018; **15**: 136–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
43. Möllmann U, Heinisch L, Bauernfeind A, *et al.*: Siderophores as drug delivery agents: application of the “Trojan Horse” strategy. *Biomaterials.* 2009; **22**(4): 615–24.
[PubMed Abstract](#) | [Publisher Full Text](#)
44. Dobias J, Dénervaud-Tendon V, Poirer L, *et al.*: Activity of the novel siderophore cephalosporin cefiderocol against multidrug-resistant Gram-negative pathogens. *Eur J Clin Microbiol Infect Dis.* 2017; **36**(12): 2319–27.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
45. Ito A, Kohira N, Bouchillon SK, *et al.*: *In vitro* antimicrobial activity of S-649266, a catechol-substituted siderophore cephalosporin, when tested against non-fermenting Gram-negative bacteria. *J Antimicrob Chemother.* 2016; **71**(3): 670–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
46. Nguyen L, Garcia J, Gruenberg K, *et al.*: Multidrug-Resistant *Pseudomonas* Infections: Hard to Treat, But Hope on the Horizon? *Curr Infect Dis Rep.* 2018; **20**(8): 23.
[PubMed Abstract](#) | [Publisher Full Text](#)
47. Karlowsky JA, Lob SH, Young K, *et al.*: Activity of imipenem/relebactam against *Pseudomonas aeruginosa* with antimicrobial-resistant phenotypes from seven global regions: SMART 2015–2016. *J Glob Antimicrob Resist.* 2018; **15**: 140–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
48. Martin-Loeches I, Dale GE, Torres A: Murepavadin: a new antibiotic class in the pipeline. *Expert Rev Anti Infect Ther.* 2018; **16**(4): 259–68.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
49. Sader HS, Flamm RK, Dale GE, *et al.*: Murepavadin activity tested against contemporary (2016–17) clinical isolates of XDR *Pseudomonas aeruginosa*. *J Antimicrob Chemother.* 2018; **73**(9): 2400–4.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
50. Serota DP, Sexton ME, Kraft CS, *et al.*: Severe Community-Acquired Pneumonia due to *Acinetobacter baumannii* in North America: Case Report and Review of the Literature. *Open Forum Infect Dis.* 2018; **5**(3): ofy044.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
51. Cohen B, Liu J, Larson E: Changes in the incidence and antimicrobial susceptibility of healthcare-associated infections in a New York hospital system, 2006–2012. *J Prev Med Hyg.* 2017; **58**(4): E294–E301.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
52. Guillaumet CV, Vazquez R, Noe J, *et al.*: A cohort study of bacteremic pneumonia: The importance of antibiotic resistance and appropriate initial

- therapy? *Medicine (Baltimore)*. 2016; **95**(35): e4708.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
53. **F** Bickenbach J, Schöneis D, Marx G, *et al.*: **Impact of multidrug-resistant bacteria on outcome in patients with prolonged weaning.** *BMC Pulm Med*. 2018; **18**(1): 141.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 54. Zilberberg MD, Nathanson BH, Sulham K, *et al.*: **Multidrug resistance, inappropriate empiric therapy, and hospital mortality in *Acinetobacter baumannii* pneumonia and sepsis.** *Crit Care*. 2016; **20**(1): 221.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 55. Garnacho-Montero J, Dimopoulos G, Poulakou G, *et al.*: **Task force on management and prevention of *Acinetobacter baumannii* infections in the ICU.** *Intensive Care Med*. 2015; **41**(12): 2057–75.
[PubMed Abstract](#) | [Publisher Full Text](#)
 56. Khawcharoenporn T, Pruetpongpun N, Tiamsak P, *et al.*: **Colistin-based treatment for extensively drug-resistant *Acinetobacter baumannii* pneumonia.** *Int J Antimicrob Agents*. 2014; **43**(4): 378–82.
[PubMed Abstract](#) | [Publisher Full Text](#)
 57. **F** Alp E, Eren E, Elay G, *et al.*: **Efficacy of loading dose of colistin in *Acinetobacter baumannii* ventilator-associated pneumonia.** *Infez Med*. 2017; **25**(4): 311–319.
[PubMed Abstract](#) | [F1000 Recommendation](#)
 58. Lu Q, Luo R, Bodin L, *et al.*: **Efficacy of High-dose Nebulized Colistin in Ventilator-associated Pneumonia Caused by Multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii*.** *Anesthesiology*. 2012; **117**(6): 1335–47.
[PubMed Abstract](#) | [Publisher Full Text](#)
 59. Pérez-Pedrero MJ, Sánchez-Casado M, Rodríguez-Villar S: **Utilización de la colistina nebulizada en la colonización e infección respiratoria por *Acinetobacter baumannii* en pacientes críticos.** *Medicina Intensiva*. 2011; **35**(4): 226–31.
[Reference Source](#)
 60. Li X, Liu L, Ji J, *et al.*: **Tigecycline resistance in *Acinetobacter baumannii* mediated by frameshift mutation in plnC, encoding 1-acyl-sn-glycerol-3-phosphate acyltransferase.** *Eur J Clin Microbiol Infect Dis*. 2015; **34**(3): 625–31.
[PubMed Abstract](#) | [Publisher Full Text](#)
 61. **F** Wright H, Bonomo RA, Paterson DL: **New agents for the treatment of infections with Gram-negative bacteria: Restoring the miracle or false dawn?** *Clin Microbiol Infect*. 2017; **23**(10): 704–712.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 62. **F** Hackel MA, Tsuji M, Yamano Y, *et al.*: **In Vitro Activity of the Siderophore Cephalosporin, Cefiderocol, against Carbapenem-Nonsusceptible and Multidrug-Resistant Isolates of Gram-Negative Bacilli Collected Worldwide in 2014 to 2016.** *Antimicrob Agents Chemother*. 2018; **62**(2): pii: e01968-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 63. **F** Falagas ME, Skolidis T, Vardakas KZ, *et al.*: **Activity of cefiderocol (S-649266) against carbapenem-resistant Gram-negative bacteria collected from inpatients in Greek hospitals.** *J Antimicrob Chemother*. 2017; **72**(6): 1704–1708.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 64. Bassetti M, Vena A, Castaldo N, *et al.*: **New antibiotics for ventilator-associated pneumonia.** *Curr Opin Infect Dis*. 2018; **31**(2): 177–186.
[PubMed Abstract](#) | [Publisher Full Text](#)
 65. Livermore DM, Mushtaq S, Warner M, *et al.*: **In Vitro Activity of Eravacycline against Carbapenem-Resistant Enterobacteriaceae and *Acinetobacter baumannii*.** *Antimicrob Agents Chemother*. 2016; **60**(6): 3840–4.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 66. Shen F, Han Q, Di Xie, *et al.*: **Efficacy and safety of tigecycline for the treatment of severe infectious diseases: an updated meta-analysis of RCTs.** *Int J Infect Dis*. 2015; **39**: 25–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
 67. García-Salguero C, Rodríguez-Avial I, Picazo JJ, *et al.*: **Can Plazomicin Alone or in Combination Be a Therapeutic Option against Carbapenem-Resistant *Acinetobacter baumannii*?** *Antimicrob Agents Chemother*. 2015; **59**(10): 5959–66.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 68. Connolly LE, Jubbs AM, Keeffe BO, *et al.*: **Plazomicin is associated with improved survival and safety compared to colistin in serious carbapenem-resistant Enterobacteriaceae (CRE) infections: results of the CARE study.** Presented at: ASM Microbe; 2017; New Orleans, LA.
[Reference Source](#)
 69. **F** Moya B, Barcelo IM, Bhagwat S, *et al.*: **Potent β -Lactam Enhancer Activity of Zidebactam and WCK 5153 against *Acinetobacter baumannii*, Including Carbapenemase-Producing Clinical Isolates.** *Antimicrob Agents Chemother*. 2017; **61**(11): pii: e01238-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 70. **F** Smith PA, Koehler MFT, Gírgis HS, *et al.*: **Optimized arylomycins are a new class of Gram-negative antibiotics.** *Nature*. 2018; **561**(7722): 189–94.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 71. **F** Domalaon R, Idowu T, Zhanel GG, *et al.*: **Antibiotic Hybrids: the Next Generation of Agents and Adjuvants against Gram-Negative Pathogens?** *Clin Microbiol Rev*. 2018; **31**(2): pii: e00077-17.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 72. Watkins RR: **A formidable foe: carbapenem-resistant *Acinetobacter baumannii* and emerging nonantibiotic therapies.** *Expert Rev Anti Infect Ther*. 2018; **16**(8): 591–3.
[PubMed Abstract](#) | [Publisher Full Text](#)
 73. Bober JR, Beisel CL, Nair NU: **Synthetic Biology Approaches to Engineer Probiotics and Members of the Human Microbiota for Biomedical Applications.** *Annu Rev Biomed Eng*. 2018; **20**: 277–300.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Open Peer Review

Current Referee Status:  

Editorial Note on the Review Process

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty and are edited as a service to readers. In order to make these reviews as comprehensive and accessible as possible, the referees provide input before publication and only the final, revised version is published. The referees who approved the final version are listed with their names and affiliations but without their reports on earlier versions (any comments will already have been addressed in the published version).

The referees who approved this article are:

Version 1

- 1 **Dean Winslow** Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California, USA
Competing Interests: No competing interests were disclosed.
- 2 **Frank Schweizer** Department of Medical Microbiology and Infectious Diseases, University of Manitoba, Winnipeg, Manitoba, Canada
Competing Interests: No competing interests were disclosed.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research