

Origin and Therapies of Osteosarcoma

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Simple Summary: Osteosarcoma is the most common malignant bone tumor in children, with a 5-year survival rate ranging from 70% to 20% depending on the aggressiveness of the disease. The current treatments have not evolved over the past four decades due in part to the genetic complexity of the disease and its heterogeneity. This review will summarize the current knowledge of OS origin, diagnosis and therapies.

Abstract: Osteosarcoma (OS) is the most frequent primary bone tumor, mainly affecting children and young adults. Despite therapeutic advances, the 5-year survival rate is 70% but drastically decreases to 20–30% for poor responders to therapies or for patients with metastasis. No real evolution of the survival rates has been observed for four decades, explained by poor knowledge of the origin, difficulties related to diagnosis and the lack of targeted therapies for this pediatric tumor. This review will describe a non-exhaustive overview of osteosarcoma disease from a clinical and biological point of view, describing the origin, diagnosis and therapies.

Keywords: osteosarcoma; origin; therapy

1. Introduction

Osteosarcoma (OS) is the most common primary malignant bone tumor. It is also the third most common type of cancer affecting children and adolescents after lymphomas and brain tumors [1]. OS cells derive from the mesenchymal lineage and are able to produce osteoid substances and/or immature bone [2,3].

OS mainly affects children or young adults. The incidence of OS increases from two to three million per year in the general population to eight to eleven million per year when only the 15–19-year-old age group is considered. OS represents 3–6% of childhood cancers (15% of extra-cranial solid cancers) and 1% of adult cancers. After the peak of incidence at 15–19 years of age, another peak of incidence occurs in elderly populations (over 65 years of age) and occurs as a result of pre-existing bone pathology or fractures. Young males are 1.4 times more likely to develop the disease than young females. However, the risks are equivalent between the sexes in people older than 65 years [4,5].

The causes of OS are still poorly understood. Its appearance in young populations during growth and its location at the ends of the long bones suggest the involvement of rapid bone production [6]. This hypothesis is supported by a higher incidence of OS in large dogs compared to small dogs [7]. In older patients, risk factors include radiation and chemotherapy for treatment of pre-existing malignancies. A history of Paget's disease is also present in approximately one third of adult OS cases. OS can also arise from hereditary genetic disorders, such as Li–Fraumeni syndrome, Rothmund–Thomson syndrome, or Werner syndrome [8,9].



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2. Clinical Features

OS is often diagnosed after persistent local pains, often mistakenly attributed to growth in young people, or to physical activity. Patients may also exhibit a palpable mass and reduced joint range of motion. Systemic symptoms (weight loss, fatigue, fever...) may also be present. The tumor will most frequently be located in the metaphysis of long bones (femur, tibia, humerus), with 50% of cases located near the knee [10–12]. The tumor can also be localized to the axial skeleton, most commonly the pelvis, in adults [12]. Approximately 15–20% of patients develop metastases at the time of diagnosis, particularly in the lungs (85%) or bone (8–10%) [13]. In rare cases, adenopathy may be indicative of metastasis to lymph nodes [11,13]. Presence of metastases is a clear sign of poor prognosis and drastically decreases the survival rate by 70% to 20% at 5 years [1].

3. Diagnosis

Diagnosis is made after palpation to determine the presence of a soft mass near the bone. Radiographically, calcifications resulting from ectopic bone formation may be seen within the soft mass, as well as a "sunburst" appearance. Ectopic bone formation is often associated with osteolysis [14]. Increased lactate dehydrogenase or alkaline phosphatase in the blood is associated with a poor prognosis [2,15]. Finally, a PET (positron emission tomography) scan can detect any potential metastases.

Because biopsy can identify OS with an accuracy level of 90%, histological analysis is often performed to confirm the diagnosis. It allows to determine the level of proliferation of the tumor cells, to classify the tumor according to the stage of severity.

4. Classification of OS

OS is classified into secondary or primary OS. Secondary OS occurs after pre-existing events, such as diseases (Paget's disease, for example) or irradiation [16,17]. Conversely, primary OS is primitive OS and is subdivided into different categories based on histological appearance. The conventional intramedullary/central high-grade type is the most common. It is subdivided into osteoblastic (50%), chondroblastic (25%) or fibroblastic (25%) types differentiated by the secreted matrix. The osteoblastic type is characterized by the secretion of a bone matrix, the chondroblastic type secretes a cartilage matrix, and the fibroblastic type is characterized by collagen-secreting spindle cells. The other types of primary OS are composed of telangiectatic OS that have blood-filled cysts and multinucleated giant cells. Parosteal OS show cartilage production in about 50% of cases, but also bone production and cells with fibroblastic morphology. Central low-grade OS present cells with fibroblast-like morphology and are organized in bundles. Finally, periosteal OS show mature dense bone but also atypical hyaline cartilage [17,18].

There are other subtypes of OS, including sclerosing osteoblastic, chondromyxoid fibroma-like, chondroblastoma-like, clear cell, giant cell, epithelioid, osteoblastoma-like or malignant fibrous histiocytoma-like OS. However, these types of OS are considered subtypes of the conventional type due to their similarities and behaviors to the conventional type [17].

The grade, used to characterize OS, takes into account the severity of the disease and the presence of metastases. Grade of OS helps to adjust treatment, estimate patient prognosis, assess treatment outcomes and facilitate communication between professionals [19,20]. Several disease classification systems exist:

The Musculoskeletal Tumor Society (MSTS) or Enneking System [19,21]:

Three main pieces of information are given by this system: grade, extension of tumor and presence or absence of metastasis (Table 1).

Grade (G): indicates the potential of the tumor to grow and spread. It is based on the histological appearance of the tumors. The G1 grade refers to low-grade tumors (morphology similar to healthy cells). They are less likely to grow quickly and metastasize. The G2 grade refers to high-grade tumors that are abnormal in morphology, rapidly dividing and likely to metastasize.

- Extent of the primary tumor (T): T1 refers to a tumor that is confined to the bone (intra-compartmental), and T2 refers to a tumor that affects surrounding structures (extra-compartmental).
- Metastasis (M): M1 indicates that a tumor has spread to nearby lymph nodes. M0 indicates no spread to lymph nodes.

Table 1. MSTS (Musculoskeletal Tumor Society) staging system for osteosarcoma. Each stage is characterized by a grade (low grade = G1; high grade = G2), the tumor extension (T1 = intra-compartmental; T2 = extra-compartmental) and presence of metastasis (M0 = no metastasis; M1 = presence of metastasis).

Stage	Grade	Tumor	Metastasis
IA	G1	T1	M0
IB	G1	Τ2	M0
IIA	G2	T1	M0
IIB	G2	Τ2	M0
III	G1 or G2	T1 or T2	M1

- The TNM (Size, Spread, Metastasis) System [19,22]:

The TNM classification system was created by the AJCC (American Joint Committee on Cancer): the T parameter describes the size of the primary tumor as well as its presence or absence in different regions of the bone; the N parameter describes the spread to nearby lymph nodes; the M parameter indicates whether the cancer has metastasized to other organs in the body (lung, breast, bone, etc.).

The AJCC classification contains 7 stages (IA, IB, IIA, IIB, III, IVA, IVB), which are subdivided into about 30 stages, each of which accurately describes the tumor. A simplified version of this classification is shown in Table 2.

Table 2. AJCC (American Joint Committee on Cancer) staging system for osteosarcoma. Each stage is characterized by a grade (low, high), the primary tumor size and presence of regional lymph nodes (N0 = no regional lymph nodes metastasis; N1 = regional lymph nodes metastasis) and distant metastasis (M0 = no distant metastasis; M1a = lung metastasis; any M = lung and other distant sites). Adapted from Ritter et Bielack [2].

Stage	Grade	Primary Tumor Size	Lymph Nodes Metastasis	Distant Metastasis
IA	Low	<8 cm	N0	M0
IB	Low	>8 cm	N0	M0
IIA	High	<8 cm	N0	M0
IIB	High	>8 cm	N0	M0
III	Any grade	Any size	N0	M0
IVA	Any grade	Any size	N0	M1a
IVB	Any grade	Any size	N1	Any M

5. Current Therapies for OS

Patients with OS are treated with a multidisciplinary approach established by Rosen et al. in the 1970s [23]. This protocol combines chemotherapy (neoadjuvant and adjuvant, for a duration of 6 to 8 months) and surgery with limb preservation. This strategy results in 5-year survival for approximately 70% of patients [10,24,25]. Radiotherapy is rarely used for the treatment of OS because of the radioresistance of the tumor [26]. Its use is reserved for specific situations, such as the impossibility of complete resection of a tumor located in a high-risk region (head, spine, etc.), or the persistence of small tumor foci after resection [26,27].

Furthermore, radiotherapy could be used to delay tumor growth and to reduce symptoms, such as pain, in the case that surgery is not possible, and could lead to possible side effects (skin reaction, nausea, diarrhea, slow bone growth in children, lung and heart function ...). The used radioactive drugs are radium-233 or samarium153-EDTMP (ethylenediaminetetramethylene phosphonic acid), which have high affinity for bone tissue and, thus, selectively deliver radiation to osteoblastic lesions [28,29]. Samarium153-EDTMP was approved by the FDA in 1998 for palliative treatment to control pain in patients with bone metastases and then was used alone or as a combined agent in high-risk OS patients. Indeed, a high dose of samarium153-EDTMP administered followed by hematopoietic stem cell rescue was tolerable, with hematological toxicity, and controlled the pain [29–31].

5.1. Chemotherapy

Chemotherapy is used before surgery to reduce the size of the tumor and create the best conditions for limb salvage surgery and after surgery to eliminate residual lesions and metastases [32]. The duration of chemotherapy is usually between 6 and 12 months and combines several agents with high efficacy, including doxorubicin (Adriamycin, ADM), cisplatin (DDP), ifosfamide (IFO), etoposide (to a lesser extent) or high-dose methotrex-ate (HDMTX), which are used in varying combinations [33–39]. Indeed, adjuvant MAP (Methotrexate, Adriamycin and Cisplatine combination) chemotherapy is the most common treatment for patients with resectable tumors, generally with two to six cycles of preoperative chemotherapy, followed by additional cycles for postsurgical adjuvant chemotherapy [40,41]. Preoperative chemotherapy is given 8–10 weeks prior to surgery [1,42]. Postoperative chemotherapy is given up to 21 days after surgery for 12 to 29 weeks of treatment [1,43]. Due to the adverse effects of multidrug therapy (cardiac, atrial dysfunction, renal and liver toxicity) [44–46], follow-up examinations should be implemented during and after therapy (echocardiography, audiogram, toxicity tests) [2]. Additional treatments can limit the side effects of chemotherapy (e.g., antiemetics or opiates) [47].

5.2. Surgery

Surgery for OS aims for the complete removal of the tumor, with a focus on large resection margins that include the tumor and the surrounding healthy tissue. However, the local recurrence rate could be up to 25% if the tumor removal is not complete [48]. There is no consensus on the definition and comparison of these resection margins between surgical teams, making it difficult to standardize practices [49,50]. Nowadays, the effectiveness of preoperative chemotherapy prevents amputation of affected limbs [51,52], which conserves physical patient integrity and motricity [3]. Nevertheless, in some cases, amputation should sometimes be considered as a superior option to limb salvage [53]. However, resection of OS involving the axial skeleton remains very difficult, and MRI (magnetic resonance imaging) of the skeleton can be used to plan surgery. If pulmonary metastases are suspected, a thoracotomy can be performed to localize them. Moreover, tumor resection can lead to soft tissue and bone defects that need to be reconstructed. Advances in bone tissue engineering allow to reconstruct the bone defect with different methods, including autologous or allogeneic bone transplantation, allograft prosthetic composite reconstruction or endoprosthetic replacement with restoration of limb function [54].

After preoperative chemotherapy, patients are classified as good or poor responders based on the number of viable cells remaining in the resection specimen [55]. The method used is called the Huvos and Rosen score [56]. Good responders have more than 90% necrosis (stage III = 91–99% necrosis and grade IV = 100% necrosis). Poor responders have less than 90% necrosis (grade II = less than 90% necrosis; grade I = less than 50% necrosis) [57,58]. The Huvos score is used to adapt postoperative chemotherapy [59].

6. Limitations of Current Treatments

Despite treatment, 30 to 40% of patients will have a recurrence within 2 to 3 years after the end of treatment. At the time of recurrence, the metastases, mostly pulmonary

(90%), must be surgically removed in order to increase patients' survival. However, the 5-year survival after the second diagnosis is about 20%. The current treatments cause severe side effects that diminish patients' quality of life [13,41]. While surgery has been improved and allows limb salvage in 80–95% of patients with limb functionality, the surgery remains invasive and often necessitates reconstruction [60]. The surgical reconstruction also presents some complications, such as infection, graft fracture, implant reject or local recurrence, which could compromise the quality of life of the patients [60–62].

Despite the fact that conventional therapies have lowered the frequency of amputations, the most commonly used drugs date from the 1970s, and the survival rates have remained relatively constant since then. This lack of progress might be attributed to the disease's rarity as well as its wide variability.

7. Genetic Disorders in OS

OS is a complex and heterogeneous tumor characterized by a high level of genomic instability, aneuploidy and genomic rearrangements, with gains of portions of chromosomes (1p, 1q, 6p, 8q and 17p) or losses of portions of chromosomes (3q, 6q, 9, 10, 13,17p and 18q) in conventional disease, generally corresponding to regions where oncogenes and tumor suppressor genes are located, respectively [63]. OS can also arise from inherited genetic disorders, such as Li–Fraumeni syndrome (p53 mutation), or mutation of the gene encoding Rb, but also Rothmund–Thomson (RECQL4 gene mutation), Bloom (BLM) and Werner (WRN) syndromes [8,9]. However, other acquired genetic changes have been described in pathology. Indeed, with the explosion of high-throughput sequencing methods in recent years, many studies have attempted to identify "driver gene mutations", i.e., mutations that confer a proliferative advantage to cells [64–67]. They can either inhibit/inactivate tumor suppressor genes or amplify/facilitate oncogene activity [68–71].

Tumor suppressor genes inhibit tumor growth, for example, by regulating the cell cycle. Many genes belonging to this category are affected in OS. For example, the TP53 gene is the most mutated gene in human tumors [72], encoding the p53 protein, a transcription factor regulating cell cycle and apoptosis. In OS, p53 is inactivated by gene mutation or chromosomal rearrangement [63,64,71,73]. Between 65% and 90% of OS cases have a mutation in TP53 (point mutation, allelic loss or rearrangement) [9,74,75]. Another cell cycle regulator Rb (Retinoblastoma protein) is frequently mutated in OS. Rb regulates the transition from G1 to S phase of the cell cycle by sequestering E2F family transcription factors. Its loss, therefore, results in the disappearance of this cell cycle checkpoint [70]. Loss-of-function mutations in Rb occur in approximately 30% of OS [64,73,76–78]. However, the evidence of the contribution of the Rb mutation in OS is not so clear since several studies reported that germline Rb mutation did not cause OS development in mice [79–81]. Walkley et al. confirmed this result in an Osx-Cre Rb^{fl/fl} mouse model, in which neither OS development nor skeletal abnormalities were observed, concluding that Rb mutation is not sufficient to induce OS development [82].

The functions of p53 and Rb can also be affected by the mutation of CDKN2A (cyclindependent kinase Inhibitor 2A). This gene encodes two proteins, p14Arf and p16INK4, which activate p53 and Rb, respectively. P14 prevents the degradation of p53 mediated by the E3 ubiquitin ligase MDM2 (mouse double minute 2 homolog). P16 inhibits CDK4 (Cyclin-dependent kinase 4), a protein capable to inactivate Rb by phosphorylation. The CDKN2A locus (9p21) is altered in 5–21% of OS [63,83–85]. Hypermethylation of p14ARF and p16INK4 promoters have also been described in OS, resulting in decreased transcription [86,87]. Hypermethylation of promoters of anti-tumor genes is a frequently cited mechanism in OS. This is the case for the promoters of GADD45 (growth arrest and DNA damage) or HIC1 (hypermethylated in cancer 1), both involved in the response to DNA damage signals [88,89].

ATRX is another protein frequently mutated in OS (29% of tumors). It is part of a multiprotein complex regulating chromatin remodeling and telomere maintenance [78]. ATRX is a known tumor suppressor gene, and its mutations lead to alternative telomere

lengthening (ALT) phenomena [68,90,91]. Other examples of tumor suppressor genes described as mutated in OS include the β -catenin regulatory protein APC (Adenomatous polyposis coli), the metalloproteinase inhibitor TIMP3 (Metalloproteinase inhibitor 3) and the Wnt pathway inhibitor WIF-1 (Wnt inhibitory factor 1) [78,92–97].

Oncogenes confer a proliferative advantage to tumor cells. Their expression or activity is increased in cancers. In OS, gain-of-function mutations are observed, for example, in E2F3 (60% of tumors), a transcription factor of the E2F family involved in many cellular processes, including replication, DNA repair and apoptosis. Its inhibition in cancers notably decreases tumor growth by disrupting the cell cycle [98]. CDK4 (cyclin-dependent kinase4) is involved in the transition from G1 to S phase of the cell cycle. It is an inhibitor of the tumor suppressor Rb. In OS, its gene is mutated in about 10% of tumors [99–101]. CDK4 is often co-amplified with MDM2 in various cancers [100]. As previously mentioned, MDM2 is a p53 inhibitory ubiquitin ligase. MDM2 is amplified in 3–25% of OS tumors, resulting in p53 inactivation [102].

C-Myc is an oncogene described in several types of cancers [103–105]. This transcription factor induces cell proliferation, particularly through the regulation of CDKs, including CDK4 [106,107]. Furthermore, via activation of mTOR (mammalian target of rapamycin) and subsequent phosphorylation of 4EBP1 (eukaryotic translation initiation factor 4E binding protein 1), c-Myc increases protein synthesis in cancers [108]. C-Myc also regulates cell death as well as angiogenesis and metastatic processes [109,110]. In OS, the c-Myc gene is amplified in 7–67% of tumors and is overexpressed in 34% of cases [111–113]. The 17p11.2~p12 region has also been described as amplified in 13–32% of high-grade OS [114]. This region contains, among others, the gene encoding PMP22 (peripheral myelin protein 22) [114], a protein involved in tumor proliferation, migration and invasion, including via the MAPK (Mitogen-activated protein kinase) pathway [115,116]. Other oncogenes, such as CDC5L (cell division cycle 5-like involved in G2 cell cycle progression and tumor growth) or RUNX2 (Runt-related transcription factor 2 in osteoblastic differentiation and known oncogene in OS), are also recurrently amplified in OS [114,117,118].

Deletions of Methylthioadenosine phophorylase (MTAP) gene coding for the MTAP, a key enzyme in the salvage of cellular adenine and methionine synthesis, is frequently observed in OS patients. Indeed, deletion of at least one exon of MTAP gene was observed in 37.5% of the patients in a cohort of 96 patients with high-grade OS [119]. No expression of the MTAP protein was observed in 27.5% of the patients in a Japenese OS cohort [120]. These MTAP deletions could be exploited as alternative therapy in MTAP-negative OS patients with the use of 6-thioguanine (6-TG), showing good results in leukemia or in metastatic prostate cancer models [121], or other inhibitors of de novo purine synthesis. However, the use of L-alanosine, a potent inhibitor of adenine biosynthesis, failed to show anti-cancer efficacy in a phase II clinical trial enrolling 65 patients, including 7 OS (2 OS patients with stable disease and 5 OS patients with progressive disease) [122].

Beyond proteins, other cellular actors are involved in genetic abnormalities in OS. Thus, miRNAs (microRNAs), lncRNAs (long non-coding RNAs) and non-protein coding ribonucleotide sequences also have deregulated expression in OS. For example, Pasic and colleagues have shown that the lncRNA loc285194, known to be a p53-controlled tumor suppressor, has its locus deleted in OS. This deletion is associated with poor prognosis in patients [123,124]. Studies have also shown the involvement of miRNAs in OS. For example, Zhou and colleagues showed that miR-340 acts as an inhibitor of tumor growth and metastatic spread by targeting ROCK1 (Rho-associated coil containing protein kinase 1), while Song et al. showed the involvement of miR-140 in chemoresistance [125,126].

Thus, OS is highly heterogeneous genomically, genetically and epigenetically. As an example, Poos et al. identified 911 proteins and 81 miRNAs described as associated with OS [127]. This tumor heterogeneity, coupled with the small number of cases (due to the rarity of the disease), make it difficult to obtain statistically significant results in clinical studies [128]. Nevertheless, the use of in vitro or in vivo models allows to evaluate the efficacy of new therapeutic approaches.

8. Osteosarcoma Models

The use of experimental models makes it possible to understand the effects of new therapeutic strategies before their use in humans. Several types of models are currently used to mimic the pathology. Cellular models from patients with OS have been used routinely in the laboratory for decades (Table 3). They are used to test potential treatments or to identify genetic abnormalities in OS. These cellular models are essential, especially from an ethical point of view, before the use of more complete in vivo models.

Cells	Origin	Gender	Age	Location	Model in Mice	p53 Status	Reference
MG-63	human	male	14	Bone	Yes	rearranged	[129]
Saos-2	human	female	11	Bone	Yes	null	[130]
U2OS	human	female	15	Tibia	Yes	WT inactivated	[131]
MMNG- HOS	human	female	13	Femur	Yes	R156P;F270L	[132]
143-B HOS	human	female	13	Femur	Yes	R156P;F270L	[133]
CAL-72	human	male	10	Knee recurrence	No	WT	[134]
G-292	human	female	9	Bone	ND		[135]
SJSA-1	human	male	19	Femur	ND	P53 and MDM2 amplification	[136]
K7M2	BALB/c mice	ND	ND	ND	syngenic		[137]
POS-1	C3H/He mice	ND	ND	ND	syngenic	WT	[138]
MOS-J	C57BL/6J mouse	ND	ND	ND	syngenic		[139]
OSRGA	Rat (Sprague Dawley)	ND	ND	ND	Syngenic in rat	WT	[140]
UMR-106	Rat (Sprague Dawley)	ND	ND	ND	Syngenic in rat	WT	[141]

Table 3. Most of the used OS cell lines (ND: no details).

The mouse (Mus musculus) is used as an in vivo model in many pathologies. One of the oldest in vivo xenograft models used in cancer is the transplantation of human OS cell lines to mice [142–146].

Other OS animal models exist, such as dogs (Canis familiaris), especially large ones, which spontaneously develop OS with a higher incidence than in humans, although the disease remains rare [147,148]. As in humans, canine OS produces bone or osteoid substances. The osteoblastic type is the majority in both species [149,150]. Tumor location is also similar in both species, with a preponderance in the appendicular skeleton [151]. The similarities between human and canine OS make the latter a good animal model to complement the previously described in vitro and mouse models [7].

The zebrafish model (Danio rerio) is a model used to study tumor growth, migration and invasion of tumor cells in cancers [152]. After injection of fluorescent human tumor cells, a few days are sufficient to observe the processes of proliferation, migration and invasion without the need for special equipment.

Animal models, although imperfect or restrictive, are essential for the development of new and effective treatments.

9. The Vicious Cycle Hypothesis in OS

It has been shown that the tumor is able to hijack the bone remodeling mechanism to ensure its growth. This phenomenon, called a vicious cycle, is found in OS but also in other tumors metastasizing to bone, such as breast or prostate cancer [143,153,154]. In these tumors, the cancer cells synthesize and secrete growth factors, such as PTHrP (parathyroid hormone related protein), IGF, FGF (fibroblast growth factor), VEGF (vascular endothelial growth factor) but also Wnt, which have activating effects on osteoblasts. Osteoblasts will, in turn, promote the differentiation and resorption activity of osteoclasts by producing RANKL (receptor activator of NF-kB ligand). The tumor may also activate osteoclasts through the production of IL-6, IL-11 or IL-1 β [117,143,155–157]. The combined action of osteoblasts and tumor cells results in exaggerated bone matrix resorption by osteoclasts. This resorption leads to the release of BMP, TGF- β , FGF and PDGF (platelet-derived growth factor) into the environment, which will contribute to the survival and proliferation of tumor cells (Figure 1). At the organism level, bone resorption can lead to hypercalcemia as well as fractures due to osteolysis [158–160].



Figure 1. The vicious cycle in osteosarcoma: the cancer cell secretes growth factors that activate osteoblasts. These, together with the tumor cell, promote osteoclastic differentiation and exaggerated resorption of the bone matrix. This resorption leads to the release into the microenvironment of factors involved in tumor survival and proliferation. PTH/rP: parathyroid hormone-related protein, IGF: insulin-like growth factor, FGF: fibroblast growth factor, VEGF: vascular endothelial growth factor, IL-(1, 6, 11): interleukin-(1, 6, 11, RANKL: receptor activator of NF-kB ligand, c-fms: colony stimulating factor 1 receptor, TGF-ß: transforming growth factor **ß**; BMP: bone morphogenetic protein, PDGF: platelet-derived growth factor, M-CSF: macrophage colony-stimulating factor, Ca²⁺: calcium ions.

10. New Therapeutic Approaches in OS

10.1. *Immunotherapy*

Immunomodulation is the modification of immunity or the immune response, with the use of interferons (IFN) as an example. These are glycoproteins of the cytokine family that are produced by immune cells in response to viral infections, showing some efficacy in cancers [161]. In OS, IFN- α -2b inhibits growth of tumor cells and PDX tumors (Table 4) [162]. The efficacy of pegylated recombinant human IFN- α 2b (IFN- α 2b) was studied in patients with OS in an international randomized controlled trial. The patients were treated with combination chemotherapy (methotrexate, doxorubicin and cisplatin: MAP) with or without pegylated IFN- α 2b and the overall survival rate was not significantly different between MAP + IFN- α 2b and MAP [25]. Activation of lymphocytes and their

differentiation into lymphokine-activated killer (LAK) cells can be induced by interleukin-2 (IL-2). LAK recognize and eliminate various tumor cells [163]. The addition of IL-2 to the standard treatment of OS has been studied with LAK reinfusion and surgery in children with metastatic OS, showing 3-year event survival rates of 34% and overall survival rates of 45%, highlighting the interest of IL2 and LAK cells in immunotherapy [164]. However, severe side effects were observed after treatment with a high-dose of IL2, such as fever and influenza-like symptoms in all patients. In some of them, increases in white blood cells, creatinine, gamma-glutamyltransferase, C-reactive protein, glucose and body weight and decreases in red blood cells, platelets, protein, albumin and cholinesterase were observed. Nevertheless, two of the four patients with OS, included in a study of high-dose IL-2 treatment, showed a complete response [165]. Stimulation of the immune system can also be achieved by synthetic molecules, such as mifamurtide (immune modulator liposomal muramyl tripeptide), which is a synthetic derivative of a peptidoglycan that makes up the plasma membrane of bacillus Calmette–Guerin. In cancer, mifamurtide induces the activation of monocytes and macrophages against tumors via the secretion of IL-6, $TNF\alpha$ and increased anti-tumor activity of infiltrating immune cells [166]. The addition of mifamurtide showed an increase in event-free life span and overall survival [167,168] but without a significant difference between non metastatic and metastatic patients [169]. The European Medicine Agency approved the use of mifamurtide in treatment of OS patients between 2 and 30 years of age [170]. Regarding other immunotherapy approaches based on the infusion of antitumor immune-effectors, recent studies reported cytokine-induced killer cells (CIK) are able to kill sarcoma cancer stem cells (sCSC) in vitro [171,172]. CIK are an ex vivo expanded mix of T lymphocytes and T-NK cell-like phenotype that showed therapeutic efficacy in sarcoma including OS, even in chemo-resistant sCSC [173], which provides a therapeutic alternative in the clinic, such as the phase 1 clinical trial CRX100 in patients with refractory solid tumors, including OS patients, which is a dose-escalation study combining CIK cells with an oncolytic virus (NCT04282044).

10.1.1. Targeting Cell Surface Proteins

One of the immunotherapy strategies is to target the cell surface proteins using monoclonal antibodies, which bind specific antigens on the surface of tumor cells, and then activate NK cells and macrophages (Table 4). A phase II clinical trial investigated the efficacy of monoclonal antibody trastuzumab, targeting the human epidermal growth factor receptor 2 (HER2) protein, in metastatic OS patients. While trastuzumab was well tolerated, it failed to significantly show any difference in the outcome between the HER2-positive and HER2-negative patients [174]. A new phase II trial, which started in February 2021, studies the effects of trastuzumab deruxtecan in treating new diagnosed or recurrent HER2 positive OS patients (NCT04616560). The monoclonal antibody trastuzumab is linked to deruxtecan, a chemotherapeutic drug. High HER2 expression in OS was also used to evaluate the efficacy of HER2-specific chimeric antigen receptor modified T cells (CAR T cells) in a clinical trial with 16 enrolled HER2-positive tumor in-patients with recurrent or refractory OS [175]. Infusion of CAR T cells was well tolerated but showed modest anti-tumor activity. Indeed, only four of sixteen patients had stable disease for 12 weeks to 14 months [175].

Monoclonal antibodies have also been used to target the growth hormone receptor insulin-like growth factor-1 (IGF-1). A first phase II clinical trial including 38 OS patients showed that R1507, a recombinant human monoclonal antibody, is well tolerated but has limited efficacy in patients with relapsed or refractory bone and soft tissue sarcomas (NCT00642941) [176]. The efficacy of the monoclonal antibody cixutumumab, which targets IGF1R, was evaluated in a phase II clinical trial in children with relapsed and refractory solid tumors, showing that cixutumumab was well tolerated but with limited efficacy as a single agent [177,178].

Disialoganglioside (GD2) is highly expressed in more than 95% of OS and is involved in cell proliferation, motility, migration, adhesion, and invasion, amounting tumor develop-

ment and malignant phenotypes [179,180]. The efficacy of anti-GD2 antibodies (monoclonal or BiTE antibodies) are evaluated in several clinical trials. Indeed, in a phase II clinical, the combination of the monoclonal antibody dinutuximab with sargramostim (recombinant granulocyte-macrophage colony-stimulating factor (GM-CSF)) is evaluated in the treatment of 39 patients with recurrent OS. While dinutuximab is well tolerated, it failed to demonstrate sufficient efficacy in the disease control rate (NCT02484443) [181]. Other antibodies composed of two single-chain variable fragments linked by a flexible coupler can be used, such as bispecific T cell activators (BiTE). The use of bispecific antibodies recognizing an antigen on the tumor target close to CD3 receptor of T cells, leading to T cells activation and thus cytolysis of tumor cells [182,183]. The efficacy of activated T cells armed with a bispecific GD2 antibody is evaluated in cancer cell lines, PDX models [184], and in a phase I/II clinical trial in children and young adults with neuroblastoma and OS (NCT02173093). A Phase I/II clinical trials is investigating the safety and efficacy of the humanized anti-GD2 and anti-CD3 bispecific antibody 3F8 (Hu3F8-BsAb) in patients with relapsed and refractory neuroblastoma, osteosarcoma and other solid tumors (NCT03860207).

Leucine rich repeat containing 15 (LRRC15) is a membrane protein highly expressed on the cell surface of stromal fibroblasts in many solid tumors including OS and its expression is induced by TGF[®]. This protein has a role in cell–cell and cell–extracellular matrix interaction. It is a novel mesenchymal protein and stromal target for monoclonal antibodydrug conjugates [185,186]. A phase I clinical trial was designed to evaluate the efficacy, the safety and pharmacokinetics of ABBV-085 (Samrotamab vedotin), an antibody drug conjugate, in solid tumors, especially sarcomas (NCT02565758). ABBV-085 was safe and well tolerable with promising antitumor activity in OS patients [187].

B7 Homolog 3 (B7-H3), also known as CD276, is a protein whose main role is to inhibit adaptive immunity by suppressing T cell activation and proliferation [188]. A phase I clinical trial is still active since 2004 and evaluates the effects of Omburtamab (antibody 8H9), a radiolabeled monoclonal antibody in patients with sarcoma and other cancers (NCT00089245). A multicenter phase I/II clinical trial is currently recruiting patients with advanced solid malignant tumors and is investigating the effect of DS-7300a, a novel B7-H3-targeting antibody–drug conjugate with a DNA topoisomerase I inhibitor DXd (NCT04145622).

Other immunoconjugates have been tested in OS using toxin or radionuclide as a linker joined with a carrier antibody and have shown promising results [189]. Indeed, an antigp72 mAb 791T/36 conjugated with methotrexate and ricin toxin A chain (RTA) showed encouraging results by inhibiting OS cells proliferation associated with immunotoxin internalization [190,191]. In the same way, TP-3 mAb, recognizing an antigen present on the surface of OS cells, was conjugated with pokeweed antiviral protein (PAP) [192]. TP-3-PAP showed promising results by reducing tumor cell growth and the number of lung metastases in an OS model [192].

CD146, overexpressed in OS, was used to develop an anti-CD-146 murine antibody named OI-3 coupled with Iodine-125 or Lutetium-177 and was evaluated in an OS xenograft model, showing promising results with therapeutically relevant biodistribution of the radionuclide [193]. Insulin-like growth factor 2 receptor (IGF2R) is overexpressed in OS [194,195], which has been used to develop radiolabeled antibody targeting IGF2R with Indium-111, Lutetium-177 and Bismuth-213 and showing delayed tumor growth in vivo in an OS model [196].

10.1.2. Checkpoint Inhibitors

Checkpoints restrict overly aggressive immune responses and, in some cases, block T cells from killing tumor cells (Table 4). T lymphocytes are better able to kill tumor cells when these checkpoints are blocked [197]. The checkpoint inhibitors targeting anticytotoxic T lymphocyte antigen-4 (CTLA-4), programmed death receptor1 (PD-1) and its ligand (PD-L1) are the most studied in OS [182]. CTLA-4 is a transmembrane glycoprotein receptor, also known as CD152 (cluster of differentiation 152), and overexpressed in patients with OS [198]. CTLA-4 receptor is expressed by activated T cells and regulatory T cells (Tregs) [199] and able to bind CD80 (B7-1) and CD86 (B7-2) expressed by dendritic cells, leading to functional inhibition [199]. Most checkpoint inhibitors targeting CTLA-4 are not living up to expectations and are indeed less effective in treating solid tumors, including

OS, compared with other malignant tumors, such as melanoma [200,201], without clear explanations. Ipilimumab, a CTLA-4 inhibitor, was evaluated in a phase I clinical trial in children with relapsed solid tumors, including patients with OS. No anti-tumor response was reported, while ipilimumab increased activation and levels of cycling of CTLs but not Tregs [202].

PD-1, also known as CD279, is a transmembrane member of the immunoglobulin family expressed on activated cytotoxic CD8+ T cells and natural killer cells [203]. PD-1 ligand (PD-L1) binds to PD-1 receptor, leading to inactivation of T cells [204]. The expression level PD-L1, also called B7-H1 or CD274, is expressed in OS cells and immune cells, especially tumor-infiltrating lymphocytes (TILs) [205,206]. In addition, high PD-L1 expression is associated with poorer 5-year event-free survival in OS compared with patients with low PD-L1 expression [207]. It was reported that PD-L1 was more highly expressed in pediatric metastatic OS tissues compared with primary OS samples [208]. PD-1 and PD-L1 checkpoint inhibitors have shown promising results in basic and preclinical research [209,210]. PD1/PD-L1 inhibitor pembrolizumab was evaluated in a multicenter phase 2 clinical in advanced soft-tissue sarcoma and bone sarcoma including 22 patients with OS and showed a poor response in OS [211]. Indeed, one patient out of 22 (5%) with recurrent OS treated with pembrolizumab had a partial response, 6 patients (27%) had stable disease and 15 patients (68%) had disease progression [211]. The efficacy of avelumab, an anti-PD-L1 antibody, was evaluated in a phase II clinical trial in patients with recurrent OS and did not demonstrate any activity (NCT03006848) [212]. In the SARC038 phase II clinical trial, the efficacy of nivolumab combined with regorafenib, a receptor of tyrosine kinase inhibitor, is currently being evaluated in patients with refractory or recurrent OS (NCT04803877).

However, the clinical evaluation of the checkpoint inhibitors in sarcoma was not a success, as observed in melanoma as an example, probably due to the immunosuppressive role of the microenvironment [213,214].

10.2. Bone Resorption Inhibition

Some therapeutic strategies are based on the inhibition of osteoclasts (Table 4). By doing so, they reduce pathological bone remodeling and its consequences (osteolysis, hypercalcemia, etc.) and directly impact the vicious cycle. In this category, we find the RANK/RANKL inhibitors. The RANK receptor and its ligand RANKL, as well as the decoy receptor OPG, are closely associated with bone remodeling. They control osteoclast differentiation via activation of NF-kB and JNK (Jun N-terminal kinase). RANK is expressed by OS cells, and this expression negatively affects treatment response and survival [215,216]. Several studies have described beneficial effects to targeting the RANK/RANKL/OPG triad in OS [143,217–221]. Denosumab, for example, is a monoclonal antibody directed against RANKL. It inhibits the binding of RANKL to RANK, thus mimicking the physiological decoy receptor role of OPG, inhibiting osteoclastic differentiation [217,222]. Denosumab is currently in a phase II clinical trial in OS, in patients resistant to conventional therapies or in relapse (NCT02470091).

Other bone resorption inhibitors have shown promising results. Indeed, bisphosphonates (e.g., zoledronate) are molecules capable of inhibiting bone resorption. They are pyrophosphate analogues capable of binding to the hydroxyapatite constituting the bone matrix and of inducing apoptosis of osteoclasts [158]. They have been used for the treatment of osteoporosis for 40 years, but also in Paget's disease, breast cancer and prostate cancer [223–226]. In OS, zoledronate has shown encouraging results. In vitro, it inhibits OS cell proliferation and induces cell death [227,228]. In addition, zoledronate sensitizes OS cells to radiation treatment by increasing oxidative stress and inhibiting DNA

repair mechanisms [229]. In vivo, zoledronate reduces tumor growth, osteolysis, angiogenesis, tumor cell invasion and lung metastasis [230–233]. A phase III clinical trial (OS2006, NCT00470223) using zoledronate in combination with chemotherapy, however, did not show an improvement in survival (overall or event-free) of OS patients [55]. A phase I clinical trial (METZOLIMOS, NCT02517918) is ongoing in OS, combining zoledronate with methotrexate, cyclophosphamide and the mTOR inhibitor sirolimus.

10.3. Targeting Receptor Tyrosine Kinases and Intracellular Signaling

Receptor tyrosine kinases (RTKs) are transmembrane receptors with an extracellular domain that binds the ligand and an intracellular domain with kinase activity (e.g., IGF-R: insulin-like growth factor 1 receptor, PDGFR: platelet-derived growth factor receptor...). RTKs are involved in cell growth, proliferation and survival, in particular through their involvement in signaling pathways, such as the PI3K (phosphoinositide 3-kinase) pathway or the MAPK pathway. About 30% of RTKs are mutated or overexpressed in cancer, including osteosarcoma. As a result, the expression of certain RTKs is associated with poor prognosis for patients [234]. Six subfamilies, among the 20 RTK subfamilies, are particularly associated with cancer pathologies. These are the EGFR/ErbB (epidermal growth factor receptor) families, PDGF, FGF, VEGF, HGF (hepatocyte growth factor), and IGF receptors [235]. Indeed, it has been observed that high expression of VEGF, IGF1R or AXL is associated with a decreased overall survival in osteosarcoma [236–238]. For example, IGF expression is associated with higher OS aggressiveness [239]. The AXL receptor is overexpressed in OS, and its expression is associated to a poor prognostic factor for patients [237], while AXL inactivation induces apoptosis of OS cells [240]. Dysfunction of RTKs is generally related to neovascularization, invasion, metastasis and chemotherapy resistance of tumors [241,242]. In OS, it has been shown that RTK KIT is expressed in 46.15% of patients with a poor response to chemotherapy [243]. RET RTK overexpression is also associated with resistance to cisplatin and bortezomib [244,245]. FGFR1 amplification, meanwhile, is observed in 18% of patients resistant to neo-adjuvant chemotherapy [246]. Regarding cell proliferation and metastasis development, it has been shown that deregulation of FGFR found in OS plays an important role in its formation as well as in the development of lung metastasis [247]. The PDGFs/PDGFRs signaling pathway also plays a central role in OS proliferation and migration [248,249]. Inhibition of signaling pathways from aberrantly activated MET and AXL RTKs increases the rate of apoptosis and suppresses migration, proliferation and invasion of OS cells [240,250–252]. IGF-1R expression is also related to the presence of distant metastasis in patients [253]. In addition, RET and MET RTKs are described for their involvement in the development of malignancy in OS cells. Indeed, it has been shown that MET overexpression leads to the malignant transformation of primary osteoblastic cells and that RET overexpression increases the stem-cell-like properties of OS cells [254,255]. RTKs are, therefore, therapeutic targets, which are of major interest in the treatment of OS.

Since the FDA approved imatinib for the treatment of chronic myeloid leukemia in 2001, many potent and well-tolerated tyrosine kinase inhibitors have been developed and have contributed significantly to advances in cancer treatment [256]. Therefore, several therapeutic approaches targeting RTKs have thus been developed but have proven relatively ineffective in light of the emergence of resistance mechanisms. This is notably the case of the VEGFRs inhibitor bevacizumab, tested in addition to conventional MAP chemotherapy (NCT00667342), which showed no progression regarding the histological response or the outcome of patients with localized OS [257]. More recent approaches based on RTKi (RTK inhibitors) favor simultaneous targeting of multiple RTKs or constitutive RTK activation pathways to circumvent resistance mechanisms [258]. Imatinib (STI571), for example, is an RTK inhibitor that targets multiple receptors (PDGFR α and β , cKIT, AXL, RYK (related to receptor tyrosine kinase), EGFR, EphA (ephrin type-A receptor) 2 and 10, IGF1R) [259]. Imatinib induces apoptosis of OS cells in vitro and reduces tumor growth in vivo in mouse OS models [259,260]. However, Imatinib failed to show effective anticancer activity in phase II clinical trials (in OS, Ewing's sarcoma, neuroblastoma and desmoplastic small round

cell tumors) [261–263]. The same finding resulted for cixutumumab, an IGF-1R inhibitor, which showed very limited effects in young patients with refractory solid tumors [177]. Dasatinib (c-KIT, Epha2 and PDGFR β RTK inhibitor) has shown similar results. It is used for the treatment of chronic myeloid leukemia and acute lymphoblastic leukemia [162]. In OS, dasatinib has shown anti-metastatic effects but failed to inhibit primary tumor growth [264,265]. Two studies are ongoing (NCT00464620, NCT00788125) with dasatinib alone or in combination with other molecules. An inhibitor of Met (HGF receptor) activity called PF-2341066 reduces tumor growth in a mouse model of OS [266].

The intertwining of different signaling pathways downstream of RTKs is responsible for the rapid emergence of compensatory mechanisms that negate the effect of molecules targeting a limited number of RTKs. Thus, multi-tyrosine kinase inhibitors (mTKIs) have emerged and have shown effects on patients in relapse and/or with unresectable OS. Five molecules stood out for their major effects: Sorafenib, Regorafenib, Cabozantib, Lenvatinib and Pazopanib. Sorafenib, an inhibitor of VEGFR, PDGFR, RET and c-Kit, has shown antiangiogenic and anti-metastatic effects in preclinical models [267]. In the clinical trial NCT00889057, sorafenib showed encouraging results in 35 patients with a PFS of 46% at 4 months and a PR of 9%, although treatment had to be reduced or briefly discontinued in 46% of patients due to toxicity [268]. The results from another phase II clinical trial in OS in patients with advanced or metastatic OS after failure of initial therapy indicate that sorafenib inhibits tumor progression at 6 months in half of the patients [269]. Two clinical trials also demonstrated the efficacy and safety of Regorafenib (targeting VEGFR, PDGFR, KIT, FGFR and RET) in patients with advanced or metastatic OS after failure of prior therapy, showing a PFS of 62% at 12 weeks and a PR of 8% (NCT02048371 and NCT02389244) [270,271]. Cabozantinib is an inhibitor of the VEGFR, KIT, RET, AXL and PDGFR RTKs that was studied in a multicenter phase II clinical trial of 42 patients with advanced or metastatic OS after failure of other systemic therapy (NCT02243605). Further, 12% of the patients developed a PR, and the PFS of 33.3% at 6 months was the best result obtained by RTKi in the treatment of OS to date [272]. A clinical trial is also underway to test the activity of Lenvatinib (targeting VEGFR, PDGFR, KIT, FGFR and RET) in relapsed patients (NCT02432274). The first published results from 31 patients show a 4-month PFS of 29% [273]. Finally, Pazopanib, a second-generation MTKi targeting VEGF, PDGFR and KIT, showed positive effects in 3 patients with relapsed OS for the second time, appearing to stabilize disease progression and thus prolong patient survival [274].

Inhibition of RTKs is promising and actively studied, but other approaches aim to inhibit intracellular signaling downstream of RTKs (Table 4). Indeed, given the importance of signaling pathways in cellular processes and in the development of cancers, including OS, many inhibitors targeting members of these pathways have been developed. For example, there are inhibitors of members of the SFK (Src family kinase), proteins that integrate and regulate the signaling of many RTKs (EGFR, PDGFR, IGF1R, VEGRF, HER2...). Through their targets or partners, SFK family members regulate cell survival, angiogenesis, cell mobility... [275]. Src (steroid receptor co-activator) kinase belonging to this family is notably involved in the activation of osteoclasts under physiological conditions [276]. Src is overexpressed in OS and other types of cancers, and this overexpression correlates with lower patient survival [277]. Saracatinib (AZD0530), a selective Src kinase inhibitor, has been tested in 18 subjects with recurrent OS localized to the lung. The study demonstrated an increase in the median PFS in the treated group from 8.6 months in the placebo treatment group to 19.4 months [278].

Inhibitors of mTOR have also been developed. mTOR is a serine/threonine kinase involved in the deregulated PI3K/Akt pathway in most cancers [279,280]. mTOR is involved in protein synthesis, cell cycle or survival and is overexpressed in OS [281–283]. In addition to the sirolimus, the class of mTOR inhibitors also includes ridaforolimus, a rapamycin analog tested in a phase III clinical trial in metastatic bone sarcoma. It showed weak but statistically significant inhibition of tumor progression in patients [284,285]. CC-115 is an analog of thalidomide [286] inhibiting mTOR, but also the serine/threonine kinase

DNA-PK (DNA-dependent protein kinase) involved in DNA repair. Treatment of OS cells with CC-115 increases their sensitivity to cisplatin and etoposide chemotherapy [287]. A phase I clinical trial is underway in several tumors, including OS (NCT01353625).

The YAP/TAZ (yes-associated protein/paralog transcriptional coactivator with PDZbinding motif) signaling pathway was recently described to be involved in OS with aberrations in the Hippo signaling pathway [288,289]. Indeed, several immunohistochemistry studies demonstrated that 60% to 85% of OS exhibit high YAP expression, with only 30% to 46% of cases showing YAP nuclear expression, with no more frequency of YAP nuclear expression in metastatic patients [290–294]. However, Morice et al. demonstrated that high YAP expression in primary tumors in OS is higher in metastatic patients versus nonmetastatic patients at diagnosis and associated with poor prognosis [295]. Inhibition of YAP using shRNA or chemical inhibitors (verteporfin or CA3) in OS cell lines delays tumor proliferation in vitro and in vivo and blocks migration and invasion [292,294,296]. Indeed, inhibition of the primary tumor growth is explained by the ability of YAP to bind TEAD, while inhibition of lung metastasis is due to the inhibition of YAP/smad3 interaction in the TGF-ß signaling pathway [295,297–299].

Thus, many therapeutic targets are currently being studied for the treatment of OS (Table 4). Often very targeted, they may come up against tumor heterogeneity. Indeed, the tumor is composed of a set of clonogenic populations that have evolved and present different characteristics and gene expressions. The use of therapeutic approaches targeted to a restricted tumor population can lead to recurrences due to the proliferation of cell populations not affected by the treatment. In this context, the strategy of targeting multiple mechanisms shared only by tumor cells has a clear advantage: impacting all tumor cells while sparing healthy cells.

Target Cell, Gene or Protein	Agent Used	Reference		
Targeting Receptor Tyrosine Kinases and Intracellular Signaling				
PDGFR α and β , cKIT, Axl, RYK, EGFR, EphA 2 and 10, IGF1R	Imatinib (STI571)	[259,261–263]		
c-KIT, Epha2 and PDGFRβ RTK inhibitor	Dasatinib	[264,265]		
Met (HGFr)	PF-2341066	[266]		
VEGFR, PDGFR, RET and c-Kit	Sorafenib	[267–269]		
VEGFRs	Bevacizumab	[257]		
IGF-1R	Cixutumumab	[177]		
VEGFR, PDGFR, KIT, FGFR and RET	Regorafenib	[270,271]		
VEGFR, KIT, RET, AXL and PDGFR	Cabozantinib	[272]		
VEGFR, PDGFR, KIT, FGFR and RET	Lenvatinib	[273]		
VEGF, PDGFR and KIT	Pazopanib	[274]		
Src	Saracatinib (AZD0530)	[278]		
mTOR	Ridaforolimus	[284,285]		
mTOR and DNA-PK	CC-115	[286,287]		
Immunomodulation				
immune system	IFN-α-2b	[25,162]		
lymphocytes	IL-2	[164,165]		
Monocytes and macrophages	Mifamurtide	[167,168]		

Table 4. New therapeutic approaches in OS.

Target Cell, Gene or Protein	Agent Used	Reference
	Targeting surface proteins	
LIED2	Trastuzumab	[174]
HEK2	Trastuzumab deruxtecan	NCT04616560
IGF-1/IGF-1R	R1507	NCT00642941 [176]
	Cixutumumab	[177,178]
	Dinutuximab + Sargramostim	NCT02484443 [181]
GD2	humanized bispecific anti-GD2 antibody 3F8 (Hu3F8-BsAb)	NCT03860207
	activated T cells armed with a bispecific GD2 antibody	NCT02173093 [184]
LRRC15	ABBV-085	NCT02565758 [187]
	Omburtanab	NCT00089245
B7-H3	DS-7300a	NCT04145622
	Checkpoint inhibitors	
CTLA-4	Ipilimumab	[202]
PD-1	Pembrolizumab	[211]
	Nivolumab	NCT04803877 [210]
PD-L1	Avelumab	NCT03006848 [212]
	Bone resorption inhibition	
RANKL	Denosumab	[217,222]
	Zoledronate	[227,228,230,232,233,281]
Hydroxyapatite	Zoledronate + chemotherapy	[55]
	Zoledronate + sirolimus	NCT02517918 (METZOLIMOS)

Table 4. Cont.

11. Conclusions

Development of therapeutic targets in OS is very complex due to the heterogeneity of this pathology, which limits the effectiveness of treatments and favors tumor recurrence and the emergence of drug resistance. Indeed, in the last few decades, only a few new therapies have shown a clinically significant impact for patients with OS. To supplement the current knowledge and uncover possible ways to improve patient outcomes, fundamental, translational and clinical research must cooperate, thus allowing the development of new prognostic markers and new therapeutic targets in OS.

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References

- 1. Luetke, A.; Meyers, P.A.; Lewis, I.; Juergens, H. Osteosarcoma treatment—Where do we stand? A state of the art review. *Cancer Treat. Rev.* 2014, 40, 523–532. [CrossRef]
- 2. Ritter, J.; Bielack, S.S. Osteosarcoma. Ann. Oncol. 2010, 21 (Suppl. S7), vii320–vii325. [CrossRef]
- 3. Simpson, E.; Brown, H.L. Understanding osteosarcomas. JAAPA 2018, 31, 15–19. [CrossRef] [PubMed]
- Mirabello, L.; Troisi, R.J.; Savage, S.A. International osteosarcoma incidence patterns in children and adolescents, middle ages and elderly persons. *Int. J. Cancer* 2009, 125, 229–234. [CrossRef] [PubMed]
- 5. Stiller, C.A.; Bielack, S.S.; Jundt, G.; Steliarova-Foucher, E. Bone tumours in European children and adolescents, 1978–1997. Report from the Automated Childhood Cancer Information System project. *Eur. J. Cancer* 2006, *42*, 2124–2135. [CrossRef] [PubMed]
- 6. Troisi, R.; Masters, M.N.; Joshipura, K.; Douglass, C.; Cole, B.F.; Hoover, R.N. Perinatal factors, growth and development, and osteosarcoma risk. *Br. J. Cancer* 2006, *95*, 1603–1607. [CrossRef]
- Simpson, S.; Dunning, M.D.; de Brot, S.; Grau-Roma, L.; Mongan, N.P.; Rutland, C.S. Comparative review of human and canine osteosarcoma: Morphology, epidemiology, prognosis, treatment and genetics. *Acta Vet. Scand.* 2017, 59, 71. [CrossRef]
- Li, F.P.; Fraumeni, J.F., Jr. Soft-tissue sarcomas, breast cancer, and other neoplasms. A familial syndrome? Ann. Intern. Med. 1969, 71, 747–752. [CrossRef]
- 9. Rickel, K.; Fang, F.; Tao, J. Molecular genetics of osteosarcoma. *Bone* 2017, 102, 69–79. [CrossRef]
- 10. Bielack, S.; Jurgens, H.; Jundt, G.; Kevric, M.; Kuhne, T.; Reichardt, P.; Zoubek, A.; Werner, M.; Winkelmann, W.; Kotz, R. Osteosarcoma: The COSS experience. *Cancer Treat. Res.* **2009**, *152*, 289–308.
- 11. Bielack, S.S.; Kempf-Bielack, B.; Delling, G.; Exner, G.U.; Flege, S.; Helmke, K.; Kotz, R.; Salzer-Kuntschik, M.; Werner, M.; Winkelmann, W.; et al. Prognostic factors in high-grade osteosarcoma of the extremities or trunk: An analysis of 1702 patients treated on neoadjuvant cooperative osteosarcoma study group protocols. *J. Clin. Oncol.* **2002**, *20*, 776–790. [CrossRef] [PubMed]
- 12. Ozaki, T.; Flege, S.; Liljenqvist, U.; Hillmann, A.; Delling, G.; Salzer-Kuntschik, M.; Jurgens, H.; Kotz, R.; Winkelmann, W.; Bielack, S.S. Osteosarcoma of the spine: Experience of the Cooperative Osteosarcoma Study Group. *Cancer* **2002**, *94*, 1069–1077. [CrossRef]
- Zhao, X.; Wu, Q.; Gong, X.; Liu, J.; Ma, Y. Osteosarcoma: A review of current and future therapeutic approaches. *Biomed. Eng.* Online 2021, 20, 24. [CrossRef]
- 14. Ohba, T.; Cole, H.A.; Cates, J.M.; Slosky, D.A.; Haro, H.; Ando, T.; Schwartz, H.S.; Schoenecker, J.G. Bisphosphonates inhibit osteosarcoma-mediated osteolysis via attenuation of tumor expression of MCP-1 and RANKL. *J. Bone Miner. Res.* **2014**, *29*, 1431–1445. [CrossRef]
- 15. Shimose, S.; Kubo, T.; Fujimori, J.; Furuta, T.; Ochi, M. A novel assessment method of serum alkaline phosphatase for the diagnosis of osteosarcoma in children and adolescents. *J. Orthop. Sci.* 2014, *19*, 997–1003. [CrossRef] [PubMed]
- Jo, V.Y.; Fletcher, C.D. WHO classification of soft tissue tumours: An update based on the 2013 (4th) edition. *Pathology* 2014, 46, 95–104. [CrossRef] [PubMed]
- 17. Kundu, Z.S. Classification, imaging, biopsy and staging of osteosarcoma. Indian J. Orthop. 2014, 48, 238–246. [CrossRef]
- Doyle, L.A. Sarcoma classification: An update based on the 2013 World Health Organization Classification of Tumors of Soft Tissue and Bone. *Cancer* 2014, 120, 1763–1774. [CrossRef]
- 19. Jawad, M.U.; Scully, S.P. In brief: Classifications in brief: Enneking classification: Benign and malignant tumors of the musculoskeletal system. *Clin. Orthop. Relat. Res.* 2010, *468*, 2000–2002. [CrossRef]
- Gomez-Brouchet, A.; Mascard, E.; Siegfried, A.; de Pinieux, G.; Gaspar, N.; Bouvier, C.; Aubert, S.; Marec-Berard, P.; Piperno-Neumann, S.; Marie, B.; et al. Assessment of resection margins in bone sarcoma treated by neoadjuvant chemotherapy: Literature review and guidelines of the bone group (GROUPOS) of the French sarcoma group and bone tumor study group (GSF-GETO/RESOS). Orthop. Traumatol. Surg. Res. 2019, 105, 773–780. [CrossRef]
- Enneking, W.F.; Spanier, S.S.; Goodman, M.A. A system for the surgical staging of musculoskeletal sarcoma. *Clin. Orthop. Relat. Res.* 1980, 153, 106–120. [CrossRef]
- Amin, M.B.; Greene, F.L.; Edge, S.B.; Compton, C.C.; Gershenwald, J.E.; Brookland, R.K.; Meyer, L.; Gress, D.M.; Byrd, D.R.; Winchester, D.P. The Eighth Edition AJCC Cancer Staging Manual: Continuing to build a bridge from a population-based to a more "personalized" approach to cancer staging. CA. *Cancer J. Clin.* 2017, 67, 93–99. [CrossRef]
- 23. Rosen, G.; Tan, C.; Sanmaneechai, A.; Beattie, E.J., Jr.; Marcove, R.; Murphy, M.L. The rationale for multiple drug chemotherapy in the treatment of osteogenic sarcoma. *Cancer* **1975**, *35* (Suppl. S3), 936–945. [CrossRef]
- 24. Arndt, C.A.; Crist, W.M. Common musculoskeletal tumors of childhood and adolescence. *N. Engl. J. Med.* **1999**, 341, 342–352. [CrossRef]
- 25. Bielack, S.S.; Smeland, S.; Whelan, J.S.; Marina, N.; Jovic, G.; Hook, J.M.; Krailo, M.D.; Gebhardt, M.; Papai, Z.; Meyer, J.; et al. Methotrexate, Doxorubicin, and Cisplatin (MAP) Plus Maintenance Pegylated Interferon Alfa-2b Versus MAP Alone in Patients With Resectable High-Grade Osteosarcoma and Good Histologic Response to Preoperative MAP: First Results of the EURAMOS-1 Good Response Randomized Controlled Trial. *J. Clin. Oncol.* 2015, 33, 2279–2287. [PubMed]
- Schwarz, R.; Bruland, O.; Cassoni, A.; Schomberg, P.; Bielack, S. The role of radiotherapy in oseosarcoma. *Cancer Treat. Res.* 2009, 152, 147–164. [PubMed]
- DeLaney, T.F.; Park, L.; Goldberg, S.I.; Hug, E.B.; Liebsch, N.J.; Munzenrider, J.E.; Suit, H.D. Radiotherapy for local control of osteosarcoma. Int. J. Radiat. Oncol. Biol. Phys. 2005, 61, 492–498. [CrossRef]

- Resche, I.; Chatal, J.F.; Pecking, A.; Ell, P.; Duchesne, G.; Rubens, R.; Fogelman, I.; Houston, S.; Fauser, A.; Fischer, M.; et al. A dose-controlled study of 153Sm-ethylenediaminetetramethylenephosphonate (EDTMP) in the treatment of patients with painful bone metastases. *Eur. J. Cancer* 1997, 33, 1583–1591. [CrossRef]
- 29. Berger, M.; Grignani, G.; Giostra, A.; Ferrari, S.; Ferraresi, V.; Tamburini, A.; Cefalo, G.; Carnevale-Schianca, F.; Vassallo, E.; Picci, P.; et al. 153Samarium-EDTMP administration followed by hematopoietic stem cell support for bone metastases in osteosarcoma patients. *Ann. Oncol.* **2012**, *23*, 1899–1905. [CrossRef]
- 30. Anderson, P.M.; Wiseman, G.A.; Dispenzieri, A.; Arndt, C.A.; Hartmann, L.C.; Smithson, W.A.; Mullan, B.P.; Bruland, O.S. High-dose samarium-153 ethylene diamine tetramethylene phosphonate: Low toxicity of skeletal irradiation in patients with osteosarcoma and bone metastases. *J. Clin. Oncol.* **2002**, *20*, 189–196. [CrossRef]
- 31. Anderson, P.M.; Wiseman, G.A.; Erlandson, L.; Rodriguez, V.; Trotz, B.; Dubansky, S.A.; Albritton, K. Gemcitabine radiosensitization after high-dose samarium for osteoblastic osteosarcoma. *Clin. Cancer Res.* **2005**, *11*, 6895–6900. [CrossRef] [PubMed]
- 32. Ferrari, S.; Serra, M. An update on chemotherapy for osteosarcoma. *Expert. Opin. Pharmacother.* **2015**, *16*, 2727–2736. [CrossRef] [PubMed]
- Bacci, G.; Longhi, A.; Fagioli, F.; Briccoli, A.; Versari, M.; Picci, P. Adjuvant and neoadjuvant chemotherapy for osteosarcoma of the extremities: 27 year experience at Rizzoli Institute, Italy. *Eur. J. Cancer* 2005, *41*, 2836–2845. [CrossRef] [PubMed]
- 34. Bishop, M.W.; Janeway, K.A.; Gorlick, R. Future directions in the treatment of osteosarcoma. *Curr. Opin. Pediatr.* **2016**, *28*, 26–33. [CrossRef] [PubMed]
- Cortes, E.P.; Holland, J.F.; Wang, J.J.; Sinks, L.F.; Blom, J.; Senn, H.; Bank, A.; Glidewell, O. Amputation and adriamycin in primary osteosarcoma. N. Engl. J. Med. 1974, 291, 998–1000. [CrossRef]
- Harris, M.B.; Cantor, A.B.; Goorin, A.M.; Shochat, S.J.; Ayala, A.G.; Ferguson, W.S.; Holbrook, T.; Link, M.P. Treatment of osteosarcoma with ifosfamide: Comparison of response in pediatric patients with recurrent disease versus patients previously untreated: A Pediatric Oncology Group study. *Med. Pediatr. Oncol.* 1995, 24, 87–92. [CrossRef]
- Jaffe, N.; Paed, D.; Farber, S.; Traggis, D.; Geiser, C.; Kim, B.S.; Das, L.; Frauenberger, G.; Djerassi, I.; Cassady, J.R. Favorable response of metastatic osteogenic sarcoma to pulse high-dose methotrexate with citrovorum rescue and radiation therapy. *Cancer* 1973, *31*, 1367–1373. [CrossRef]
- Ochs, J.J.; Freeman, A.I.; Douglass, H.O., Jr.; Higby, D.S.; Mindell, E.R.; Sinks, L.F. cis-Dichlorodiammineplatinum (II) in advanced osteogenic sarcoma. *Cancer Treat. Rep.* 1978, 62, 239–245.
- Gaspar, N.; Occean, B.V.; Pacquement, H.; Bompas, E.; Bouvier, C.; Brisse, H.J.; Castex, M.P.; Cheurfa, N.; Corradini, N.; Delaye, J.; et al. Results of methotrexate-etoposide-ifosfamide based regimen (M-EI) in osteosarcoma patients included in the French OS2006/sarcome-09 study. *Eur. J. Cancer* 2018, *88*, 57–66. [CrossRef]
- Eaton, B.R.; Schwarz, R.; Vatner, R.; Yeh, B.; Claude, L.; Indelicato, D.J.; Laack, N. Osteosarcoma. *Pediatr. Blood. Cancer* 2021, 68 (Suppl. S2), e28352. [CrossRef]
- Meltzer, P.S.; Helman, L.J. New Horizons in the Treatment of Osteosarcoma. N. Engl. J. Med. 2021, 385, 2066–2076. [CrossRef] [PubMed]
- 42. Anninga, J.K.; Gelderblom, H.; Fiocco, M.; Kroep, J.R.; Taminiau, A.H.; Hogendoorn, P.C.; Egeler, R.M. Chemotherapeutic adjuvant treatment for osteosarcoma: Where do we stand? *Eur. J. Cancer* **2011**, *47*, 2431–2445. [CrossRef] [PubMed]
- Imran, H.; Enders, F.; Krailo, M.; Sim, F.; Okuno, S.; Hawkins, D.; Neglia, J.; Randall, R.L.; Womer, R.; Mascarenhas, L.; et al. Effect of time to resumption of chemotherapy after definitive surgery on prognosis for non-metastatic osteosarcoma. *J. Bone Joint. Surg. Am.* 2009, *91*, 604–612. [CrossRef] [PubMed]
- 44. Russo, C.; Lavorgna, M.; Cesen, M.; Kosjek, T.; Heath, E.; Isidori, M. Evaluation of acute and chronic ecotoxicity of cyclophosphamide, ifosfamide, their metabolites/transformation products and UV treated samples. *Environ. Pollut.* **2018**, *233*, 356–363. [CrossRef] [PubMed]
- 45. Schwartz, C.L.; Wexler, L.H.; Krailo, M.D.; Teot, L.A.; Devidas, M.; Steinherz, L.J.; Goorin, A.M.; Gebhardt, M.C.; Healey, J.H.; Sato, J.K.; et al. Intensified Chemotherapy With Dexrazoxane Cardioprotection in Newly Diagnosed Nonmetastatic Osteosarcoma: A Report From the Children's Oncology Group. *Pediatr. Blood. Cancer* 2016, 63, 54–61. [CrossRef]
- 46. Aznab, M.; Hematti, M. Evaluation of clinical process in osteosarcoma patients treated with chemotherapy including cisplatin, adriamycin, ifosfamide, and etoposide and determination of the treatment sequels in a long-term 11-year follow-up. *J. Cancer Res. Ther.* **2017**, *13*, 291–296. [CrossRef]
- Alvarez, O.; Freeman, A.; Bedros, A.; Call, S.K.; Volsch, J.; Kalbermatter, O.; Halverson, J.; Convy, L.; Cook, L.; Mick, K.; et al. Randomized double-blind crossover ondansetron-dexamethasone versus ondansetron-placebo study for the treatment of chemotherapy-induced nausea and vomiting in pediatric patients with malignancies. *J. Pediatr. Hematol. Oncol.* 1995, 17, 145–150. [CrossRef]
- 48. Kim, H.J.; Chalmers, P.N.; Morris, C.D. Pediatric osteogenic sarcoma. Curr. Opin. Pediatr. 2010, 22, 61–66. [CrossRef]
- 49. Heymann, M.F.; Brown, H.K.; Heymann, D. Drugs in early clinical development for the treatment of osteosarcoma. *Expert. Opin. Investig. Drugs* **2016**, 25, 1265–1280. [CrossRef]
- 50. Hasley, I.; Gao, Y.; Blevins, A.E.; Miller, B.J. The Significance of a "Close" Margin in Extremity Sarcoma: A Systematic Review. *Iowa Orthop. J.* 2018, *38*, 123–130.
- 51. Gosheger, G.; Gebert, C.; Ahrens, H.; Streitbuerger, A.; Winkelmann, W.; Hardes, J. Endoprosthetic reconstruction in 250 patients with sarcoma. *Clin. Orthop. Relat. Res.* 2006, 450, 164–171. [CrossRef] [PubMed]

- 52. Grimer, R.J. Surgical options for children with osteosarcoma. Lancet Oncol. 2005, 6, 85–92. [CrossRef]
- 53. Levin, A.S.; Arkader, A.; Morris, C.D. Reconstruction Following Tumor Resections in Skeletally Immature Patients. J. Am. Acad. Orthop. Surg. 2017, 25, 204–213. [CrossRef] [PubMed]
- 54. Misaghi, A.; Goldin, A.; Awad, M.; Kulidjian, A.A. Osteosarcoma: A comprehensive review. SICOT. J. 2018, 4, 12. [CrossRef]
- 55. Piperno-Neumann, S.; Le Deley, M.C.; Redini, F.; Pacquement, H.; Marec-Berard, P.; Petit, P.; Brisse, H.; Lervat, C.; Gentet, J.C.; Entz-Werle, N.; et al. Zoledronate in combination with chemotherapy and surgery to treat osteosarcoma (OS2006): A randomised, multicentre, open-label, phase 3 trial. *Lancet Oncol.* 2016, *17*, 1070–1080. [CrossRef]
- 56. Rosen, G.; Marcove, R.C.; Caparros, B.; Nirenberg, A.; Kosloff, C.; Huvos, A.G. Primary osteogenic sarcoma: The rationale for preoperative chemotherapy and delayed surgery. *Cancer* **1979**, *43*, 2163–2177. [CrossRef]
- O'Kane, G.M.; Cadoo, K.A.; Walsh, E.M.; Emerson, R.; Dervan, P.; O'Keane, C.; Hurson, B.; O'Toole, G.; Dudeney, S.; Kavanagh, E.; et al. Perioperative chemotherapy in the treatment of osteosarcoma: A 26-year single institution review. *Clin. Sarcoma. Res.* 2015, *5*, 17. [CrossRef]
- 58. Vijayanarasimha, D.; Nayanar, S.K.; Vikram, S.; Patil, V.M.; Babu, S.; Satheesan, B. Clinico-pathological Study of Limb Salvage Surgery for Osteosarcoma: Experience in a Rural Cancer Center. *Indian. J. Surg. Oncol.* **2017**, *8*, 136–141. [CrossRef]
- Crenn, V.; Biteau, K.; Amiaud, J.; Dumars, C.; Guiho, R.; Vidal, L.; Nail, L.L.; Heymann, D.; Moreau, A.; Gouin, F.; et al. Bone microenvironment has an influence on the histological response of osteosarcoma to chemotherapy: Retrospective analysis and preclinical modeling. *Am. J. Cancer Res.* 2017, *7*, 2333–2349.
- 60. Grinberg, S.Z.; Posta, A.; Weber, K.L.; Wilson, R.J. Limb Salvage and Reconstruction Options in Osteosarcoma. *Adv. Exp. Med. Biol.* **2020**, 1257, 13–29.
- 61. Mangat, K.S.; Jeys, L.M.; Carter, S.R. Latest developments in limb-salvage surgery in osteosarcoma. *Expert Rev. Anticancer Ther.* **2011**, *11*, 205–215. [CrossRef] [PubMed]
- Perrot, P.; Rousseau, J.; Bouffaut, A.L.; Redini, F.; Cassagnau, E.; Deschaseaux, F.; Heymann, M.F.; Heymann, D.; Duteille, F.; Trichet, V.; et al. Safety concern between autologous fat graft, mesenchymal stem cell and osteosarcoma recurrence. *PLoS ONE* 2010, 5, e10999. [CrossRef]
- 63. Martin, J.W.; Squire, J.A.; Zielenska, M. The genetics of osteosarcoma. Sarcoma 2012, 2012, 627254. [CrossRef] [PubMed]
- 64. Bousquet, M.; Noirot, C.; Accadbled, F.; Sales de Gauzy, J.; Castex, M.P.; Brousset, P.; Gomez-Brouchet, A. Whole-exome sequencing in osteosarcoma reveals important heterogeneity of genetic alterations. *Ann. Oncol.* **2016**, *27*, 738–744. [CrossRef] [PubMed]
- Kovac, M.; Blattmann, C.; Ribi, S.; Smida, J.; Mueller, N.S.; Engert, F.; Castro-Giner, F.; Weischenfeldt, J.; Kovacova, M.; Krieg, A.; et al. Exome sequencing of osteosarcoma reveals mutation signatures reminiscent of BRCA deficiency. *Nat. Commun.* 2015, 6, 8940. [CrossRef] [PubMed]
- Vogelstein, B.; Papadopoulos, N.; Velculescu, V.E.; Zhou, S.; Diaz, L.A., Jr.; Kinzler, K.W. Cancer genome landscapes. *Science* 2013, 339, 1546–1558. [CrossRef]
- Savage, S.A.; Mirabello, L.; Wang, Z.; Gastier-Foster, J.M.; Gorlick, R.; Khanna, C.; Flanagan, A.M.; Tirabosco, R.; Andrulis, I.L.; Wunder, J.S.; et al. Genome-wide association study identifies two susceptibility loci for osteosarcoma. *Nat. Genet.* 2013, 45, 799–803. [CrossRef]
- 68. Napier, C.E.; Huschtscha, L.I.; Harvey, A.; Bower, K.; Noble, J.R.; Hendrickson, E.A.; Reddel, R.R. ATRX represses alternative lengthening of telomeres. *Oncotarget* 2015, *6*, 16543–16558. [CrossRef]
- Scheel, C.; Schaefer, K.L.; Jauch, A.; Keller, M.; Wai, D.; Brinkschmidt, C.; van Valen, F.; Boecker, W.; Dockhorn-Dworniczak, B.; Poremba, C. Alternative lengthening of telomeres is associated with chromosomal instability in osteosarcomas. *Oncogene* 2001, 20, 3835–3844. [CrossRef]
- 70. van Harn, T.; Foijer, F.; van Vugt, M.; Banerjee, R.; Yang, F.; Oostra, A.; Joenje, H.; te Riele, H. Loss of Rb proteins causes genomic instability in the absence of mitogenic signaling. *Genes Dev.* **2010**, *24*, 1377–1388. [CrossRef]
- Weiss, M.B.; Vitolo, M.I.; Mohseni, M.; Rosen, D.M.; Denmeade, S.R.; Park, B.H.; Weber, D.J.; Bachman, K.E. Deletion of p53 in human mammary epithelial cells causes chromosomal instability and altered therapeutic response. *Oncogene* 2010, 29, 4715–4724. [CrossRef] [PubMed]
- 72. Thoenen, E.; Curl, A.; Iwakuma, T. TP53 in bone and soft tissue sarcomas. *Pharmacol. Ther.* **2019**, 202, 149–164. [CrossRef] [PubMed]
- 73. Serra, M.; Hattinger, C.M. The pharmacogenomics of osteosarcoma. *Pharmacogen. J.* 2017, 17, 11–20. [CrossRef] [PubMed]
- 74. Andreassen, A.; Oyjord, T.; Hovig, E.; Holm, R.; Florenes, V.A.; Nesland, J.M.; Myklebost, O.; Hoie, J.; Bruland, O.S.; Borresen, A.L.; et al. p53 abnormalities in different subtypes of human sarcomas. *Cancer Res.* **1993**, *53*, 468–471. [PubMed]
- 75. Toguchida, J.; Yamaguchi, T.; Dayton, S.H.; Beauchamp, R.L.; Herrera, G.E.; Ishizaki, K.; Yamamuro, T.; Meyers, P.A.; Little, J.B.; Sasaki, M.S.; et al. Prevalence and spectrum of germline mutations of the p53 gene among patients with sarcoma. *N. Engl. J. Med.* **1992**, 326, 1301–1308. [CrossRef] [PubMed]
- Miller, C.W.; Aslo, A.; Won, A.; Tan, M.; Lampkin, B.; Koeffler, H.P. Alterations of the p53, Rb and MDM2 genes in osteosarcoma. J. Cancer Res. Clin. Oncol. 1996, 122, 559–565. [CrossRef] [PubMed]
- Alonso, J.; Garcia-Miguel, P.; Abelairas, J.; Mendiola, M.; Pestana, A. A microsatellite fluorescent method for linkage analysis in familial retinoblastoma and deletion detection at the RB1 locus in retinoblastoma and osteosarcoma. *Diagn. Mol. Pathol.* 2001, 10, 9–14. [CrossRef]

- Chen, X.; Bahrami, A.; Pappo, A.; Easton, J.; Dalton, J.; Hedlund, E.; Ellison, D.; Shurtleff, S.; Wu, G.; Wei, L.; et al. Recurrent somatic structural variations contribute to tumorigenesis in pediatric osteosarcoma. *Cell. Rep.* 2014, 7, 104–112. [CrossRef]
- 79. Jacks, T.; Fazeli, A.; Schmitt, E.M.; Bronson, R.T.; Goodell, M.A.; Weinberg, R.A. Effects of an Rb mutation in the mouse. *Nature* **1992**, *359*, 295–300. [CrossRef]
- Clarke, A.R.; Maandag, E.R.; van Roon, M.; van der Lugt, N.M.; van der Valk, M.; Hooper, M.L.; Berns, A.; te Riele, H. Requirement for a functional Rb-1 gene in murine development. *Nature* 1992, 359, 328–330. [CrossRef]
- 81. Lee, E.Y.; Chang, C.Y.; Hu, N.; Wang, Y.C.; Lai, C.C.; Herrup, K.; Lee, W.H.; Bradley, A. Mice deficient for Rb are nonviable and show defects in neurogenesis and haematopoiesis. *Nature* **1992**, *359*, 288–294. [CrossRef] [PubMed]
- Walkley, C.R.; Qudsi, R.; Sankaran, V.G.; Perry, J.A.; Gostissa, M.; Roth, S.I.; Rodda, S.J.; Snay, E.; Dunning, P.; Fahey, F.H.; et al. Conditional mouse osteosarcoma, dependent on p53 loss and potentiated by loss of Rb, mimics the human disease. *Genes Dev.* 2008, 22, 1662–1676. [CrossRef] [PubMed]
- 83. Kansara, M.; Thomas, D.M. Molecular pathogenesis of osteosarcoma. DNA Cell. Biol. 2007, 26, 1–18. [CrossRef] [PubMed]
- Khanna, C.; Khan, J.; Nguyen, P.; Prehn, J.; Caylor, J.; Yeung, C.; Trepel, J.; Meltzer, P.; Helman, L. Metastasis-associated differences in gene expression in a murine model of osteosarcoma. *Cancer Res.* 2001, *61*, 3750–3759.
- Toguchida, J.; Ishizaki, K.; Sasaki, M.S.; Ikenaga, M.; Sugimoto, M.; Kotoura, Y.; Yamamuro, T. Chromosomal reorganization for the expression of recessive mutation of retinoblastoma susceptibility gene in the development of osteosarcoma. *Cancer Res.* 1988, 48, 3939–3943.
- 86. Badal, V.; Menendez, S.; Coomber, D.; Lane, D.P. Regulation of the p14ARF promoter by DNA methylation. *Cell Cycle* 2008, 7, 112–119. [CrossRef]
- 87. Oh, J.H.; Kim, H.S.; Kim, H.H.; Kim, W.H.; Lee, S.H. Aberrant methylation of p14ARF gene correlates with poor survival in osteosarcoma. *Clin. Orthop. Relat. Res.* 2006, 442, 216–222. [CrossRef]
- Al-Romaih, K.; Somers, G.R.; Bayani, J.; Hughes, S.; Prasad, M.; Cutz, J.C.; Xue, H.; Zielenska, M.; Wang, Y.; Squire, J.A. Modulation by decitabine of gene expression and growth of osteosarcoma U2OS cells in vitro and in xenografts: Identification of apoptotic genes as targets for demethylation. *Cancer Cell. Int.* 2007, 7, 14. [CrossRef]
- Rathi, A.; Virmani, A.K.; Harada, K.; Timmons, C.F.; Miyajima, K.; Hay, R.J.; Mastrangelo, D.; Maitra, A.; Tomlinson, G.E.; Gazdar, A.F. Aberrant methylation of the HIC1 promoter is a frequent event in specific pediatric neoplasms. *Clin. Cancer Res.* 2003, 9, 3674–3678.
- Cheung, N.K.; Zhang, J.; Lu, C.; Parker, M.; Bahrami, A.; Tickoo, S.K.; Heguy, A.; Pappo, A.S.; Federico, S.; Dalton, J.; et al. Association of age at diagnosis and genetic mutations in patients with neuroblastoma. *JAMA* 2012, 307, 1062–1071. [CrossRef]
- 91. Heaphy, C.M.; de Wilde, R.F.; Jiao, Y.; Klein, A.P.; Edil, B.H.; Shi, C.; Bettegowda, C.; Rodriguez, F.J.; Eberhart, C.G.; Hebbar, S.; et al. Altered telomeres in tumors with ATRX and DAXX mutations. *Science* **2011**, *333*, 425. [CrossRef] [PubMed]
- Entz-Werle, N.; Lavaux, T.; Metzger, N.; Stoetzel, C.; Lasthaus, C.; Marec, P.; Kalifa, C.; Brugieres, L.; Pacquement, H.; Schmitt, C.; et al. Involvement of MET/TWIST/APC combination or the potential role of ossification factors in pediatric high-grade osteosarcoma oncogenesis. *Neoplasia* 2007, 9, 678–688. [CrossRef]
- Harada, K.; Toyooka, S.; Maitra, A.; Maruyama, R.; Toyooka, K.O.; Timmons, C.F.; Tomlinson, G.E.; Mastrangelo, D.; Hay, R.J.; Minna, J.D.; et al. Aberrant promoter methylation and silencing of the RASSF1A gene in pediatric tumors and cell lines. *Oncogene* 2002, 21, 4345–4349. [CrossRef] [PubMed]
- Hou, P.; Ji, M.; Yang, B.; Chen, Z.; Qiu, J.; Shi, X.; Lu, Z. Quantitative analysis of promoter hypermethylation in multiple genes in osteosarcoma. *Cancer* 2006, 106, 1602–1609. [CrossRef] [PubMed]
- 95. Kansara, M.; Tsang, M.; Kodjabachian, L.; Sims, N.A.; Trivett, M.K.; Ehrich, M.; Dobrovic, A.; Slavin, J.; Choong, P.F.; Simmons, P.J.; et al. Wnt inhibitory factor 1 is epigenetically silenced in human osteosarcoma, and targeted disruption accelerates osteosarcomagenesis in mice. J. Clin. Investig. 2009, 119, 837–851. [CrossRef]
- 96. Shao, Y.W.; Wood, G.A.; Lu, J.; Tang, Q.L.; Liu, J.; Molyneux, S.; Chen, Y.; Fang, H.; Adissu, H.; McKee, T.; et al. Cross-species genomics identifies DLG2 as a tumor suppressor in osteosarcoma. *Oncogene* **2019**, *38*, 291–298. [CrossRef]
- Yamaguchi, T.; Toguchida, J.; Yamamuro, T.; Kotoura, Y.; Takada, N.; Kawaguchi, N.; Kaneko, Y.; Nakamura, Y.; Sasaki, M.S.; Ishizaki, K. Allelotype analysis in osteosarcomas: Frequent allele loss on 3q, 13q, 17p, and 18q. *Cancer Res.* 1992, 52, 2419–2423.
- Lee, M.; Oprea-Ilies, G.; Saavedra, H.I. Silencing of E2F3 suppresses tumor growth of Her2+ breast cancer cells by restricting mitosis. Oncotarget 2015, 6, 37316–37334. [CrossRef]
- Martin, J.W.; Yoshimoto, M.; Ludkovski, O.; Thorner, P.S.; Zielenska, M.; Squire, J.A.; Nuin, P.A. Analysis of segmental duplications, mouse genome synteny and recurrent cancer-associated amplicons in human chromosome 6p21-p12. *Cytogenet. Genome Res.* 2010, 128, 199–213. [CrossRef]
- Mejia-Guerrero, S.; Quejada, M.; Gokgoz, N.; Gill, M.; Parkes, R.K.; Wunder, J.S.; Andrulis, I.L. Characterization of the 12q15 MDM2 and 12q13-14 CDK4 amplicons and clinical correlations in osteosarcoma. *Genes Chromosom. Cancer* 2010, 49, 518–525.
- 101. Smida, J.; Baumhoer, D.; Rosemann, M.; Walch, A.; Bielack, S.; Poremba, C.; Remberger, K.; Korsching, E.; Scheurlen, W.; Dierkes, C.; et al. Genomic alterations and allelic imbalances are strong prognostic predictors in osteosarcoma. *Clin. Cancer Res.* 2010, 16, 4256–4267. [CrossRef] [PubMed]
- 102. Lonardo, F.; Ueda, T.; Huvos, A.G.; Healey, J.; Ladanyi, M. p53 and MDM2 alterations in osteosarcomas: Correlation with clinicopathologic features and proliferative rate. *Cancer* **1997**, *79*, 1541–1547. [CrossRef]

- Li, X.X.; Shi, L.; Zhou, X.J.; Wu, J.; Xia, T.S.; Zhou, W.B.; Sun, X.; Zhu, L.; Wei, J.F.; Ding, Q. The role of c-Myc-RBM38 loop in the growth suppression in breast cancer. J. Exp. Clin. Cancer Res. 2017, 36, 49. [CrossRef]
- 104. Lin, X.; Sun, R.; Zhao, X.; Zhu, D.; Zhao, X.; Gu, Q.; Dong, X.; Zhang, D.; Zhang, Y.; Li, Y.; et al. C-myc overexpression drives melanoma metastasis by promoting vasculogenic mimicry via c-myc/snail/Bax signaling. J. Mol. Med. 2017, 95, 53–67. [CrossRef] [PubMed]
- 105. Pennanen, M.; Hagstrom, J.; Heiskanen, I.; Sane, T.; Mustonen, H.; Arola, J.; Haglund, C. C-myc expression in adrenocortical tumours. *J. Clin. Pathol.* **2018**, *71*, 129–134. [CrossRef]
- 106. Dang, C.V. MYC on the path to cancer. Cell 2012, 149, 22-35. [CrossRef]
- 107. Steiner, P.; Philipp, A.; Lukas, J.; Godden-Kent, D.; Pagano, M.; Mittnacht, S.; Bartek, J.; Eilers, M. Identification of a Mycdependent step during the formation of active G1 cyclin-cdk complexes. *EMBO J.* **1995**, *14*, 4814–4826. [CrossRef]
- Pourdehnad, M.; Truitt, M.L.; Siddiqi, I.N.; Ducker, G.S.; Shokat, K.M.; Ruggero, D. Myc and mTOR converge on a common node in protein synthesis control that confers synthetic lethality in Myc-driven cancers. *Proc. Natl. Acad. Sci. USA* 2013, 110, 11988–11993. [CrossRef]
- Baudino, T.A.; McKay, C.; Pendeville-Samain, H.; Nilsson, J.A.; Maclean, K.H.; White, E.L.; Davis, A.C.; Ihle, J.N.; Cleveland, J.L. c-Myc is essential for vasculogenesis and angiogenesis during development and tumor progression. *Genes Dev.* 2002, 16, 2530–2543. [CrossRef]
- 110. Chen, B.J.; Wu, Y.L.; Tanaka, Y.; Zhang, W. Small molecules targeting c-Myc oncogene: Promising anti-cancer therapeutics. *Int. J. Biol. Sci.* **2014**, *10*, 1084–1096. [CrossRef]
- 111. Gamberi, G.; Benassi, M.S.; Bohling, T.; Ragazzini, P.; Molendini, L.; Sollazzo, M.R.; Pompetti, F.; Merli, M.; Magagnoli, G.; Balladelli, A.; et al. C-myc and c-fos in human osteosarcoma: Prognostic value of mRNA and protein expression. *Oncology* **1998**, 55, 556–563. [CrossRef] [PubMed]
- 112. Sadikovic, B.; Thorner, P.; Chilton-Macneill, S.; Martin, J.W.; Cervigne, N.K.; Squire, J.; Zielenska, M. Expression analysis of genes associated with human osteosarcoma tumors shows correlation of RUNX2 overexpression with poor response to chemotherapy. BMC Cancer 2010, 10, 202. [CrossRef] [PubMed]
- Squire, J.A.; Pei, J.; Marrano, P.; Beheshti, B.; Bayani, J.; Lim, G.; Moldovan, L.; Zielenska, M. High-resolution mapping of amplifications and deletions in pediatric osteosarcoma by use of CGH analysis of cDNA microarrays. *Genes Chromosom. Cancer* 2003, 38, 215–225. [CrossRef] [PubMed]
- 114. van Dartel, M.; Cornelissen, P.W.; Redeker, S.; Tarkkanen, M.; Knuutila, S.; Hogendoorn, P.C.; Westerveld, A.; Gomes, I.; Bras, J.; Hulsebos, T.J. Amplification of 17p11.2 approximately p12, including PMP22, TOP3A, and MAPK7, in high-grade osteosarcoma. *Cancer Genet. Cytogenet.* 2002, 139, 91–96. [CrossRef]
- Li, X.; Jiang, H.; Xiao, L.; Wang, S.; Zheng, J. miR-200bc/429 Inhibits Osteosarcoma Cell Proliferation and Invasion by Targeting PMP22. *Med. Sci. Monit.* 2017, 23, 1001–1008. [CrossRef]
- 116. Liu, S.; Chen, Z. The Functional Role of PMP22 Gene in the Proliferation and Invasion of Osteosarcoma. *Med. Sci. Monit.* **2015**, *21*, 1976–1982.
- 117. Lamoureux, F.; Baud'huin, M.; Rodriguez Calleja, L.; Jacques, C.; Berreur, M.; Redini, F.; Lecanda, F.; Bradner, J.E.; Heymann, D.; Ory, B. Selective inhibition of BET bromodomain epigenetic signalling interferes with the bone-associated tumour vicious cycle. *Nat. Commun.* 2014, *5*, 3511. [CrossRef]
- 118. Lu, X.Y.; Lu, Y.; Zhao, Y.J.; Jaeweon, K.; Kang, J.; Xiao-Nan, L.; Ge, G.; Meyer, R.; Perlaky, L.; Hicks, J.; et al. Cell cycle regulator gene CDC5L, a potential target for 6p12-p21 amplicon in osteosarcoma. *Mol. Cancer Res.* 2008, *6*, 937–946. [CrossRef]
- 119. Garcia-Castellano, J.M.; Villanueva, A.; Healey, J.H.; Sowers, R.; Cordon-Cardo, C.; Huvos, A.; Bertino, J.R.; Meyers, P.; Gorlick, R. Methylthioadenosine phosphorylase gene deletions are common in osteosarcoma. *Clin. Cancer Res.* **2002**, *8*, 782–787.
- 120. Miyazaki, S.; Nishioka, J.; Shiraishi, T.; Matsumine, A.; Uchida, A.; Nobori, T. Methylthioadenosine phosphorylase deficiency in Japanese osteosarcoma patients. *Int. J. Oncol.* **2007**, *31*, 1069–1076.
- 121. Munshi, P.N.; Lubin, M.; Bertino, J.R. 6-thioguanine: A drug with unrealized potential for cancer therapy. *Oncologist* 2014, 19, 760–765. [CrossRef] [PubMed]
- Kindler, H.L.; Burris, H.A., 3rd; Sandler, A.B.; Oliff, I.A. A phase II multicenter study of L-alanosine, a potent inhibitor of adenine biosynthesis, in patients with MTAP-deficient cancer. *Investig. New. Drugs* 2009, 27, 75–81. [CrossRef] [PubMed]
- Liu, Q.; Huang, J.; Zhou, N.; Zhang, Z.; Zhang, A.; Lu, Z.; Wu, F.; Mo, Y.Y. LncRNA loc285194 is a p53-regulated tumor suppressor. Nucleic Acids Res. 2013, 41, 4976–4987. [CrossRef] [PubMed]
- 124. Pasic, I.; Shlien, A.; Durbin, A.D.; Stavropoulos, D.J.; Baskin, B.; Ray, P.N.; Novokmet, A.; Malkin, D. Recurrent focal copy-number changes and loss of heterozygosity implicate two noncoding RNAs and one tumor suppressor gene at chromosome 3q13.31 in osteosarcoma. *Cancer Res.* 2010, 70, 160–171. [CrossRef]
- 125. Song, B.; Wang, Y.; Xi, Y.; Kudo, K.; Bruheim, S.; Botchkina, G.I.; Gavin, E.; Wan, Y.; Formentini, A.; Kornmann, M.; et al. Mechanism of chemoresistance mediated by miR-140 in human osteosarcoma and colon cancer cells. *Oncogene* 2009, 28, 4065–4074. [CrossRef]
- 126. Zhou, X.; Wei, M.; Wang, W. MicroRNA-340 suppresses osteosarcoma tumor growth and metastasis by directly targeting ROCK1. *Biochem. Biophys. Res. Commun.* **2013**, 437, 653–658. [CrossRef] [PubMed]
- 127. Poos, K.; Smida, J.; Nathrath, M.; Maugg, D.; Baumhoer, D.; Neumann, A.; Korsching, E. Structuring osteosarcoma knowledge: An osteosarcoma-gene association database based on literature mining and manual annotation. *Database* 2014, 2014, 1–9. [CrossRef]

- Morrow, J.J.; Khanna, C. Osteosarcoma Genetics and Epigenetics: Emerging Biology and Candidate Therapies. Crit. Rev. Oncog. 2015, 20, 173–197. [CrossRef]
- 129. Billiau, A.; Edy, V.G.; Heremans, H.; Van Damme, J.; Desmyter, J.; Georgiades, J.A.; De Somer, P. Human interferon: Mass production in a newly established cell line, MG-63. *Antimicrob. Agents Chemother.* **1977**, *12*, 11–15. [CrossRef]
- Fogh, J.; Fogh, J.M.; Orfeo, T. One hundred and twenty-seven cultured human tumor cell lines producing tumors in nude mice. J. Natl. Cancer Inst. 1977, 59, 221–226. [CrossRef]
- 131. Ponten, J.; Saksela, E. Two established in vitro cell lines from human mesenchymal tumours. *Int. J. Cancer* **1967**, *2*, 434–447. [CrossRef] [PubMed]
- 132. Rhim, J.S.; Park, D.K.; Arnstein, P.; Huebner, R.J.; Weisburger, E.K.; Nelson-Rees, W.A. Transformation of human cells in culture by N-methyl-N'-nitro-N-nitrosoguanidine. *Nature* 1975, 256, 751–753. [CrossRef] [PubMed]
- Samid, D.; Mandler, R. Human osteosarcoma cells transformed by ras-oncogenes: A new model for in vivo studies of pulmonary metastasis. *Clin. Biotechnol.* 1989, 1, 21–26.
- 134. Rochet, N.; Dubousset, J.; Mazeau, C.; Zanghellini, E.; Farges, M.F.; de Novion, H.S.; Chompret, A.; Delpech, B.; Cattan, N.; Frenay, M.; et al. Establishment, characterisation and partial cytokine expression profile of a new human osteosarcoma cell line (CAL 72). *Int. J. Cancer* 1999, *82*, 282–285. [CrossRef]
- 135. Peebles, P.; Trisch, T.; Papageorge, A. 727 isolation of four unusual pediatric solid tumor cell lines. *Pediatr. Res.* **1978**, *12*, 485. [CrossRef]
- 136. Roberts, W.M.; Douglass, E.C.; Peiper, S.C.; Houghton, P.J.; Look, A.T. Amplification of the gli gene in childhood sarcomas. *Cancer Res.* **1989**, *49*, 5407–5413.
- Schmidt, J.; Strauss, G.P.; Schon, A.; Luz, A.; Murray, A.B.; Melchiori, A.; Aresu, O.; Erfle, V. Establishment and characterization of osteogenic cell lines from a spontaneous murine osteosarcoma. *Differentiation* 1988, 39, 151–160. [CrossRef]
- Nitto, H.; Koshino, T.; Mitsugi, N.; Hiruma, T. Growth of a murine osteosarcoma-derived cell sarcoma increases serum immunosuppressive acidic protein levels. *Cancer J.* 1998, 11, 254–258.
- Joliat, M.J.; Umeda, S.; Lyons, B.L.; Lynes, M.A.; Shultz, L.D. Establishment and characterization of a new osteogenic cell line (MOS-J) from a spontaneous C57BL/6J mouse osteosarcoma. *In Vivo* 2002, *16*, 223–228.
- 140. Jasmin, C.; Allouche, M.; Jude, J.G.; Klein, B.; Thiery, J.P.; Perdereau, B.; Gongora, R.; Gongora, G.; Mazabraud, A. An experimental model of osteosarcomas in rats. *Sem. Hop.* **1982**, *58*, 1684–1689.
- 141. Martin, T.J.; Ingleton, P.M.; Underwood, J.C.; Michelangeli, V.P.; Hunt, N.H.; Melick, R.A. Parathyroid hormone-responsive adenylate cyclase in induced transplantable osteogenic rat sarcoma. *Nature* **1976**, *260*, 436–438. [CrossRef] [PubMed]
- 142. Fujiwara, T.; Uotani, K.; Yoshida, A.; Morita, T.; Nezu, Y.; Kobayashi, E.; Yoshida, A.; Uehara, T.; Omori, T.; Sugiu, K.; et al. Clinical significance of circulating miR-25-3p as a novel diagnostic and prognostic biomarker in osteosarcoma. *Oncotarget* 2017, *8*, 33375–33392. [CrossRef] [PubMed]
- 143. Lamoureux, F.; Richard, P.; Wittrant, Y.; Battaglia, S.; Pilet, P.; Trichet, V.; Blanchard, F.; Gouin, F.; Pitard, B.; Heymann, D.; et al. Therapeutic relevance of osteoprotegerin gene therapy in osteosarcoma: Blockade of the vicious cycle between tumor cell proliferation and bone resorption. *Cancer Res.* 2007, 67, 7308–7318. [CrossRef] [PubMed]
- 144. Liao, D.; Zhong, L.; Duan, T.; Zhang, R.H.; Wang, X.; Wang, G.; Hu, K.; Lv, X.; Kang, T. Aspirin Suppresses the Growth and Metastasis of Osteosarcoma through the NF-kappaB Pathway. *Clin. Cancer Res.* **2015**, *21*, 5349–5359. [CrossRef] [PubMed]
- 145. Martins-Neves, S.R.; Paiva-Oliveira, D.I.; Fontes-Ribeiro, C.; Bovee, J.; Cleton-Jansen, A.M.; Gomes CMF. IWR-1, a tankyrase inhibitor, attenuates Wnt/beta-catenin signaling in cancer stem-like cells and inhibits in vivo the growth of a subcutaneous human osteosarcoma xenograft. *Cancer Lett.* 2018, 414, 1–15. [CrossRef] [PubMed]
- 146. Ory, B.; Baud'huin, M.; Verrecchia, F.; Royer, B.B.; Quillard, T.; Amiaud, J.; Battaglia, S.; Heymann, D.; Redini, F.; Lamoureux, F. Blocking HSP90 Addiction Inhibits Tumor Cell Proliferation, Metastasis Development, and Synergistically Acts with Zoledronic Acid to Delay Osteosarcoma Progression. *Clin. Cancer Res.* 2016, *22*, 2520–2533. [CrossRef]
- 147. Anfinsen, K.P.; Grotmol, T.; Bruland, O.S.; Jonasdottir, T.J. Breed-specific incidence rates of canine primary bone tumors–a population based survey of dogs in Norway. *Can. J. Vet. Res.* **2011**, *75*, 209–215.
- 148. Misdorp, W. Skeletal osteosarcoma. Animal model: Canine osteosarcoma. Am. J. Pathol. 1980, 98, 285–288.
- 149. Klein, M.J.; Siegal, G.P. Osteosarcoma: Anatomic and histologic variants. Am. J. Clin. Pathol. 2006, 125, 555–581. [CrossRef]
- 150. Loukopoulos, P.; Robinson, W.F. Clinicopathological relevance of tumour grading in canine osteosarcoma. *J. Comp. Pathol.* 2007, 136, 65–73. [CrossRef]
- 151. Morello, E.; Martano, M.; Buracco, P. Biology, diagnosis and treatment of canine appendicular osteosarcoma: Similarities and differences with human osteosarcoma. *Vet. J.* **2011**, *189*, 268–277. [CrossRef] [PubMed]
- Marques, I.J.; Weiss, F.U.; Vlecken, D.H.; Nitsche, C.; Bakkers, J.; Lagendijk, A.K.; Partecke, L.I.; Heidecke, C.D.; Lerch, M.M.; Bagowski, C.P. Metastatic behaviour of primary human tumours in a zebrafish xenotransplantation model. *BMC Cancer* 2009, *9*, 128. [CrossRef] [PubMed]
- Sohail, A.; Sherin, L.; Butt, S.I.; Javed, S.; Li, Z.; Iqbal, S.; Be'g, O.A. Role of key players in paradigm shifts of prostate cancer bone metastasis. *Cancer Manag. Res.* 2018, 10, 1619–1626. [CrossRef] [PubMed]
- 154. Yang, Y.; Wang, B. PTH1R-CaSR Cross Talk: New Treatment Options for Breast Cancer Osteolytic Bone Metastases. *Int. J. Endocrinol.* 2018, 2018, 7120979. [CrossRef]

- 155. Lamora, A.; Talbot, J.; Mullard, M.; Brounais-Le Royer, B.; Redini, F.; Verrecchia, F. TGF-beta Signaling in Bone Remodeling and Osteosarcoma Progression. J. Clin. Med. 2016, 5, 96. [CrossRef]
- 156. Guise, T.A.; Mohammad, K.S.; Clines, G.; Stebbins, E.G.; Wong, D.H.; Higgins, L.S.; Vessella, R.; Corey, E.; Padalecki, S.; Suva, L.; et al. Basic mechanisms responsible for osteolytic and osteoblastic bone metastases. *Clin. Cancer Res.* 2006, 12, 6213s–6216s. [CrossRef]
- 157. Akiyama, T.; Dass, C.R.; Choong, P.F. Novel therapeutic strategy for osteosarcoma targeting osteoclast differentiation, boneresorbing activity, and apoptosis pathway. *Mol. Cancer Ther.* **2008**, *7*, 3461–3469. [CrossRef]
- 158. Clemons, M.; Gelmon, K.A.; Pritchard, K.I.; Paterson, A.H. Bone-targeted agents and skeletal-related events in breast cancer patients with bone metastases: The state of the art. *Curr. Oncol.* **2012**, *19*, 259–268. [CrossRef]
- Kim, W.; Takyar, F.M.; Swan, K.; Jeong, J.; VanHouten, J.; Sullivan, C.; Dann, P.; Yu, H.; Fiaschi-Taesch, N.; Chang, W.; et al. Calcium-Sensing Receptor Promotes Breast Cancer by Stimulating Intracrine Actions of Parathyroid Hormone-Related Protein. *Cancer Res.* 2016, 76, 5348–5360. [CrossRef]
- 160. Maurizi, A.; Rucci, N. The Osteoclast in Bone Metastasis: Player and Target. Cancers 2018, 10, 218. [CrossRef]
- Tarhini, A.A.; Kirkwood, J.M. How much of a good thing? What duration for interferon alfa-2b adjuvant therapy? J. Clin. Oncol. 2012, 30, 3773–3776. [CrossRef] [PubMed]
- 162. Shaikh, A.B.; Li, F.; Li, M.; He, B.; He, X.; Chen, G.; Guo, B.; Li, D.; Jiang, F.; Dang, L.; et al. Present Advances and Future Perspectives of Molecular Targeted Therapy for Osteosarcoma. *Int. J. Mol. Sci.* **2016**, *17*, 506. [CrossRef] [PubMed]
- 163. Stern, J.B.; Smith, K.A. Interleukin-2 induction of T-cell G1 progression and c-myb expression. *Science* **1986**, 233, 203–206. [CrossRef] [PubMed]
- 164. Meazza, C.; Cefalo, G.; Massimino, M.; Daolio, P.; Pastorino, U.; Scanagatta, P.; Morosi, C.; Podda, M.; Ferrari, A.; Terenziani, M.; et al. Primary metastatic osteosarcoma: Results of a prospective study in children given chemotherapy and interleukin-2. *Med. Oncol.* 2017, 34, 191. [CrossRef]
- 165. Schwinger, W.; Klass, V.; Benesch, M.; Lackner, H.; Dornbusch, H.J.; Sovinz, P.; Moser, A.; Schwantzer, G.; Urban, C. Feasibility of high-dose interleukin-2 in heavily pretreated pediatric cancer patients. *Ann. Oncol.* 2005, *16*, 1199–1206. [CrossRef] [PubMed]
- Harris, M.A.; Hawkins, C.J. Recent and Ongoing Research into Metastatic Osteosarcoma Treatments. Int. J. Mol. Sci. 2022, 23, 3817. [CrossRef]
- MacEwen, E.G.; Kurzman, I.D.; Rosenthal, R.C.; Smith, B.W.; Manley, P.A.; Roush, J.K.; Howard, P.E. Therapy for osteosarcoma in dogs with intravenous injection of liposome-encapsulated muramyl tripeptide. J. Natl. Cancer Inst. 1989, 81, 935–938. [CrossRef]
- 168. Meyers, P.A.; Schwartz, C.L.; Krailo, M.D.; Healey, J.H.; Bernstein, M.L.; Betcher, D.; Ferguson, W.S.; Gebhardt, M.C.; Goorin, A.M.; Harris, M.; et al. Osteosarcoma: The addition of muramyl tripeptide to chemotherapy improves overall survival—A report from the Children's Oncology Group. J. Clin. Oncol. 2008, 26, 633–638. [CrossRef]
- 169. Chou, A.J.; Kleinerman, E.S.; Krailo, M.D.; Chen, Z.; Betcher, D.L.; Healey, J.H.; Conrad, E.U., 3rd; Nieder, M.L.; Weiner, M.A.; Wells, R.J.; et al. Addition of muramyl tripeptide to chemotherapy for patients with newly diagnosed metastatic osteosarcoma: A report from the Children's Oncology Group. *Cancer* 2009, 115, 5339–5348. [CrossRef]
- Meyers, P.A. Muramyl Tripeptide-Phosphatidyl Ethanolamine Encapsulated in Liposomes (L-MTP-PE) in the Treatment of Osteosarcoma. Adv. Exp. Med. Biol. 2020, 1257, 133–139.
- 171. Sangiolo, D.; Mesiano, G.; Gammaitoni, L.; Leuci, V.; Todorovic, M.; Giraudo, L.; Cammarata, C.; Dell'Aglio, C.; D'Ambrosio, L.; Pisacane, A.; et al. Cytokine-induced killer cells eradicate bone and soft-tissue sarcomas. *Cancer Res.* 2014, 74, 119–129. [CrossRef] [PubMed]
- 172. Gammaitoni, L.; Giraudo, L.; Leuci, V.; Todorovic, M.; Mesiano, G.; Picciotto, F.; Pisacane, A.; Zaccagna, A.; Volpe, M.G.; Gallo, S.; et al. Effective activity of cytokine-induced killer cells against autologous metastatic melanoma including cells with stemness features. *Clin. Cancer Res.* 2013, *19*, 4347–4358. [CrossRef] [PubMed]
- 173. Mesiano, G.; Grignani, G.; Fiorino, E.; Leuci, V.; Rotolo, R.; D'Ambrosio, L.; Salfi, C.; Gammaitoni, L.; Giraudo, L.; Pisacane, A.; et al. Cytokine Induced Killer cells are effective against sarcoma cancer stem cells spared by chemotherapy and target therapy. Oncoimmunology 2018, 7, e1465161. [CrossRef]
- 174. Ebb, D.; Meyers, P.; Grier, H.; Bernstein, M.; Gorlick, R.; Lipshultz, S.E.; Krailo, M.; Devidas, M.; Barkauskas, D.A.; Siegal, G.P.; et al. Phase II trial of trastuzumab in combination with cytotoxic chemotherapy for treatment of metastatic osteosarcoma with human epidermal growth factor receptor 2 overexpression: A report from the children's oncology group. *J. Clin. Oncol.* 2012, 30, 2545–2551. [CrossRef] [PubMed]
- 175. Ahmed, N.; Brawley, V.S.; Hegde, M.; Robertson, C.; Ghazi, A.; Gerken, C.; Liu, E.; Dakhova, O.; Ashoori, A.; Corder, A.; et al. Human Epidermal Growth Factor Receptor 2 (HER2) -Specific Chimeric Antigen Receptor-Modified T Cells for the Immunotherapy of HER2-Positive Sarcoma. *J. Clin. Oncol.* 2015, *33*, 1688–1696. [CrossRef]
- 176. Pappo, A.S.; Vassal, G.; Crowley, J.J.; Bolejack, V.; Hogendoorn, P.C.; Chugh, R.; Ladanyi, M.; Grippo, J.F.; Dall, G.; Staddon, A.P.; et al. A phase 2 trial of R1507, a monoclonal antibody to the insulin-like growth factor-1 receptor (IGF-1R), in patients with recurrent or refractory rhabdomyosarcoma, osteosarcoma, synovial sarcoma, and other soft tissue sarcomas: Results of a Sarcoma Alliance for Research Through Collaboration study. *Cancer* 2014, *120*, 2448–2456.
- 177. Weigel, B.; Malempati, S.; Reid, J.M.; Voss, S.D.; Cho, S.Y.; Chen, H.X.; Krailo, M.; Villaluna, D.; Adamson, P.C.; Blaney, S.M. Phase 2 trial of cixutumumab in children, adolescents, and young adults with refractory solid tumors: A report from the Children's Oncology Group. *Pediatr. Blood. Cancer* 2014, 61, 452–456. [CrossRef]

- 178. Malempati, S.; Weigel, B.; Ingle, A.M.; Ahern, C.H.; Carroll, J.M.; Roberts, C.T.; Reid, J.M.; Schmechel, S.; Voss, S.D.; Cho, S.Y.; et al. Phase I/II trial and pharmacokinetic study of cixutumumab in pediatric patients with refractory solid tumors and Ewing sarcoma: A report from the Children's Oncology Group. *J. Clin. Oncol.* **2012**, *30*, 256–262. [CrossRef]
- 179. Poon, V.I.; Roth, M.; Piperdi, S.; Geller, D.; Gill, J.; Rudzinski, E.R.; Hawkins, D.S.; Gorlick, R. Ganglioside GD2 expression is maintained upon recurrence in patients with osteosarcoma. *Clin. Sarcoma Res.* **2015**, *5*, 4. [CrossRef]
- Nazha, B.; Inal, C.; Owonikoko, T.K. Disialoganglioside GD2 Expression in Solid Tumors and Role as a Target for Cancer Therapy. Front. Oncol. 2020, 10, 1000. [CrossRef]
- 181. Hingorani, P.; Krailo, M.D.; Buxton, A.; Hutson, P.R.; Davis, J.; Janeway, K.A.; Gorlick, R.G.; Isakoff, M. Phase II study of antidisialoganglioside antibody, dinutuximab, in combination with GM-CSF in patients with recurrent osteosarcoma (AOST1421): A report from the Children's Oncology Group. J. Clin. Oncol. 2020, 38, 10508. [CrossRef]
- 182. Wedekind, M.F.; Wagner, L.M.; Cripe, T.P. Immunotherapy for osteosarcoma: Where do we go from here? *Pediatr. Blood Cancer* **2018**, 65, e27227. [CrossRef]
- 183. Majzner, R.G.; Heitzeneder, S.; Mackall, C.L. Harnessing the Immunotherapy Revolution for the Treatment of Childhood Cancers. *Cancer Cell* **2017**, *31*, 476–485. [PubMed]
- Park, J.A.; Santich, B.H.; Xu, H.; Lum, L.G.; Cheung, N.V. Potent ex vivo armed T cells using recombinant bispecific antibodies for adoptive immunotherapy with reduced cytokine release. *J. Immunother. Cancer* 2021, *9*, e002222. [CrossRef] [PubMed]
- 185. Purcell, J.W.; Tanlimco, S.G.; Hickson, J.; Fox, M.; Sho, M.; Durkin, L.; Uziel, T.; Powers, R.; Foster, K.; McGonigal, T.; et al. LRRC15 Is a Novel Mesenchymal Protein and Stromal Target for Antibody-Drug Conjugates. *Cancer Res.* 2018, 78, 4059–4072. [CrossRef]
- 186. Wang, Y.; Liu, Y.; Zhang, M.; Lv, L.; Zhang, X.; Zhang, P.; Zhou, Y. LRRC15 promotes osteogenic differentiation of mesenchymal stem cells by modulating p65 cytoplasmic/nuclear translocation. *Stem Cell Res. Ther.* **2018**, *9*, 65. [CrossRef]
- 187. Demetri, G.D.; Luke, J.J.; Hollebecque, A.; Powderly, J.D., 2nd; Spira, A.I.; Subbiah, V.; Naumovski, L.; Chen, C.; Fang, H.; Lai, D.W.; et al. First-in-Human Phase I Study of ABBV-085, an Antibody-Drug Conjugate Targeting LRRC15, in Sarcomas and Other Advanced Solid Tumors. *Clin. Cancer Res.* 2021, 27, 3556–3566. [CrossRef]
- 188. Dong, P.; Xiong, Y.; Yue, J.; Hanley, S.J.B.; Watari, H. B7H3 As a Promoter of Metastasis and Promising Therapeutic Target. *Front. Oncol.* **2018**, *8*, 264. [CrossRef]
- 189. Polito, L.; Calafato, G.; Bortolotti, M.; Chiarelli Olivari, C.; Maiello, S.; Bolognesi, A. Antibody Conjugates for Sarcoma Therapy: How Far along Are We? *Biomedicines* **2021**, *9*, 978. [CrossRef]
- Byers, V.S.; Pawluczyk, I.Z.; Hooi, D.S.; Price, M.R.; Carroll, S.; Embleton, M.J.; Garnett, M.C.; Berry, N.; Robins, R.A.; Baldwin, R.W. Endocytosis of immunotoxin-791T/36-RTA by tumor cells in relation to its cytotoxic action. *Cancer Res.* 1991, 51, 1990–1995.
- 191. Garnett, M.C.; Baldwin, R.W. An improved synthesis of a methotrexate-albumin-791T/36 monoclonal antibody conjugate cytotoxic to human osteogenic sarcoma cell lines. *Cancer Res.* **1986**, *46*, 2407–2412. [PubMed]
- Anderson, P.M.; Meyers, D.E.; Hasz, D.E.; Covalcuic, K.; Saltzman, D.; Khanna, C.; Uckun, F.M. In vitro and in vivo cytotoxicity of an anti-osteosarcoma immunotoxin containing pokeweed antiviral protein. *Cancer Res.* 1995, 55, 1321–1327. [PubMed]
- 193. Westrom, S.; Bonsdorff, T.B.; Abbas, N.; Bruland, O.S.; Jonasdottir, T.J.; Maelandsmo, G.M.; Larsen, R.H. Evaluation of CD146 as Target for Radioimmunotherapy against Osteosarcoma. *PLoS ONE* **2016**, *11*, e0165382. [CrossRef]
- Hassan, S.E.; Bekarev, M.; Kim, M.Y.; Lin, J.; Piperdi, S.; Gorlick, R.; Geller, D.S. Cell surface receptor expression patterns in osteosarcoma. *Cancer* 2012, 118, 740–749. [CrossRef] [PubMed]
- 195. Broqueza, J.; Prabaharan, C.B.; Andrahennadi, S.; Allen, K.J.H.; Dickinson, R.; MacDonald-Dickinson, V.; Dadachova, E.; Uppalapati, M. Novel Human Antibodies to Insulin Growth Factor 2 Receptor (IGF2R) for Radioimmunoimaging and Therapy of Canine and Human Osteosarcoma. *Cancers* 2021, *13*, 2208. [CrossRef] [PubMed]
- 196. Karkare, S.; Allen, K.J.H.; Jiao, R.; Malo, M.E.; Dawicki, W.; Helal, M.; Godson, D.L.; Dickinson, R.; MacDonald-Dickinson, V.; Yang, R.; et al. Detection and targeting insulin growth factor receptor type 2 (IGF2R) in osteosarcoma PDX in mouse models and in canine osteosarcoma tumors. *Sci. Rep.* 2019, *9*, 11476. [CrossRef] [PubMed]
- 197. Wagner, L.M.; Adams, V.R. Targeting the PD-1 pathway in pediatric solid tumors and brain tumors. *OncoTargets. Ther.* **2017**, *10*, 2097–2106. [CrossRef]
- 198. Hingorani, P.; Maas, M.L.; Gustafson, M.P.; Dickman, P.; Adams, R.H.; Watanabe, M.; Eshun, F.; Williams, J.; Seidel, M.J.; Dietz, A.B. Increased CTLA-4(+) T cells and an increased ratio of monocytes with loss of class II (CD14(+) HLA-DR(lo/neg)) found in aggressive pediatric sarcoma patients. *J. Immunother. Cancer* 2015, *3*, 35. [CrossRef]
- 199. Callahan, M.K.; Postow, M.A.; Wolchok, J.D. CTLA-4 and PD-1 Pathway Blockade: Combinations in the Clinic. *Front. Oncol.* 2014, 4, 385. [CrossRef]
- Rodig, S.J.; Gusenleitner, D.; Jackson, D.G.; Gjini, E.; Giobbie-Hurder, A.; Jin, C.; Chang, H.; Lovitch, S.B.; Horak, C.; Weber, J.S.; et al. MHC proteins confer differential sensitivity to CTLA-4 and PD-1 blockade in untreated metastatic melanoma. *Sci. Transl. Med.* 2018, *10*, 450. [CrossRef]
- 201. Ma, L.; Dichwalkar, T.; Chang, J.Y.H.; Cossette, B.; Garafola, D.; Zhang, A.Q.; Fichter, M.; Wang, C.; Liang, S.; Silva, M.; et al. Enhanced CAR-T cell activity against solid tumors by vaccine boosting through the chimeric receptor. *Science* 2019, 365, 162–168. [CrossRef] [PubMed]
- Merchant, M.S.; Wright, M.; Baird, K.; Wexler, L.H.; Rodriguez-Galindo, C.; Bernstein, D.; Delbrook, C.; Lodish, M.; Bishop, R.; Wolchok, J.D.; et al. Phase I Clinical Trial of Ipilimumab in Pediatric Patients with Advanced Solid Tumors. *Clin. Cancer Res.* 2016, 22, 1364–1370. [CrossRef] [PubMed]

- 203. Pauken, K.E.; Wherry, E.J. Overcoming T cell exhaustion in infection and cancer. *Trends. Immunol.* 2015, *36*, 265–276. [CrossRef] [PubMed]
- Nowicki, T.S.; Anderson, J.L.; Federman, N. Prospective immunotherapies in childhood sarcomas: PD1/PDL1 blockade in combination with tumor vaccines. *Pediatr. Res.* 2016, 79, 371–377. [CrossRef] [PubMed]
- 205. Shen, J.K.; Cote, G.M.; Choy, E.; Yang, P.; Harmon, D.; Schwab, J.; Nielsen, G.P.; Chebib, I.; Ferrone, S.; Wang, X.; et al. Programmed cell death ligand 1 expression in osteosarcoma. *Cancer Immunol. Res.* 2014, 2, 690–698. [CrossRef]
- 206. Palmerini, E.; Agostinelli, C.; Picci, P.; Pileri, S.; Marafioti, T.; Lollini, P.L.; Scotlandi, K.; Longhi, A.; Benassi, M.S.; Ferrari, S. Tumoral immune-infiltrate (IF), PD-L1 expression and role of CD8/TIA-1 lymphocytes in localized osteosarcoma patients treated within protocol ISG-OS1. *Oncotarget* 2017, *8*, 111836–111846. [CrossRef]
- 207. Koirala, P.; Roth, M.E.; Gill, J.; Piperdi, S.; Chinai, J.M.; Geller, D.S.; Hoang, B.H.; Park, A.; Fremed, M.A.; Zang, X.; et al. Immune infiltration and PD-L1 expression in the tumor microenvironment are prognostic in osteosarcoma. *Sci. Rep.* 2016, *6*, 30093. [CrossRef]
- Lussier, D.M.; O'Neill, L.; Nieves, L.M.; McAfee, M.S.; Holechek, S.A.; Collins, A.W.; Dickman, P.; Jacobsen, J.; Hingorani, P.; Blattman, J.N. Enhanced T-cell immunity to osteosarcoma through antibody blockade of PD-1/PD-L1 interactions. *J. Immunother.* 2015, 38, 96–106. [CrossRef]
- 209. Roberts, S.S.; Chou, A.J.; Cheung, N.K. Immunotherapy of Childhood Sarcomas. Front. Oncol. 2015, 5, 181. [CrossRef] [PubMed]
- 210. Zheng, B.; Ren, T.; Huang, Y.; Sun, K.; Wang, S.; Bao, X.; Liu, K.; Guo, W. PD-1 axis expression in musculoskeletal tumors and antitumor effect of nivolumab in osteosarcoma model of humanized mouse. *J. Hematol. Oncol.* **2018**, *11*, 16.
- 211. Tawbi, H.A.; Burgess, M.; Bolejack, V.; Van Tine, B.A.; Schuetze, S.M.; Hu, J.; D'Angelo, S.; Attia, S.; Riedel, R.F.; Priebat, D.A.; et al. Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): A multicentre, two-cohort, single-arm, open-label, phase 2 trial. *Lancet Oncol.* 2017, 18, 1493–1501. [CrossRef]
- 212. Bishop, M.W.; Kaste, S.C.; Sykes, A.; Pan, H.; Cruz, F.S.D.; Whittle, S.; Mascarenhas, L.; Thomas, P.G.; Youngblood, B.; Harman, J.L.; et al. OSTPDL1: A phase II study of avelumab, a monoclonal antibody targeting programmed death-ligand 1 (PD-L1) in adolescent and young adult patients with recurrent or progressive osteosarcoma. *J. Clin. Oncol.* 2020, *38*, 10521. [CrossRef]
- 213. Maki, R.G.; Jungbluth, A.A.; Gnjatic, S.; Schwartz, G.K.; D'Adamo, D.R.; Keohan, M.L.; Wagner, M.J.; Scheu, K.; Chiu, R.; Ritter, E.; et al. A Pilot Study of Anti-CTLA4 Antibody Ipilimumab in Patients with Synovial Sarcoma. *Sarcoma* 2013, 2013, 168145. [CrossRef] [PubMed]
- 214. Kim, J.R.; Moon, Y.J.; Kwon, K.S.; Bae, J.S.; Wagle, S.; Kim, K.M.; Park, H.S.; Lee, H.; Moon, W.S.; Chung, M.J.; et al. Tumor infiltrating PD1-positive lymphocytes and the expression of PD-L1 predict poor prognosis of soft tissue sarcomas. *PLoS ONE* 2013, *8*, e82870. [CrossRef] [PubMed]
- 215. Lee, J.A.; Jung, J.S.; Kim, D.H.; Lim, J.S.; Kim, M.S.; Kong, C.B.; Song, W.S.; Cho, W.H.; Jeon, D.G.; Lee, S.Y.; et al. RANKL expression is related to treatment outcome of patients with localized, high-grade osteosarcoma. *Pediatr. Blood. Cancer* 2011, 56, 738–743. [CrossRef] [PubMed]
- Trieb, K.; Windhager, R. Receptor activator of nuclear factor kappaB expression is a prognostic factor in human osteosarcoma. Oncol. Lett. 2015, 10, 1813–1815. [CrossRef]
- de Groot, A.F.; Appelman-Dijkstra, N.M.; van der Burg, S.H.; Kroep, J.R. The anti-tumor effect of RANKL inhibition in malignant solid tumors—A systematic review. *Cancer Treat. Rev.* 2018, 62, 18–28. [CrossRef]
- 218. Heymann, D. Anti-RANKL therapy for bone tumours: Basic, pre-clinical and clinical evidences. *J. Bone Oncol.* 2012, *1*, 2–11. [CrossRef]
- Kupas, V.; Weishaupt, C.; Siepmann, D.; Kaserer, M.L.; Eickelmann, M.; Metze, D.; Luger, T.A.; Beissert, S.; Loser, K. RANK is expressed in metastatic melanoma and highly upregulated on melanoma-initiating cells. *J. Investig. Dermatol.* 2011, 131, 944–955. [CrossRef]
- 220. Lamoureux, F.; Trichet, V.; Chipoy, C.; Blanchard, F.; Gouin, F.; Redini, F. Recent advances in the management of osteosarcoma and forthcoming therapeutic strategies. *Expert. Rev. Anticancer Ther.* **2007**, *7*, 169–181. [CrossRef]
- 221. Stopeck, A.T.; Lipton, A.; Body, J.J.; Steger, G.G.; Tonkin, K.; de Boer, R.H.; Lichinitser, M.; Fujiwara, Y.; Yardley, D.A.; Viniegra, M.; et al. Denosumab compared with zoledronic acid for the treatment of bone metastases in patients with advanced breast cancer: A randomized, double-blind study. J. Clin. Oncol. 2010, 28, 5132–5139. [CrossRef] [PubMed]
- Savvidou, O.D.; Bolia, I.K.; Chloros, G.D.; Papanastasiou, J.; Koutsouradis, P.; Papagelopoulos, P.J. Denosumab: Current Use in the Treatment of Primary Bone Tumors. *Orthopedics* 2017, 40, 204–210. [CrossRef] [PubMed]
- 223. Dhesy-Thind, S.; Fletcher, G.G.; Blanchette, P.S.; Clemons, M.J.; Dillmon, M.S.; Frank, E.S.; Gandhi, S.; Gupta, R.; Mates, M.; Moy, B.; et al. Use of Adjuvant Bisphosphonates and Other Bone-Modifying Agents in Breast Cancer: A Cancer Care Ontario and American Society of Clinical Oncology Clinical Practice Guideline. J. Clin. Oncol. 2017, 35, 2062–2081. [CrossRef]
- 224. Fleisch, H. Development of bisphosphonates. *Breast Cancer Res.* **2002**, *4*, 30–34. [CrossRef]
- 225. Hortobagyi, G.N.; Van Poznak, C.; Harker, W.G.; Gradishar, W.J.; Chew, H.; Dakhil, S.R.; Haley, B.B.; Sauter, N.; Mohanlal, R.; Zheng, M.; et al. Continued Treatment Effect of Zoledronic Acid Dosing Every 12 vs. 4 Weeks in Women With Breast Cancer Metastatic to Bone: The OPTIMIZE-2 Randomized Clinical Trial. JAMA Oncol. 2017, 3, 906–912. [CrossRef]
- 226. Landre, T.; Guetz, G.D.; Chouahnia, K.; Fossey-Diaz, V.; Taleb, C.; Culine, S. Is There a Benefit of Addition Docetaxel, Abiraterone, Celecoxib, or Zoledronic Acid in Initial Treatments for Patients Older Than 70 Years With Hormone-sensitive Advanced Prostate Cancer? A Meta-analysis. *Clin. Genitourin. Cancer* 2019, 17, e806–e813. [CrossRef]

- 227. Ory, B.; Blanchard, F.; Battaglia, S.; Gouin, F.; Redini, F.; Heymann, D. Zoledronic acid activates the DNA S-phase checkpoint and induces osteosarcoma cell death characterized by apoptosis-inducing factor and endonuclease-G translocation independently of p53 and retinoblastoma status. *Mol. Pharmacol.* **2007**, *71*, 333–343. [CrossRef]
- 228. Muraro, M.; Mereuta, O.M.; Carraro, F.; Madon, E.; Fagioli, F. Osteosarcoma cell line growth inhibition by zoledronate-stimulated effector cells. *Cell. Immunol.* 2007, 249, 63–72. [CrossRef]
- Kim, E.H.; Kim, M.S.; Lee, K.H.; Koh, J.S.; Jung, W.G.; Kong, C.B. Zoledronic acid is an effective radiosensitizer in the treatment of osteosarcoma. Oncotarget 2016, 7, 70869–70880. [CrossRef]
- Heymann, D.; Ory, B.; Blanchard, F.; Heymann, M.F.; Coipeau, P.; Charrier, C.; Couillaud, S.; Thiery, J.P.; Gouin, F.; Redini, F. Enhanced tumor regression and tissue repair when zoledronic acid is combined with ifosfamide in rat osteosarcoma. *Bone* 2005, 37, 74–86. [CrossRef] [PubMed]
- Ory, B.; Heymann, M.F.; Kamijo, A.; Gouin, F.; Heymann, D.; Redini, F. Zoledronic acid suppresses lung metastases and prolongs overall survival of osteosarcoma-bearing mice. *Cancer* 2005, 104, 2522–2529. [CrossRef] [PubMed]
- 232. Dass, C.R.; Choong, P.F. Zoledronic acid inhibits osteosarcoma growth in an orthotopic model. *Mol. Cancer Ther.* 2007, *6*, 3263–3270. [CrossRef] [PubMed]
- Koto, K.; Horie, N.; Kimura, S.; Murata, H.; Sakabe, T.; Matsui, T.; Watanabe, M.; Adachi, S.; Maekawa, T.; Fushiki, S.; et al. Clinically relevant dose of zoledronic acid inhibits spontaneous lung metastasis in a murine osteosarcoma model. *Cancer Lett.* 2009, 274, 271–278. [CrossRef] [PubMed]
- Tian, Z.; Niu, X.; Yao, W. Receptor Tyrosine Kinases in Osteosarcoma Treatment: Which Is the Key Target? Front. Oncol. 2020, 10, 1642. [CrossRef]
- 235. Segaliny, A.I.; Tellez-Gabriel, M.; Heymann, M.F.; Heymann, D. Receptor tyrosine kinases: Characterisation, mechanism of action and therapeutic interests for Bone cancers. *J. Bone Oncol.* **2015**, *4*, 1–12. [CrossRef]
- 236. Chen, H.X.; Sharon, E. IGF-1R as an anti-cancer target-trials and tribulations. Chin. J. Cancer 2013, 32, 242–252. [CrossRef]
- Han, J.; Tian, R.; Yong, B.; Luo, C.; Tan, P.; Shen, J.; Peng, T. Gas6/Axl mediates tumor cell apoptosis, migration and invasion and predicts the clinical outcome of osteosarcoma patients. *Biochem. Biophys. Res. Commun.* 2013, 435, 493–500. [CrossRef] [PubMed]
- Wang, K.; Zhuang, Y.; Liu, C.; Li, Y. Inhibition of c-Met activation sensitizes osteosarcoma cells to cisplatin via suppression of the PI3K-Akt signaling. Arch. Biochem. Biophys. 2012, 526, 38–43. [CrossRef]
- Jentzsch, T.; Robl, B.; Husmann, M.; Bode-Lesniewska, B.; Fuchs, B. Worse prognosis of osteosarcoma patients expressing IGF-1 on a tissue microarray. *Anticancer Res.* 2014, 34, 3881–3889.
- Zhang, Y.; Tang, Y.J.; Man, Y.; Pan, F.; Li, Z.H.; Jia, L.S. Knockdown of AXL receptor tyrosine kinase in osteosarcoma cells leads to decreased proliferation and increased apoptosis. *Int. J. Immunopathol. Pharmacol.* 2013, 26, 179–188. [CrossRef]
- 241. Jiao, Q.; Bi, L.; Ren, Y.; Song, S.; Wang, Q.; Wang, Y.S. Advances in studies of tyrosine kinase inhibitors and their acquired resistance. *Mol. Cancer* 2018, 17, 36. [CrossRef] [PubMed]
- Knosel, T.; Kampmann, E.; Kirchner, T.; Altendorf-Hofmann, A. Tyrosine kinases in soft tissue tumors. *Pathologe* 2014, 35 (Suppl. S2), 198–201. [PubMed]
- Miiji, L.N.; Petrilli, A.S.; Di Cesare, S.; Odashiro, A.N.; Burnier, M.N., Jr.; de Toledo, S.R.; Garcia, R.J.; Alves, M.T. C-kit expression in human osteosarcoma and in vitro assays. *Int. J. Clin. Exp. Pathol.* 2011, 4, 775–781. [PubMed]
- Luo, J.; Xia, Y.; Yin, Y.; Luo, J.; Liu, M.; Zhang, H.; Zhang, C.; Zhao, Y.; Yang, L.; Kong, L. ATF4 destabilizes RET through nonclassical GRP78 inhibition to enhance chemosensitivity to bortezomib in human osteosarcoma. *Theranostics* 2019, 9, 6334–6353. [CrossRef] [PubMed]
- 245. Kim, M.; Jung, J.Y.; Choi, S.; Lee, H.; Morales, L.D.; Koh, J.T.; Kim, S.H.; Choi, Y.D.; Choi, C.; Slaga, T.J.; et al. GFRA1 promotes cisplatin-induced chemoresistance in osteosarcoma by inducing autophagy. *Autophagy* 2017, 13, 149–168. [CrossRef]
- 246. Fernanda Amary, M.; Ye, H.; Berisha, F.; Khatri, B.; Forbes, G.; Lehovsky, K.; Frezza, A.M.; Behjati, S.; Tarpey, P.; Pillay, N.; et al. Fibroblastic growth factor receptor 1 amplification in osteosarcoma is associated with poor response to neo-adjuvant chemotherapy. *Cancer Med.* 2014, 3, 980–987. [CrossRef]
- 247. Weekes, D.; Kashima, T.G.; Zandueta, C.; Perurena, N.; Thomas, D.P.; Sunters, A.; Vuillier, C.; Bozec, A.; El-Emir, E.; Miletich, I.; et al. Regulation of osteosarcoma cell lung metastasis by the c-Fos/AP-1 target FGFR1. *Oncogene* **2016**, *35*, 2852–2861. [CrossRef]
- 248. Takagi, S.; Takemoto, A.; Takami, M.; Oh-Hara, T.; Fujita, N. Platelets promote osteosarcoma cell growth through activation of the platelet-derived growth factor receptor-Akt signaling axis. *Cancer Sci.* **2014**, *105*, 983–988. [CrossRef]
- Zhang, D.; Cui, G.; Sun, C.; Lei, L.; Lei, L.; Williamson, R.A.; Wang, Y.; Zhang, J.; Chen, P.; Wang, A.; et al. Hypoxia promotes osteosarcoma cell proliferation and migration through enhancing platelet-derived growth factor-BB/platelet-derived growth factor receptor-beta axis. *Biochem. Biophys. Res. Commun.* 2019, 512, 360–366. [CrossRef]
- Coltella, N.; Manara, M.C.; Cerisano, V.; Trusolino, L.; Di Renzo, M.F.; Scotlandi, K.; Ferracini, R. Role of the MET/HGF receptor in proliferation and invasive behavior of osteosarcoma. *FASEB J.* 2003, 17, 1162–1164. [CrossRef]
- Zhou, Q.; Hu, T.; Xu, Y. Anticancer potential of TUG1 knockdown in cisplatin-resistant osteosarcoma through inhibition of MET/Akt signalling. J. Drug Target 2020, 28, 204–211. [CrossRef] [PubMed]
- 252. Xie, W.; Xiao, J.; Wang, T.; Zhang, D.; Li, Z. MicroRNA-876-5p inhibits cell proliferation, migration and invasion by targeting c-Met in osteosarcoma. *J. Cell. Mol. Med.* 2019, 23, 3293–3301. [CrossRef] [PubMed]

- 253. Wang, Y.H.; Han, X.D.; Qiu, Y.; Xiong, J.; Yu, Y.; Wang, B.; Zhu, Z.Z.; Qian, B.P.; Chen, Y.X.; Wang, S.F.; et al. Increased expression of insulin-like growth factor-1 receptor is correlated with tumor metastasis and prognosis in patients with osteosarcoma. *J. Surg. Oncol.* 2012, 105, 235–243. [CrossRef] [PubMed]
- 254. Patane, S.; Avnet, S.; Coltella, N.; Costa, B.; Sponza, S.; Olivero, M.; Vigna, E.; Naldini, L.; Baldini, N.; Ferracini, R.; et al. MET overexpression turns human primary osteoblasts into osteosarcomas. *Cancer Res.* 2006, 66, 4750–4757. [CrossRef] [PubMed]
- 255. Chen, Y.; Huang, W.; Sun, W.; Zheng, B.; Wang, C.; Luo, Z.; Wang, J.; Yan, W. LncRNA MALAT1 Promotes Cancer Metastasis in Osteosarcoma via Activation of the PI3K-Akt Signaling Pathway. Cell Physiol. Biochem. 2018, 51, 1313–1326. [CrossRef] [PubMed]
- 256. Huang, L.; Jiang, S.; Shi, Y. Tyrosine kinase inhibitors for solid tumors in the past 20 years (2001–2020). *J. Hematol. Oncol.* 2020, 13, 143. [CrossRef] [PubMed]
- 257. Navid, F.; Santana, V.M.; Neel, M.; McCarville, M.B.; Shulkin, B.L.; Wu, J.; Billups, C.A.; Mao, S.; Daryani, V.M.; Stewart, C.F.; et al. A phase II trial evaluating the feasibility of adding bevacizumab to standard osteosarcoma therapy. *Int. J. Cancer* 2017, 141, 1469–1477. [CrossRef]
- 258. Fleuren, E.D.; Versleijen-Jonkers, Y.M.; Boerman, O.C.; van der Graaf, W.T. Targeting receptor tyrosine kinases in osteosarcoma and Ewing sarcoma: Current hurdles and future perspectives. *Biochim. Biophys. Acta* 2014, 1845, 266–276. [CrossRef]
- Gobin, B.; Moriceau, G.; Ory, B.; Charrier, C.; Brion, R.; Blanchard, F.; Redini, F.; Heymann, D. Imatinib mesylate exerts antiproliferative effects on osteosarcoma cells and inhibits the tumour growth in immunocompetent murine models. *PLoS ONE* 2014, 9, e90795. [CrossRef]
- McGary, E.C.; Weber, K.; Mills, L.; Doucet, M.; Lewis, V.; Lev, D.C.; Fidler, I.J.; Bar-Eli, M. Inhibition of platelet-derived growth factor-mediated proliferation of osteosarcoma cells by the novel tyrosine kinase inhibitor STI571. *Clin. Cancer Res.* 2002, *8*, 3584–3591. [CrossRef]
- Bond, M.; Bernstein, M.L.; Pappo, A.; Schultz, K.R.; Krailo, M.; Blaney, S.M.; Adamson, P.C. A phase II study of imatinib mesylate in children with refractory or relapsed solid tumors: A Children's Oncology Group study. *Pediatr. Blood. Cancer* 2008, 50, 254–258. [CrossRef]
- Chao, J.; Budd, G.T.; Chu, P.; Frankel, P.; Garcia, D.; Junqueira, M.; Loera, S.; Somlo, G.; Sato, J.; Chow, W.A. Phase II clinical trial of imatinib mesylate in therapy of KIT and/or PDGFRalpha-expressing Ewing sarcoma family of tumors and desmoplastic small round cell tumors. *Anticancer Res.* 2010, 30, 547–552.
- 263. Chugh, R.; Wathen, J.K.; Maki, R.G.; Benjamin, R.S.; Patel, S.R.; Meyers, P.A.; Priebat, D.A.; Reinke, D.K.; Thomas, D.G.; Keohan, M.L.; et al. Phase II multicenter trial of imatinib in 10 histologic subtypes of sarcoma using a bayesian hierarchical statistical model. *J. Clin. Oncol.* 2009, 27, 3148–3153. [CrossRef]
- 264. Aplenc, R.; Blaney, S.M.; Strauss, L.C.; Balis, F.M.; Shusterman, S.; Ingle, A.M.; Agrawal, S.; Sun, J.; Wright, J.J.; Adamson, P.C. Pediatric phase I trial and pharmacokinetic study of dasatinib: A report from the children's oncology group phase I consortium. *J. Clin. Oncol.* 2011, 29, 839–844. [CrossRef] [PubMed]
- 265. Hingorani, P.; Zhang, W.; Gorlick, R.; Kolb, E.A. Inhibition of Src phosphorylation alters metastatic potential of osteosarcoma in vitro but not in vivo. *Clin. Cancer Res.* 2009, *15*, 3416–3422. [CrossRef]
- 266. Sampson, E.R.; Martin, B.A.; Morris, A.E.; Xie, C.; Schwarz, E.M.; O'Keefe, R.J.; Rosier, R.N. The orally bioavailable met inhibitor PF-2341066 inhibits osteosarcoma growth and osteolysis/matrix production in a xenograft model. *J. Bone Miner. Res.* 2011, 26, 1283–1294. [CrossRef] [PubMed]
- 267. Pignochino, Y.; Grignani, G.; Cavalloni, G.; Motta, M.; Tapparo, M.; Bruno, S.; Bottos, A.; Gammaitoni, L.; Migliardi, G.; Camussi, G.; et al. Sorafenib blocks tumour growth, angiogenesis and metastatic potential in preclinical models of osteosarcoma through a mechanism potentially involving the inhibition of ERK1/2, MCL-1 and ezrin pathways. *Mol. Cancer* 2009, *8*, 118. [CrossRef] [PubMed]
- 268. Grignani, G.; Palmerini, E.; Dileo, P.; Asaftei, S.D.; D'Ambrosio, L.; Pignochino, Y.; Mercuri, M.; Picci, P.; Fagioli, F.; Casali, P.G.; et al. A phase II trial of sorafenib in relapsed and unresectable high-grade osteosarcoma after failure of standard multimodal therapy: An Italian Sarcoma Group study. Ann. Oncol. 2012, 23, 508–516. [CrossRef]
- 269. Grignani, G.; Palmerini, E.; Ferraresi, V.; D'Ambrosio, L.; Bertulli, R.; Asaftei, S.D.; Tamburini, A.; Pignochino, Y.; Sangiolo, D.; Marchesi, E.; et al. Sorafenib and everolimus for patients with unresectable high-grade osteosarcoma progressing after standard treatment: A non-randomised phase 2 clinical trial. *Lancet Oncol.* 2015, *16*, 98–107. [CrossRef]
- Davis, L.E.; Bolejack, V.; Ryan, C.W.; Ganjoo, K.N.; Loggers, E.T.; Chawla, S.; Agulnik, M.; Livingston, M.B.; Reed, D.; Keedy, V.; et al. Randomized Double-Blind Phase II Study of Regorafenib in Patients With Metastatic Osteosarcoma. *J. Clin. Oncol.* 2019, 37, 1424–1431. [CrossRef] [PubMed]
- 271. Duffaud, F.; Mir, O.; Boudou-Rouquette, P.; Piperno-Neumann, S.; Penel, N.; Bompas, E.; Delcambre, C.; Kalbacher, E.; Italiano, A.; Collard, O.; et al. Efficacy and safety of regorafenib in adult patients with metastatic osteosarcoma: A non-comparative, randomised, double-blind, placebo-controlled, phase 2 study. *Lancet Oncol.* 2019, 20, 120–133. [CrossRef]
- 272. Italiano, A.; Mir, O.; Mathoulin-Pelissier, S.; Penel, N.; Piperno-Neumann, S.; Bompas, E.; Chevreau, C.; Duffaud, F.; Entz-Werle, N.; Saada, E.; et al. Cabozantinib in patients with advanced Ewing sarcoma or osteosarcoma (CABONE): A multicentre, single-arm, phase 2 trial. *Lancet Oncol.* 2020, 21, 446–455. [CrossRef]
- 273. Gaspar, N.; Campbell-Hewson, Q.; Gallego Melcon, S.; Locatelli, F.; Venkatramani, R.; Hecker-Nolting, S.; Gambart, M.; Bautista, F.; Thebaud, E.; Aerts, I.; et al. Phase I/II study of single-agent lenvatinib in children and adolescents with refractory or relapsed solid malignancies and young adults with osteosarcoma (ITCC-050). *ESMO Open* **2021**, *6*, 100250. [CrossRef]

- 274. Umeda, K.; Kato, I.; Saida, S.; Okamoto, T.; Adachi, S. Pazopanib for second recurrence of osteosarcoma in pediatric patients. *Pediatr. Int.* **2017**, *59*, 937–938. [CrossRef] [PubMed]
- Mayer, E.L.; Krop, I.E. Advances in targeting SRC in the treatment of breast cancer and other solid malignancies. *Clin. Cancer Res.* 2010, 16, 3526–3532. [CrossRef]
- Miyazaki, T.; Sanjay, A.; Neff, L.; Tanaka, S.; Horne, W.C.; Baron, R. Src kinase activity is essential for osteoclast function. J. Biol. Chem. 2004, 279, 17660–17666. [CrossRef]
- Hu, C.; Deng, Z.; Zhang, Y.; Yan, L.; Cai, L.; Lei, J.; Xie, Y. The prognostic significance of Src and p-Src expression in patients with osteosarcoma. *Med. Sci. Monit.* 2015, 21, 638–645.
- 278. Baird, K.; Glod, J.; Steinberg, S.M.; Reinke, D.; Pressey, J.G.; Mascarenhas, L.; Federman, N.; Marina, N.; Chawla, S.; Lagmay, J.P.; et al. Results of a Randomized, Double-Blinded, Placebo-Controlled, Phase 2.5 Study of Saracatinib (AZD0530), in Patients with Recurrent Osteosarcoma Localized to the Lung. *Sarcoma* 2020, 2020, 7935475. [CrossRef]
- 279. Dancey, J.E. Inhibitors of the mammalian target of rapamycin. *Expert Opin. Investig. Drugs* **2005**, *14*, 313–328. [CrossRef]
- 280. Shaw, R.J.; Cantley, L.C. Ras, PI(3)K and mTOR signalling controls tumour cell growth. Nature 2006, 441, 424–430. [CrossRef]
- Ory, B.; Moriceau, G.; Redini, F.; Heymann, D. mTOR inhibitors (rapamycin and its derivatives) and nitrogen containing bisphosphonates: Bi-functional compounds for the treatment of bone tumours. *Curr. Med. Chem.* 2007, 14, 1381–1387. [CrossRef] [PubMed]
- 282. Perry, J.A.; Kiezun, A.; Tonzi, P.; Van Allen, E.M.; Carter, S.L.; Baca, S.C.; Cowley, G.S.; Bhatt, A.S.; Rheinbay, E.; Pedamallu, C.S.; et al. Complementary genomic approaches highlight the PI3K/mTOR pathway as a common vulnerability in osteosarcoma. *Proc. Natl. Acad. Sci. USA* 2014, 111, E5564–E5573. [CrossRef]
- 283. Zhou, Q.; Deng, Z.; Zhu, Y.; Long, H.; Zhang, S.; Zhao, J. mTOR/p70S6K signal transduction pathway contributes to osteosarcoma progression and patients' prognosis. *Med. Oncol.* 2010, 27, 1239–1245. [CrossRef] [PubMed]
- 284. Chawla, S.P.; Staddon, A.P.; Baker, L.H.; Schuetze, S.M.; Tolcher, A.W.; D'Amato, G.Z.; Blay, J.Y.; Mita, M.M.; Sankhala, K.K.; Berk, L.; et al. Phase II study of the mammalian target of rapamycin inhibitor ridaforolimus in patients with advanced bone and soft tissue sarcomas. J. Clin. Oncol. 2012, 30, 78–84. [CrossRef] [PubMed]
- 285. Demetri, G.D.; Chawla, S.P.; Ray-Coquard, I.; Le Cesne, A.; Staddon, A.P.; Milhem, M.M.; Penel, N.; Riedel, R.F.; Bui-Nguyen, B.; Cranmer, L.D.; et al. Results of an international randomized phase III trial of the mammalian target of rapamycin inhibitor ridaforolimus versus placebo to control metastatic sarcomas in patients after benefit from prior chemotherapy. *J. Clin. Oncol.* 2013, *31*, 2485–2492. [CrossRef]
- 286. Corral, L.G.; Muller, G.W.; Moreira, A.L.; Chen, Y.; Wu, M.; Stirling, D.; Kaplan, G. Selection of novel analogs of thalidomide with enhanced tumor necrosis factor alpha inhibitory activity. *Mol. Med.* **1996**, *2*, 506–515. [CrossRef] [PubMed]
- Li, X.; Tian, J.; Bo, Q.; Li, K.; Wang, H.; Liu, T.; Li, J. Targeting DNA-PKcs increased anticancer drug sensitivity by suppressing DNA damage repair in osteosarcoma cell line MG63. *Tumour Biol.* 2015, *36*, 9365–9372. [CrossRef]
- Morice, S.; Danieau, G.; Redini, F.; Brounais-Le-Royer, B.; Verrecchia, F. Hippo/YAP Signaling Pathway: A Promising Therapeutic Target in Bone Paediatric Cancers? *Cancers* 2020, 12, 645. [CrossRef]
- Kovar, H.; Bierbaumer, L.; Radic-Sarikas, B. The YAP/TAZ Pathway in Osteogenesis and Bone Sarcoma. *Cells* 2020, 9, 972.
 [CrossRef]
- Zucchini, C.; Manara, M.C.; Cristalli, C.; Carrabotta, M.; Greco, S.; Pinca, R.S.; Ferrari, C.; Landuzzi, L.; Pasello, M.; Lollini, P.L.; et al. ROCK2 deprivation leads to the inhibition of tumor growth and metastatic potential in osteosarcoma cells through the modulation of YAP activity. J. Exp. Clin. Cancer Res. 2019, 38, 503. [CrossRef]
- Bouvier, C.; Macagno, N.; Nguyen, Q.; Loundou, A.; Jiguet-Jiglaire, C.; Gentet, J.C.; Jouve, J.L.; Rochwerger, A.; Mattei, J.C.; Bouvard, D.; et al. Prognostic value of the Hippo pathway transcriptional coactivators YAP/TAZ and beta1-integrin in conventional osteosarcoma. *Oncotarget* 2016, 7, 64702–64710. [CrossRef] [PubMed]
- 292. Zhang, Y.H.; Li, B.; Shen, L.; Shen, Y.; Chen, X.D. The role and clinical significance of YES-associated protein 1 in human osteosarcoma. *Int. J. Immunopathol. Pharmacol.* 2013, 26, 157–167. [CrossRef]
- 293. Chan, L.H.; Wang, W.; Yeung, W.; Deng, Y.; Yuan, P.; Mak, K.K. Hedgehog signaling induces osteosarcoma development through Yap1 and H19 overexpression. *Oncogene* **2014**, *33*, 4857–4866. [CrossRef] [PubMed]
- 294. Yang, Z.; Zhang, M.; Xu, K.; Liu, L.; Hou, W.K.; Cai, Y.Z.; Xu, P.; Yao, J.F. Knockdown of YAP1 inhibits the proliferation of osteosarcoma cells in vitro and in vivo. *Oncol. Rep.* 2014, 32, 1265–1272. [CrossRef]
- 295. Morice, S.; Danieau, G.; Tesfaye, R.; Mullard, M.; Brion, R.; Dupuy, M.; Ory, B.; Brounais-Le Royer, B.; Corre, I.; Redini, F.; et al. Involvement of the TGF-beta Signaling Pathway in the Development of YAP-Driven Osteosarcoma Lung Metastasis. *Front. Oncol.* 2021, 11, 765711. [CrossRef]
- 296. Morice, S.; Mullard, M.; Brion, R.; Dupuy, M.; Renault, S.; Tesfaye, R.; Brounais-Le Royer, B.; Ory, B.; Redini, F.; Verrecchia, F. The YAP/TEAD Axis as a New Therapeutic Target in Osteosarcoma: Effect of Verteporfin and CA3 on Primary Tumor Growth. *Cancers* 2020, 12, 3847. [CrossRef] [PubMed]
- Chai, J.; Xu, S.; Guo, F. TEAD1 mediates the oncogenic activities of Hippo-YAP1 signaling in osteosarcoma. *Biochem. Biophys. Res. Commun.* 2017, 488, 297–302. [CrossRef] [PubMed]

- 298. Fujii, M.; Nakanishi, H.; Toyoda, T.; Tanaka, I.; Kondo, Y.; Osada, H.; Sekido, Y. Convergent signaling in the regulation of connective tissue growth factor in malignant mesothelioma: TGFbeta signaling and defects in the Hippo signaling cascade. *Cell Cycle* **2012**, *11*, 3373–3379. [CrossRef]
- 299. Grannas, K.; Arngarden, L.; Lonn, P.; Mazurkiewicz, M.; Blokzijl, A.; Zieba, A.; Soderberg, O. Crosstalk between Hippo and TGFbeta: Subcellular Localization of YAP/TAZ/Smad Complexes. *J. Mol. Biol.* **2015**, *427*, 3407–3415. [CrossRef]