

Case Report



A Novel *VPS33B* Variant Identified by Exome Sequencing in a Patient with Arthrogryposis-Renal Dysfunction-Cholestasis Syndrome

Min Ju Lee , Chae Ri Suh , Jeong Hee Shin , Jee Hyun Lee , Yoon Lee , Baik-Lin Eun , Kee Hwan Yoo , and Jung Ok Shim

Department of Pediatrics, Korea University College of Medicine, Seoul, Korea

OPEN ACCESS

Received: Jun 20, 2019

Accepted: Aug 31, 2019

Correspondence to

Jung Ok Shim

Division of Pediatric Gastroenterology and Hepatology, Department of Pediatrics, Korea University Guro Hospital, 148 Gurodong-ro, Guro-gu, Seoul 08308, Korea.
E-mail: shimjo@korea.ac.kr

Copyright © 2019 by The Korean Society of Pediatric Gastroenterology, Hepatology and Nutrition

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ORCID iDs

Min Ju Lee
<https://orcid.org/0000-0002-8900-5953>
Chae Ri Suh
<https://orcid.org/0000-0003-1975-4323>
Jeong Hee Shin
<https://orcid.org/0000-0003-3718-968X>
Jee Hyun Lee
<https://orcid.org/0000-0002-4318-2487>
Yoon Lee
<https://orcid.org/0000-0001-9521-3575>
Baik-Lin Eun
<https://orcid.org/0000-0001-8735-292X>
Kee Hwan Yoo
<https://orcid.org/0000-0001-6490-4293>
Jung Ok Shim
<https://orcid.org/0000-0001-6449-7819>

ABSTRACT

Arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome is a rare autosomal recessive multisystemic disease that is associated with the liver, kidney, skin, and central nervous and musculoskeletal systems. ARC occurs as a result of mutations in the *VPS33B* (Vacuolar protein sorting 33 homolog B) or *VIPAR* (VPS33B interacting protein, apical-basolateral polarity regulator) genes. A female infant presented with neonatal cholestasis with a severe clinical outcome. She was diagnosed with ARC syndrome using targeted exome sequencing (TES). Exome sequencing revealed compound heterozygous mutations, c.707A>T and c.239+5G>A, in *VPS33B*, where c.707A>T was a novel variant; the resultant functional protein defects were predicted via *in silico* analysis. c.239+5G>A, a pathogenic mutation that affects splicing, is found in less than 0.1% of the general population. Invasive techniques, such as liver biopsies, did not contribute to a differential diagnosis of ARC syndrome; thus, early TES together with clinical presentations constituted an apparently accurate diagnostic procedure.

Keywords: Neonatal cholestasis; *VIPAR*; *VPS33B*; Mutation

INTRODUCTION

Arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome (MIM 208085) is a rare autosomal recessive disorder first reported in 1973 by Lutz-Richter and Landolt in the offspring of a marriage between close relatives [1]. Its characteristic clinical features are arthrogryposis, renal tubular acidosis, and neonatal cholestatic jaundice. ARC syndrome is sometimes associated with additional presentations, including ichthyosis, agenesis of the corpus callosum, congenital cardiovascular abnormalities, nephrogenic diabetes insipidus, hypothyroidism, recurrent sepsis, deafness, and platelet abnormality [2]. The locus that causes this disorder is located on chromosome 15q26.1, and germline mutations were identified in vacuolar protein sorting 33 homolog B (*VPS33B*) and VPS33B-interacting protein, apical-basolateral polarity regulator (*VIPAR*) genes [3]. ARC syndrome is lethal, with death generally occurring in the first year of life. Mild, atypical symptoms at birth and in the first few weeks after birth result in delayed treatment of this disorder. In this case study, we describe an infant who was diagnosed with ARC using targeted exome sequencing (TES) before symptoms became apparent.

Conflict of Interest

The authors have no financial conflicts of interest.

The Institutional Review Board (IRB) of Korea University Guro Hospital approved this study (No. 2019GR0272). Written consent was waived under the IRB's approval.

CASE REPORT

A female neonate was born at the gestational age of 41 weeks and 2 days via cesarean section because of progression failure. Her birth weight was 3.36 kg and the parents were not consanguineous. There was no family history of hepatobiliary disease. She had a coarse face, a high arched palate, ichthyosis, hepatosplenomegaly, and arthrogryposis with bilateral dislocation of the hips, flexion contracture of the knee joints, and a vertical talus (**Fig. 1**). She suffered from generalized hypotonia and respiratory distress, and a nasogastric tube (NG) was placed immediately after birth because of impaired sucking and swallowing.

At birth, the patient's hemoglobin level was 14.9 g/dL, her leucocyte level was 2,900/ μ L, and her platelet level was 157,000/ μ L (normal platelet morphology). Her initial electrolyte profile was normal. Levels of serum total bilirubin and direct bilirubin were 1.71 mg/dL and 0.55 mg/dL, respectively. At 7 day after birth, laboratory evaluation revealed cholestasis (total bilirubin 15.91 mg/dL, direct bilirubin 4.05 mg/dL) with a normal gamma glutamyl aminotransferase (GGT) level (32 IU/L). Serum protein and albumin levels were 5.1 and 3.3 g/dL, respectively. Prothrombin and partial thromboplastin times were normal. At 12 d after birth, the total bilirubin level had increased to 25.94 mg/dL. Hepatitis viral markers and neonatal metabolic screening test results were negative.

Magnetic resonance cholangiopancreatography (MRCP) revealed poor delineation of the biliary tree. Liver biopsy revealed paucity of bile ducts, giant cell transformation, and some lipofuscin pigments, suggestive of biliary atresia (BA) or progressive familial intrahepatic cholestasis (PFIC) (**Fig. 2**). Regardless of the MRCP and biopsy findings, genetic studies were conducted for dysmorphism and normal GGT levels to exclude disorders such as ARC syndrome. TES was performed using peripheral blood samples from the parents and the

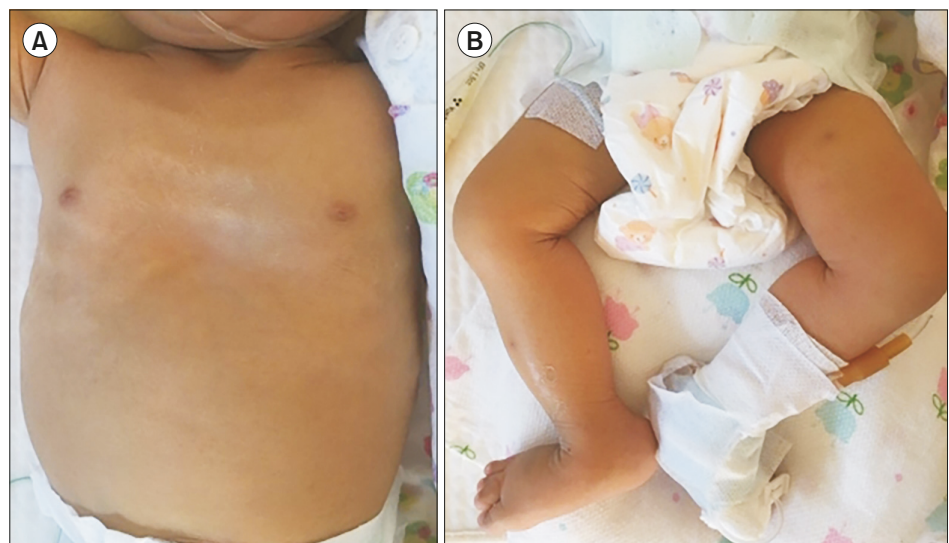


Fig. 1. The dysmorphic features associated with arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome. An infant with ARC syndrome presenting symptoms of (A) ichthyosis and (B) arthrogryposis manifested by bilaterally dislocated hips, flexion contracture of the knees, and vertical talus.

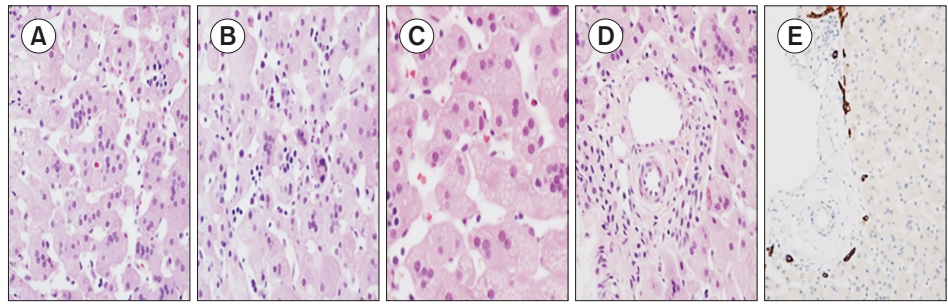


Fig. 2. Micrographs from the liver biopsy of arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome patient. Micrographs from the histological examination (liver biopsy) of patient with ARC syndrome showing (A) giant cell hepatitis, (B) intrahepatic hematopoiesis, (C) lipofuscin deposition, and (D, E) a paucity of bile ducts (A-D: Sections were stained with hematoxylin and eosin and viewed under 400 \times magnification; E: Sections were stained with CK19 and viewed under 200 \times magnification).

patient after obtaining written informed consent. All sequences were BLAST searched against the TruSight One Sequencing Panel (https://support.illumina.com/sequencing/sequencing_kits/trusight_one_kit.html) to determine variations, and single nucleotide polymorphisms were excluded using the Genome Aggregation Database [4], Exome Aggregation Consortium [5], 1,000 Genome Project [6], and Korean Reference Genome Database (<http://coda.nih.go.kr/coda/KRGDB/index.jsp>). Exome sequencing together with Sanger sequencing revealed compound heterozygous mutations, c.707A>T and c.239+5G>A, in *VPS33B*. c.707A>T, inherited from the father, is a novel variant. c.239+5G>A, inherited from the mother, is a rare variant found in less than 0.1% of the general population (**Fig. 3**).

At 15 day, urine analysis indicated proteinuria, global aminoaciduria, glucosuria, hypercalciuria, hyperuricosuria, and hyperphosphaturia with metabolic acidosis (normal anion gap), suggesting renal tubular acidosis. There was hydronephrosis of both kidneys, but no nephrocalcinosis was observed on abdominal ultrasonography. Brain magnetic resonance imaging suggested agenesis of the corpus callosum. Thyroid function evaluation indicated levels of free thyroxine at 1.15 ng/dL and thyroid stimulating hormone levels greater than 100 μ IU/mL. Congenital hypothyroidism with an ectopic thyroid gland was diagnosed. Neonatal hearing screening revealed sensorineural hearing loss on both sides.

She was discharged from the neonatal intensive care unit at 82 days of age but was readmitted frequently because of recurrent infections. At 6 months of age, growth was still faltering despite adequate calorie intake via NG tubes. While her cholestasis improved slightly, her renal function was impaired because of severe renal Fanconi syndrome with signs of nephrogenic diabetes insipidus. Despite continuous renal replacement therapy, she died at the age of 9 months from renal failure secondary to sepsis.

DISCUSSION

Our patient exhibited all the classical clinical features of ARC syndrome, with comorbid presentations of ichthyosis, agenesis of corpus callosum, deafness, hypothyroidism, and nephrogenic diabetes insipidus. Previous cases suggested a usual pattern of liver histology including paucity of bile ducts, lipofuscin deposition, and giant cell hepatitis [1,7-12]. Eastham et al. [13] argued that absence of such changes might be unimportant, because histopathology depends on the timing and site of biopsy. The pathology of cholestasis is

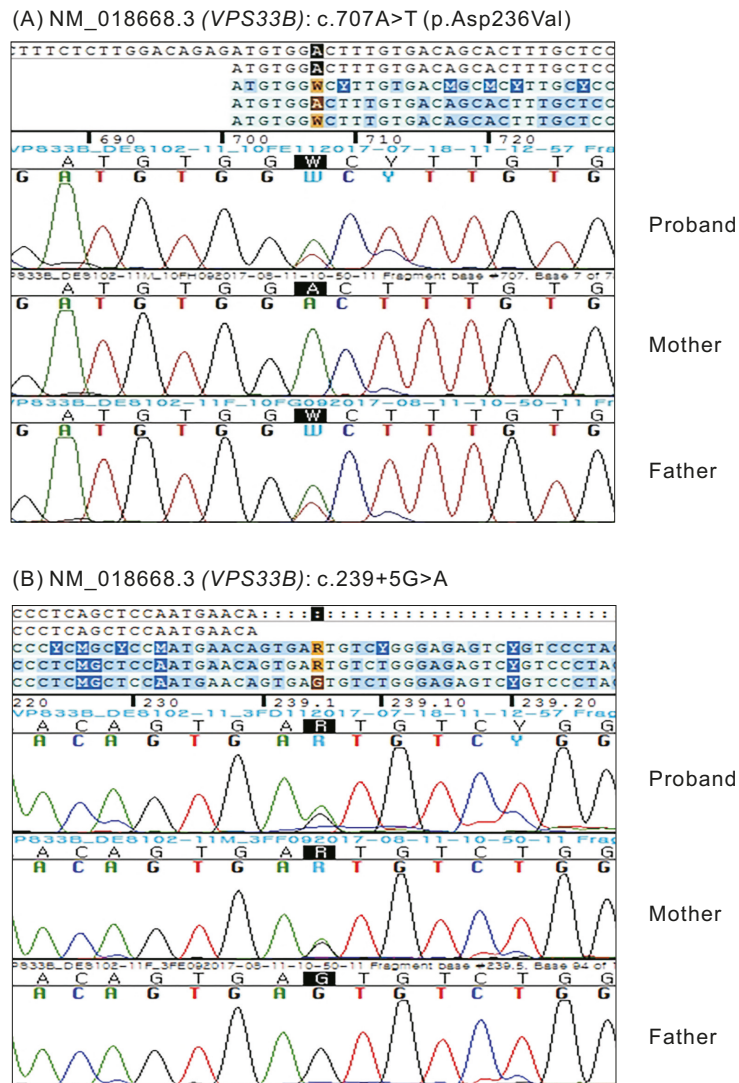


Fig. 3. Missense mutations found in a neonate with cholestasis. Molecular analysis of a neonate with cholestasis revealed compound heterozygous mutations in vacuolar protein sorting 33 homolog B. The DNA chromatograms highlighting the missense mutations: (A) c.707A>T (p.Asp236Val) and (B) c.239+5G>A. The c.707A>T mutation was a novel variant that was acquired from the father.

nonspecific; therefore, pathologists adhere to rigorous criteria to avoid over interpretation of histological findings. Additional diagnoses (i.e., TES) are required before pathogenesis can be confirmed. Furthermore, ARC syndrome patients might have platelet abnormalities and are vulnerable to coagulation. Thus, liver biopsy can lead to the risk of lethal bleeding.

In differential diagnosis, BA, which is first considered in neonates with cholestasis, may be excluded by the presence of normal bile ducts on diagnostic images and normal serum GT levels [14]. Similar clinical and laboratory results might be evident in ARC syndrome, PFIC, and bile acid synthesis disorders (BASDs). PFIC is an autosomal recessive disorder of cholestasis, which causes cholestasis and hepatocellular damage resulting from bile acid transport defects. BASDs are a group of metabolic disorders characterized by defects in the production of normal bile acids, and the accumulation of unusual bile acids and intermediary

metabolites. If clinical symptoms are not fully manifested in the patient, additional genetic testing should be considered to distinguish ARC syndrome from other diseases [15].

Family history, classical clinical presentations, and genetic mutations should be evaluated for accurate diagnosis and early initiation of tailored treatment [14]. In our patient, invasive techniques, such as liver biopsy, did not contribute to a differential diagnosis of ARC syndrome, whereas early TES, together with the clinical presentations, constituted an apparently accurate diagnostic procedure.

ARC syndrome was mapped to chromosome 15q26.1 and germline mutations in *VPS33B* were identified by Gissen et al. [3] in 14 kindreds with ARC syndrome. *VIPAR* is another causative gene of ARC syndrome. *VPS33B* is crucial in intracellular vesicular trafficking pathways, while *VIPAR* exerts pleiotropic effects on polarity and apical membrane protein restriction via the formation of *VPS33B-VIPAR* complexes, ensuring a normal cellular structure. These proteins are found in many parts of the body, including the kidneys, liver, heart, lungs, brain, skin, and skeletal muscles, accounting for the multisystemic symptoms characteristic of the ARC clinical phenotype [16-19].

To allow researchers easy access to updated information on global genetic epidemiology, the online, locus-specific Leiden Open-Source Variation Database for ARC syndrome was created in 2011. This database includes a total of 228 unique variants in *VPS33B* and 34 unique variants in *VIPAR*, of which sequence mutations are categorized as 'pathogenic,' 'probably pathogenic,' 'no known pathogenicity,' 'probably no pathogenicity,' and 'effect unknown,' depending on their predicted effects on the protein and the clinical phenotype [20]. In our case, exome sequencing revealed compound heterozygous mutations of c.707A>T and c.239+5G>A in *VPS33B*.

Notably, c.707A>T is a novel variant that has not been reported among the 123,136 variants in the general population. According to the PolyPhen-2 program, which predicts protein functional defects, a c.707A>T score of '1.000' was 'probably damaging.' Scores obtained using SIFT, FATHMM, LRT, MutationTaster, MutationAssessor, PROVEAN, and VEST3 (<http://asia.ensembl.org/info/genome/variation/index.html>) via *in silico* analyses were '0.000,' '-4.030,' '0.000,' '1.000,' '3.560,' '-8.600,' and '0.988,' respectively, indicating that this mutation might cause functional damage.

Furthermore, c.239+5G>A in *VPS33B* is rare, being found in less than 0.1% of the general population. Adaptive boosting and random forest scores from dbSNV19 (<http://asia.ensembl.org/info/genome/variation/index.html>) were '1.000' and '0.996,' respectively, suggesting that the mutation was pathogenic and affected splicing. Functional analyses should be conducted to determine the effect of this missense mutation on protein function and patient phenotypes.

In this study, we reported a patient with ARC carrying a novel *VPS33B* mutation, determined using TES. Liver biopsy was non-specific and did not contribute to the differential diagnosis. Early TES, together with clinical presentations, constituted an apparently accurate diagnostic procedure. No specific treatment currently exists for ARC, therefore, early recognition and genetic diagnosis is essential to predict and prepare for the course of this disease.

REFERENCES

1. Lutz-Richner AR, Landolt RF. Familiäre gallengansmissbildungen mit tubularer neiereninsuffizienz. *Helv Paediatr Acta* 1973;28:1-12.
2. Zhou Y, Zhang J. Arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome: from molecular genetics to clinical features. *Ital J Pediatr* 2014;40:77.
[PUBMED](#) | [CROSSREF](#)
3. Gissen P, Tee L, Johnson CA, Genin E, Caliebe A, Chitayat D, et al. Clinical and molecular genetic features of ARC syndrome. *Hum Genet* 2006;120:396-409.
[PUBMED](#) | [CROSSREF](#)
4. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, et al. Variation across 141,456 human exomes and genomes reveals the spectrum of loss-of-function intolerance across human protein-coding genes. *BioRxiv* 531210 [Preprint]. 2019 [cited 2019 Jan 28]. Available from: <http://dx.doi.org/10.1101/531210>.
5. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, et al.; Exome Aggregation Consortium. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 2016;536:285-91.
[PUBMED](#) | [CROSSREF](#)
6. 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO. A global reference for human genetic variation. *Nature* 2015;526:68-74.
[PUBMED](#) | [CROSSREF](#)
7. Saraiva JM, Lemos C, Gonçalves I, Carneiro F, Mota HC. Arthrogryposis multiplex congenita with renal and hepatic abnormalities in a female infant. *J Pediatr* 1990;117:761-3.
[PUBMED](#) | [CROSSREF](#)
8. Di Rocco M, Callea F, Pollice B, Faraci M, Campiani F, Borrone C. Arthrogryposis, renal dysfunction and cholestasis syndrome: report of five patients from three Italian families. *Eur J Pediatr* 1995;154:835-9.
[PUBMED](#) | [CROSSREF](#)
9. Coleman RA, Van Hove JL, Morris CR, Rhoads JM, Summar ML. Cerebral defects and nephrogenic diabetes insipidus with the ARC syndrome: additional findings or a new syndrome (ARCC-NDI)? *Am J Med Genet* 1997;72:335-8.
[PUBMED](#) | [CROSSREF](#)
10. Nezelof C, Dupart MC, Jaubert F, Eliachar E. A lethal familial syndrome associating arthrogryposis multiplex congenita, renal dysfunction, and a cholestatic and pigmentary liver disease. *J Pediatr* 1979;94:258-60.
[PUBMED](#) | [CROSSREF](#)
11. Horslen SP, Quarrell OW, Tanner MS. Liver histology in the arthrogryposis multiplex congenita, renal dysfunction, and cholestasis (ARC) syndrome: report of three new cases and review. *J Med Genet* 1994;31:62-4.
[PUBMED](#) | [CROSSREF](#)
12. Mikati MA, Barakat AY, Sulh HB, Der Kaloustian VM. Renal tubular insufficiency, cholestatic jaundice, and multiple congenital anomalies--a new multisystem syndrome. *Helv Paediatr Acta* 1984;39:463-71.
[PUBMED](#)
13. Eastham KM, McKiernan PJ, Milford DV, Ramani P, Wyllie J, Van't Hoff W, et al. ARC syndrome: an expanding range of phenotypes. *Arch Dis Child* 2001;85:415-20.
[PUBMED](#) | [CROSSREF](#)
14. Ilhan O, Ozer EA, Ozdemir SA, Akbay S, Memur S, Kanar B, et al. Arthrogryposis-renal tubular dysfunction-cholestasis syndrome: a cause of neonatal cholestasis. case report. *Arch Argent Pediatr* 2016;114:e9-12.
[PUBMED](#) | [CROSSREF](#)
15. Knisely AS. Progressive familial intrahepatic cholestasis in children. In: Dhawan A, ed. *Concise pediatric and adolescent hepatology*. pediatric and adolescent medicine 16. Basel: Karger, 2012:30-7.
16. Carim L, Sumoy L, Andreu N, Estivill X, Escarceller M. Cloning, mapping and expression analysis of VPS33B, the human orthologue of rat Vps33b. *Cytogenet Cell Genet* 2000;89:92-5.
[PUBMED](#) | [CROSSREF](#)
17. Cullinane AR, Straatman-Iwanowska A, Zaucker A, Wakabayashi Y, Bruce CK, Luo G, et al. Mutations in VIPAR cause an arthrogryposis, renal dysfunction and cholestasis syndrome phenotype with defects in epithelial polarization. *Nat Genet* 2010;42:303-12.
[PUBMED](#) | [CROSSREF](#)
18. Gissen P, Johnson CA, Morgan NV, Stapelbroek JM, Forshew T, Cooper WN, et al. Mutations in VPS33B, encoding a regulator of SNARE-dependent membrane fusion, cause arthrogryposis-renal dysfunction-cholestasis (ARC) syndrome. *Nat Genet* 2004;36:400-4.
[PUBMED](#) | [CROSSREF](#)

19. Matthews RP, Plumb-Rudewiez N, Lorent K, Gissen P, Johnson CA, Lemaigre F, et al. Zebrafish *vps33b*, an ortholog of the gene responsible for human arthrogryposis-renal dysfunction-cholestasis syndrome, regulates biliary development downstream of the onecut transcription factor *hnf6*. *Development* 2005;132:5295-306.
[PUBMED](#) | [CROSSREF](#)
20. Smith H, Galmes R, Gogolina E, Straatman-Iwanowska A, Reay K, Banushi B, et al. Associations among genotype, clinical phenotype, and intracellular localization of trafficking proteins in ARC syndrome. *Hum Mutat* 2012;33:1656-64.
[PUBMED](#) | [CROSSREF](#)