



Draft Genome Sequence of *Brevundimonas* sp. Strain SH203, Producing Cellouronate (β -1,4-Linked Polyglucuronate) Lyase

Tomohiro Suzuki,^a Masako Kikuchi,^b Naotake Konno,^{a,b} Naoto Habu^b

Center for Bioscience Research and Education, Utsunomiya University, Utsunomiya, Tochigi, Japan^a;
Department of Applied Biological Chemistry, School of Agriculture, Utsunomiya University, Utsunomiya, Tochigi, Japan^b

ABSTRACT In this study, we report the draft genome sequence of *Brevundimonas* sp. strain SH203, which was previously isolated from natural soil and has the ability to degrade β -1,4-polyglucuronate (cellouronate). This genomic information may provide new insight into the mechanisms by which cellouronate is degraded.

A bacterial strain with the ability to degrade cellouronate and isolated from natural soil was identified as *Brevundimonas* sp. strain SH203 by comparing its 16S rRNA gene sequence with that in the GenBank database (1). *Brevundimonas* sp. SH203 was reported to degrade β -1,4-polyglucuronate (cellouronate), which is artificially prepared from regenerated cellulose by TEMPO (2,2,6,6-tetramethylpiperidine-1-oxyl radical)-mediated oxidation. We have purified two types of cellouronate lyases from the strain, CUL-I and CUL-II, which catalyze the depolymerization of cellouronate by cleaving glycoside bonds through β -elimination (1–4).

Brevundimonas sp. SH203 was cultivated in 10 ml of LB broth at 30°C for 2 days, and genomic DNA extracted using the DNeasy blood and tissue kit, followed by RNase A digestion. The sheared genomic DNA, resulting in fragments with an average size of 550 bp, and a paired-end DNA library were constructed using the TruSeq DNA PCR-free library preparation kit, according to the manufacturer's instructions. The DNA library then sequenced on a MiSeq instrument at the Center for Bioscience Research and Education, Utsunomiya University, Japan. The raw sequencing data (2 × 301 bp in length) were trimmed to remove low-quality ends (<15) and adapters, using Trimmomatic (version 0.36) (5). To remove contaminant sequences or sequence errors, reads with low coverage (<5×) were removed using khmer (version 2.0) (6), resulting in 1,084,068 high-quality reads (542,034 pairs) totaling 266 Mb (~84-fold coverage). The high-quality reads were further assembled using SPAdes version 3.9.0 (7), with the "careful" option selected, and contigs of less than 200 bp were eliminated. The draft genome of *Brevundimonas* sp. SH203 assembled into 16 contigs, with a total length of 3,145,540 bp (N_{50} , 497,772 bp) and 67.6% G+C content.

The resultant draft genome sequence was annotated using Prokka version 1.11 (8). Furthermore, tRNA genes and rRNA sequences were predicted using tRNAscan-SE version 1.3.1 (9) and RNAmmer version 1.2 (10), respectively. The annotated genome contains 2,961 protein-coding sequences, 47 tRNA genes, and 3 rRNA sequences. Functional annotation of the predicted proteins was conducted using Clusters of Orthologous Groups (COG) (11) and Pfam (12). Among 2,961 proteins, 2,496 (84.2%) and 2,469 (83.3%) proteins were annotated by COG and Pfam, respectively. From the results of Pfam analysis, one alginate-lyase (PF05426) and three β -eliminating lyase (PF01212) domain-containing sequences were detected which might be involved in

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Address correspondence to Tomohiro Suzuki, suzukit@cc.utsunomiya-u.ac.jp, or Naotake Konno, konno@cc.utsunomiya-u.ac.jp.

cellouronate degradation. This genomic information may provide new insight into the mechanisms by which cellouronate is degraded.

Accession number(s). The draft genome sequence of *Brevundimonas* sp. strain SH203 been deposited to the DDBJ/EMBL/GenBank database under the accession no. [BDM00000000](https://doi.org/10.1093/nar/gkm160).

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REFERENCES

- Konno N, Habu N, Maeda I, Azuma N, Isogai A. 2006. Cellouronate (β -1,4-linked polyglucuronate) lyase from *Brevundimonas* sp. SH203: purification and characterization. *Carbohydr Polym* 64:589–596. <https://doi.org/10.1016/j.carbpol.2005.11.015>.
- Konno N, Habu N, Iihashi N, Isogai A. 2008. Purification and characterization of exo-type cellouronate lyase. *Cellulose* 15:453–463. <https://doi.org/10.1007/s10570-007-9195-z>.
- Konno N, Igarashi K, Habu N, Samejima M, Isogai A. 2009. Cloning of the *Trichoderma reesei* cDNA encoding a glucuronan lyase belonging to a novel polysaccharide lyase family. *Appl Environ Microbiol* 75:101–107. <https://doi.org/10.1128/AEM.01749-08>.
- Konno N, Ishida T, Igarashi K, Fushinobu S, Habu N, Samejima M, Isogai A. 2009. Crystal structure of polysaccharide lyase family 20 endo- β -1,4-glucuronan lyase from the filamentous fungus *Trichoderma reesei*. *FEBS Lett* 583:1323–1326. <https://doi.org/10.1016/j.febslet.2009.03.034>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Crusoe MR, Alameldin HF, Awad S, Boucher E, Caldwell A, Cartwright R, Charbonneau A, Constantinides B, Edverson G, Fay S, Fenton J, Fenzl T, Fish J, Garcia-Gutierrez L, Garland P, Gluck J, Gonzalez I, Guermond S, Guo J, Gupta A, Herr JR, Howe A, Hyer A, Harpfer A, Irber L, Kidd R, Lin D, Lippi J, Mansour T, McA'Nulty P, McDonald E, Mizzi J, Murray KD, Nahum JR, Nanlohy K, Nederbragt AJ, Ortiz-Zuazaga H, Ory J, Pell J, Pepe-Ranney C, Russ ZN, Schwarz E, Scott C, Seaman J, Sievert S, Simpson J, Skennerton CT, Spencer J, Srinivasan R, Standage D, et al. 2015. The Khmer software package: enabling efficient nucleotide sequence analysis. *F1000Res* 4:900.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
- Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS Web servers for the detection of tRNAs and snoRNAs. *Nucleic Acids Res* 33:W686–W689. <https://doi.org/10.1093/nar/gki366>.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <https://doi.org/10.1093/nar/gkm160>.
- Galperin MY, Makarova KS, Wolf YI, Koonin EV. 2015. Expanded microbial genome coverage and improved protein family annotation in the COG database. *Nucleic Acids Res* 43:D261–D269. <https://doi.org/10.1093/nar/gku1223>.
- Finn RD, Bateman A, Clements J, Coghill P, Eberhardt RY, Eddy SR, Heger A, Hetherington K, Holm L, Mistry J, Sonnhammer ELL, Tate J, Punta M. 2014. Pfam: the protein families database. *Nucl Acids Res* 42:D222–D230. <https://doi.org/10.1093/nar/gkt1223>.