



# Draft Genome Sequence of a Strain of *Bacillus intestinalis* sp. nov., a New Member of Sporobiota Isolated from the Small Intestine of a Single Patient with Intestinal Cancer

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**ABSTRACT** We report here the draft genome sequence of *Bacillus intestinalis* strain 1731, a novel spore-forming bacterium isolated from the small intestine of a patient with intestinal cancer. The genome comprised 4,047,276 bp, with 43.9% G+C content. There were 3,913 predicted protein-coding genes, including those associated with antibiotic resistance and virulence.

*Bacillus intestinalis* strain 1731 is an aerobic, spore-forming, motile, Gram-positive, rod-shaped bacterium that was isolated from the small intestine of a patient with intestinal malignancy. The members of the *Bacillaceae* family are well-known representatives of the human gut microbiota and are implicated in a number of pathologies (1–3). Using combined culture and genetic workflow, we have previously identified the unexplored diversity of endospore-forming bacteria within humans; this identification necessitated the characterization of spore-forming bacteria, such as *Sporobiota* spp., and the collection of their genes as a sporobiome, owing to their unique common characteristics (4–6).

The 16S rRNA gene sequences of *B. intestinalis* strain 1731 shared 99% similarity with those of various *Bacillus* strains, including *B. subtilis*, *B. amyloliquefaciens*, and *B. atrophaeus*.

An *in silico* DNA-DNA hybridization (DDH) analysis, using the genome-to-genome distance calculator (GGDC2.1) algorithm, produced a highest DDH value of 50.80%, which indicates that *B. intestinalis* strain 1731 is a new species belonging to the *Bacillus* genus (7).

The genome of *B. intestinalis* strain 1731 was sequenced using an Illumina HiSeq sequencing platform (GA IIx; Illumina, CA). Library preparation, sequencing, and runs were performed in accordance with the manufacturer's instructions. *De novo* assembly was performed with SPAdes version 3.9.0 (8), and annotation was performed via the NCBI and RAST servers (9, 10). The final draft genome assembly consisted of 164 contigs and 4,047,276 bp (G+C content of 43.9%), and the total coverage over the genome was 145-fold. It comprises 3,913 gene-coding sequences and 106 predicted RNA genes (86 tRNA, 15 rRNA, and 5 ncRNA genes). Genome analysis using the NCBI and CARD databases revealed genes coding for resistance to the antibiotics tunicamycin, fosfomycin, and bleomycin and class A and B beta-lactamases; the genes *mprF*, *aadK*, and *tmrB* (determinant of resistance to peptide, aminoglycoside, and nucleoside antibiotics, respectively); major facilitator superfamily (MFS) transporters; and the organic hydroperoxide resistance protein OhrA (11).

The genome contains virulence factors, including hemolysin D, serine protease, zinc metalloprotease HtpX, peptidases, alpha-amylase, amidohydrolase, phospholipase,

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phenolic acid decarboxylase, and subtilisin A exodeoxyribonuclease III and VII capsular, flagellar, and sporulation proteins (12, 13). In addition, we identified superoxide dismutases that are considered to possess carcinogenic properties in some bacteria (14).

The availability of the *B. intestinalis* genome sequence facilitates further analysis of human sporobiota and its possible implications in intestinal malignization.

**Accession number(s).** The complete genome sequence has been deposited in the NCBI database under the accession no. [MWZB00000000](https://doi.org/10.1097/IPC.0000000000000335).

## REFERENCES

- Shih AF, Grant M, Sankey C. 2016. *Bacillus cereus* bacteremia and matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. *Infect Dis Clin Practice* 24:14–17. <https://doi.org/10.1097/IPC.0000000000000335>.
- Ha J, Park YJ, Kim YJ, Oh HC, Kim YA. 2016. Late prosthetic joint infection and bacteremia by *Bacillus cereus* confirmed by 16S rRNA sequencing and hip joint tissue pathology. *Ann Clin Microbiol* 19:54. <https://doi.org/10.5145/ACM.2016.19.2.54>.
- Gherardi G. 2016. *Bacillus cereus* disease other than food-borne poisoning, p 93–107. *In* Savini D (ed), *The diverse faces of Bacillus cereus*. Academic Press, Cambridge, MA.
- Tetz G, Tetz V, Vecherkovskaya M. 2016. Genomic characterization and assessment of the virulence and antibiotic resistance of the novel species *Paenibacillus* sp. strain VT-400, a potentially pathogenic bacterium in the oral cavity of patients with hematological malignancies. *Gut Pathog* 8:6. <https://doi.org/10.1186/s13099-016-0089-1>.
- Tetz G, Tetz V. 2015. Complete genome sequence of *Bacilli bacterium* strain VT-13-104 isolated from the intestine of a patient with duodenal cancer. *Genome Announc* 3(4):e00705-15. <https://doi.org/10.1128/genomeA.00705-15>.
- Tetz G, Tetz V. 2017. Draft genome sequence of *Bacillus obstructivus* VT-16-70 isolated from the bronchoalveolar lavage fluid of a patient with chronic obstructive pulmonary disease. *Genome Announc* 5(9):e01754-16. <https://doi.org/10.1128/genomeA.01754-16>.
- Auch AF, von Jan M, Klenk HP, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. *Stand Genomic Sci* 2:117–134. <https://doi.org/10.4056/sigs.531120>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Ciufu S, Li W. 2013. Prokaryotic genome annotation pipeline. *In* Beck J, Benson D, Coleman J, Hoepfner M, Johnson M, Maglott D, Mizrahi I, Morris R, Ostell J, Pruitt K, Rubinstein W, Sayers E, Sirotkin K, Tatusova T (ed), *The NCBI handbook*, 2nd ed. National Center for Biotechnology Information, Bethesda, MD.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Busch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Jia B, Raphenya AR, Alcock B, Waglechner N, Guo P, Tsang KK, Lago BA, Dave BM, Pereira S, Sharma AN, Doshi S, Courtot M, Lo R, Williams LE, Frye JG, Elsayegh T, Sardar D, Westman EL, Pawlowski AC, Johnson TA, Brinkman FS, Wright GD, McArthur AG. 2017. CARD 2017: expansion and model-centric curation of the comprehensive antibiotic resistance database. *Nucleic Acids Res* 45:D566–D573. <https://doi.org/10.1093/nar/gkw1004>.
- Cui W, Chen L, Huang T, Gao Q, Jiang M, Zhang N, Zheng L, Feng K, Cai Y, Wang H. 2013. Computationally identifying virulence factors based on KEGG pathways. *Mol Biosyst* 9:1447–1452. <https://doi.org/10.1039/c3mb70024k>.
- Liaudet L, Szabó C, Evgenov OV, Murthy KG, Pacher P, Virág L, Mabley JG, Marton A, Soriano FG, Kirov MY, Bjertnaes LJ, Salzman AL. 2003. Flagellin from Gram-negative bacteria is a potent mediator of acute pulmonary inflammation in sepsis. *Shock* 19:131–137. <https://doi.org/10.1097/00024382-200302000-00008>.
- Smith DG, Lawson GH. 2001. *Lawsonia intracellularis*: getting inside the pathogenesis of proliferative enteropathy. *Vet Microbiol* 82:331–345. [https://doi.org/10.1016/S0378-1135\(01\)00397-2](https://doi.org/10.1016/S0378-1135(01)00397-2).