



Cytokine Storm as a Cellular Response to dsDNA Breaks: A New Proposal

OPEN ACCESS

Edited by:

Claire Isabelle Vanpouille-Box, Weill Cornell Medicine, United States

Reviewed by:

Arjun Rustagi, Stanford University, United States Jurnana Saleh, Sultan Qaboos University, Oman

> *Correspondence: Indraneel Mittra imittra@actrec.gov.in

Specialty section:

This article was submitted to Inflammation, a section of the journal Frontiers in Immunology

Received: 29 October 2020 Accepted: 07 January 2021 Published: 01 February 2021

Citation:

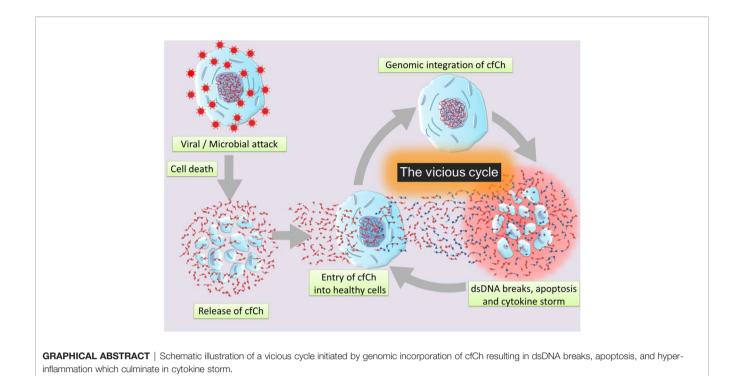
Shabrish S and Mittra I (2021) Cytokine Storm as a Cellular Response to dsDNA Breaks: A New Proposal. Front. Immunol. 12:622738. doi: 10.3389/fimmu.2021.622738

Snehal Shabrish and Indraneel Mittra*

Translational Research Laboratory, Advanced Centre for Treatment, Research and Education in Cancer, Tata Memorial Centre, and Homi Bhabha National Institute, Mumbai, India

Pathogenesis of cytokine storm is poorly understood. In this article we propose a new mechanism and suggest innovative therapeutic avenues for its prevention. We have reported that particles of cell-free chromatin (cfCh) that are released from the billions of cells that die in the body everyday can illegitimately integrate into genomes of healthy cells to trigger dsDNA breaks. The latter leads to apoptosis and/or intense activation of inflammatory cytokines in the affected cells. We hypothesise that a similar phenomenon of dsDNA breaks and inflammation is involved in cytokine storm. The abundant cfCh particles that are released from dying host cells following viral/microbial invasion initiate a cascading effect of more cell death resulting in a vicious cycle of further DNA damage, apoptosis and hyper-inflammation which culminate in cytokine storm. We propose that this unrelenting vicious cycle of cellular DNA damage and cytokine storm may be the underlying cause of high mortality from severe COVID-19. We discuss results of our preclinical studies wherein we have shown that endotoxin induced cytokine storm in mice can be reversed by three different agents that have the ability to inactivate cfCh. These agents may be worthy of investigation in clinical trials to reduce mortality from COVID-19.

Keywords: Cytokine storm, dsDNA breaks, COVID-19, inflammation, apoptosis, resveratrol, copper, free radicals



INTRODUCTION

Cytokine storm is a condition characterized by an overwhelming and uncontrolled inflammation with major implications for global health (1). Cytokine storm is a critical component of the current COVID-19 pandemic, and is associated with severity of the disease and high mortality (2). In the worldwide flu pandemic of 1918, cytokine storm was a major cause of high death rate (3). Cytokine storm has also been described in other pandemics such as H1N1 swine flu (4), H5N1 avian flu (5) and severe acute respiratory syndrome coronavirus (SARS-CoV) (6). It is associated with sepsis in general which affects 48.9 million people worldwide every year of which 11 million die (7). Death from sepsis accounts for 19.7% of all global deaths, especially in poorer countries of the world (7). Several experimental studies and clinical trials have suggested that cytokine storm correlates directly with tissue injury, DNA damage and severity of the disease (1). In spite of intensive research, pathogenesis of the cytokine storm remains poorly understood, hindering development of effective therapies.

CYTOKINE STORM: SUMMARY OF CURRENT KNOWLEDGE

The innate immune response is activated by pattern recognition receptors (PRRs) in response to pathogen-associated molecular patterns (PAMPs) and/or damage-associated molecular patterns (DAMPs) (8). Activated immune response triggers intracellular signalling cascades in immune cells leading to production of inflammatory cytokines by various cells including macrophages, natural killer (NK) cells, dendritic cells, T cells, mast cells, endothelial and epithelial cells (8–10). The immune response is highly regulated and sequentially and temporally orchestrated (8). However, in certain pathological conditions, a profusion of PAMPs or DAMPs are released in response to cell death and stress (11–15). This causes hyper-stimulation of immune cells leading to intense secretion of inflammatory cytokines which results in the cytokine storm (13, 14). This hyper-inflammation triggering cytokine storm can either be in response to PAMPs which activates pathogen-induced hyper-inflammation, or to DAMPs which are self-molecules derived from host cells itself triggering auto-inflammatory response. Although it is widely accepted that these DAMPs and PAMPs are key molecules that trigger an inflammatory response (11–15), the precise nature of these molecules has not been characterized (16–18).

Recently, there has been a spurt of publications associating genomic stress and DNA damage in activation of inflammation (19-22). According to these reports, DNA that accumulates in the cytoplasm following DNA damage and/or microbial infection, acts as DAMPs and activates the DNA sensing GMP-AMP synthase-stimulator of interferon genes (cGAS-STING)-mediated pathway (19-22). The latter triggers an innate immune response by activating pro-inflammatory cytokines (19-22). In addition to microbial DNA and self-DNA from the nucleus, cGAS-STING pathway is also activated by cytosolic mitochondrial DNA (mtDNA) (23, 24). The latter has the potential to induce inflammatory responses and organ injuries in various diseases including cancer (25), diabetes (26), cardiovascular diseases (27) and trauma (28). Elevated levels of mtDNA in circulation has also been reported to be associated with severity of sepsis (29). Recent reports have also implicated presence of cytoplasmic chromatin fragments (CCF) in immune activation (30-32). CCF that are pinched off from nuclei during

cellular senescence (33) are recognized by cGAS to stimulate an inflammatory response *via* STING (30–32). However, whether presence of DNA and/or CCF in the cytoplasm has the potential to trigger the cytokine storm or whether some other mechanism(s) is involved in triggering hyper-inflammation remains unknown. Thus, although the cytokine storm has been known for more than a century (1, 3), and much has been reported on its pathological consequences (1, 5), the trigger for the cytokine storm continues to remain elusive, hindering the development of effective therapies (34). Herein we put forward the hypothesis that cell-free chromatin (cfCh) particles (nucleosomes) released from dying host cells may contribute to the cytokine storm.

CELL FREE CHROMATIN (cfCh) AS A NOVEL TRIGGER FOR THE CYTOKINE STORM

Origin and Structure of Cell-Free Chromatin (cfCh)

It has been estimated that $10^9 - 10^{12}$ cells die in the body, primarily by apoptosis, every day (35). Apoptosis is characterized by nuclear and chromatin condensation followed by fragmentation of DNA by endogenous nucleases, especially caspase-3 and activated DNase (36). Although not demonstrated, it is likely that cfCh particles are also released following other forms of cell death such as necrosis, NETosis, pyroptosis (37). In spite of the body's best efforts to get rid of cfCh (38, 39), a significant amount escapes into the extracellular compartments as well as into the blood circulation (40, 41). cfCh in circulation are cleared by the body by several mechanisms. These include: 1) phagocytosis by macrophages (42); 2) degradation by DNase I present in circulation (43), and 3) liver continuously removing cfCh resulting in a turnover half-life of 10-15 min (38, 39). Low baseline levels of cfCh in healthy individuals play a critical role in maintaining an efficient immune environment. However, elevated cfCh levels as seen in various clinical conditions may lead to runaway inflammation. These conditions have included autoimmune diseases (44), severe infections (45), trauma (46), burns (47), deep vein thrombosis (DVT) (48), cerebral stroke (49), malignancy (50). Increasing cfCh levels positively correlate with age (51).

The Hypothesis

Our hypothesis is based on our recent finding that cfCh particles that are released from the hundreds of millions of cells that die in the body daily to enter into the blood stream can illegitimately integrate into genomes of healthy cells to damage their DNA by inducing dsDNA breaks (52, 53). Such events may also occur locally following cell death in tissues with release of cfCh which integrates into genomes of bystander cells in the neighbourhood (53, 54). Genomic integration of cfCh can have catastrophic consequences, especially since the DNA damage is repaired by the error-prone non-homologous end joining (NHEJ) mechanism, which further accentuates genomic mutations in the form of deletions, insertions, re-arrangements and chromosomal damage which may often cause apoptosis of the cells. The hypothesis also incorporates our recent observation that dsDNA breaks resulting from cfCh integration leads to intense activation of inflammatory cytokines (54, 55). Since cell death is markedly increased following viral or bacterial invasion, we hypothesise that illegitimate genomic integration of cfCh particles that are released from the dying host cells trigger a vicious cycle of more dsDNA breaks, apoptosis and hyper-inflammation which culminate in the cytokine storm (**Graphical Abstract**). We propose that the abundant cfCh that arise following viral/microbial invasion act as DAMPs and activate systemic inflammation. This proposal is supported by reports that circulating levels of cfCh are markedly elevated in patients admitted to ICU with severe sepsis (34). Since the latter is usually associated with the cytokine storm (56), it leads to the possibility that cfCh may be an important factor that contributes to the cytokine storm in sepsis.

Can Cell-Free DNA and/or Free Histones Explain the Cytokine Storm?

Cell-free DNA (cfDNA) and free histones have been shown to have pro-inflammatory properties (57, 58). However, the immune stimulatory effects induced individually by cfDNA and free histones are different when compared to that induced when they are complexed in the form of cfCh (59). Furthermore, the question as to whether naked DNA and/or free histones are indeed present in circulation is in doubt. Apoptotic cell death results in chromosomal condensation and fragmentation with release of chromatin fragments and not of cfDNA or free histones (60). The existence of cfCh in serum and/or plasma can be easily detected by ELISA (61), while the demonstration of cfDNA requires DNA to be extracted from plasma/serum using Proteinase-K treatment. Therefore, the possibility that the isolated cfDNA has, in fact, been derived from circulating cfCh cannot be excluded. Reports of the existence of a direct and strong correlation between circulating cfCh and cfDNA would support such a possibility (62). Similarly, with respect to studies reporting immune-stimulatory effects of free histones (59, 63, 64), the methodologies used to quantify histones did not make a distinction between free histones and cfCh (59). Therefore, whether the latter are present in circulation also remains unclear (59). This uncertainty may have been put to rest by our recent observations made in relation to lipopolysaccharide (LPS) induced sepsis in a mouse model (65). Using confocal microscopy of histological sections of mouse vital organs after staining with fluorescent antibodies against DNA and histone H4, we have shown that it is cfCh, and not free DNA or histones, that are extruded from dying host cells following LPS treatment (65). Therefore, it is likely that cfCh, rather than cfDNA or free histones, is the agent responsible for initiating the cytokine storm in severe infection.

cfCh in Circulation Integrate Into Genomes of Healthy Cells

Although existence of circulating cfCh particles has been known since 1990 (66), whether they have any patho-physiological role to play in the host has only recently been addressed (51, 52). Isolation of cfCh from sera of cancer patients followed by examination under electron microscope revealed particles of

variable sizes (~10 nm >1000 nm) having a beads-on-a-string appearance characteristic of chromatin (52). When cfCh particles isolated from serum where fluorescently labelled and added to cultured mouse fibroblast cells, numerous cfCh particles could be detected in nuclei of recipient cells within 6h (52). The up-taken cfCh rapidly associated themselves with chromosomes of host cells which was followed by activation of an intense DNA damage repair response (DDR) followed by their incorporation into the host cell genomes (52). The activated DDR proteins included H2AX, ATR, ATM, P-p53, P-p21, MDC-1, GADD-34, RAD-50, NIBRIN, MRE-11, DNA-PKcs and DNA ligase IV (52). In addition, apoptotic pathway proteins namely, JC-1, cytochrome-C and caspase 3, were also activated (52) indicating that many of the affected cells were destined to undergo apoptotic cell death. Next generation sequencing detected tens of thousands of DNA reads of human origin in single cell clones developed from the cfCh treated mouse cells; while PCR amplification revealed presence of multiple human Alu sequences (52). cfCh integration resulted in dsDNA breaks as indicated by activation of H2AX which was seen both in vitro and in vivo (52). A unique mechanism was proposed by which cfCh particles integrate themselves into genomes of healthy cells, and which is facilitated by premature activation of DDR (discussed below).

cfCh Released From Dying Cells Integrates Into Genomes of Bystander Cells

We have reported that cfCh released from dying host cells can also integrate into genomes of surrounding healthy bystander cells (54, 55). When human cancer cells were treated with ionizing radiation and co-cultured with mouse fibroblasts, human DNA (cfCh) signals could be detected in the nuclei of mouse cells by FISH (55). Confirmation that cfCh had actually integrated into the genomes was confirmed by detection of multiple human Alu sequences in the mouse cells (55). Bystander uptake and genomic integration of cfCh released from dying cells was also shown to occur in distant organs (55). When mice were delivered focused mini-beam irradiation (20 Gy) to the umbilical region, intense activation of H2AX, caspase 3, NF κ B and IL-6 was detected in brain cells (55).

cfCh Integrates Into the Genome by a Unique Mechanism

How does cfCh enter the cell and integrate themselves into the genome? Our microarray studies have revealed that pathways related to phagocytosis are maximally up-regulated as early as at 6h in mouse fibroblast cells in response to cfCh particles that are released from co-cultured dying Jurkat cells (54). This finding would suggest phagocytosis or pinocytosis to be one of the important mechanisms by which the cell ingests extraneous cfCh. Once inside the cell, cfCh particles integrate themselves into the genome of the host cell by a mechanism which is unique in being the opposite of the classical model of DNA damage, DDR is activated after the DNA damage occurs in response to agents such as ionizing and UV radiation and radiomimetic chemicals (67). According to the proposed new model, entry of cfCh into the cell misleads the cell into perceiving them as broken

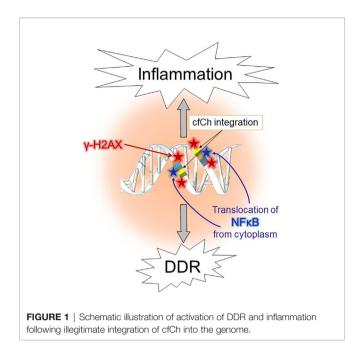
fragments of its own chromosomes containing dsDNA breaks at both ends (52). This prompts the cell to activate a premature DDR much before any damage to DNA having actually occurred. The activated DDR includes repair proteins such as DNA PKc, DNA ligase IV which link up the multiple heterogenous cfCh fragments into concatamers of different sizes. The latter, containing a mosaic of multiple discontinuous DNA segments in the form of conctamers, now form new substrates for incorporation into the genome of host cells, by non-homologous recombination (NHR). The resulting DNA damage is repaired by the error-prone NHEJ mechanism (68) which creates further mutations in the form of insertions, deletions, genetic rearrangements as well as chromosomal damage (52). Thus, paradoxically, DDR which is supposed to protect the integrity of the genome ends up damaging it by its premature activation. The formation of intracellular concatamers is supported by the argument that since the threshold for detection of FISH signals is of the order of 30-50 kilo bases (69), presence of human DNA signals in mouse cells detected by FISH indicates that relatively long human DNA sequences, rather than discrete cfCh particles, incorporate themselves into the mouse cell genomes. Genomic integration of cfCh concatemers by NHR leads to intense activation of inflammatory cytokines (discussed below).

Genomic Integration of cfCh Leads to Somatic Mosaicism

Illegitimate genomic integration of cfCh, derived from the billions of cells that die in the body every day may result in dsDNA breaks and repair by NHEJ. These damaging events occurring repeatedly throughout life may generate multiple genomic polymorphisms which are likely to increase with age (53). Rapid and cumulative effects of DNA damage may exceed the adaptive capacity of the human genome in aging populations which leads to increased mutagenesis and development of various diseases, including cancer. This would be in accordance with the exploding literature fueled by advances in Next generation sequencing on the discovery of somatic mosaicism in healthy cells (70, 71). Somatic mosaicism is related to aging (72), cardiovascular diseases (73), Alzheimer's disease (74) and cancer (75). The above discussion would suggest that approaches to retard aging would need to take into account accumulating dsDNA breaks that result from life-long and repeated genomic integration of cfCh.

cfCh Integration, dsDNA Breaks, and Activation of Inflammatory Cytokines

The aforementioned co-culture experiment of irradiated dying cancer cells of human origin with mouse fibroblasts, led to activation not only of H2AX but also of multiple inflammatory cytokines (54). The latter included NF κ B, IL-6, TNF- α and IFN- γ , all of which were activated simultaneously by 6h (54), and their activation coincided with point of the maximal induction of H2AX (6h) (54). Co-activation of dsDNA breaks and inflammatory cytokines suggested an interrelationship between the two, which was further substantiated by microarray analysis which revealed activation of multiple pathways related to inflammation concurrently with those that accompany DNA damage and cell



cycle at 6h (54). Injection of irradiated dying cancer cells prelabelled with BrdU intravenously into mice led to uptake and genomic integration of BrdU labelled cfCh particles into nuclei of vital organs accompanied by activation of H2AX (54). Genomic integration of cfCh led to intense activation of multiple inflammatory cytokines to include NF κ B, IL-6, TNF- α and IFN- γ . These experiments made the additional novel observation that fluorescent signals of YH2AX co-localized strictly with those of the transcription factor NF κ B in the nuclei of vital organs (54). The inactivated form of NFkB normally remains sequestered in the cytoplasm (76) and trans-locates to the nucleus upon activation by stressful stimuli such as DNA damage (77). Although several nuclear translocation sites for NFKB have been described (78), the finding that YH2AX and NFKB fluorescence signals co-localize has led to the proposal that, following the catastrophic event of integration of cfCh into the genome and the consequent dsDNA breaks, NFKB is strongly activated, followed by its translocation from the cytoplasm to the sites of cfCh integration (79, 80). This finding indicated that inflammation might be a direct consequence of dsDNA breaks inflicted by integration of cfCh (80). It also suggests that cfCh acts as a major form of DAMPs. A schematic model to represent the relationship between cfCh induced dsDNA breaks and inflammation is given in Figure 1.

Inactivation of cfCh Can Prevent the Cytokine Storm

We have identified several cfCh inactivating agents that can prevent the cytokine storm (55, 65, 81). These have included 1) anti-histone antibody complexed nanoparticles (CNPs) which inactivate cfCh by binding to histones; DNase I which inactivates cfCh by degrading its DNA component; and 3) a newly described pro-oxidant combination of the well-researched nutraceuticals Resveratrol and metallic Copper (R-Cu) which degrades cfCh through the medium of free radicals. We have recently reported that these cfCh inactivating/degrading agents can reverse the cytokine storm following endotoxin sepsis, chemotherapy and radiation therapy in mice. Details of these studies are given in the subsequent paragraphs.

Inactivation of cfCh Can Prevent the Cytokine Storm in Endotoxin Sepsis

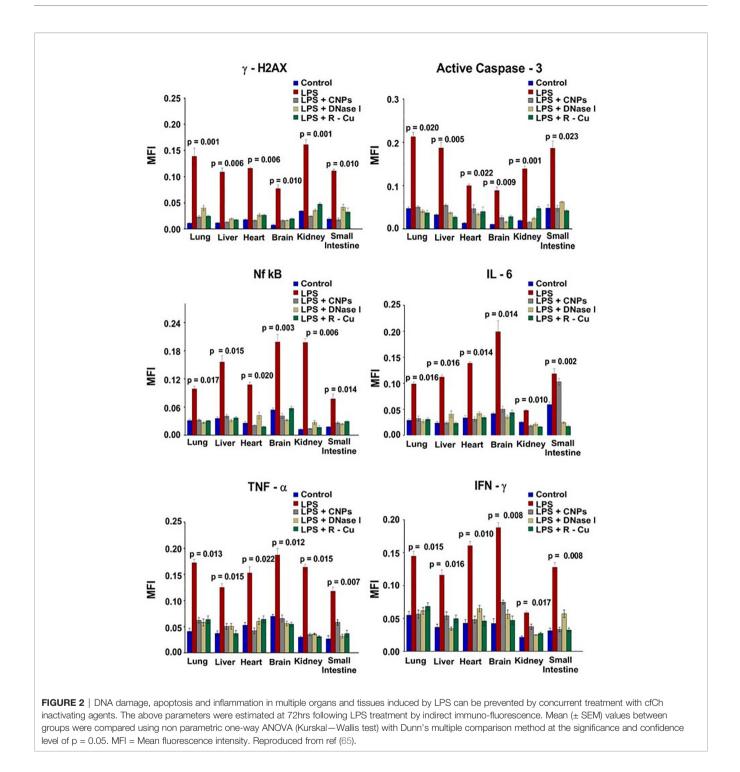
The International Sepsis Forum defines sepsis as "a lifethreatening condition that arises when the body's response to an infection injures its own tissues and organs" (82). This definition implies that hyper-inflammation and immune suppression in sepsis is a result of body's own response against the pathogen and not due to the pathogen *per se* (83). We have recently shown in an endotoxin induced sepsis model that cfCh particles that are released from dying host cells following viral/ microbial infection may be the agents that injure the body's own tissues and organs that leads to sepsis - a finding which would be consistent with the above definition of the International Forum (65).

Several studies have reported that not only PAMPs, but also DAMPs, are recognized by pattern recognizing receptors (PRRs) expressed on immune-reactive cells (84–86). DAMPs are nuclear or cytoplasmic non-microbial molecules, released from the host cells following tissue injury which includes histones, cfDNA, chromatin, HMGB1, etc. (86). Clinical studies have shown a positive correlation of levels of DAMPs, especially of histones and nucleosomes, with sepsis severity (45, 87, 88).

In our study, sepsis was induced in mice by injecting lipopolysaccharide (LPS), a bacterial membrane antigen, which led to extensive cell death and copious release of cfCh particles into extracellular spaces of vital organs and into the circulation (65) (Figure 2). cfCh particles thus released followed by their integration into genomes of surviving cells led to extensive dsDNA breaks and apoptosis in cells of multiple organs viz., lung, liver, heart, brain, kidney and small intestine (65), as well as those of immune related organs such as thymus and spleen. cfCh integration and dsDNA breaks led to intense activation of inflammatory cytokines CRP, IL-6, IL-1β, TNF-α, and IFN-γ in multiple organs as well as release of these cytokines in circulation. The extensive DNA damage also led to immune suppression, coagulopathy, fibrinolysis, thrombocytopenia, multi- organ failure and death. All the above pathologies could be abrogated by administration of the cfCh inactivating agents to mice concurrently with LPS. This data provided strong evidence for a relationship between cfCh integration, dsDNA breaks, cytokine storm and sepsis.

Inactivation of cfCh Can Prevent the Cytokine Storm Associated With Chemotherapy and Radiation Therapy

Cancer treatments involving chemotherapy and radiation therapy are known to trigger intense activation of pro-inflammatory cytokines (89, 90). The latter is thought to be activated by unidentified molecules which act as DAMPs and stimulate immune cells to release pro-inflammatory cytokines (91).



However, the nature of these DAMPs continues to remain unidentified (17, 92). We have shown that, as in the case of sepsis, cfCh released from chemo- or radio therapy induced dying cells are the elusive DAMPs. Therapy induced cell death and cfCh release triggers a cascading effect of more cell death leading to a vicious cycle of further rounds of DNA damage, apoptosis and inflammation which perpetuate and amplify the toxic effects of these cancer therapies (55, 81). We have further reported that administration of the above three cfCh inactivating agents interrupted this vicious cycle thereby preventing the toxic effects of cancer treatment (55, 81). This reinforces the conclusion reached above, with respect to endotoxin sepsis, that copious release of cfCh particles following cell death resulting from chemotherapy and radiation therapy act as DAMPs to trigger the cytokine storm.

Is Cell Free Chromatin Implicated in Pathogenesis of COVID-19?

Pathogenesis of COVID-19 is not well understood. The disease primarily affects the lungs leading to hypoxemic respiratory failure, secondary bacterial pneumonia and direct tissue damage. The disease is also associated with the cytokine storm with excessive release of inflammatory cytokines which can cause multi-organ damage (93). The other organs that are affected include heart, nerves, brain, vessels, kidneys and skin. We have already alluded to the potential role of cfCh in the cytokine storm, but, currently, literature on direct measurement of cfCh levels in COVID-19 patients is lacking. Elevated levels of cfCh in these patients is to be expected since sepsis forms a major manifestations of the SARS-CoV-2 viral infection (94), and there is abundant literature to show that cfCh levels are elevated in sepsis (45, 87, 88, 95). As the title of current article suggests, and discussed extensively above, the cytokine storm is a likely consequence of DNA and cellular damage inflicted by cfCh. We propose that cfCh induced tissue/organ damage can not only explain the aetiology of the cytokine storm, but also help to explain the multi-organ injury that is associated with COVID-19 as a direct consequence of cfCh induced cellular DNA damage.

CONCLUSION AND FUTURE PROSPECTS

In this article we have proposed that inflammation may be a direct consequence of dsDNA breaks inflicted by genomic integration of cfCh released from dying host cells, and that cfCh may be the key instigators of the cytokine storm (54, 80). cfCh particles released from dying host cells following viral/microbial infection may trigger a cascading effect of more host cell death leading to a vicious cycle of further rounds of DNA damage, apoptosis and inflammation which perpetuate and amplify the pathological effects of the offending agent culminating in the cytokine storm. Although, currently, literature on direct measurement of cfCh levels in COVID-19 patients is lacking, we hypothesise that the high mortality in severe COVID-19 may be due to the cytokine

REFERENCES

- Tisoncik JR, Korth MJ, Simmons CP, Farrar J, Martin TR, Katze MG. Into the Eye of the Cytokine Storm. *Microbiol Mol Biol Rev* (2012) 76(1):16–32. doi: 10.1128/MMBR.05015-11
- Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'Cytokine Storm' in COVID-19.' J Infection (2020) 80:607–13. doi: 10.1016/j.jinf.2020.03.037
- Morens DM, Taubenberger JK, Harvey HA, Memoli MJ. The 1918 influenza pandemic: Lessons for 2009 and the future. *Crit Care Med* (2010) 38(SUPPL. 4):e10. doi: 10.1097/CCM.0b013e3181ceb25b
- Guo X, Zhi J, Thomas PG. New fronts emerge in the influenza cytokine storm. Semin Immunopathol (2017) 39:541–50. doi: 10.1007/s00281-017-0636-y
- Liu Q, Zhou YH, Yang ZQ. The cytokine storm of severe influenza and development of immunomodulatory therapy. *Cell Mol Immunol Chin Soc Immunol* (2016) 13:3–10. doi: 10.1038/cmi.2015.74
- Huang KJ, Su IJ, Theron M, Wu YC, Lai SK, Liu CC, et al. An interferon-γrelated cytokine storm in SARS patients. J Med Virol (2005) 75(2):185–94. doi: 10.1002/jmv.20255

storm related sepsis. The latter being perpetuated by the vicious cycle triggered by profuse release of cfCh particles that result from Corona virus induced cell death. The implication of such a suggestion is that, while eliminating the virus may result in resolution of disease in asymptomatic or early symptomatic COVID-19 patients, once the vicious cycle sets in, elimination of the virus may not prevent death in patients with severe disease. Indeed, a recent study has reported that effects of the cytokine storm can persist for a long time after the virus has been eliminated from the body (96). Furthermore, the observation that elderly patients and those with underlying ageing related co-morbidities such as diabetes (97) and cardio-vascular diseases (98) are more prone to COVID-19 related complications, may be attributable to elevated levels of cfCh in these conditions (51, 99, 100). We propose that treatment of severe COVID-19 should include cfCh inactivating agents to prevent death, and that these agents are worthy of investigation in clinical trials in patients suffering from severe COVID-19.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

IM conceptualized and wrote the article. SS wrote the article and prepared the figures. All authors contributed to the article and approved the submitted version.

FUNDING

This study was supported by the Department of Atomic Energy, Government of India, through its grant CTCTMC to Tata Memorial Centre awarded to IM.

- Rudd KE, Johnson SC, Agesa KM, Shackelford KA, Tsoi D, Kievlan DR, et al. Global, regional, and national sepsis incidence and mortality, 1990–2017: analysis for the Global Burden of Disease Study. *Lancet* (2020) 395 (10219):200–11. doi: 10.1016/S0140-6736(19)32989-7
- Chaplin DD. Overview of the immune response. J Allergy Clin Immunol (2010) 125(2 SUPPL. 2):S3. doi: 10.1016/j.jaci.2009.12.980
- 9. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* (2006) 124:783–801. doi: 10.1016/j.cell.2006.02.015
- Zhang JM, An J. Cytokines, inflammation, and pain. Int Anesthesiol Clinics (2007) 45:27–37. doi: 10.1097/AIA.0b013e318034194e
- Sharma SK, Naidu G. The role of danger-associated molecular patterns (DAMPs) in trauma and infections. J Thoracic Dis (2016) 8:1406–9. doi: 10.21037/jtd.2016.05.22
- Relja B, Land WG. Damage-associated molecular patterns in trauma. Eur J Trauma Emergency Surg (2019) 46(4):1–25. doi: 10.1007/s00068-019-01235-w
- Land WG. The role of damage-associated molecular patterns in human diseases: Part I - Promoting inflammation and immunity. Sultan Qaboos Univ Med J (2015) 15:9–21.

- Roh JS, Sohn DH. Damage-associated molecular patterns in inflammatory diseases. *Immune Netw Korean Assoc Immunol* (2018) 18:1–14. doi: 10.4110/ in.2018.18.e27
- Hernandez C, Huebener P, Schwabe RF. Damage-associated molecular patterns in cancer: A double-edged sword. *Oncogene* (2016) 35:5931–41. doi: 10.1038/onc.2016.104
- Vénéreau E, Ceriotti C, Bianchi ME. DAMPs from cell death to new life. Front Immunol (2015) 6:422. doi: 10.3389/fimmu.2015.00422
- Schaefer L. Complexity of danger: The diverse nature of damage-associated molecular patterns. J Biol Chem (2014) 289:35237–45. doi: 10.1074/jbc.R114.619304
- Matzinger P. The danger model: A renewed sense of self. Science (2002) 296:301-5. doi: 10.1126/science.1071059
- Sun L, Wu J, Du F, Chen X, Chen ZJ. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. *Sci* (80) (2013) 339 (6121):786–91. doi: 10.1126/science.1232458
- Li T, Chen ZJ. The cGAS-cGAMP-STI NG pathway connects DNA damage to inflammation, senescence, and cancer. J Exp Med (2018) 215:1287–99. doi: 10.1084/jem.20180139
- Paludan SR, Bowie AG. Immune Sensing of DNA. *Immunity* (2013) 38:870– 80. doi: 10.1016/j.immuni.2013.05.004
- Härtlova A, Erttmann SF, Raffi FAM, Schmalz AM, Resch U, Anugula S, et al. DNA Damage Primes the Type I Interferon System via the Cytosolic DNA Sensor STING to Promote Anti-Microbial Innate Immunity. *Immunity* (2015) 42(2):332–43. doi: 10.1016/j.immuni.2015.01.012
- West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, et al. Mitochondrial DNA stress primes the antiviral innate immune response. *Nature* (2015) 520(7548):553–7. doi: 10.1038/nature14156
- Kanneganti T-D, Kundu M, Green DR. Innate Immune Recognition of mtDNA— An Undercover Signal? *Cell Metab* (2015) 21(6):793–4. doi: 10.1016/ j.cmet.2015.05.019
- Arnold RS, Fedewa SA, Goodman M, Osunkoya AO, Kissick HT, Morrissey C, et al. Bone metastasis in prostate cancer: Recurring mitochondrial DNA mutation reveals selective pressure exerted by the bone microenvironment. *Bone* (2015) 78:81–6. doi: 10.1016/j.bone.2015.04.046
- Alvarado-Vásquez N. Circulating cell-free mitochondrial DNA as the probable inducer of early endothelial dysfunction in the prediabetic patient. *Exp Gerontol* (2015) 69:70–8. doi: 10.1016/j.exger.2015.05.010
- Bray AW, Ballinger SW. Mitochondrial DNA mutations and cardiovascular disease. *Curr Opin Cardiol* (2017) 32(3):267–74. doi: 10.1097/HCO. 000000000000383
- Gu X, Yao Y, Wu G, Lv T, Luo L, Song Y. The Plasma Mitochondrial DNA Is an Independent Predictor for Post-Traumatic Systemic Inflammatory Response Syndrome. *PloS One* (2013) 8(8):e72834. doi: 10.1371/ journal.pone.0072834
- Yan HP, Li M, Lu XL, Zhu YM, Ou-yang W, Xiao ZH, et al. Use of plasma mitochondrial DNA levels for determining disease severity and prognosis in pediatric sepsis: a case control study. *BMC Pediatr* (2018) 18(1):267–74. doi: 10.1186/s12887-018-1239-z
- Glück S, Guey B, Gulen MF, Wolter K, Kang TW, Schmacke NA, et al. Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence. *Nat Cell Biol* (2017) 19(9):1061–70. doi: 10.1038/ncb3586
- Yang H, Wang H, Ren U, Chen Q, Chena ZJ. CGAS is essential for cellular senescence. Proc Natl Acad Sci U S A (2017) 114(23):E4612–20. doi: 10.1073/ pnas.1705499114
- Dou Z, Ghosh K, Vizioli MG, Zhu J, Sen P, Wangensteen KJ, et al. Cytoplasmic chromatin triggers inflammation in senescence and cancer. *Nature* (2017) 550(7676):402–6. doi: 10.1038/nature24050
- Adams PD, Ivanov A, Pawlikowski J, Manoharan I, Van Tuyn J, Nelson DM, et al. Lysosome-mediated processing of chromatin in senescence. J Cell Biol (2013) 202(1):129–43. doi: 10.1083/jcb.201212110
- Trzeciak A, Pietropaoli AP, Kim M. Biomarkers and Associated Immune Mechanisms for Early Detection and Therapeutic Management of Sepsis. *Immune Netw* (2020) 20(3):e23. doi: 10.4110/in.2020.20.e23
- Maślińska D. [Apoptosis: physiological cell death and its role in pathogenesis of diseases]. Neurol Neurochir Pol (2003) 37(2):315–26.
- Kerr JFR, Wyllie AH, Currie AR. Apoptosis: A basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer* (1972) 26 (4):239–57. doi: 10.1038/bjc.1972.33

- Tang D, Kang R, Vanden Berghe T, Vandenabeele P, Kroemer G. The molecular machinery of regulated cell death. *Cell Res* (2019) 29(5):347–64. doi: 10.1038/s41422-019-0164-5
- Gauthier VJ, Tyler LN, Mannik M. Blood clearance kinetics and liver uptake of mononucleosomes in mice. J Immunol (1996) 156(3):1151–6.
- Du Clos TW, Volzer MA, Hahn FF, Xiao R, Mold C, Searles RP. Chromatin clearance in C57B1/10 mice: Interaction with heparan sulphate proteoglycans and receptors on Kupffer cells. *Clin Exp Immunol* (1999) 117(2):403–11. doi: 10.1046/j.1365-2249.1999.00976.x
- Suzuki N, Kamataki A, Yamaki J, Homma Y. Characterization of circulating DNA in healthy human plasma. *Clin Chim Acta* (2008) 387(1–2):55–8. doi: 10.1016/j.cca.2007.09.001
- Van Der Vaart M, Pretorius PJ. The origin of circulating free DNA [1]. Clin Chem (2007) 53:2215. doi: 10.1373/clinchem.2007.092734
- 42. Savill J. CELL BIOLOGY: Eat Me or Die. Sci (80) (2003) 302(5650):1516–7. doi: 10.1126/science.1092533
- Tamkovich SN, Cherepanova AV, Kolesnikova EV, Rykova EY, Pyshnyi DV, Vlassov VV, et al. Circulating DNA and DNase activity in human blood. In: *Annals of the New York Academy of Sciences*. Blackwell Publishing Inc (2006) 1075:191–6. doi: 10.1196/annals.1368.026
- 44. Holdenrieder S, Eichhorn P, Beuers U, Samtleben W, Schoenermarck U, Zachoval R, et al. Nucleosomal DNA Fragments in Autoimmune Diseases. *Ann N Y Acad Sci* (2006) 1075(1):318–27. doi: 10.1196/annals.1368.043
- Chen Q, Ye L, Jin Y, Zhang N, Lou T, Qiu Z, et al. Circulating nucleosomes as a predictor of sepsis and organ dysfunction in critically ill patients. *Int J Infect Dis* (2012) 16(7):e558–64. doi: 10.1016/j.ijid.2012.03.007
- 46. Hayakwa M, Ooyasu T, Sadamoto Y, Saito T, Yoshida T, Katabami K, et al. Microparticles and Nucleosomes Are Released From Parenchymal Cells Destroyed After Injury in a Rat Model of Blunt Trauma. *Clin Appl Thromb* (2020) 26:1–8. doi: 10.1177/1076029620950825
- Kaufman T, Magosevich D, Moreno MC, Guzman MA, D'Atri LP, Carestia A, et al. Nucleosomes and neutrophil extracellular traps in septic and burn patients. *Clin Immunol* (2017) 183:254–62. doi: 10.1016/j.clim. 2017.08.014
- Van Montfoort ML, Stephan F, Lauw MN, Hutten BA, Van Mierlo GJ, Solati S, et al. Circulating nucleosomes and neutrophil activation as risk factors for deep vein thrombosis. *Arterioscler Thromb Vasc Biol* (2013) 33(1):147–51. doi: 10.1161/ ATVBAHA.112.300498
- Geiger S, Holdenrieder S, Stieber P, Hamann GF, Bruening R, Ma J, et al. Nucleosomes in serum of patients with early cerebral stroke. *Cerebrovasc Dis* (2006) 21(1–2):32–7. doi: 10.1159/000089591
- Holdenrieder S, Nagel D, Schalhorn A, Heinemann V, Wilkowski R, Von Pawel J, et al. Clinical relevance of circulating nucleosomes in cancer. In: *Annals of the New York Academy of Sciences*. Blackwell Publishing Inc (2008) 1137:180–9. doi: 10.1196/annals.1448.012
- Mittra I, Nair NK, Mishra PK. Nucleic acids in circulation: Are they harmful to the host? J Biosci (2012) 37:301–12. doi: 10.1007/s12038-012-9192-8
- Mittra I, Kumar Khare N, Venkata Raghuram G, Chaubal R, Khambatti F, Gupta D, et al. Circulating nucleic acids damage DNA of healthy cells by integrating into their genomes. J Biosci (2015) 40:91–111. doi: 10.1007/ s12038-015-9508-6
- Raghuram GV, Chaudhary S, Johari S, Mittra I. Illegitimate and repeated genomic integration of cell-free chromatin in the aetiology of somatic mosaicism, ageing, chronic diseases and cancer. *Genes* (2019) 10:407–25. doi: 10.3390/genes10060407
- 54. Mittra I, Samant U, Sharma S, Raghuram GV, Saha T, Tidke P, et al. Cell-free chromatin from dying cancer cells integrate into genomes of bystander healthy cells to induce DNA damage and inflammation. *Cell Death Discovery* (2017) 3(1):1–14. doi: 10.1038/cddiscovery.2017.15
- 55. Kirolikar S, Prasannan P, Raghuram GV, Pancholi N, Saha T, Tidke P, et al. Prevention of radiation-induced bystander effects by agents that inactivate cell-free chromatin released from irradiated dying cells. *Cell Death Dis* (2018) 9(12):1142–59. doi: 10.1038/s41419-018-1181-x
- Chousterman BG, Swirski FK, Weber GF. Cytokine storm and sepsis disease pathogenesis. Semin Immunopathol (2017) 39:517–28. doi: 10.1007/s00281-017-0639-8
- 57. Stortz JA, Hawkins RB, Holden DC, Raymond SL, Wang Z, Brakenridge SC, et al. Cell-free nuclear, but not mitochondrial, DNA concentrations correlate

with the early host inflammatory response after severe trauma. *Sci Rep* (2019) 9(1):13648–57. doi: 10.1038/s41598-019-50044-z

- Silk E, Zhao H, Weng H, Ma D. The role of extracellular histone in organ injury. Cell Death Dis (2017) 8(5):e2812. doi: 10.1038/cddis.2017.52
- Marsman G, Zeerleder S, Luken BM. Extracellular histones, cell-free DNA, or nucleosomes: Differences in immunostimulation. *Cell Death Dis* (2016) 7: e2518. doi: 10.1038/cddis.2016.410
- 60. Van Nieuwenhuijze AEM, Van Lopik T, Smeenk RJT. Time between onset of apoptosis and release of nucleosomes from apoptotic cells: putative implications for systemic lupus erythematosus. *Ann Rheum Dis* (2003) 62 (1):10–4. doi: 10.1136/ard.62.1.10
- Holdenrieder S, Stieber P, Bodenmüller H, Fertig G, Fürst H, Schmeller N, et al. Nucleosomes in serum as a marker for cell death. *Clin Chem Lab Med* (2001) 39(7):596–605. doi: 10.1515/CCLM.2001.095
- Holdenrieder S, Stieber P, Chan LYS, Geiger S, Kremer A, Nagel D, et al. Cell-free DNA in serum and plasma: Comparison of ELISA and quantitative PCR. *Clin Chem* (2005) 51(8):1544–6. doi: 10.1373/clinchem. 2005.049320
- Allam R, Kumar SVR, Darisipudi MN, Anders HJ. Extracellular histones in tissue injury and inflammation. J Mol Med (2014) 92:465–72. doi: 10.1007/ s00109-014-1148-z
- Gould TJ, Lysov Z, Liaw PC. Extracellular DNA and histones: Double-edged swords in immunothrombosis. J Thromb Haemostasis (2015) 13:S82–91. doi: 10.1111/jth.12977
- 65. Mittra I, Pal K, Pancholi N, Tidke P, Siddiqui S, Rane B, et al. Cell-free chromatin particles released from dying host cells are global instigators of endotoxin sepsis in mice. Mukhopadhyay P, editor. *PloS One* (2020) 15(3): e0229017. doi: 10.1371/journal.pone.0229017
- Rumore PM, Steinman CR. Endogenous circulating DNA in systemic lupus erythematosus. Occurrence as multimeric complexes bound to histone. J Clin Invest (1990) 86(1):69–74. doi: 10.1172/JCI114716
- Giglia-Mari G, Zotter A, Vermeulen W. DNA Damage Response. Cold Spring Harb Perspect Biol (2011) 3(1):a000745-a000745. doi: 10.1101/ cshperspect.a000745
- Rodgers K, Mcvey M. Error-Prone Repair of DNA Double-Strand Breaks. J Cell Physiol (2016) 231:15–24. doi: 10.1002/jcp.25053
- Frengen E, de Jong PJ, Brede G, Solheim J, Thomsen PD, Prydz H. The gene cluster containing the LCAT gene is conserved between human and pig. *Cytogenet Genome Res* (1997) 76(1–2):53–7. doi: 10.1159/000134515
- Gajecka M. Unrevealed mosaicism in the next-generation sequencing era. Mol Genet Genomics (2016) 291:513–30. doi: 10.1007/s00438-015-1130-7
- 71. King DA, Sifrim A, Fitzgerald TW, Rahbari R, Hobson E, Homfray T, et al. Detection of structural mosaicism from targeted and whole-genome sequencing data. *Genome Res* (2017) 27(10):1704–14. doi: 10.1101/ gr.212373.116
- 72. Forsberg LA, Rasi C, Razzaghian HR, Pakalapati G, Waite L, Thilbeault KS, et al. Age-related somatic structural changes in the nuclear genome of human blood cells. *Am J Hum Genet* (2012) 90(2):217–28. doi: 10.1016/j.ajhg.2011.12.009
- Jaiswal S, Fontanillas P, Flannick J, Manning A, Grauman PV, Mar BG, et al. Age-related clonal hematopoiesis associated with adverse outcomes. N Engl J Med (2014) 371(26):2488–98. doi: 10.1056/NEJMoa1408617
- 74. Dumanski JP, Lambert JC, Rasi C, Giedraitis V, Davies H, Grenier-Boley B, et al. Mosaic Loss of Chromosome y in Blood Is Associated with Alzheimer Disease. Am J Hum Genet (2016) 98(6):1208–19. doi: 10.1016/ j.ajhg.2016.05.014
- Laurie CC, Laurie CA, Rice K, Doheny KF, Zelnick LR, McHugh CP, et al. Detectable clonal mosaicism from birth to old age and its relationship to cancer. *Nat Genet* (2012) 44(6):642–50. doi: 10.1038/ng.2271
- Pahl HL. Activators and target genes of Rel/NF-κB transcription factors. Oncogene (1999) 18:6853–66. doi: 10.1038/sj.onc.1203239
- Hadian K, Krappmann D. Signals from the nucleus: Activation of NF-κB by cytosolic ATM in the DNA damage response. *Sci Signaling* (2011) 4:pe2. doi: 10.1126/scisignal.2001712
- Wan F, Lenardo MJ. Specification of DNA binding activity of NF-kappaB proteins. *Cold Spring Harbor Perspect Biol* (2009) 1:1–11. doi: 10.1101/ cshperspect.a000067

- Chaudhary S, Mittra I. Cell-free chromatin: A newly described mediator of systemic inflammation. J Biosci (2019) 44:1–6. doi: 10.1007/s12038-019-9849-7
- Chaudhary S, Raghuram GV, Mittra I. Is inflammation a direct response to dsDNA breaks? *Mutat Res Fundam Mol Mech Mutagen* (2018) 808:48–52. doi: 10.1016/j.mrfmmm.2018.02.002
- Mittra I, Pal K, Pancholi N, Shaikh A, Rane B, Tidke P, et al. Prevention of chemotherapy toxicity by agents that neutralize or degrade cell-free chromatin. Ann Oncol Off J Eur Soc Med Oncol (2017) 28(9):2119–27. doi: 10.1093/annonc/mdx318
- Czura CJ. "Merinoff symposium 2010: Sepsis"-speaking with one voice. Mol Med (2011) 17(1–2):2–3. doi: 10.2119/molmed.2010.00001.commentary
- Hotchkiss RS, Karl IE. The pathophysiology and treatment of sepsis. New Engl J Med (2003) 348:138–50. doi: 10.1056/NEJMra021333
- Rajaee A, Barnett R, Cheadle WG. Pathogen- A nd Danger-Associated Molecular Patterns and the Cytokine Response in Sepsis. Surg Infect (Larchmt) (2018) 19(2):107-16. doi: 10.1089/sur.2017.264
- Gentile LF, Moldawer LL. DAMPs, PAMPs, and the origins of SIRS in bacterial sepsis. *Shock* (2013) 39:113–4. doi: 10.1097/SHK.0b013e318277109c
- Denning NL, Aziz M, Gurien SD, Wang P. Damps and nets in sepsis. Front Immunol (2019) 10:2536. doi: 10.3389/fimmu.2019.02536
- Walborn AT, Patel P, Hoppensteadt D, Mosier M, Statz S, Rondina M, et al. Plasma Nucleosomes Correlate with Markers of Inflammation and Thrombosis in Patients with Sepsis-Associated Coagulopathy. *FASEB J* (2018) 31:674.15–5.
- Zeerleder S, Zwart B, Wuillemin WA, Aarden LA, Groeneveld ABJ, Caliezi C, et al. Elevated nucleosome levels in systemic inflammation and sepsis. *Crit Care Med* (2003) 31(7):1947–51. doi: 10.1097/01.CCM.0000074719.40109.95
- Filippou PS, Karagiannis GS. Cytokine storm during chemotherapy: A new companion diagnostic emerges? Oncotarg Impact J LLC (2020) 11:213–5. doi: 10.18632/oncotarget.27442
- Schaue D, Micewicz ED, Ratikan JA, Xie MW, Cheng G, McBride WH. Radiation and Inflammation. *Semin Radiat Oncol* (2015) 25:4–10. doi: 10.1016/j.semradonc.2014.07.007
- Barker CA, Kim SK, Budhu S, Matsoukas K, Daniyan AF, D'Angelo SP. Cytokine release syndrome after radiation therapy: Case report and review of the literature. *J Immunother Cancer* (2018) 6(1):1–7. doi: 10.1186/s40425-017-0311-9
- Krysko O, Aaes TL, Bachert C, Vandenabeele P, Krysko DV. Many faces of DAMPs in cancer therapy. *Cell Death Dis* (2013) 4:e631–1. doi: 10.1038/ cddis.2013.156
- Hossain MF, Hasana S, Mamun A, Uddin MS, Wahed MII, Sarker S, et al. COVID-19 Outbreak: Pathogenesis, Current Therapies, and Potentials for Future Management. *Front Pharmacol* (2020) 11:1–25. doi: 10.3389/ fphar.2020.563478
- Beltrán-García J, Osca-Verdegal R, Pallardó FV, Ferreres J, Rodríguez M, Mulet S, et al. Sepsis and Coronavirus Disease 2019: Common Features and Anti-Inflammatory Therapeutic Approaches. *Crit Care Med* (2020) 48(12):1– 4. doi: 10.1097/CCM.000000000004625
- Ribon M, Mussard J, Semerano L, Singer BB, Decker P. Extracellular chromatin triggers release of soluble CEACAM8 upon activation of neutrophils. *Front Immunol* (2019) 10(JUN):1-10. doi: 10.3389/ fimmu.2019.01346
- Kaneko N, Kuo H-H, Boucau J, Padera RF, Farmer JR, Allard-Chamard H, et al. Loss of Bcl-6-Expressing T Follicular Helper Cells and Germinal Centers in COVID-19. *Cell* (2020) 183:143–57. doi: 10.1016/j.cell.2020.08.025
- Lim S, Bae JH, Kwon H-S, Nauck MA. COVID-19 and diabetes mellitus: from pathophysiology to clinical management. *Nat Rev Endocrinol* (2021) 17 (1):11–30. doi: 10.1038/s41574-020-00435-4
- Bansal M. Cardiovascular disease and COVID-19. Diabetes Metab Syndr Clin Res Rev (2020) 14(3):247–50. doi: 10.1016/j.dsx.2020.03.013
- Butt AN, Shalchi Z, Hamaoui K, Samadhan A, Powrie J, Smith S, et al. Circulating nucleic acids and diabetic complications. In: *Annals of the New York Academy of Sciences*. Blackwell Publishing Inc (2006) 1075:258–70. doi: 10.1196/annals.1368.034
- 100. Borissoff JI, Joosen IA, Versteylen MO, Brill A, Fuchs TA, Savchenko AS, et al. Elevated Levels of Circulating DNA and Chromatin Are Independently Associated With Severe Coronary Atherosclerosis and a Prothrombotic

State. Arterioscler Thromb Vasc Biol (2013) 33(8):2032-40. doi: 10.1161/ ATVBAHA.113.301627

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Shabrish and Mittra. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.