# Articles

# Risk of strong antibody decline in dialysis and transplant patients after SARS-CoV-2mRNA vaccination: Six months data from the observational Dia-Vacc study

Julian Stumpf,<sup>a,b</sup> Jörg Schwöbel,<sup>c</sup> Tom Lindner,<sup>d</sup> Leona Anders,<sup>e</sup> Torsten Siepmann,<sup>f</sup> Claudia Karger,<sup>g</sup> Jan Hüther,<sup>h</sup> Heike Martin,<sup>i</sup> Petra Müller,<sup>j</sup> Robert Faulhaber-Walter,<sup>k</sup> Torsten Langer,<sup>1</sup> Holger Schirutschke,<sup>m</sup> Thomas Stehr,<sup>n</sup> Frank Meistring,<sup>o</sup> Annegret Pietzonka,<sup>p</sup> Kirsten Anding-Rost,<sup>q</sup> Katja Escher,<sup>r</sup> Frank Pistrosch,<sup>s</sup> Jens Schewe,<sup>t</sup> Harald Seidel,<sup>u</sup> Kerstin Barnett,<sup>v</sup> Thilo Pluntke,<sup>w</sup> Simon Cerny,<sup>x</sup> Alexander Paliege,<sup>a</sup> Ingolf Bast,<sup>e</sup> Anne Steglich,<sup>a</sup> Florian Gembardt,<sup>a</sup> Friederike Kessel,<sup>a</sup> Hannah Kröger,<sup>a</sup> Patrick Arndt,<sup>a</sup> Jan Sradnick,<sup>a</sup> Kerstin Frank,<sup>y</sup> Anna Klimova,<sup>z</sup> René Mauer,<sup>aa</sup> Xina Grählert,<sup>ab</sup> Torsten Tonn,<sup>ac,ad</sup> and Christian Hugo<sup>a,b,1\*</sup>

<sup>a</sup>Medizinische Klinik und Poliklinik III, Universitätsklinikum, Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany

- <sup>b</sup>KfH-Nierenzentrum Dresden, Dresden, Germany
- <sup>c</sup>Dialysezentrum Chemnitz, Chemnitz, Germany
- <sup>d</sup>Division of Nephrology, University Hospital Leipzig, Leipzig, Germany
- <sup>e</sup>Dialysepraxis Leipzig, Leipzig, Germany
- <sup>f</sup>KfH-Nierenzentrum am Klinikum Chemnitz, Krankenhaus Küchwald, Chemnitz, Germany
- <sup>9</sup>KfH-Nierenzentrum am Klinikum St. Georg, Leipzig, Germany
- <sup>h</sup>Nephrocare GmbH Döbeln, Döbeln, Germany
- <sup>i</sup>Nephrologisches Zentrum Zwickau, Zwickau, Germany
- <sup>j</sup>PHV Dialysezentrum Dresden-Johannstadt, Dresden, Germany
- <sup>k</sup>Nephrologisches Zentrum Freiberg, Freiberg, Germany
- <sup>1</sup>Dialysezentrum Annaberg, Annaberg-Buchholz, Germany
- <sup>m</sup>PHV Dialysezentrum Dresden Friedrichstadt, Dresden, Germany
- <sup>n</sup>KfH-Nierenzentrum Bautzen, Bautzen, Germany
- °KfH-Nierenzentrum am Städtischen Klinikum Görlitz, Görlitz, Germany
- <sup>p</sup>Via medis Nierenzentrum Dresden MVZ GmbH, Dresden, Germany
- <sup>q</sup>KfH-Nierenzentrum Bischofswerda, Bischofswerda, Germany
- <sup>r</sup>KfH-Gesundheitszentrum Aue, Aue-Bad-Schlema, Germany
- <sup>s</sup>Nephrologisches Zentrum Hoyerswerda, Hoyerswerda, Germany
- <sup>t</sup>Dialyse- und Nierenambulanz Sebnitz, Sebnitz, Germany
- <sup>u</sup>KfH-Nierenzentrum am Vogtland Krankenhaus Plauen, Plauen, Germany
- <sup>v</sup>Dialyse Heidenau, Heidenau, Germany
- <sup>w</sup>KfH-Nierenzentrum Grimma, Grimma, Germany
- <sup>x</sup>ELBLAND Dialyse Großenhain, Großenhain, Germany
- <sup>y</sup>Institut für Transfusionsmedizin Plauen, DRK-Blutspendedienst Nord-Ost Gemeinnützige GmbH, Plauen, Germany
- <sup>z</sup>National Center for Tumor Diseases (NCT) Partner Site Dresden, Dresden, Germany
- <sup>aa</sup>Faculty of Medicine Carl Gustav Carus, Institute for Medical Informatics and Biometry (IMB), Technische Universität, Dresden, Germany
- <sup>ab</sup>Coordinating Centre for Clinical Trials, Dresden, Germany
- <sup>ac</sup>Institute for Transfusion Medicine, German Red Cross Blood Donation Service North-East, Dresden, Germany
- <sup>ad</sup>Faculty of Medicine Carl Gustav Carus, Transfusion Medicine, Technische Universität, Dresden, Germany

\*Corresponding author at: Medizinische Klinik und Poliklinik III, Universitätsklinikum, Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany.

E-mail address: christian.hugo@uniklinikum-dresden.de (C. Hugo).

<sup>1</sup> Present address: Division of Nephrology, Department of Internal Medicine III, University Hospital Carl Gustav Carus of the Technische Universität Dresden, 01307 Dresden, Germany.



The Lancet Regional Health - Europe 2022;17: 100371 Published online xxx https://doi.org/10.1016/j. lanepe.2022.100371

# Summary

**Background** Vulnerable dialysis and kidney transplant patients show impaired seroconversion rates compared to medical personnel eight weeks after SARS-CoV-2mRNA vaccination.

**Methods** We evaluated six months follow up data in our observational Dia-Vacc study exploring specific cellular (interferon-γ release assay) or/and humoral immune responses after 2x SARS-CoV-2mRNA vaccination in 1205 participants including medical personnel (125 MP), dialysis patients (970 DP) and kidney transplant recipients (110 KTR) with seroconversion (*de novo* IgA or IgG antibody positivity by ELISA) after eight weeks.

**Findings** Six months after vaccination, seroconversion remained positive in 98% of MP, but 91%/87% of DP/KTR (p = 0.005), respectively. Receptor binding domain-IgG (RBD-IgG) antibodies were positive in 98% of MP, but only 68%/57% of DP/KTR (p < 0.001), respectively. Compared to MP, DP and KTR were at risk for a strong IgG or RBD-IgG decline (p < 0.001). Within the DP but not KTR group male gender, peritoneal dialysis, short time on dialysis, BNT162b2mRNA vaccine, immunosuppressive drug use and diabetes mellitus were independent risk factors for a strong decline of IgG or RBD antibodies. The percentage of cellular immunity decline was similar in all groups.

**Interpretation** Both vulnerable DP and KTR groups are at risk for a strong decline for IgG and RBD antibodies. In KTR, antibody titres peak at a markedly lower level and accelerated antibody decline is mixed with a delayed/increasing IgG, RBD-IgG, or cellular immune response in a 16% fraction of patients. In both populations, immune monitoring should be used for early timing of additional booster vaccinations.

Funding This study was funded by the Else Kröner Fresenius Stiftung, Bad Homburg v. d. H., grant number Fördervertrag EKFS 2021\_EKSE.27.

**Copyright** © 2022 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

Keywords: SARS-CoV-2mRNA vaccination; Antibody fading; Medical personnel; Dialysis patients; Kidney transplant recipients

#### **Research in context**

#### Evidence before this study

The vulnerable dialysis patients (DP) and kidney transplant recipients (KTR) experience a markedly increased COVID-19 disease related mortality rate compared to normal population. The most powerful mRNA vaccines BNT162b2mRNA or mRNA-1273 reach seroconversion and efficacy rates of about 95% in the general population. Immunocompromised DP unexpectedly showed vaccine-related seroconversion rates close to general population, but successful seroconversion in immunosuppressed KTR varies between 30% and 50% only. Even in successfully vaccinated persons, breakthrough COVID-19 infections and disease are possible and apparently play a substantial role in the pandemia. While COVID-19 breakthrough infections may be dependent on vaccination-related immune response quality and quantity, little information is known regarding humoral and/or cellular immunity fading in risk populations such as DP and KTR. To assess the availability of the data, results from database source National Library of Medicine(https://pubmed.ncbi.nlm.nih.gov) have been searched. For our search, following terms have been used: COVID-19, vaccination, SARS-CoV-2, BNT162b2, mRNA-1273, immunosuppression, antibody, humoral, T cells.

#### Added value of this study

Six months follow up data of our prospective Dia-Vacc study exploring specific cellular (interferon- $\gamma$  release assay) and humoral immune responses after SARS-CoV-2mRNA boost vaccination in 1205 participants including medical personnel (MP, 125), dialysis patients (970) and kidney transplant recipients (110) with successful de novo seroconversion after eight weeks were studied. Six months after vaccination start, seroconversion remained positive in 98% of MP and 91%/87% of DP/ KTR, respectively. Receptor binding domain (RBD-lgG-IgG) antibodies were positive in 98% of MP, but only 68%/57% of DP/KTR, respectively. Using 20% as a margin, only 41%/24% of MP but 68%/68% of DP, and 57%/ 55% of KTR showed decreased anti S1 IgG or RBD-IgG-IgG antibody titres between two and six months, respectively. 0%/1% of MP/KTR but 12-16% of transplant recipients showed IgG or RBD-IgG-IgG antibody increases up to six months. Similar results were found for the time course of cellular immunity measurements by the interferon-gamma release assay. DP and KTR were at high risk for a strong IgG or RBD-IgG-IgG decline. Within the DP but not KTR group male gender, peritoneal dialysis, BNT162b2mRNA vaccine, short time on dialysis, immunosuppressive drug use and diabetes mellitus were independent risk factors for a strong decline of IgG or RBD-IgG antibodies. Nevertheless, in all three study groups IgG and RBD-IgG antibody titres

at six months were higher in patients immunized with 2x mRNA-1273 compared to 2x BNT162b2mRNA. The positive effect of individual monitor guided early reboostering following a weak seroconversion response could be demonstrated in our KTR<sub>boost</sub> group (n = 20), where almost all patients with extremely low titre sero-conversion reacted with a marked antibody increase (instead of a decline) in both IgG and RBD-IgG-IgG antibodies up to six months.

#### Implications of all the available evidence

After successful SARS-CoV-2 mRNA vaccination, DP and KTR compared to MP are at specific risk for a strong antibody decline. In DP but not KTR, this humoral response is accelerated by male gender, diabetes mellitus as comorbidity, peritoneal dialysis, short time on dialysis, immunosuppressive drug use, and BNT162b2mRNA compared to mRNA-1273. In contrast, despite a delayed immune response with rising levels in a 16% patient fraction, successfully vaccinated KTR reach on average immunity levels markedly below MP and DP throughout the monitoring period of six months after first mRNA vaccination, but can be successfully boostered by a third vaccination when titres are quite low. This data indicate the value of immune monitoring especially in high-risk populations such as DP and KTR to define the best strategy and (early) timing of additional booster vaccinations even after positive seroconversion.

# Introduction

COVID-19 continues to pose an unprecedented challenge to global public health. Immunity conferred by vaccination can limit breakout of any infection and is an efficacious way to fight COVID-19 pandemic. Successful vaccination is in particular critical for high risk populations such as dialysis patients (DP) and kidney transplant recipients (KTR) experiencing COVID-19 disease related increased mortality rates compared to normal population.<sup>1</sup> The mRNA vaccines BNT162b2mRNA (Pfizer/BioNTech) or mRNA-1273 (Moderna) reach seroconversion and efficacy rates of about 95% in the general population. Immunocompromised DP unexpectedly showed vaccine-related seroconversion rates close to general population, but successful seroconversion in immunosuppressed KTR varies between 30% and 50% only.<sup>2–</sup>

Even in successfully vaccinated persons, breakthrough COVID-19 infections and disease are possible and apparently play a substantial role in the pandemic. While COVID-19 breakthrough infections may be dependent on vaccination-related immune response quality and quantity,<sup>6</sup> little information is known regarding immunity fading in risk populations such as DP and KTR. We hypothesized that DP and KTR after seroconversion are at risk for a strong immunity decline compared to medical personnel. The multicentre, investigator-driven, prospective observational Dia-Vacc study investigates the SARS-CoV-2-specific humoral as well as cellular immune response in DP and KTR and medical personnel at defined intervals after appropriate vaccination using 2x BNTI62b2mRNA or 2x mRNA-1273 as basic vaccination.<sup>4</sup> Here, we report the time course of cellular and/or humoral immunity measures in seroconverted study participants up to 6 months after vaccination.

# **Methods**

# Study design

The investigator-driven, multicentre, non-interventional, prospective, observational Dia-Vacc study (NCT number: 04799808) started with SARS-CoV-2 vaccination using either 2x BNT162b2mRNA or 2  $\times$  1273mRNA in 26 nephrology centres from January 15th to February 24th, exploring the time course of a specific cellular or/and humoral immune response to disease and/or SARS-CoV-2 vaccination in MP, DP, and KTR.4 Of all 36 dialysis centres in Saxony, being incorporated in a COVID-19 network since march 2020 as described,<sup>4</sup> the first 26 committing dialysis centres providing 3101 study participants were accepted for the DIA-Vacc study. Later requests could not be considered due to funding restrictions. Study start (To) was immediately before first vaccination. Further monitoring time points were three (BNT162b2mRNA) to four (mRNA-1273) weeks later before second vaccination (T1), about eight weeks after study start (T2; five to four weeks after the second vaccination respectively) and six months after study start (T3). By vaccine availability during January (BNT162b2mRNA) and February (mRNA-1273) 2021 only the first four dialysis centres being assigned the vaccination campaign, to received BNT162b2mRNA, while all other following dialysis centres received mRNA-1273 vaccine for both vaccinations. Neither any dialysis centre nor any participant nor the study centre (Dresden) had a choice or influence regarding the type of vaccine, which was assigned in the order of contacting the central vaccination institute in Saxony. Information to all dialysis centres about the start of the vaccination campaign was distributed by the central vaccination institute via email at the same time.<sup>4</sup>

For the six months (T<sub>3</sub>) study results reported here, the "pure vaccination cohort" with a positive seroconversion eight weeks (T<sub>2</sub>) (as defined by de novo IgA or IgG antibody positivity by ELISA) after 2 x vaccination were followed up to six months after vaccination start without any additional boostering in a total of 1205 participants including 125 MP, 970 DP, and 110 KTR. In addition, a subgroup of 20 seroconverted (at T<sub>2</sub>) KTR was evaluated, in which by individual decision an additional mRNA vaccine booster was given between T<sub>2</sub> and T<sub>3</sub> (KTR<sub>boost</sub>). For clinical outcome assessment all PCR- positive, symptomatically and asymptomatically (NCP seroconversion) COVID-19 diseased participants between T2 and T3 were counted. On the basis of immunity monitoring, all symptomatically and asymptomatically (NCP seroconversion) COVID-19 diseased participants before, during and after vaccination (up to T3) were excluded to assess a purely vaccination-related immune response. Patients were tested for SARS-CoV-2 infection by RT-PCR, in the dialysis centres, if they presented one of the classic symptoms (fever, cough, shortness of breath, myalgias, diarrhea, or other symptoms consistent with such an infection) or if they were in contact with a person with RT-PCR-confirmed disease. Routine PCR screening without a cause was not part of good medical practice of the dialysis centres.

In all study participants (eligibility if > 18 years old and signed informed consent) at To (vaccination start), T2, and T3, SARS-CoV-2 specific IgG- or IgA-antibody reactions (Euroimmun<sup>7–10</sup>) against the Spike protein subunit S1 and IgG-antibodies against the nucleocapsid protein subunit (NCP) were analysed. In addition, the receptor binding domain (RBD-IgG) antibody formation suggesting neutralising activity against the SARS-CoV-2 virus was also examined at T2 and T3.<sup>11</sup> For all antibody measurements Euroimmun ELISAs on Euroimmun analysers were used.

To provide characterisation of the cellular SARS-CoV-2 immune response, a SARS-CoV-2 specific interferon- $\gamma$  release assay (IGRA<sup>12</sup>) was performed at To, T<sub>2</sub>, and T<sub>3</sub> in representative subgroups.<sup>4</sup> Further details on procedures and analysis are found elsewhere.<sup>4</sup>

#### Ethic declaration

According to the professional code of conduct for doctors (§15) the clinical study was submitted to the ethical institutional review boards at Technische Universität Dresden (TU Dresden) responsible for the coordinating investigator (BO-EK-45012021), as well as at the University of Leipzig (046/21-lk) and Saxon Medical Association (Sächsische Landesärztekammer – EK-BR-10/21-1) responsible for further participating study sites.

## **End points**

The primary end point is the positive humoral immune response after vaccination as defined by *de novo* positivity of either IgG- or IgA- anti-SpikeS1 antibodies without development of virus-specific NCP antibodies. Secondary end points were the development of vaccination-induced *de novo* T-cellular immunity, the clinical outcome (COVID-19 disease), as well as serological and cellular immune response parameters and titres.

To investigate the time course (T2 to T3) of established vaccination – related *de novo* immunity reaction for the different tests, a 20% margin for (increased/ equal/decreased) antibody and IGRA titre/value development was used and the percentage of patients within each margin was calculated for each group and each time point.

In addition, the time course was analysed on the interval scale. The detectable ranges of anti-SI-IgG and RBD-IgG antibody values were categorised into five intervals, labeled from 0 to 4 (referred to as "levels" in the data analysis), and the change in levels, varying from -4 to +4, was calculated for each patient. Patients whose level decreased from T<sub>2</sub> to T<sub>3</sub> by more than one (or  $\geq$  two) unit were defined as a "strong decline" (Supplementary methods, SI Figure).

#### Statistical analysis

In the descriptive analysis of main study endpoints, categorical variables were summarised as absolute frequencies or percentages, and continuous variables were summarised using mean and standard deviation or median and interquartile range (IQR). Time trends in IgG and RBD-IgG responses as well as between-group differences were analysed either by the Wilcoxon signed-rank test, Mann-Whitney U test, or the chisquared test, as appropriate. The analysis of risk factors of patients with a strong antibody decline was carried out using multiple logistic regression. First, we fitted a logistic regression model to each study group separately. Because, as was observed in a number of studies,<sup>13,14</sup> a substantial difference in seroconversion response may occur after administering different vaccines, in each logistic regression model, we included the vaccine type as a risk factor of a strong antibody decline. Other potential risk factors, common to all study groups, were gender, age, and body mass index (BMI).<sup>2</sup> The age distribution differed considerably between study groups (see Table 1), and, therefore, in order to reduce a possible confounding effect, adjusting for age in all models was especially important. While no additional risk factors were considered for the MP group, the models for DP and KTR contained the effects of immunosuppression, hepatitis B vaccination failure, diabetes mellitus diagnosis, as well as group-specific effects: time on transplantation (KTR) and time on dialysis (DP). Because the immunosuppression is prevalent in KTR, with many patients taking several IS medications, the number of IS drugs was used as a covariate in the logistic-regression model for KTR. On the other hand, the IS prevalence in DP is relatively low, and, therefore, only an indicator of presence of IS was included in the logistic regression for DP. In order to estimate the group effect, we fitted a logistic regression model for all groups jointly, where only common risk factors were included. To investigate the effect of dialysis type on strong decline rates, we carried out a propensity score matching procedure for PD and HD patients (1:4 ratio, without resampling, logistic regression scores), based on the primary vaccine type and antibody

Variable	Category	МР	DP	KTR
Number	evaluable	125	970	110
Age (years)	$mean\pmSD$	$47.8 \pm 11.7$	$67.5 \pm 13.9$	$57.6 \pm 13$
Male Sex	n / %	28 / 22·4	636 / 65.6	69 / 62.7
BMI (kg/m²)	$mean\pmSD$	$26 \cdot 1 \pm 5 \cdot 1$	$27{\cdot}8\pm5{\cdot}8$	$26{\cdot}5\pm4{\cdot}7$
Cause of end stage renal disease	n / %	n.a.	797 / 82·2	70 / 63.6
Diabetes-Hypertension-Vascular disease	n / %	n.a.	481 / 49.6	19 / 17·3
Glomerulonephritis-Interstitial nephritis	n / %	n.a.	206 / 21.2	27 / 24.5
Vasculitis	n / %	n.a.	24 / 2.5	2 / 1.8
Polycystic kidney disease	n / %	n.a.	86 / 8.9	22 / 20
Unknown	n / %	n.a.	173 / 17.8	40 / 36·4
Drug treated comorbidities	n / %	29 / 23·2	933 / 96·2	95 / 86·4
Diabetes mellitus	n / %	4/3.2	348 / 35.9	24 / 21.8
Cardiovascular disease	n / %	23 / 18·4	903 / 93.1	90 / 81.8
Lung disease	n / %	5/4	58 / 6	6 / 5.5
Liver cirrhosis	n / %	0/0	13 / 1.3	1 / 0.9
Cancer	n / %	0/0	38 / 3.9	5 / 4.5
None	n / %	96 / 76·8	37 / 3.8	15 / 13.6
Type of dialysis		n.a.	970 / 100	n.a.
Hemodialysis	n / %	n.a.	929 / 95.8	n.a.
Peritonealdialysis	n / %	n.a.	41 / 4·2	n.a.
Time on dialysis (years)	mean $\pm$ SD	n.a.	$5.8 \pm 5.8$	$5.9\pm5.9$
On transplant waiting list	n/%	n.a.	118 / 12-2	n.a.
Time on transplantation (years)	mean $\pm$ SD	n.a.	n.a.	$11.7\pm7.8$
Previous transplantation	n / %	n.a.	68 / 7	19 / 17·3
Hepatitis B vaccination failure	n / %	2 / 1.6	202 / 20.8	12 / 10.9
Flu vaccination winter 2020/21	n / %	74 / 59·2	713 / 73.5	58 / 52·
On immunosuppressive therapy	n / %	1 / 0.8	34 / 3-5	109 / 99·1
Corticosteroids	n / %	0 / 0	24 / 2.5	47 / 42.7
Calcineurin-Inhibitor	n / %	0 / 0	8 / 0.8	95 / 86·4
MMF/MPA	n / %	0 / 0	4 / 0.4	62 / 56·4
mTOR-Inhibitor	n / %	0 / 0	1 / 0.1	23 / 20.9
Belatacept	n / %	0 / 0	1 / 0.1	3 / 2.7
T-cell depleting ab	n / %	0 / 0	0 / 0	0/0
B-cell depleting ab	n / %	0 / 0	0 / 0	0/0
Other	n / %	1 / 0.8	3 / 0.3	4 / 3.6
Type of vaccine				
BNT162b2 mRNA	n / %	36 / 28.8	151 / 15.6	29 / 26·4
mRNA-1273	n / %	89 / 71·2	819 / 84-4	81 / 73.6

Table 1: Baseline characteristics of SARS-CoV-2 unexposed, but at T2 seroconverted persons / patients of the DIA-Vacc pure vaccination cohort.

For this evaluation all patients with asymptomatic\* or documented symptomatic\*\* COVID-19 disease before and during vaccination up to T<sub>3</sub> (six months) were excluded. Hepatitis B vaccination failure definition - patients with unsuccessful vaccination after at least four attempts; MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; MMF-MPA = mycophenolate mofetil or mycophenolic acid;

\*Asymptomatic COVID-19 disease definition - neither knowledge nor symptoms of COVID-19 disease, but IgG-antibody reaction to nucleocapsid (To, T1, T2, or T3) or to the Spike protein subunit S1 (only To) of the SARS-CoV-2 virus is positive.

\*\*Symptomatic COVID-19 disease definition - SARS-CoV-2 PCR positive patients with clinical symptoms.

values (IgG or RBD for the corresponding analyses) at T2. Also notable is a comparison between boostered and unboostered KTR was also implemented via propensity score matching (I:I ratio, without resampling, logistic regression scores, based on IgG value at T2), see Table 6. For hypothesis testing, the significance level of 5% (two-sided) was chosen. A Bonferroni correction was applied during posthoc testing of group effects.

To check the robustness of our main results against missing data, we applied a multiple imputation procedure, namely, multiple imputation with chained equations (MICE),<sup>15</sup> to impute IgG antibody values of 102 patients (Figure 1) who missed blood sampling at T3. For each cohort separately and for all participants pulled together, we generated ten data sets of the corresponding cohort size, that is, 40 data sets in total. The



variables that were considered as risk factors for antibody decline, see Tables 4A-4D were used for imputation of missing IgG values at T3. Table S1 displays the proportions of patients by cohort whose IgG values remained at the same level/decreased/ increased with respect to 20% margin. We then proceeded with categorization of IgG values according to our interval scale and identified patients who experienced a strong decline in IgG. After that, to each imputed data set we applied exactly the same multiple logistic regression model as the one used for the complete data in the corresponding analysis cohort (MP, DP, KTR, all patients together). For each cohort, the ten effect estimates for each risk factor were averaged on the log scale and then converted to odds ratios. The resulting estimates, with a 95% confidence interval, are provided along with the summary obtained from complete data, see Tables 4A-4D. Antibody values for patients who died between T2 and T3 time points were not included in imputations, because within our study scope, it is not possible to estimate a confounding effect of a particular death reason, while ignoring such an effect could be a potential source of bias.

Data analysis was implemented in the R Environment for Statistical Computing,<sup>16</sup> version 4.0.4. R script can be provided upon request.

# Results

# Basic study cohort characteristics (Table 1)

Of more than 3100 original study participants, a total of 1225 participants fulfilled all "pure vaccination cohort" requirements combined with a positive de novo seroconversion at eight weeks, of whom 125 MP, 970 DP, 110 KTR, and 20 KTR<sub>boost</sub> were monitored up to six months after vaccination. A study flow chart indicates very few group changes and reasons for patient exclusions between T2 and T3 (Figure 1). Patient characteristics of the three large subgroups are shown at Table 1. For the following results it needs to be considered that de novo seroconversion definition at T2 included either IgA or IgG positivity and also a different proportion of RBD-IgG/IGRA positivity within the groups (100%/86% of MP, 95%/78% of DP but only 67%/30% of KTR), respectively.<sup>4</sup> IgA antibody levels are calculated by a ratio and only represent semiquantitative values. The results of subgroup KTR<sub>boost</sub> are summarised separately.

#### Study end points

Immune response rates six months after vaccination (T3) in the seroconverted pure vaccination cohort (Table 2). At the six month time point, seroconversion of IgG or IgA anti-SI antibodies as our primary study endpoint remained positive in 98% of MP, 91% of DP,

and 87% of KTR (p = 0.006), respectively (Table 2). Hereby, the IgG antibody positivity was with 98% of MP, 89% of DP, and 84% of KTR the predominant denominator of a sustained seroconversion reaction up to month 6 (Table 2). In contrast, only 77% of MP, 57%% of DP, and 59% of KTR still showed antibody positivity for anti-SI IgA antibodies at this time point (p < 0.001). RBD-IgG antibodies stayed positive in 98% of MP, but only in 68%/57% of DP/KTR (p < 0.001), respectively. T-cellular immunity to vaccination as indicated by a positive IGRA response was seen in 79% of MP, 62% in DP, and 37% of KTR at the six month time point (p < 0.001) being comparable to rates at two months.

Using 20% as a margin, only 41%/24% of MP but 68%/68% of DP, and 57%/55% of KTR showed decreased anti SI IgG or RBD-IgG antibody titres between two and six months, respectively (p < 0.001). 0%/1% of MP/DP but 12-16% of transplant recipients showed IgG or RBD-IgG antibody increases up to six months (p < 0.001). This delayed antibody response is also reflected by 6% of seroconverted KTR, who are characterised by a de novo RBD-IgG positivity between two and six months, respectively. Therefore, 59%/76% of MP, but only 31%/31% of DP and 27%/34% of KTR remained at an equal level of IgG or RBD-IgG antibodies against S1 protein between month two and six, respectively (p < 0.001). IgA anti S1 protein antibody ratios showed a stronger decline than IgG/RBD-IgG, since 98% of MP, 97% of DP and 95% of KTR demonstrated decreased values at six compared to two months. Increased or de novo IgA antibody ratios between two and six months were rare events in all three groups.

Cellular immunity monitoring via IGRA measurements showed similar rates of decreasing titres around 60% in all groups. Substantial rates (35%) of IGRA titre increases were solely seen in KTR up to six months indicating the delayed response of KTR also in regard to Tcell immunity (Table 2). Only up to 6% of KTR showed *de novo* positivity for IGRA measurements at six months.

The time course of anti-SpikeS1 protein IgG or RBD-IgG antibody titres of all seroconverted Dia-Vacc study participants differs between the study groups. Table 3A summarises the average titre levels at two and six months (median  $\pm$  interquartile range) within the different groups demonstrating largely different antibody and IGRA titres already at T<sub>2</sub> but also different degrees of immunity fading up to T<sub>3</sub>. The higher percentage of anti-S<sub>1</sub> IgG antibody levels below the positivity limit of 35·2 BAU/ml in the DP and KTR groups compared to the MP group can be appreciated in Figure 2A. Solely in the KTR group, antibody decline is mixed with IgG titre increases or some *de novo* positivity in up to 16% of patients between T<sub>2</sub> and T<sub>3</sub> (Figure 2A). A very similar

Variable	Category	Medical personnel	Dialysis patients	Kidney transplant recipients	<i>p</i> -value (chi-squared test)
Patient number	n	125	970	110	<u> </u>
Humoral responses	n of total n (%)				
IgG-Ab or IgA-Ab Spike S1 positive	n of total n (%)	123 / 125 (98·4%)	879 / 970 (90.6%)	96 / 110 (87·3%)	0.005
IgA-Ab Spike S1 positive	n of total n (%)	96 / 125 (76·8%)	537 / 938 (57-2%)	65 / 110 (59.1%)	< 0.001
IgA-Ab Spike S1 increasing	n of total n (%)	1 / 125 (0.8%)	20 / 970 (2.1%)	4 / 110 (3.6%)	0.313
lgA-Ab Spike S1 equal	n of total n (%)	2 / 125 (1.6%)	11 / 970 (1.1%)	2 / 110 (1.8%)	0.771
IgA-Ab Spike S1 decreasing	n of total n (%)	122 / 125 (97.6%)	939 / 970 (96-8%)	104 / 110 (94.5%)	0.379
De novo IgA-Ab positivity	n of total n (%)	0 / 125 (0%)	5 / 970 (0.5%)	2 / 110 (1.8%)	0.156
(T2 negative, T3 positive)					
IgG-Ab Spike S1 positive	n of total n (%)	123 / 125 (98-4%)	865 / 970 (89-2%)	92 / 110 (83.6%)	0.001
IgG-Ab Spike S1 increasing	n of total n (%)	0 / 125 (0%)	13 / 970 (1.3%)	17 / 110 (15.5%)	< 0.001
lgG-Ab Spike S1 equal	n of total n (%)	74 / 125 (59·2%)	297 / 970 (30.6%)	30 / 110 (27.3%)	< 0.001
IgG-Ab Spike S1 decreasing	n of total n (%)	51 / 125 (40.8%)	660 / 970 (68%)	63 / 110 (57·3%)	< 0.001
De novo IgG-Ab positivity	n of total n (%)	0 / 125 (0%)	1 / 970 (0.1%)	3 / 110 (2.7%)	< 0.001
(T2 negative, T3 positive)					
RBD positive	n of total n (%)	122 / 125 (97.6%)	656 / 970 (67.6%)	63 / 110 (57.3%)	< 0.001
RBD increasing	n of total n (%)	0 / 116 (0%)	7 / 908 (0.8%)	12 / 104 (11.5%)	< 0.001
RBD equal	n of total n (%)	88 / 116 (75·9%)	282 / 908 (31.1%)	35 / 104 (33.7%)	< 0.001
RBD decreasing	n of total n (%)	28 / 116 (24.1%)	619 / 908 (68-2%)	57 / 104 (54.8%)	< 0.001
De novo RBD positive	n of total n (%)	0 / 116 (0%)	3 / 908 (0.3%)	6 / 104 (5.8%)	< 0.001
(T2 negative, T3 positive)					
Interferon- $\gamma$ release assay (IGRA)– T	-cellular response				
IGRA positive	n of total n (%)	22 / 28 (78.6%)	72 / 116 (62·1%)	15 / 41 (36.6%)	0.001
IGRA increasing	n of total n (%)	1 / 29 (3·4%)	7 / 102 (6·9%)	12 / 34 (35·3%)	< 0.001
IGRA equal	n of total n (%)	10 / 29 (34.5 %)	33 / 102 (32·4%%)	1 / 34 (2·9%)	0.002
IGRA decreasing	n of total n (%)	18 / 29 (62·1%)	62 / 102 (60.8%)	21 / 34 (61.8%)	0.989
De novo positivity for IGRA	n of total n (%)	0 / 29 (0%)	2 / 102 (2%)	2 / 34 (5·9%)	0.282

Table 2: Immune response rates six months after vaccination (T3) in the seroconverted pure vaccination cohort.

MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; Interferon- $\gamma$  release assay = IGRA;

Humoral vaccination responses were assessed as positive, when de novo production of the antibody to the Spike SI (IgA or IgG) protein or RBD (IgG) subunit was above positivity level. A positive T-cellular response to vaccination as assessed by interferon- $\gamma$  release assay (IGRA) turned from a negative result on To to positive on T<sub>3</sub>, respectively ( $\geq 100 \text{ mIU/ml}$ , as being recommended by the manufactures). Using 20% as a margin, the time course of antibody or IGRA tires at T<sub>3</sub> compared to T<sub>2</sub> time point were categorized into increased (> 20%), equal (within 20% range), and decreased (< 20%). For this evaluation, all participants with asymptomatic of documented symptomatic\*\* COVID-19 disease before and during vaccination up to T<sub>3</sub> (six months) were excluded. \*Asymptomatic COVID-19 disease definition - neither knowledge nor symptoms of COVID-19 disease, but IgG-antibody reaction to nucleocapsid (To, T<sub>2</sub> or

T<sub>3</sub>) or to the Spike protein subunit S<sub>1</sub> (only T<sub>0</sub>) of the SARS-CoV-2 virus is positive.

\*\*Symptomatic COVID-19 disease definition - SARS-CoV-2 PCR positive patients with clinical symptoms.

pattern can be observed for the time course of vaccination-related RBD-IgG antibodies in the different study groups (Figure 2B).

Since the primary seroconversion rates in DP and KTR were markedly vaccine dependent, Figure 2 C/D as well as Table 3B shows averaged titre levels of IgG/RBD-IgG antibodies at two and six months separated by both mRNA vaccines, respectively. In MP, the average IgG levels were similar for both vaccines at two months, but remained higher at six months for mRNA-1273 compared with BNT162b2mRNA (Figure 2C, Table 3B). In DP but not KTR (only a trend) groups, mean IgG titres were higher for mRNA-1273 than for BNT162b2mRNA at two and remained even more noticeably different at six months (Figure 2C, Table 3B). A very similar vaccine-dependent pattern in regard to both levels and time

courses was seen for RBD-IgG or IgA antibodies in each study group (Figure 2D, Table 3B). In contrast, IGRA levels did not differ in a vaccine dependent matter in all groups (Table 3B).

Multivariate analysis of risk factors of a strong antibody decline in DP and KTR cohorts compared to the MP cohort. In order to further explore risk factors of a decline in humoral response, an interval categorization of anti-SI protein IgG and RBD antibody values was introduced. These categories, labelled from o to 4 and referred to as "levels" in the data analysis, where level o was assigned to IgG and RBD values below the positivity threshold of 35.2 BAU/ml and 35%, respectively, and the remaining values were split uniformly into four



Figure 2. Time course of anti-SARS-CoV-2 IgG (A/C) or receptor binding domain (B/D) antibodies in seroconverted Dia-Vacc

intervals of approximately equal length. Such a categorisation takes into account the truncated structure of IgG and RBD-IgG distributions and allows for a more comparable interpretation of IgG and RBD-IgG results. Under this approach, the quantification of change between T<sub>2</sub> and T<sub>3</sub> in IgG and RBD-IgG values can be performed on the same scale, simply by calculating the difference between the patient's level at T<sub>3</sub> and his/her level at T<sub>2</sub>. With all changes ranging from -4 to +4, a

study participants. Figure 2A: IgG against S1 protein in different study groups. Each thin line corresponds the anti-Spike S1 protein IgG antibody values (QuantiVac, Euroimmun) of a study participant from T0 (vaccination start) via T2 (eight weeks after vaccination start) to T3 (six months after vaccination start). Only patients with successful de novo seroconversion at T2 (IgA or IgG antibody positivity against the SARS-CoV-2 S1 protein) after 2x mRNA vaccination and without SARS-CoV-2 nucleocapsid (NCP) antibodies were considered. All three patient groups are represented (MP-green, DP-red, KTR-blue). The area shaded grey designates IgG range below positivity level. The vertical axis is depicted on log<sub>10</sub> scale with corresponding unit BAU/ml. Figure 2B: Receptor binding domain antibodies against S1 protein in different study groups. Each thin line corresponds the anti-Spike S1 protein RBD-lgG antibody values (Euroimmun) of a study participant from T2 (eight weeks after vaccination start) to T3 (six months after vaccination start). Only patients with successful de novo seroconversion at T2 (IgA or IgG antibody positivity against the SARS-CoV-2 S1 protein) after 2x mRNA vaccination and without SARS-CoV-2 nucleocapsid (NCP) antibodies were considered. All three patient groups are represented (MP-green, DP-red, KTR-blue). The area shaded grey designates IgG range below positivity level. The vertical axis is depicted on log<sub>10</sub> scale with corresponding unit % inhibition. Figure 2C: IgG against S1 protein dependent on vaccine type. Each thin line corresponds the anti-Spike S1 protein IgG antibody values (QuantiVac, Euroimmun) of a study participant from T0 (vaccination start) via T2 (eight weeks after vaccination start) to T3 (six months after vaccination start). The thicker lines represent average responses by vaccine type (red-BNT162b2mRNA and blue 1273-mRNA) in each group. Only patients with successful de novo seroconversion at T2 (IgA or IgG antibody positivity against the SARS-CoV-2 S1 protein) after 2x mRNA vaccination and without SARS-CoV-2 nucleocapsid (NCP) antibodies were considered. The area shaded grey designates IgG range below positivity level. The vertical axis is depicted on log<sub>10</sub> scale with corresponding unit BAU/ml. Figure 2D: Receptor binding domain antibody against S1 protein dependent on vaccine type. Each thin line corresponds the anti-Spike S1 protein RBD-IgG antibody values (Euroimmun) of a study participant from T0 (vaccination start) via T2 (eight weeks after vaccination start) to T3 (six months after vaccination start). The thicker lines represent average responses by vaccine type (red-BNT162b2mRNA and blue 1273-mRNA) in each group. Only patients with successful de novo seroconversion at T2 (IgA or IgG antibody positivity against the SARS-CoV-2 S1 protein) after 2x mRNA vaccination and without SARS-CoV-2 nucleocapsid (NCP) antibodies were considered. The area shaded grey designates IgG range below positivity level. The vertical axis is depicted on log<sub>10</sub> scale with corresponding unit % inhibition.

Humoral responses           IgA-Ab Spike S1         MP         Median (interquartile range)         9 (5-8–9)         3·31 (1·5–5·9)         < 0.001           IgA-Ab Spike S1         DP         Median (interquartile range)         7·87 (4·0–9)         1.84 (0·8–4·3)         < 0.001           IgA-Ab Spike S1         DP         Median (interquartile range)         7·87 (4·0–9)         1.84 (0·8–4·1)         < 0.001           IgA-Ab Spike S1         KTR         Median (interquartile range)         4·6 (2–9)         1·8 (0·8–4·1)         < 0.001           IgG-Ab Spike S1         MP         Median (interquartile range)         384 (384–384)         351·1 (239–384)         < 0.001           IgG-Ab Spike S1         DP         Median (interquartile range)         384 (384–384)         192·9 (86–356)         < 0.001           IgG-Ab Spike S1         KTR         Median (interquartile range)         329·8 (110·7–384)         121·1 (51·8–295·6)         < 0.001           IgG-Ab Spike S1         KTR         Median (interquartile range)         99·33 (98·7–99·6)         89 (79·2–95·9)         < 0.001           RBD         MP         Median (interquartile range)         98·1 (91·2–99·4)         57·7 (24·8–84·9)         < 0.001           RBD         DP         Median (interquartile range)         98·1 (91·2–99·4) </th <th>Variable</th>	Variable
IgA-Ab Spike S1         MP         Median (interquartile range)         9 (5·8–9)         3·31 (1·5–5·9)         < 0.001           IgA-Ab Spike S1         DP         Median (interquartile range)         7·87 (4·0–9)         1·84 (0·8–4·3)         < 0.001	Humoral responses
IgA-Ab Spike S1         DP         Median (interquartile range)         7.87 (4.0-9)         1.84 (0.8-4.3)         < 0.001           IgA-Ab Spike S1         KTR         Median (interquartile range)         4.6 (2-9)         1.8 (0.8-4.1)         < 0.001	lgA-Ab Spike S1
IgA-Ab Spike S1         KTR         Median (interquartile range)         4-6 (2-9)         1-8 (0-8-4-1)         < 0.001           IgG-Ab Spike S1         MP         Median (interquartile range)         384 (384-384)         351-1 (239-384)         < 0.001	lgA-Ab Spike S1
IgG-Ab Spike S1         MP         Median (interquartile range)         384 (384–384)         351·1 (239–384)         < 0.001           IgG-Ab Spike S1         DP         Median (interquartile range)         384 (384–384)         192·9 (86–356)         < 0.001	lgA-Ab Spike S1
IgG-Ab Spike S1         DP         Median (interquartile range)         384 (384–384)         192-9 (86–356)         < 0.001           IgG-Ab Spike S1         KTR         Median (interquartile range)         329-8 (110-7–384)         121·1 (51.8–295.6)         < 0.001	lgG-Ab Spike S1
IgG-Ab Spike S1         KTR         Median (interquartile range)         329-8 (110.7-384)         121.1 (51.8-295.6)         < 0.001           RBD         MP         Median (interquartile range)         99.33 (98.7-99.6)         89 (79.2-95.9)         < 0.001	lgG-Ab Spike S1
RBD         MP         Median (interquartile range)         99-33 (98-7-99-6)         89 (79-2-95-9)         < 0.001           RBD         DP         Median (interquartile range)         98-1 (91-2-99-4)         57-7 (24-8-84-9)         < 0.001	lgG-Ab Spike S1
RBD         DP         Median (interquartile range)         98-1 (91-2-99-4)         57-7 (24-8-84-9)         < 0.001           PRD         KTP         Median (interquartile range)         69.6 (32-2-95-5)         40.1 (18.7-71.9)         < 0.001	RBD
PRD KTP Modian (interguartile range) 60.6 (38.3–05.5) (0.1 (18.7–71.0) < 0.001	RBD
	RBD
IGRA- T-cellular response	IGRA— T-cellular resp
IGRA MP Median (interquartile range) 2282·1 (874·6-2487·7) 554·6 (307·2-2461·5) < 0.001	IGRA
IGRA DP Median (interquartile range) 888-7 (270-4–2479) 335-0 (79-5–1161-4) < 0-001	IGRA
IGRA         KTR         Median (interquartile range)         113·3 (14·3-326·4)         48·0 (15·9-176·1)         0·055	IGRA

Table 3A: Antibody and IGRA titres two (T2) and six months (T3) after vaccination in the seroconverted pure vaccination cohort.

MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; Interferon- $\gamma$  release assay = IGRA;

This table compares the average titre levels (median/interquartile range) on T3 with T2 (different columns) for the different anti-Spike S1 IgA, IgG, RBD-IgG antibodies as well as for cellular immunity via IGRA measurements. For this evaluation, all participants with asymptomatic\* or documented symptomatic\*\* COVID-19 disease before and during vaccination up to T3 (six months) were excluded.

\*Asymptomatic COVID-19 disease definition - neither knowledge nor symptoms of COVID-19 disease, but IgG-antibody reaction to nucleocapsid (To, T2 or T3) or to the Spike protein subunit S1 (only To) of the SARS-CoV-2 virus is positive.

\*\*Symptomatic COVID-19 disease definition - SARS-CoV-2 PCR positive patients with clinical symptoms.

"strong antibody decline" in either IgG or RBD-IgG was defined as a decrease by at least two levels between T2 and T3. See Supplementary Appendix for a more detailed description.

The analysis of between-group differences and potential risk factors of strong antibody decline was performed using logistic regression, applied to the study groups jointly (all seroconverted study participants) as well as to each group separately. The group effect in IgG decline was found to be highly significant ( $\chi^2(2) = 22.258$ , p < 0.001). In particular, the chance of strong decline in IgG antibodies for DP or KTR was found to be about three or two times higher than for MP participants, respectively. In addition, the DP group showed a more pronounced trend towards antibody decline than the KTR group (p = 0.097, after adjustment for multiple comparison) (Table 4A).

Separate analyses by study group produced the following results: In the MP group, the vaccine type was found to be the only risk factor of strong antibody decline, with BNT162b2mRNA being associated with a five fold higher risk as compared to mRNA-1273 (p = 0.004) (Table 4B). Within the DP but not KTR group, male gender, BNT162b2mRNA (compared to mRNA-1273) vaccine, immunosuppressive drug use, diabetes mellitus as comorbidity, and a short time on dialysis were found to be independent risk factors of a strong IgG decline after vaccination (Table 4C). None of risk factors, which were included in the logistic regression analysis for the KTR group, had a statistically significant effect on strong antibody decline of IgG antibodies (Table 4D). A detailed summary of fit results is given in Tables 4A–4D. One can also notice the coherence between effect estimates obtained before and after imputation. Lower odds ratios of group effects are likely due to the imputed IgG values being closer to the average, and therefore, while a strong decline is associated to extreme cases.

The same approach was used to investigate potential risk factors of strong RBD-IgG antibody decline (Table 5). The group effect was again found to be highly significant  $(\chi^2(2) = 40.478, p < 0.001, Table 5A)$ . Similar to IgG, the chance of a strong decline in RBD-IgG antibodies for DP and KTR, was found to be about five or three times higher than for MP participants (p < 0.001, Table 5A), respectively. The strong antibody decline in DP was more than two times higher than in KTR (p = 0.004). Minor influences/trends for the vaccine type or age were found in MP. Within the DP but not KTR group again male gender and BNT162b2mRNA vaccine, and a short time on dialysis were risk factors of a strong RBD-IgG decline after vaccination. In contrast to IgG, immunosuppressive drug use in DP did not appear to be a risk factor of strong RBD-IgG decline. As in the IgG case for KTR, none of considered potential risk factors (including immunosuppressive drug number) of a strong RBD-IgG antibody decline were found statistically significant. A detailed summary is provided in Tables 5A–5D.

To investigate the effect of dialysis type on strong decline rates, we carried out a propensity score matching procedure for PD and HD patients. In the matched data, PD patients had three times higher chances than HD patients to experience strong decline in IgG

Variable	Group	Time point	Category	BNT162b2 mRNA	mRNA-1273	<i>p</i> -value (Mann-Whitney test)
Humoral response	s					
lgA-Ab Spike S1	MP	T2	Median (interquartile range)	4-3 (2-3-6-8)	9 (8-4-9)	< 0.001
lgA-Ab Spike S1	MP	T3	Median (interquartile range)	1.7 (1.0-3.3)	4-2 (2-3-6)	< 0.001
lgA-Ab Spike S1	DP	T2	Median (interquartile range)	3.7 (1.5-7.1)	8.8 (5.1–9)	< 0.001
lgA-Ab Spike S1	DP	T3	Median (interquartile range)	0.9 (0.4-1.8)	2.2 (1.0-4.8)	< 0.001
lgA-Ab Spike S1	KTR	T2	Median (interquartile range)	2.9 (1.5-5.7)	5.8 (2.6–9)	0.007
lgA-Ab Spike S1	KTR	T3	Median (interquartile range)	1.6 (0.6–2.8)	2.0 (0.9-4.7)	0.051
lgG-Ab Spike S1	MP	T2	Median (interquartile range)	384 (384–384)	384 (384–384)	0.12
lgG-Ab Spike S1	MP	Т3	Median (interquartile range)	241.5 (101.1-325.2)	384 (266-7-384)	< 0.001
lgG-Ab Spike S1	DP	T2	Median (interquartile range)	384 (194-3-384)	384 (384–384)	< 0.001
lgG-Ab Spike S1	DP	T3	Median (interquartile range)	68.6 (25.1-133.5)	224.3 (108.7-384)	< 0.001
lgG-Ab Spike S1	KTR	T2	Median (interquartile range)	201.9 (90.1-384)	384 (126-9-384)	0.086
lgG-Ab Spike S1	KTR	T3	Median (interquartile range)	99.8 (31.9-184.5)	125.0 (65.6–344.8)	0.051
RBD	MP	T2	Median (interquartile range)	98.7 (974–99.4)	99.4 (99.1–99.6)	< 0.001
RBD	MP	T3	Median (interquartile range)	82.1 (69.3–91.6)	91.6 (83.1-97.2)	0.001
RBD	DP	T2	Median (interquartile range)	80.6 (57.8–95.3)	98.4 (93.5-99.4)	< 0.001
RBD	DP	T3	Median (interquartile range)	27.8 (9.9-47.5)	63.8 (31.6-87.5)	< 0.001
RBD	KTR	T2	Median (interquartile range)	56.9 (32.6-77.8)	76.5 (40.5–97.5)	0.018
RBD	KTR	T3	Median (interquartile range)	35.9 (16.9–48.4)	45.5 (21.7-81.5)	0.072
IGRA- T-cellular r	esponse					
IGRA	MP	T2	Median (interquartile range)	1593.3 (740.2-2474.5)	2413.8 (1454.7-2490.3)	0.198
IGRA	MP	T3	Median (interquartile range)	453.1 (200.9–1183.5)	700.0 (481.2-2461.5)	0.144
IGRA	DP	T2	Median (interquartile range)	732.6 (154.5–2479.1)	1021.17 (366.4–2477.4)	0.364
IGRA	DP	T3	Median (interquartile range)	256.9 (34.7-1387.1)	428.9 (101.6-1161.4)	0.485
IGRA	KTR	T2	Median (interquartile range)	138-4 (14-5-326-4)	43.8 (11.1–282.6)	0.554
IGRA	KTR	Т3	Median (interquartile range)	75.6 (12.8–173.8)	25.9 (16.0-244.4)	0.627

# Table 3B: Antibody and IGRA titres two (T2) and six months (T3) after vaccination in the seroconverted pure vaccination cohort dependent on vaccine type.

MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; Interferon- $\gamma$  release assay = IGRA;

This table compares the average titre levels (median/interquartile range) of 2x BNT162b2mRNA or 2×1273-mRNA (different columns) vaccinated study participants on T2 (two months) and T3 (six months) (different columns) for the different anti-Spike S1 IgA, IgG, RBD-IgG antibodies as well as for cellular immunity via IGRA measurements. For this evaluation, all participants with asymptomatic\* or documented symptomatic\*\* COVID-19 disease before and during vaccination up to T3 (six months) were excluded.

\*Asymptomatic COVID-19 disease definition - neither knowledge nor symptoms of COVID-19 disease, but IgG-antibody reaction to nucleocapsid (To, T2 or T3) or to the Spike protein subunit S1 (only T0) of the SARS-CoV-2 virus is positive.

\*\*Symptomatic COVID-19 disease definition - SARS-CoV-2 PCR positive patients with clinical symptoms.

OR	95% CI	<i>p</i> -value	OR, 95%CI (MICE)
1.007	[0.998,1.016]	0.142	1.008[1.007,1.009]
1.412	[1.095,1.820]	0.008	1.421[1.383,1.460]
2.546	[1.855,3.496]	<0.001	2.535[2.483,2.587]
3.237	[1.897,5.525]	<0.001	2.953[2.766,3.153]
1.926	[1.018,3.644]	0.044	1.702[1.615,1.793]
1.304	[1.005,1.691]	0.045	1.289[1.268,1.310]
	OR 1.007 1.412 2.546 3.237 1.926 1.304	OR         95% CI           1.007         [0.998,1.016]           1.412         [1.095,1.820]           2.546         [1.855,3.496]           3.237         [1.897,5.525]           1.926         [1.018,3.644]           1.304         [1.005,1.691]	OR         95% Cl         p-value           1.007         [0.998,1.016]         0.142           1.412         [1.095,1.820]         0.008           2.546         [1.855,3.496]         <0.001

 Table 4A: Multiple logistic regression analysis of a strong decline of vaccination-specific anti-S1 IgG antibodies in seroconverted

 participants of the DIA-Vacc pure vaccination cohort between T2 and T3. Table 4A. Strong IgG decline for all participants (n = 1205).

 MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; Ref. = reference category; a "strong IgG strong fading response antibody

 decline" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material.

 Comparator is the MP cohort. MICE means multiple imputation with chained equations.

 Model fit: AIC = 1556.70; BIC = 1592-35.

Age         1.037         [0.993,1.082]         0.101         1.028[1.021,1.034]           Sex (Ref. = female)         1.611         [0.515,5.042]         0.413         1.930[1.710,2.180]           Vaccine type (Ref. = mRNA-1273)         4.567         [1.638,12.735]         0.004         5.213[4.636,5.863]	Risk factor	Estimate	95% CI	<i>p</i> -value	OR, 95%CI (MICE)
Sex (Ref. = female)         1.611         [0.515,5.042]         0.413         1.930[1.710,2.180]           Vaccine type (Ref. = mRNA-1273)         4.567         [1.638,12.735]         0.004         5.213[4.636,5.863]	Age	1.037	[0.993,1.082]	0.101	1.028[1.021,1.034]
Vaccine type (Ref. = mRNA-1273)         4-567         [1-638,12-735]         0-004         5.213[4.636,5.863]	Sex (Ref. = female)	1.611	[0.515,5.042]	0.413	1.930[1.710,2.180]
	Vaccine type (Ref. = mRNA-1273)	4.567	[1.638,12.735]	0.004	5.213[4.636,5.863]
BMI 0.929 [0.819,1.052] 0.245 0.925[0.911,0.939]	BMI	0.929	[0.819,1.052]	0.245	0.925[0.911,0.939]

#### Table 4B: Strong IgG antibody decline for medical personnel (n = 125).

Ref. = reference category; a "strong IgG antibody declinestrong fading response" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. BMI means body mass index. MICE means multiple imputation with chained equations.

Model fit: AIC = 107.44; BIC = 121.42.

Risk factor	OR	95% CI	<i>p</i> -value	OR,95%CI (MICE)
Age	1.005	[0.995,1.014]	0.367	1.003[1.002,1.004]
Sex (Ref. = female)	1.445	[1.089,1.916]	0.011	1.421[1.401,1.442]
Vaccine type (Ref. = mRNA-1273)	2.684	[1.836,3.923]	< 0.001	2.677[2.591,2.767]
BMI	0.990	[0.967,1.014]	0.428	0.988[0.986,0.990]
IS drugs (Ref. = no drugs)	3.052	[1.360,6.849]	0.007	3.340[3.107,3.590]
Time on dialysis	0.977	[0.953,1.001]	0.058	0.977[0.975,0.979]
Hep B vacc failure (Ref. = no)	1.002	[0.723,1.388]	0.990	0.965[0.945,0.986]
Diabetes mellitus (Ref. = none)	1.386	[1.041,1.845]	0.025	1.389[1.356,1.424]

#### Table 4C: Strong IgG antibody decline for dialysis patients (n = 970).

Ref. = reference category; a "strong IgG antibody declinestrong fading response" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. Comparator is the MP cohort. IS means immunosuppressiveon; BMI means body mass index; Hep B vacc failure definition - patients with unsuccessful vaccination after at least four attempts. MICE means multiple imputation with chained equations.

Model fit: AIC = 1277.30; BIC = 1325.86.

Risk factor	OR	95% CI	<i>p</i> -value	OR, 95%CI (MICE)
Age	1.038	[0.997,1.080]	0.069	1.034[1.027,1.041]
Sex (Ref. = female)	1.008	[0.403,2.523]	0.987	0.882[0.815,0.955]
Time after transplantation	0.987	[0.925,1.052]	0.686	1.000[0.994,1.007]
Vaccine type (Ref. = mRNA-1273)	1.556	[0.585,4.133]	0.376	1.361[1.247,1.485]
BMI	1.049	[0.954,1.153]	0.327	1.019[1.004,1.035]
IS drugs number	0.988	[0.505,1.933]	0.972	0.853[0.801,0.909]
Diabetes mellitus	0.630	[0.221,1.797]	0.388	0.592[0.561,0.625]

Table 4D: Strong IgG antibody decline for kidney transplant recipients (n = 110).

Ref. = reference category; a "strong IgG antibody declinestrong fading response" between T<sub>2</sub> (two months after first vaccination) and T<sub>3</sub> (six months) was defined as described in Results and in more detail in Supplementary material. IS means immunosuppressive drug; BMI means body mass index. MICE means multiple imputation with chained equations.

Model fit: AIC = 144.18; BIC = 168.06

(OR = 3·10 [1·06; 10·39], Fisher test, p = 0.024). RBD strong decline in PD group was numerically but not statistically about two times more likely, in comparison to HD (OR = 2·46 [0·68, 11·38], Fisher test, p = 0.176).

#### Clinical outcome between T2 and T3

Only one dialysis and one transplant patient experienced mild symptomatic and nine dialysis patients asymptomatic COVID-19 disease (NCP seroconversion) between two and six months after vaccination. No single patient had to be hospitalised or died due to COVID-19 disease.

#### Results of the KTR<sub>boost</sub> group

All patients of this group showed very low seroconversion values at T2 with either IgA positivity and no IgG positivity or very low/negative levels of RBD-IgG antibody values stimulating the individual decision for an additional boostering at  $4\cdot 2 \pm 1$  months despite

Between T2 and T3, only very few group changes took

place (Figure 1). No allograft failure was reported between

two and six months. 35 deaths not related to COVID-19

were noted between two and six months (Figure 1).

Risk factor	OR	95% CI	<i>p</i> -value
Age	1.012	[1.002,1.021]	0.013
Sex (Ref. = female)	1.463	[1.125,1.902]	0.005
Vaccine type (Ref. = mRNA-1273)	2.147	[1.470,3.136]	<0.001
DP (Ref. = MP)	5.435	[2·876,10·271]	<0.001
KTR (Ref. = MP)	2.533	[1·213,5·290]	0.013
Diabetes Mellitus	1.214	[0.929,1.587]	0.155

Tables 5A: Multiple logistic regression analysis of a strong decline of vaccination-specific anti-SpikeS1 receptor binding domain (RBD-lgG) antibodies in seroconverted participants of the DIA-Vacc pure vaccination cohort between T2 and T3. Table 5A. Strong RBD-lgG antibody decline for all participants (*n* = 1128).

MP = Medical Personnel; DP = Dialysis Patients; KTR = Kidney Transplant Recipient; Ref. = reference category; a "strong RBD-IgG antibody decline" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. Comparator is the MP cohort.

Model fit: AIC = 1446.97; BIC = 1482.16

Risk factor	OR	95% CI	<i>p</i> -value
Age	1.065	[1.006,1.129]	0.032
Sex (Ref. = female)	1.204	[0·299,4.853]	0.794
Vaccine type (Ref. = mRNA-1273)	3.320	[0.921,11.963]	0.067
BMI	0.927	[0.792,1.085]	0.346

Table 5B: Strong RBD-IgG antibody decline for medical personnel (n = 116).

Ref. = reference category; a "strong RBD-IgG antibody decline" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. Comparator is the MP cohort. IS means immunosuppression; BMI means body mass index. Model fit: AIC = 80.38; BIC = 93.97

Risk factor	OR	95% CI	<i>p</i> -value
Age	1.009	[0.999,1.019]	0.087
Sex (Ref. = female)	1.441	[1.081,1.922]	0.013
Vaccine type (Ref. = mRNA-1273)	2.592	[1.621,4.143]	<0.001
BMI	0.983	[0.959,1.008]	0.188
IS drugs (Ref. = no drugs)	0.894	[0.398,2.008]	0.787
Time on dialysis	0.961	[0.937,0.986]	0.003
Hep B vacc failure	1.185	[0.846,1.659]	0.323
Diabetes mellitus (Ref. = none)	1.251	[0.934,1.676]	0.134

Table 5C: Strong RBD-IgG antibody decline for dialysis patients (n = 908).

Ref. = reference category; a "strong RBD-IgG antibody decline" " between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. Comparator is the MP cohort. IS means immunosuppressive; BMI means body mass index; Hep B vacc failure definition - patients with unsuccessful vaccination after at least four attempts.

Model fit: AIC = 1211.87; BIC = 1259.81

seroconversion. The IgA antibody ratio (mean/interquartile range) changed from 1.8 (1.1–3.0) at T2 to 2.0 (1.6–6) at T3, the IgG titres from 13.7 BAU/ml (4.1 –22.3) to 290.4 BAU/ml (75.4–384), and the RBD-IgG levels from 8.1% (4.7–12.9) to 89.3% (28.7–98.6). At six months, 90% of patients were still positive for either IgA or IgG antibodies, 85%/90%/65% were positive for IgA/IgG/RBD-IgG antibodies, respectively. In contrast to the unboostered KTR group, only 10.3% qualified for a strong antibody decline between two and six months, while 65%/85%/86% showed increased (by > 20%) IgA/IgG/RBD-IgG antibody levels for this time period. No patient developed *de novo* IgA, but 75% *de novo* IgG and 64% RBD-IgG antibodies between T2 and T3. One patient experienced asymptomatic COVID-19 disease with NCP antibody conversion. To get a closer look at the booster effect, we used propensity scores to match the KTR<sub>boost</sub> group with unboostered KTR (I:I ratio, without resampling). Only their IgG values at T2 were used for matching, as it reflected on the actual selection procedure used by the clinics to select KTRs for booster vaccination. A summary of seroconversion markers for

OR	95% CI	<i>p</i> -value
1.017	[0.973,1.064]	0.458
1.559	[0.550,4.419]	0.403
0.987	[0.920,1.058]	0.703
1.492	[0.522,4.262]	0.455
1.064	[0.499,2.267]	0.872
1.096	[0.987,1.267]	0.086
4.037	[0.903,18.042]	0.068
0.717	[0·231,2·658]	0.854
	OR 1.017 1.559 0.987 1.492 1.064 1.096 4.037 0.717	OR         95% CI           1.017         [0.973,1.064]           1.559         [0.550,4.419]           0.987         [0.920,1.058]           1.492         [0.522,4.262]           1.064         [0.499,2.267]           1.096         [0.987,1.267]           4.037         [0.903,18.042]           0.717         [0.231,2.658]

#### Table 5D: Strong RBD-IgG antibody decline for kidney transplant recipients (n = 104).

Ref. = reference category; a "strong RBD-IgG antibody decline" between T2 (two months after first vaccination) and T3 (six months) was defined as described in Results and in more detail in Supplementary material. Comparator is the MP cohort. IS means immunosuppressive drug; BMI means body mass index; Hep B vacc failure definition - patients with unsuccessful vaccination after at least four attempts. Model fit: AIC = 127.16; BIC = 127.16

time	KTR <sub>boost</sub>	Matched KTR (unboostered)	<i>p</i> -value
Т2	1.8 (1.1-3.0)	1.8 (0.8–3.8)	0.766
Т3	2 (1.6–6)	1.0 (0.5–2.8)	0.019
T2	13.7 (4.1–22.3)	41.6 (6.3–61.7)	0.022
Т3	290.4 (75.4–384)	51.8 (17.2–124.6)	0.017
T2	8.1 (4.7–12.9)	19.0 (16.9–28.6)	< 0.001
Т3	89.3 (28.7-98.6)	17.8 (7.1–43.5)	0.004
	time T2 T3 T2 T3 T2 T2 T3	time         KTR <sub>boost</sub> T2         1.8 (1.1–3.0)           T3         2 (1.6–6)           T2         13.7 (4.1–22.3)           T3         290.4 (75.4–384)           T2         8.1 (4.7–12.9)           T3         89.3 (28.7–98.6)	time         KTR <sub>boost</sub> Matched KTR (unboostered)           T2         1.8 (1.1–3.0)         1.8 (0.8–3.8)           T3         2 (1.6–6)         1.0 (0.5–2.8)           T2         13.7 (4.1–22.3)         41.6 (6.3–61.7)           T3         290.4 (75.4–384)         51.8 (17.2–124.6)           T2         8.1 (4.7–12.9)         19.0 (16.9–28.6)           T3         89.3 (28.7–98.6)         17.8 (7.1–43.5)

#### Table 6: Antibody response in matched KTR cohorts.

The KTR<sub>boost</sub> group (n = 20) is defined as a very low seroconversion group with values at T<sub>2</sub> (eight weeks) with either IgA positivity and no IgG positivity or IgG/IgA positivity and low/negative levels of RBD-IgG antibody values stimulating the individual decision for an additional boostering between T<sub>2</sub> and T<sub>3</sub> (six months). Propensity scores were used to match the KTR<sub>boost</sub> group with unboostered KTR (I:I ratio). Vaccination-related antibody titre levels were compared in both groups at seroconversion (T<sub>2</sub>) and after six months (T<sub>3</sub>).

the matched groups is given in Table 6 and shows the marked differences in the time course of antibody titres of the KTR<sub>boost</sub> versus unboostered group.

# Discussion

Comparative immunity fading data are scarce but highly relevant considering especially that DP and KTR belong to the most vulnerable patient populations exposed to markedly increased COVID-19 related mortality rates.<sup>1,4</sup>

In our Dia-Vacc study six months after vaccination start, almost all study participants of the MP group with seroconversion after two months still possessed positive anti-SI IgG and RBD-IgG antibodies, but in the seroconverted DP or KTR groups, up to 16%/43% of patients were negative for anti-SI IgG/RBD-IgG antibodies at six months, respectively. This indicates severe differences of antibody development and decline in the immunocompromised and immunosuppressed risk groups of DP and KTR compared to the MP group, which we further analysed in this study.

Hereby, it needs to be considered that on average the antibody titres two months after vaccination were highest in the MP, intermediate in DP, and lowest in KTR

groups.<sup>4</sup> Using different measures (20% titre decrease or a strong decline qualification - see Methods), IgG or RBD-IgG antibody decline between two and six months was smallest in MP, highest in DP and intermediate in KTR. Interestingly, almost nobody of the MP or DP groups showed IgG or RBD-IgG titres with more than a 20% increase from two to six months, but about up to 16% of KTR did. This apparently reflects a unique delayed immune response, which appears to be aggravated in a part of the immunosuppressed transplant patients and includes even some delayed de novo positivity for RBD-IgG antibodies at six months despite IgG or IgA seroconversion at two months. Similar to this finding, about 15% of KTR without a positive seroconversion two months after vaccination experience a delayed seroconversion up to six months without any additional vaccine booster.17 These results are consistent with other studies in KTR, where compared to a healthy control group a delayed humoral response specific for IgG antibody formation and secretion was seen after vaccination against influenza<sup>18</sup> or after COVID-19 disease.<sup>19</sup> Whether the immunosuppressive drugs belatacept and MMF/MPA being predominantly responsible for impaired seroconversion rates in KTR in our Dia-Vacc<sup>4</sup> and other studies3 also cause a delayed humoral

response with accelerated antibody decline, should be explored in future studies.

Similar to the humoral response, the T-cellular immunity response as determined by IGRA measurements was most frequent and highest in the MP, intermediate in the DP, and lowest in the KTR group at two<sup>4</sup> but still also at six months. Between two and six months, IGRA titres decreased in about 60% of patients in all three groups. Solely in the KTR group, about a third of patients showed increased IGRA titres between two and six months, mimicking the delayed antibody response also at the T-cellular level.

Analysis of risk factors for a strong IgG or RBD-IgG antibody decline between two and six months identified the DP and also the KTR group at risk compared to the MP group.

While most differences of the mRNA vaccine types have been found regarding seroconversion rates in patient study groups exposed to therapeutics altering the immune response,<sup>3,4</sup> in this study BNT162b2mRNA (compared to mRNA-1273) seemed to be a risk factor of strong IgG or RBD-IgG antibody decline (besides age) in the MP group. Within the KTR group, all of the considered patient characteristics, including immunosuppressive drug number, could not be identified as risk factors of a strong antibody decline.

In the DP group, several risk factors for a strong antibody decline were identified. The vaccine type BNT162b2mRNA was associated with a 2.5 fold increased risk for both IgG and RBD-IgG decline adding another reason for using mRNA-1273 as the preferred vaccine type in DP besides higher seroconversion rates and peak antibody titres.4,20 The use of immunosuppressive drugs and diabetes mellitus as a comorbidity were risk factors of IgG but not of RBD-IgG antibody decline in the DP cohort. This difference between IgG and RBD-IgG antibody decline cannot be explained but suggests differential immune response influences under/due to immunosuppression or diabetes mellitus. Male gender and short time on dialysis, but not BMI were associated with an increased risk of a strong decline in both IgG and RBD-IgG antibodies between two and six months. While it was unexpected that short time on dialysis could be identified as a risk factor for antibody decline after seroconversion, it was also associated with increased risk for seroconversion failure.<sup>4</sup> This suggests that either the humoral immune system may adapt/recover during longer times on dialysis therapy in stable DP or that this may reflect a "survival of the fittest" effect. Consistent with hepatitis B vaccination studies,<sup>21</sup> the dialysis type was linked to a higher risk of a strong decline in both IgG and RBD-IgG antibodies, with peritoneal dialysis being associated with a four times higher risk of strong antibody decline than hemodialysis. Whether this antibody decline difference relates to potential differences in the clearance of uremia related substances is unclear. In contrast,

seroconversion rates for both SARS-CoV-2<sup>22</sup> or Hepatitis B<sup>23</sup> vaccination are equivalent in hemodialysis and peritoneal dialysis patients.

Most importantly, between month two and six after vaccination, no single seroconverted study participant experienced any severe COVID-19 disease. While this region in Germany was exposed to the third wave pandemic with high COVID-19 incidences,<sup>24</sup> only 12 breakthrough infections occurred in this large study population. Symptomatic breakthrough infections are known to be related to lower titre levels.<sup>6</sup> Nevertheless, to define general antibody cutoff values for full COVID-19 protection may not be feasible considering antibody test heterogeneity,<sup>25</sup> individual SARS-CoV-2 infectious particle load and route, different virus variants, the degree of additional T-cell immunity, and the dynamic time course of immunity.

Our study highlights great immune response variability with specific characteristics in different patient populations implicating the usefulness of antibody (and T-cell immune) monitoring especially in DP and KTR to determine timely reboostering. In contrast to our MP group only six months after the start of mRNA vaccination, two thirds of successfully seroconverted DP show a strong antibody decline, which is dependent on individual risk factors such as male gender, diabetes mellitus as comorbidity, dialysis type, immunosuppressive drug use, but also vaccine type. mRNA-1273 could be identified as the vaccine with the highest antibody levels during that follow up period. In DP, T-cell immunity fading as measured by IGRA appeared at a similar rate as in MP, but on average titre levels were bisected. In contrast, successfully vaccinated KTR are also at an increased risk for a strong antibody decline when compared with the MP group. Unique to the KTR group is the mixture of (humoral and T-cellular) immunity fading and rise, which fits to a delayed immune response in at least a 16% fraction of immunosuppressed transplant patients. Besides these two intercalating effects, KTR in general reach antibody and IGRA levels markedly below the "normal" immunocompetent or even the immunocompromised DP population during the six months monitoring period. The positive effect of individual monitor guided early reboostering following a weak seroconversion response could be demonstrated in our KTR<sub>boost</sub> group, where almost all patients with extremely low titre seroconversion reacted with a marked antibody increase (instead of a decline) in both IgG and RBD-IgG antibodies apparently providing clinical protection up to six months.

Our study has several limitations including the observational, non-randomized study character and the lack of demographic matching between different cohorts. Although the former cannot be controlled for and may lead to hidden bias, the latter was accounted for in the multivariate analyses by including demographic factors as covariates. Due to the complete lack of mRNA vaccination related immunity monitoring data in dialysis and transplant patients at study start, no sample size calculation was performed for this study. Financial resources of the study determined that only 3101 participants in 26 out of 36 available dialysis centres of the Saxonian network could be recruited as described in methods. 102 patients missed blood sampling at T3. Using a multiple imputation procedure, conclusions remained consistent across all imputed sets and to those obtained based on the pure vaccination cohort. Further limitations apply to individual determination of ethnicity, race and income/occupation as well as socio-economic position, factors known to be associated with infection risk and risk of KTR or DP (but not regarding antibody fading). In addition, only mRNA vaccines were given to all study participants excluding heterologous vaccination strategies. Many different test systems for SARS-CoV-2 related humoral or T-cellular immunity are commercially available and may not give interchangeable results to our study. Nevertheless, it can be considered a study strength that all participants have been examined centralised with the same system even on the same analysers including a complete panel of antibodies to differentiate disease and vaccination immunity. Applying de novo NCP positivity for counting and exclusion of asymptomatic COVID-19 breakthrough infections from further analysis may underestimate cases.<sup>25</sup> Finally, the T-cell response was not measured in all study participants.

In conclusion, this study shows that after successful SARS-CoV-2 mRNA vaccination, DP and KTR compared to MP are at specific risk for a strong antibody decline. In DP but not KTR, this fading response is accelerated by male gender, diabetes mellitus as comorbidity, peritoneal dialysis, immunosuppressive drug use, and BNT162b2mRNA compared to mRNA-1273. In contrast, despite a delayed immune response with rising levels in a 16% patient fraction, successfully vaccinated KTR reach on average immunity levels markedly below MP and DP throughout the monitoring period of six months, but can be successfully boostered by a third vaccination when titres are low (as in the KTR<sub>boost</sub> group). This data indicates the value of immune monitoring especially in high risk populations such as DP and KTR to define the best strategy and (early) timing of additional booster vaccinations even after positive seroconversion.

## Contributors

JS and CH contributed to study design, data collection, data interpretation, and drafting of the manuscript and contributed to data interpretation and drafting of the manuscript.

AS, FG, FK, HK, PA, JSr, KF, TT were involved in data acquisition and collection or study organization. AK, RM, XG, were involved in statistical analysis or data management of the study. JSc, TL, LA, TS, CK, JH, HM, PM, RF-W, TLa, HS, TSt, FM, AP, KA-R, KE, FP, JeS, HSe, KB, TP, SC, APa, IB were involved in patient recruitment and data collection. All authors have approved the final version for submission.

### **Declaration of interests**

JS, JSc, TL, LA, TS, CK, JH, HM, PM, RF-W, TLa, HS, TSt, FM, AP, KA-R, KE, FP, JeS, HSe, KB, TP, SC, APa, IB, AS, FG, FK, HK, PA, JSr, KF, AK, RM, XG, and TT have no conflict of interests.

## Acknowledgment

We acknowledge the support from the DIA-VACC Investigators (see full list in Appendix).

#### Further supporters

D. Michael Albrecht, Medizinischer Vorstand (Sprecher), Universitätsklinikum Carl Gustav Carus Dresden

Thomas Grünewald, Chemnitz, Vorsitzender Sächsische Impfkommission

Patricia Klein, Ärztliche Geschäftsführerin, Sächsische Landesärztekammer Körperschaft des öffentlichen Rechts

Marcus Strotkötter, Senior Operations Officer for Vaccination Center and Mobile Vaccination Teams at DRK Landesverband Sachsen e.V.

EUROIMMUN Medizinische Labordiagnostika AG, Lübeck, Germany provided antibody ELISAs and interferon-gamma release assays for this study.

#### Supplementary materials

Supplementary material associated with this article can be found in the online version at doi:10.1016/j. lanepe.2022.100371.

# Appendix A. \*DIA-Vacc- investigators

Members in alphabetical order within the corresponding centres:

<sup>1</sup>Medizinische Klinik und Poliklinik III, Universitätsklinikum, Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany

Andrea Angermann, Nicole Bunk, Susanne Dollfus, Petya Drinova, Annegret Fleischer, Chris Gebauer, Susann Graichen, Kerstin Haaser, Sabrina Heide, Gabriele Hirt, Franziska Klemt, Jens Passauer, Miriam Poser, Margrit Reinhardt, Ronny Rettig, Vladimir Todorov, Kerstin Zäumer

<sup>3</sup>Dialysezentrum Chemnitz, Chemnitz, Germany

Markus Gläser, Ulrike Richardt, Anja Richter, Kathrin Scheibner, Andreas Sohn <sup>4</sup>Division of Nephrology, University Hospital Leipzig, Leipzig, Germany

Anette Bachmann, Jonathan Christoph de Fallois, Jan Halbritter, Claudia Kreyig, Christof Mayer, Benjamin Sandner

<sup>5</sup>Dialysepraxis Leipzig, Leipzig, Germany

Daniel Grey

<sup>6</sup>KfH-Nierenzentrum am Klinikum Chemnitz, Krankenhaus Küchwald, Chemnitz, Germany

Angela Grzegorek, Markus Hallmann, Saskia Härtwig, Christiane Hintzen-Kruse, Frauke Neidt und Söhne, Susann Reissland, Bianca Seidel, Anett Speer, Peggy Wei, Christine Wiech

<sup>7</sup>KfH-Nierenzentrum am Klinikum St. Georg, Leipzig, Germany

Joachim Beige, Grit Glombig, Kathrin Heipmann, Christian Schauff, Peyo Kolev Sivenov

<sup>8</sup>Nephrocare GmbH Döbeln, Döbeln, Germany

Noureddin Ibrahim

<sup>9</sup>Nephrologisches Zentrum Zwickau. Zwickau. Germany

Sebastian Höhne, Konrad Mager, Grit Müller, Ingrid Petzold, Alexander Sämann

<sup>10</sup>PHV Dialysezentrum Dresden-Johannstadt, Dresden, Germany

Steffen Bischoff, Tatjana Seitz

<sup>11</sup>Nephrologisches Zentrum Freiberg, Freiberg, Germany Anja Sawalha, Emad Sawalha

<sup>12</sup>Dialysezentrum Annaberg, Annaberg-Buchholz, Germany

Rainer Giese

<sup>13</sup>PHV Dialysezentrum Dresden Friedrichstadt, Dresden, Germany

Roland Behnisch, Peter Grossmann, Peter Heduschka

<sup>14</sup>KfH-Nierenzentrum Bautzen, Bautzen, Germany

Norgit Meyer, Sybille Stehr, Patrice Steinert

<sup>15</sup>KfH-Nierenzentrum am Städtischen Klinikum Görlitz, Görlitz, Germany

Angela Jansen, Ellen Pirlich, Torsten Siegert

<sup>16</sup>Via medis Nierenzentrum Dresden MVZ GmbH, Dresden, Germany

Ivo Döhler, Matthias Pietzonka

<sup>17</sup>KfH-Nierenzentrum Bischofswerda, Bischofswerda, Germany

Simone Kuchinke, Eva Strakow, Valeria Uschakow

<sup>18</sup>KfH-Gesundheitszentrum Aue, Aue-Bad-Schlema, Germany

Matthias Bauer, Grit Baumgart, Anne Martin, Tino Peter Zschocke

<sup>19</sup>Dialvsiszentrum Hoyerswerda, Hoyerswerda, Germany

Jan Nawka

<sup>21</sup>*K*fH-Nierenzentrum am Vogtland Krankenhaus Plauen, Plauen, Germany

Kristin Hartmann, Monika Horbach, Sina Koch, Mike Zahn

<sup>22</sup>Dialyse-Praxis Weiwasser, Weiwasser, Germany Mato Nagel

<sup>23</sup>Dialyse Heidenau, Heidenau, Germany Sebastian Heinrich, Iris Meyer, Annett Olschewski

<sup>24</sup>KfH-Nierenzentrum Grimma, Grimma, Germany Dietmar Göhring, Michael Windgassen

<sup>25</sup>ELBLAND Dialyse Groenhain, Groenhain, Germany

Michael Dechant, Kerstin Heinze, Daniela Kschiwan <sup>26</sup>Institut für Transfusionsmedizin Plauen, DRK-Blutspendedienst Nord-Ost gemeinnützige GmbH, Plauen, Germany

Nadine Hildebrandt, Andreas Karl

<sup>29</sup>Coordinating Centre for Clinical Trials, Dresden, Germany

Stephan Friedrich

<sup>33</sup>Nieren-, Hochdruck- und Rheuma-Zentrum Dresden MVZ mit Dialyse und Apherese, Dresden, Germany Sven Hänsel, Falko Neumann, Catrin Palm, Doreen Reimann

#### References

- Jager KJ, Kramer A, Chesnaye NC, et al. Results from the ERA-EDTA Registry indicate a high mortality due to COVID-19 in dialysis patients and kidney transplant recipients across Europe. Kidney Int. 2020;98(6):1540-1548.
- Grupper A, Rabinowich L, Schwartz D, et al. Reduced humoral response to mRNA SARS-CoV-2 BNT162b2 vaccine in kidney transplant recipients without prior exposure to the virus. Am J Transplant. 2021;21(8):2719-2726. https://doi.org/10.1111 ajt.16615.
- Boyarsky BJ, Werbel WA, Avery RK, et al. Antibody response to 2-3 dose SARS-CoV-2 mRNA vaccine series in solid organ transplant recipients. JAMA. 2021;325(21):2204-2206.
- Stumpf J, Siepmann T, Lindner T, et al. Humoral and cellular immunity to \$ARS-CoV-2 vaccination in renal transplant versus dialysis patients: a prospective, multicenter observational study using mRNA-1273 or BNT162b2 mRNA vaccine. Lancet Reg Health Eur. 2021;9: 100178.
- Stumpf J, Tonnus W, Paliege A, et al. Cellular and humoral 5 immune responses after 3 doses of BNT162b2 mRNA SARS-CoV-2 vaccine in kidney transplant. Transplantation. 2021;105(11):267-269.
- 6 Dimeglio C, Herin F, Martin-Blondel G, et al. Antibody titers and protection against a SARS-CoV-2 infection. J Infect. 2021;84 (2):248-288. https://doi.org/10.1016/j.jinf.2021.09.013. S0163-4453(21)00483-7.
- Padoan A, Sciacovelli L, Basso D, et al. IgA-Ab response to spike 7 glycoprotein of SARS-CoV-2 in patients with COVID-19: a longitudinal study. Clin Chim Acta. 2020;507:164–166
- Rubio-Acero R, Castelletti N, Fingerle V, et al. In search for the SARS-CoV-2 protection correlate: a head-to-head comparison of two quantitative SI assays in a group of pre-characterized oligo-/ asymptomatic patients. medRxiv 2021: 2021.02.19.21252080.
- Müller L, Andrée M, Moskorz W, et al. Age-dependent immune response to the Biontech/Pfizer BNT162b2 COVID-19 vaccination. medRxiv 2021: 2021.03.03.21251066.
- Meyer B, Torriani G, Yerly S, et al. Validation of a commercially available SARS-CoV-2 serological immunoassay. *Clin Microbiol* 10 Infect. 2020;26(10):1386–1394. EUROIMMUN Medizinische Labordiagnostika AG aPc. SARS-
- II CoV-2 NeutraLISA. 31.03.2021 2021. https://www.coronavirus-

diagnostics.com/documents/Indications/Infections/Coronavirus/ EI\_2606\_D\_UK\_F.pdf.

- 12 Aiello A, Najafi Fard S, Petruccioli E, et al. Spike is the most recognized antigen in the whole-blood platform in both acute and convalescent COVID-19 patients. Int J Infect Dis. 2021;106:338–347.
- 13 Shaw RH, Stuart A, Greenland M, et al. Heterologous prime-boost COVID-19 vaccination: initial reactogenicity data. *Lancet.* 2021;397:2043–2046.
- Munro APS, Janani L, Cornelius V, et al. Safety and immunogenicity of seven COVID-19 vaccines as a third dose (booster) following two doses of ChAdOxi nCov-19 or BNTi62b2 in the UK (COV-BOOST): a blinded, multicentre, randomised, controlled, phase 2 trial. *Lancet.* 2021;398(10318):2258–2276. https://doi.org/10.1016/ S0140-6736(21)02717-3. S0140-6736(21)02717-3.
   Azur MJ, Stuart EA, Frangakis C, Leaf PJ. Multiple imputation by
- 15 Azur MJ, Stuart EA, Frangakis C, Leaf PJ. Multiple imputation by chained equations: what is it and how does it work? *Int J Methods Psychiatr Res.* 2011;20(1):40–49.
- 16 R Core Team. R: A Language and Environment for Statistical Computing. version 4.0.1. Vienna, Austria: R Foundation for Statistical Computing; 2020.
- 17 Stumpf J, Schwöbel J, Siepmann T, et al. Revaccination success in kidney transplant recipients with no initial humoral response is linked to basic vaccine type. (*submitted*).
- 18 Gangappa S, Wrammert J, Wang D, et al. Kinetics of antibody response to influenza vaccination in renal transplant recipients. *Transpl Immunol.* 2019;53:51–60.

- 19 Cravedi P, Ahearn P, Wang L, et al. Delayed kinetics of IgG, but not IgA, antispike antibodies in transplant recipients following SARS-CoV-2 infection. J Am Soc Nephrol. 2021;32(12):3221–3230.
- 20 Stumpf J, Hugo C. Humoral immunity to SARS-CoV-2 vaccination in haemodialysis patients—authors' reply. Lancet Reg Health Eur. 2021;10: 100244.; Nov;.
- 21 Lin SY, Liu JH, Lin CC, et al. Comparison of hepatitis B surface antibody decay rates after vaccination between hemodialysis and peritoneal dialysis patients. *Vaccine*. 2011;29(21):3738–3741. https://doi.org/10.1016/j.vaccine.2011.03.049.
- 22 Stumpf J, Klimova A, Mauer R, et al. Equivalent humoral and cellular immune response but different side effect rates following SARS-CoV-2 vaccination in peritoneal and hemodalysis patients using mRNA vaccines. *Nephrol Dial Transplant*. 2021;37(4):796– 798. https://doi.org/10.1093/ndt/gfab324.
- 23 Liu YL, Kao MT, Huang CC. A comparison of responsiveness to hepatitis B vaccination in patients on hemodialysis and peritoneal dialysis. *Vaccine*. 2005;23(30):3957–3960.
- 24 COVID-19-Inzidenzen. Robert Koch Institut, 2021. Available at: https://www.rki.de/DE/Content/InfAZ/N/Neuartiges\_Coronavi rus/Daten/Inzidenzen.html.). Accessed 4 May 2021.
- 25 Whitaker H.J., Gower C., Otter A.D., et al. Nucleocapsid antibody positivity as a marker of past SARS-CoV-2 infection in population serosurveillance studies: impact of variant, vaccination, and choice of assay cut-off. Medrxiv 2021 Oct; https://doi.org/10.1101/ 2021.10.25.21264964.