Impact of age, sex and body mass index on cortisol secretion in 143 healthy adults

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Abstract

Context: Studies on 24-h cortisol secretion are rare. The impact of sex, age and adiposity on cortisol levels, often restricted to one or a few samples, are well recognized, but conflicting.

Objective: To investigate cortisol dynamics in 143 healthy men and women, spanning 7 decades and with a 2-fold body mass index (BMI) range with different analytic tools. *Setting*: Clinical Research Unit.

Design: Cortisol concentrations in 10-min samples collected for 24h. Outcomes were mean levels, deconvolution parameters, approximate entropy (ApEn, regularity statistic) and 24-h rhythms.

Results: Total 24-h cortisol secretion rates estimated by deconvolution analysis were sex, age and BMI independent. Mean 24-h cortisol concentrations were lower in premenopausal women than those in men of comparable age (176 ± 8.2 vs 217 ± 9.4 nmol/L, P=0.02), but not in subjects older than 50 years. This was due to lower daytime levels in women, albeit similar in the quiescent overnight period. Aging increased mean cortisol by 10 nmol/L per decade during the quiescent secretory phase and advanced the acrophase of the diurnal rhythm by 24 min/decade. However, total 24-h cortisol secretion rates estimated by deconvolution analysis were sex, age and BMI independent. ApEn of 24-h profiles was higher (more random) in premenopausal women than those in men (1.048 ± 0.025 vs 0.933 ± 0.023 , P=0.001), but not in subjects older than 50 years. ApEn peaked during the daytime.

Conclusion: Sex and age jointly determine the 24-h cortisol secretory profile. Sex effects are largely restricted to age <50 years, whereas age effects elevate concentrations in the late evening and early night and advance the timing of the peak diurnal rhythm.

Key Words

- cortisol
- approximate entropy
- deconvolution
- human
- circadian rhythms

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Introduction

The hypothalamic–pituitary–adrenal (HPA) axis is the most important neuroendocrine stress-responsive system, which is of critical importance for survival in mammals (1).

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Tightly controlled regulation of the HPA axis is crucial for maintaining both mental and physical health, in as much as hypoactivity and hyperactivity of this system leads to



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disease states (2, 3, 4, 5, 6, 7, 8, 9). Although the HPA axis has been studied for several decades, published reports on the influence of age, sex and body mass on HPA dynamics are conflicting (3, 4, 5, 6, 10, 11, 12, 13, 14, 15, 16, 17, 18). The small size of most cohorts evaluated to date, the narrow age and BMI ranges encompassed and the lack of inclusion of both genders collectively make the possibility of statistical type I or type II errors high in earlier studies. As importantly, because of correlations among age, BMI and gender, multivariate regression is needed for definitive inferences. Nonetheless, multivariate analysis also is unreliable in small cohorts (19, 20). In addition, given the large concentration difference across the 24-h cycle and the marked pulsatility of cortisol, studies using a single sample, or a few blood samples, have large random variance, and are not representative of finely timed physiologically regulated cortisol secretion per se. To overcome these obstacles would require investigation of a larger number of healthy unmedicated adults, both men and women, over wide ranges of age and BMI, and utilization of suitable analytical tools to quantify the dependencies of dynamic cortisol secretion properties (mean, peak, nadir, acrophase, basal and pulsatile, 24-h secretion, pattern regularity or ApEn) on individual and/ or combined clinical characteristics. The present analyses seek to address these issues in 143 healthy individuals sampled frequently (every 10 min) for a sufficiently representative duration (24h) and analyzed with robust immunoassays.

Subjects and methods

Clinical protocol

The 109 volunteers from Leiden, The Netherlands, had participated as controls in studies on neurological disorders, obesity, longevity and ACTH-cortisol relationships (3, 4, 9, 21). In addition, 34 healthy male participants of a not yet published sleep interference study were included from Rochester, MN, USA. Informed consent was obtained from each subject after full explanation of the purpose and nature of all procedures used as approved by the Mayo Clinic Internal Review Board and Leiden University Ethics Committee. Clinical characteristics are listed in Table 1. Postmenopausal individuals studied here did not use estrogen therapy. Premenopausal women were studied in the early follicular phase of the menstrual cycle. Participants maintained conventional work and sleeping patterns and reported no recent transmeridian travel, weight change (>2 kg in 6 week), shift work,

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Table 1	Demographic	properties	of the	volunteers.
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	Men (79)	Women (64)	P value
Age (year)	47.2±1.9	45.4±1.7	0.93
BMI (kg/m²)	26.8 ± 0.6	27.9 ± 0.7	0.28
IGF-1 (nmol/L)	19.9 ± 0.7	19.4 ± 0.8	0.62
Free T4 (nmol/L)	15.8 ± 0.4	15.0 ± 0.2	0.09

Data are shown as mean and s.e.m. Statistical comparisons were done with the 2-tailed Student's t-test.

psychosocial stress, prescription medication use. including contraceptive pills in women, substance abuse, neuropsychiatric illness or acute or chronic systemic disease. A complete medical history, physical examination and screening biochemistry tests were normal. No subject had been exposed to glucocorticoids within the preceding 3 months. Volunteers were admitted to the Study Unit in the evening or stayed overnight in a nearby hotel (3). Ambulation was permitted to the lavatory only. Vigorous exercise, daytime sleep, snacks, caffeinated beverages and cigarette smoking were disallowed. Meals were provided at 09:00, 12:30 and 17:30 h, and room lights were turned off between 22:00 and 24:00h. Blood samples (2.0mL) were withdrawn at 10-min intervals for 24 h. Total blood loss was less than 430 mL. Volunteers were compensated for the time spent in the study as approved by the IRB.

Assays

Serum cortisol concentrations were measured with different assays. In the majority of the Leiden studies, cortisol was assayed by solid-phase RIA (Sorin Biomedical, Milan, Italy). Intra-assay and interassay CVs were 5.1 and 6.4%, respectively. Cortisol in the Leiden Longevity Study was measured by the electrochemiluminescence immunoassay on the Modular E-170 immunoanalyzer (Roche Diagnostics). The detection limit of the first generation of the assay was 5.5 nmol/L, and intra- and interassay CVs were 3.5-6.3% and 3.1–5.1%, respectively. The Mayo study used Immulite 2000 (Siemens Healthcare Diagnostics, Flanders, NJ, USA). This chemiluminescent assay has a detection range of 5.5-1380nmol/L with intra- and interassay coefficients of variation of 7.2-9.4% and 6.3-9.4% at respective concentrations of 10.5-122 and 10.2-113 nmol/L. All samples of each individual were measured the same assay. No samples were undetectable in any assay.

Average, peak and nadir GH cortisol concentrations

Mean (24-h and 3-h averages), median, absolute peak (single daily maximum) and absolute nadir (single daily



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minimum) cortisol concentrations were determined in each subject.

Deconvolution analysis

Cortisol concentration time series were analyzed using a recently developed automated deconvolution method. Pulse detection was empirically validated using hypothalamo pituitary sampling and simulated pulsatile time series. Sensitivity and specificity both exceed 93%. The Matlab-based algorithm first detrends the data and normalizes concentrations to the unit interval (0, 1). Second, the program creates multiple successively decremental potential pulse-onset time sets, each containing one fewer pulse by a smoothing process (a nonlinear adaptation of the heat-diffusion equation). Third, a maximum-likelihood expectation (MLE) estimation method calculates all secretion and elimination parameters simultaneously conditional on each of the candidate pulse-time sets. Deconvolution parameters comprise basal secretion, secretory-burst mass (concentration units), mode (time delay in min to maximum value) and frequency (number of bursts per sampling duration, lambda of Weibull distribution). Pulsatile secretion is the sum of secretory-burst mass and total secretion of the sum of basal and pulsatile secretion. The fast half-life of cortisol was represented as 2.41 min, constituting 37% of the decay amplitude and the slow half-life ranging between 40 and 70 min. Statistical model selection was performed using the Akaike information criterion (22, 23).

Approximate entropy

Adaptive ensemble (network-like) control of hormone release patterns was appraised via the approximate entropy statistic (ApEn) (24). ApEn provides a model-free and scale-invariant statistical estimate of relative randomness in the data, wherein larger values denote greater process randomness and less feedback control (25, 26).

Diurnal variations

Diurnal variations of cortisol levels were studied in two ways: first, by relating mean concentrations in successive 3-h windows to age, BMI and sex; and secondly, by Cosinor analysis, which quantitates the nyctohemeral variation of cortisol concentrations by using a nonlinear unweighted least-squares cosine regression. Ninety-five

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Table 2	Mean cortisol concentrations in 3-h bins in 143	;
healthy n	en and women.	

Time bin (h)	Women (64)	Men (79)	P value
9–12	240±11	266±10	0.09
12–15	190±8	224±9	0.008
15–18	154±8	200±9	< 0.0001
18–21	118 ± 10	137±11	0.22
21–24	104 ± 10	99±8	0.64
24–3	117±10	122±7	0.72
3–6	232 ± 14	244 ± 9	0.50
6–9	307±11	352±13	0.01

Statistical calculations with the two-sided *t*-test were done on un transformed data. Data are mean \pm s.E.M. Time bins correspond to clock hours.

percent statistical confidence intervals were determined for the 24-h cosineamplitude (50% of the zenith-nadir difference), mesor (rhythmic mean) and acrophase (clock time of maximal value).

Statistical analysis

The primary endpoint was the 24-h mean cortisol concentration with segmentation into 3-h bins across the 24 h. Secondary mechanistic outcomes were (1) deconvolution, (2) ApEn and (3) Cosinor parameters.

The *a priori* clinical hypothesis was that sex, age and/or BMI determine time-dependent cortisol concentrations. Comparisons between sexes were done with the Student's *t*-test for unpaired data. Data were analyzed using a General Linear Model (GLM) for repeat measurements with categorical variables, sex and



Figure 1

Mean serum cortisol concentrations in 3-h bins and stratified for age. Symbols: circles: subjects younger than 40 years, squares: subjects 40–60 years, triangles: subjects older than 60 years. Additional statistical data are given in Supplementary Table 3.



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assay method. The effect of age was analyzed by linear regression. Calculations were performed using Systat 13 (Systat Software, Inc, San Jose, CA, USA) and Matlab 8.6 (The MathWorks, Inc, Natick, MA, USA). Cosinor analysis was carried out with Sigmaplot 13 (Systat Software, Inc). P<0.05 was construed as significant for the overall study. *Post hoc* multiple comparisons used Sidak's correction.

Results

Mean 24-h cortisol concentrations were lower in premenopausal women than men of comparable age $(176\pm8.2 \text{ vs} 217\pm9.4 \text{ nmol/L}, P=0.02)$, due to lower cortisol secretory-burst mass $(220\pm17.6 \text{ and } 266\pm16.8 \text{ nmol/L}, P=0.05)$. No differences existed after age 50 years.

Mean cortisol concentrations over 24-h and in 3-h bins are shown in Table 2. Cortisol was higher in men than women during daytime only. As shown in 3 different age cohorts in Fig. 1, older individuals had higher cortisol in the late evening and early night (Table 3). Regression analyses delineated positive correlations with age between clock times 21 and 3 h (Table 4), as illustrated graphically in Fig. 2. The magnitude of the late-day cortisol increment was about 10 nmol/L per decade.

Results of deconvolution of the 24-h serum cortisol time series are shown in Table 5. Cortisol secretory-burst frequency was slightly higher, whereas cortisol half-life and basal secretion were shorter and lower, respectively, in women than men. Total daily cortisol secretion, secretoryburst mode and pulsing regularity were sex independent. Age and BMI were not significant regression factors for deconvolution parameters (Supplementary Tables 1 and 2, see section on supplementary data given at the end of this article).

Table 3 Cortisol means of 3 h bins stratified for three agegroups.

Time bin (h)	Age <40 years (60)	Age 40–60 years (38)	Age >60 years (45)
9–12	270±13	230±12	252±13
12–15	218±10	204 ± 11	201±12
15–18	202±12	153±11	173±8.9
18–21	136±12	130 ± 18	117 ± 10
21–24	85±8.6	117±13	111±11
24–3	100 ± 9.1	1269 ± 10	142 ± 12
3–6	231±14	221±15	266±13
6–9	344 ± 17	323±18	322±9.8

GLM procedure for repeat measurements: between subjects: age group: P=0.43, sex: P=0.022, BMI: P=0.041. Within subjects: time (bin): P<0.0001, time x age group: P<0.0001, time x sex: P=0.034, time x BMI: P=0.42.

http://www.endocrineconnections.org DOI: 10.1530/EC-17-0160 © 2017 The authors Published by Bioscientifica Ltd **Table 4** Linear regressions of mean cortisol and age.

Time bin (h)	R	Regression slope β	P value	<i>P</i> value log- transformed data
9–12	0.114	-0.658 ± 0.485	0.18	0.16
12–15	0.110	-0.526 ± 0.402	0.19	0.19
15–18	0.213	-1.093 ± 0.421	0.011	0.029
18–21	0.104	-0.590 ± 0.475	0.22	0.26
21–24	0.199	0.952 ± 0.394	0.017	0.001
24–3	0.249	1.152 ± 0.405	0.003	0.001
3–6	0.143	0.912 ± 0.530	0.087	0.032
6–9	0.07	-0.475 ± 0.573	0.41	0.77

The regression slopes are shown as mean ± s.p.

Cosinor analysis (Fig. 3) showed an acrophase advance (earlier clock time) of 0.40 h (24 min) per decade. Neither mesor nor amplitude was age dependent (both P>0.05). As an simple model-free measure of diurnal amplitude, the difference of 3-h cortisol means in the morning (9–12 h) and nighttime (21–24 h) decreased with age with R=0.28, P=0.001, and β =1.60±0.48.

Cortisol ApEn over 24 h was 0.921 ± 0.024 in men, and 1.019 ± 0.026 in women (P<0.0001). In premenopausal women, ApEn was 1.048 ± 0.025 and higher than men age <50 years: 0.933 ± 0.023 , P=0.001, but not in subjects aged 50 years or more (0.963 ± 0.058 vs 0.909 ± 0.043 , P=0.46). There was no BMI effect. Cortisol was higher with age; viz., borderline in the DiaSorin assay (P=0.06), but P=0.017 for the Roche assay and P=0.004 for the Immulite assay. The time pattern of cortisol ApEn variation across the 3-h bins is illustrated in Fig. 4, showing higher daytime than nighttime values (P<0.0001) and dependency on sex (P<0.0001).

The influence of cortisol assay was analyzed further. The DiaSorin assay group comprised 89 subjects, the Immulite 2000-assay group 34 subjects and the Roche E170 assay group 20 subjects. For the 3 assays, median 24-h cortisol concentrations (145 samples) were 161 ± 6.1 , 167 ± 10.5 and 185 ± 11.5 nmol/L, respectively. In the GLM procedure, assay type was not significant (*P*=0.33).

By GLM analysis, age and BMI did not influence 24-h mean cortisol concentrations (P values 0.11 and 0.36), whereas sex did with median levels in men of 176 ± 6.4 vs 146 ± 7.1 nmol/L in women (P=0.005).

GLM analysis of deconvolution parameters (Table 6) revealed no effects of cortisol assay, age, BMI or sex.

As a sensitivity analysis, leaving out the data obtained in the Roche E170 or Immulite, or both, assays did not alter conclusions.

Progressively reducing sampling intensity affected estimates of mean 24-h cortisol concentration, as portrayed in Fig. 5. The squared correlation coefficient



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500 R=0.199,P=0.017, $\beta = 0.952 + / -0.394$ 400 Cortisol (nmol/L) 300 200 100 0 10 20 60 80 90 30 40 50 70

Regression of Mean Serum Cortisol Concentrations

between 2100 h and MN on Age







Figure 2

Linear regressions of mean cortisol (3-h) concentrations on age in the clock time intervals of 21–24h (upper panel) and 24–3h (lower panel).

decreased from 0.998 (20 vs 10-min sampling) to 0.91 (120 vs 20-min sampling). Means of two samples per 24 h, when taken 12 h apart, yielded R^2 =0.51, thus explaining 51% of the variability.

Discussion

This study quantifies simple and complex measures of cortisol secretion with respect to sex, age and BMI utilizing 10-min sampling for 24h in 143 adults ages 19–78 years and validated analytical measures of putative feedback control (ApEn), pulsatile and nonpulsatile secretion (deconvolution) and diurnal rhythmicity (Cosinor analysis). Total 24-h cortisol secretion estimated

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Table 5 Deconvolution analysis of serum cortisol profiles in143 healthy volunteers.

Parameter	Men (79)	Women (64)	P value
Pulse frequency (#/24h)	16.4±0.5	18.5±0.6	0.005
Slow half-life (min)	64.4 ± 1.0	57.0±1.3	0.008
Mode day (min)	11.2 ± 0.8	14.5 ± 1.3	0.054
Mode night (min)	12.1 ± 0.7	14.5 ± 1.2	0.19
Basal secretion (nmol/L/24h)	903±66	786±94	0.007
Pulsatile secretion (nmol/L/24h)	4011±158	4950±243	0.68
Total secretion (nmol/L/24h)	4915 ± 195	4954±243	0.92
Mean pulse mass (nmol/L)	253±10	236±13	0.17
Weibull lambda (#/24h)	15.7 ± 0.4	16.9 ± 0.6	0.09
Weibull gamma (dimensionless)	1.816±0.046	1.846 ± 0.051	0.59
ApEn (dimensionless)	0.921±0.024	1.019 ± 0.026	0.007

Data are shown as mean and s.E.M. The two-tailed Student's *t*-test was used for statistical comparisons on logarithmically transformed data.

by deconvolution of intensively sampled profiles was independent of sex, age and BMI. In subanalyses, cortisol dynamics exhibited a higher pulse frequency, shorter half-life and lower basal secretion in women than men. Nycthemeral variations differed by sex and age, with higher mean cortisol in the daytime in men than women, and in late evening and early night in older than young subjects. This result seems to be at variance with a study in which cortisol levels were stimulated with low-dose and high-dose ACTH and by glucagon administration in 55 healthy individuals (38).

Acrophase of Cortisol versus Age



Figure 3

Relation between time of cortisol acrophase (maximal Cosinor-estimated 24-h concentration) and age. The apparent advance (earlier timing) with age was 24 min per decade.



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Figure 4

Approximate entropy (ApEn) of cortisol concentrations in men and women, segmented in 3-h time bins. During the morning and afternoon, ApEn was higher than in the remaining part of the diurnal cycle.

In this limited study BMI, age and sex were not significant variables. However, acute stimulation tests are strictly nonphysiological, and therefore, any comparison is hampered by these considerations. Moreover, for a fair comparison between such studies dose–response curves are required, but such expensive and labor-intensive studies are far beyond the clinical requirements.

ApEn was higher in women than men, higher in the daytime and independent of aging. The nycthemeral maximum (acrophase) was advanced to an earlier clock time by 40min/decade and increased by about 10nmol/L/ decade.

HPA axis activity ostensibly varies with age, sex, BMI, socioeconomic status, chronic illness, stress, psychiatric disorders and sleep disruption (1, 13, 18). Comparisons among studies, however, are hampered by different experimental designs, including number and spacing of

blood samples, salivary samples or urinary excretion of cortisol and/or cortisol metabolites, size of groups, sex, BMI and age distributions. Blood sampling intensity in studies in which age was related to mean serum cortisol levels varied from 4 to 72 samples/24h. The study of 145 samples/24h could not demonstrate age effects on mean (median) cortisol concentrations or distinct cortisol secretion parameters. Three studies reported an increase in cortisol area under the curve (5, 6, 13), a fourth study a decrease in mean 24-h cortisol (10), and a fifth and sixth no influence of age (10, 11, 27). The present approach differs by using 10-min sampling, 24-h data, a larger cohort of subjects, and all three of deconvolution, Cosinor and ApEn analyses. Nonetheless, the age range of 19-78 years, like most other studies, limits conclusions in the very old.

Cortisol's regulation was examined further using consecutive 3-h time windows over 24h, since higher cortisol in the late evening-early night quiescent period has been associated with sleep disorders and the metabolic syndrome (7, 28). By cohort comparisons, two studies found higher cortisol concentrations in older than young subjects in the quiescent period (6, 13), whereas one study did not (12). The present analysis across continuous ages delineates a late-day rise in mean cortisol concentrations of 10nmol/L/decade. This effect could be due to diminished feedback of cortisol on pituitary corticotropes and/or brain CRH/AVP-secreting neurons (29, 30) and may have health implications. Thus, the Leiden Longevity Study of 303 subjects disclosed lower salivary cortisol in the late evening in offspring of healthy nonagenarian siblings compared to their partners (8). In another study, hydrocortisone administration during the quiescent phase in metyrapone-clamped volunteers reduced insulin sensitivity and glucose tolerance (31). However, in this study, serum cortisol levels reached about 800 nmol/L, exceeding physiological levels. Conversely, continuous sc infusion of cortisol in Addison's patients

Table 6 Significance values of deconvolution parameters in the GLM procedure of serum cortisol profiles in 143 healthysubjects.

	Pulse frequency	Slow half-life	Mode day	Mode night	Basal secretion	Pulsatile secretion	Total secretion	MPM	Lambda	Gamma
Sex	0.27	0.21	0.42	0.27	0.27	0.46	0.30	0.11	0.32	0.66
Assay	0.17	0.87	0.78	0.27	0.578	0.377	0.37	0.92	0.47	0.55
BMI	0.77	0.91	0.37	0.83	0.69	0.14	0.16	0.41	0.78	0.51
Age	0.81	0.51	0.86	0.39	0.16	0.68	0.38	0.98	0.84	0.42
Age × assay	0.56	0.86	0.98	0.28	0.51	0.67	0.52	0.92	0.60	0.51

The used cortisol assays were DiaSorin RIA, First generation of the Roche E170 ECLIA and the Siemens Immulite 2000 assay. MPM, mean pulse mass.

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Squared Correlations between 144 Samples-based Mean and Decreasing Number of Samples.



Figure 5

Relationship between the mean cortisol concentration determined from decreasing sampling interval (*x*-axis) and mean cortisol level based on 144 samples taken at 10min intervals for 24h. The squared correlation coefficients determined by linear regressions are shown on the *y*-axis.

was sufficient to prevent the nocturnal glucose decrease (32). Direct experimental data are needed on the impact of glucocorticoid levels in this time window over days or weeks on other health indices, such as blood pressure, abdominal adiposity, diabetes control and systemic inflammation.

Few studies have assessed whether 24-h cortisol secretion is sex dependent or estradiol dependent. One small isotope dilution study in 7 men and 7 women under age 40 years estimated nearly 50% lower cortisol production in women than men (14). Another study reported lower mean 24-h cortisol concentrations in women than men, supporting the present findings in volunteers under age 50 years (13). Using 24-h sampling, we could show that cortisol concentrations were higher in men only during daytime. This insight would agree with higher single-morning total and free cortisol levels reported in men earlier (33).

The present analysis revealed that cortisol secretoryburst mass is smaller and half-life shorter in premenopausal women than men under age 50 years. Because of a higher pulse frequency, no sex difference emerged for total cortisol secretion. Although the role of (premenopausal) estrogen is not so clear (18), in older adults shortterm administration of estradiol, but not testosterone, heightens cortisol-mediated inhibition of pulsatile ACTH secretion (15). In principle, therefore, an estrogenic milieu could lower mean cortisol in premenopausal women. Higher BMI is associated with attenuated 24-h GH secretion, and a more subtly increased 24-h TSH, prolactin and ACTH secretion (4, 21, 34, 35). Although urinary levels of cortisol and its metabolites rise in obese individuals, this is not true when normalized for fat mass or BMI. Concomitantly, serum cortisol levels remain unchanged or even decrease, due to increases in distribution volume and metabolic clearance (35). In accordance with these reports, the current analysis found no significant effect of BMI along or in combination with sex and age on 24-h cortisol concentrations or multiple dynamic measures of cortisol secretion.

Cosinor analysis is a widely used tool to quantify 24-h rhythms of adrenal steroids, sex steroids, TSH, GH, LH, FSH, prolactin and deep body temperature. Herein, we found, consistent with published data, an advance in the acrophase of cortisol, defined by cortisol's peaking about 0.4h earlier in the day per decade of age (6, 10, 11). Although Cosinor analysis detected no effect of age on either the cortisol mesor or amplitude, a model-free approach revealed an amplitude decrease of about 16nmol/L per decade age. As suggested earlier (22), sensitivity of the Cosinor method is reduced when the nadir and acrophase are not 12h apart.

ApEn is a measure of serial regularity, reflecting the balance between feedforward and feedback strength (24). Our new finding that cortisol ApEn is higher in women than men is theoretically compatible with inferred increased feedforward of CRH, AVP and ACTH and diminished feedback by dexamethasone and cortisol (16, 17, 29, 30, 36, 37). Cortisol ApEn was age dependent only in sex-specific evaluations. Recent data on ApEn of GH, TSH and prolactin rises with age in both sexes, consistent with altered feedforward/feedback control. At this time, we have no explanation for possible assay dependency of cortisol ApEn.

Intensive 24-h hormone studies are expensive and therefore rare. Only two large studies on cortisol profiles are available, one by van Cauter and coworkers (13) and the present one. They differ in design, i.e., sampling interval and used analytical techniques. Aside from applying deconvolution of the profiles we used a model-free approach to quantitate particular time intervals instead of the best fit curve based on periodogram calculations. Both studies had, not unexpectedly, similarities but also differences. In the present study, mean 24-h cortisol concentration was independent of sex, age and BMI, and this result was corroborated by deconvolution, while the other study showed an age-dependent increase. On the other hand, both studies showed age-dependent



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concentration increases in the quiescent period of the 24-h cycle. In the present study, an age-dependent advance shift of the acrophase was found, but not in the study by van Cauter, although they described an advance shift in the timing of the onset of the diurnal concentration increase. Thus summarizing, differences and similarities between both studies exist, partly attributable to different analytical approaches.

For clinicians, this study has only restricted relevance. A major point to realize is that a single-morning cortisol specimen has very limited predictability of the 24-h secretion. Furthermore, mean 3-h morning cortisol levels are higher in men than women, and there is an age-related increase in levels during the quiescent hours in the late evening and early night. For diagnostic considerations, these subtle physiological adaptations are not relevant in diagnosing major endocrine disruptions such as Cushing's disease, Cushing's syndrome and primary and secondary adrenal failure. For these disorders, Endocrine Societies have formulated clear guidelines. The most challenging condition is the differential diagnosis between ectopic ACTH secretion and pituitary-dependent hypercortisolemia, when imaging and pituitary blood sampling studies are equivocal. Unfortunately, for such cases, no data are available whether extensive evaluation of plasma ACTH-cortisol profiles is helpful in making a distinction.

A potential problem inherent in retrospective studies is that different hormone assays are used. Extensive statistical GLM analyses, wherein assay type was a potential dependent variable, did not reveal statistically significant assay effects. This was confirmed by sensitivity analyses, which entailed removing one assay at a time. A similar issue exists in other large retrospective analyses, such as with data from 9 different institutions and thus different cortisol assays (13).

When the clinical goal is an accurate estimate of the mean 24-h cortisol concentration 10-min sampling is not required. An accurate 24-h mean value can be obtained from as few as 4 equally distributed (e.g. every 6h) blood samples.

Limitations of the present study include the needs to study younger adults (\leq age 18 years) and quite old individuals (\geq age 80 years); verify 24-h rhythms by 3–5 days of 10-min sampling; estimate body composition beyond BMI *per se*; quantify free cortisol concentrations and/or CBG levels and conduct similar analyses as an additional function of season. In addition, the use of a single high-sensitivity immunological cortisol assay or HPLC-tandem MS–MS is preferable.

http://www.endocrineconnections.org DOI: 10.1530/EC-17-0160 © 2017 The authors Published by Bioscientifica Ltd In summary, healthy adults maintain consistent 24-h cortisol concentrations independently of BMI. In relation to age, the cortisol acrophase advances about 24 min/ decade and late evening and early night cortisol increases about 10 nmol/L/decade. Sex differences in mean cortisol concentrations operate during the daytime in the premenopausal age range of <50 year. Cortisol ApEn is higher in the day than night and in premenopausal women than men, denoting less cortisol feedback control. ApEn was BMI and age independent. The collective outcomes should provide a better framework for interpreting cortisol physiology and pathophysiology in health and disease.

Supplementary data

This is linked to the online version of the paper at http://dx.doi.org/10.1530/ EC-17-0160.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Author contribution statement

F R, J D V and P T helped in conception/design of research. D V H, P T, R Y and F R performed experiments. F R and J D V analyzed the data. F R and J D V helped in manuscript preparation. All authors approved the final manuscript.

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