



## Data in Brief

Genome sequences of *Photorhabdus luminescens* strains isolated from entomopathogenic nematodes from southern India

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## ABSTRACT

We report here draft whole genome sequences of three novel strains of *Photorhabdus luminescens* of 5.2–5.3 Mbps in size, and with a G+C content of 42.5% (each). Symbiotic  $\gamma$ -proteobacteria belonging to the genera, *Photorhabdus* (Family: *Enterobacteriaceae*) with their natural vectors, the entomopathogenic nematodes (EPN) (Phylum: *Nematoda*; Order: *Rhabditida*; Family: *Heterorhabditidae*), have emerged as important biological control agents of insect pests, and are capable of production and delivery of diverse compounds to influence host biology [1–3]. Analysis of these genomes is expected to provide enhanced insight into mechanisms of virulence, insecticidal toxin genetic diversity, antibiotic resistance and monoxenicity. The nucleotide sequence information for the three strains NBAIL PLHb105, NBAIL HiPL101 and NBAIL H75HRPL105 has been deposited in NCBI Nucleotide database and is accessible via AZAB000000000, JTHJ000000000 and JXUR000000000 accession numbers respectively.

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## Specifications

Organism/cell line/tissue	<i>Photorhabdus luminescens</i> NBAIL PLHb105, <i>Photorhabdus luminescens</i> NBAIL HiPL101, <i>Photorhabdus luminescens</i> NBAIL H75HRPL105
Sex	N/A
Sequencer or array type	Illumina-MiSeq
Data format	Analyzed
Experimental factors	Cultures isolated from natural hosts, cultured in <i>Galleria mellonella</i>
Experimental features	Whole-genome sequences and variants from existing references
Consent	N/A
Sample source location	Karnataka, India

## 1. Direct link to deposited data

*Photorhabdus* (Family: *Enterobacteriaceae*) with their natural vectors, the entomopathogenic nematodes (EPN) (Phylum: *Nematoda*; Order: *Rhabditida*; Family: *Heterorhabditidae*), have emerged as important biological control agents of insect pests, and are capable of production and delivery of diverse compounds to influence host biology [1–3].

Table 1

Accession numbers, and direct-link URLs to data in this study.

	<i>Photorhabdus luminescens</i> NBAIL H75HRPL105	<i>Photorhabdus luminescens</i> NBAIL HiPL101	<i>Photorhabdus luminescens</i> NBAIL PLHb105
NCBI accession numbers	AZAB000000000	JTHJ000000000	JXUR000000000
URL	<a href="http://www.ncbi.nlm.nih.gov/assembly/GCF_000826725.1/">http://www.ncbi.nlm.nih.gov/assembly/GCF_000826725.1/</a>	<a href="http://www.ncbi.nlm.nih.gov/assembly/GCA_000798635.1/">http://www.ncbi.nlm.nih.gov/assembly/GCA_000798635.1/</a>	<a href="http://www.ncbi.nlm.nih.gov/assembly/GCF_000931955.1/">http://www.ncbi.nlm.nih.gov/assembly/GCF_000931955.1/</a>

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**Table 2**

Total raw reads, sequenced base-pairs, N50 value of WGS assembly and SNPs identified from *P. luminescens* NBAlI H75HRPL105, *P. luminescens* NBAlI HiPL101, and *P. luminescens* NBAlI HbPL105. \*Sequenced base pairs calculated after trimming of Illumina adapters following FASTQC analysis. N50 values calculated from WGS de novo assemblies using FASTQC-trimmed reads. \*\*Sequence variants called from reference-guided assemblies with all *P. luminescens* strains mapped against *P. luminescens* subsp. *Laumondii* TT01 reference genome (NCBI accession NC\_005126.1). SNPs—single nucleotide polymorphism. MNP—multiple nucleotide polymorphism. Indel—insertion and/or deletion mutations.

	<i>Photorhabdus luminescens</i> NBAlI H75HRPL105	<i>Photorhabdus luminescens</i> NBAlI HiPL101	<i>Photorhabdus luminescens</i> NBAlI HbPL105
Total reads	2,604,823	2,790,255	2,463,266
Total bp*	189,591,052	203,431,557	214,900,170
GC content	42.5%	42.5%	42.5%
N50	22,874	27,588	20,747
Mean contig length	8430	7454	6677
SNPs, MNPs, indels**	639	2179	6549

Raw sequencing reads, and whole-genome shotgun assemblies for three *P. luminescens* strains have been deposited at DDBJ/EMBL/GenBank under the accession numbers provided in Table 1.

Total raw reads, sequenced base-pairs, N50 value of WGS assembly and SNPs identified from *P. luminescens* NBAlI H75HRPL105, *P. luminescens* NBAlI HiPL101, and *P. luminescens* NBAlI HbPL105 have been summarized in Table 2.

## 2. Experimental design, materials and methods

*Photorhabdus luminescens* and *Xenorhabdus* sp. are symbiotic bacteria associated with soil-born *Heterorhabditis* and *Steinernema* species of entomopathogenic nematodes. Bacterial cultures were established from isolation in these natural hosts, then cultured in lab hosts *Galleria mellonella*. Finally, pure monoxenic cultures were then grown in LB media. Genomic DNA was then extracted using the Sigma Bacterial Genomic DNA isolation kit. Purity of these isolations was checked with 16s rRNA gene sequences, before they were used for whole genome sequencing. Isolated genomic DNA was then used for sequencing and library preparation using the Illumina MiSeq platform (at Chromous Biotech Ltd., Bengaluru, 560692, Karnataka, India) with paired-end libraries generated for each of the three bacterial genomes. Reads were processed, analyzed and trimmed according to FASTQC to remove Illumina adapter sequences. Trimmed reads were assembled into contigs to capture whole-genome shotgun sequences (WGS) using de novo and reference-guided methods using CLCBio Genomics

Workbench v. 7.5. All *P. luminescens* strains were mapped to the reference genome of *P. luminescens laumondii* strain TT01 (NCBI accession NC\_005126.1, for reference-guided genome assemblies) using global alignment, and trimmed where base-call confidence was less than 95%. Sequence variants (SNPs, multiple nucleotide polymorphisms and indels) were identified against the reference genome of *P. luminescens* subsp. *Laumondii* TT01 reference genome (NCBI accession NC\_005126.1) in CLCBio Genomics Workbench using the following parameters: minimum variant coverage—50, minimum variant count—9, minimum variant frequency—50%, minimum quality score neighborhood radius—13, minimum variant quality score—30, and minimum neighborhood quality score—25. Broken read pairs were also ignored.

We report draft genome sequences of three bacterial strains from India, viz., *P. luminescens* strain NBAlI H75HRPL105, *P. luminescens* strain NBAlI HiPL101, and, *P. luminescens* strain NBAlI PLHb105 isolated from the entomopathogenic nematodes, *Heterorhabditis species* strain NBAlI H75HR, *Heterorhabditis indica* strain NBAlI Hi101 and *Heterorhabditis bacteriophora* strain NBAlI Hb105, respectively.

## Conflicts of interest

The authors declare no conflicts of interest.

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## References

- [1] P.S. Grewal, R.U. Ehlers, D.I. Shapiro-Ilan, *Nematodes as Biocontrol Agents*. CABI Publishing, Wallingford UK, 2005 513.
- [2] G.O. Poinar Jr., The presence of *Achromobacter nematophilus* in the infective stage of a *Neoplectana* sp. (Steinernematidae: Nematoda). *Nematologica* 12 (1966) 105–108.
- [3] N. Boemare, A. Givaudan, M. Brechelin, C. Laumond, Symbiosis and pathogenicity of nematode-bacterium complexes. *Symbiosis* 22 (1997) 21–45.