

## RESEARCH PAPER

# Nanomolar potency and selectivity of a Ca<sup>2+</sup> release-activated Ca<sup>2+</sup> channel inhibitor against store-operated Ca<sup>2+</sup> entry and migration of vascular smooth muscle cells

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### BACKGROUND AND PURPOSE

The aim was to advance the understanding of Orai proteins and identify a specific inhibitor of the associated calcium entry mechanism in vascular smooth muscle cells (VSMCs).

### EXPERIMENTAL APPROACH

Proliferating VSMCs were cultured from human saphenous veins. Intracellular calcium was measured using fura-2, whole-cell current was recorded using patch-clamp and cell migration quantified in modified Boyden chambers. Subcellular protein localization was determined by microscopy. Isometric tension was recorded from mouse aortic rings.

### KEY RESULTS

Molecular disruption and rescue experiments indicated the importance of Orai1 in calcium entry caused by store depletion evoked passively or by platelet-derived growth factor (PDGF), suggesting the presence of Ca<sup>2+</sup> release-activated Ca<sup>2+</sup> (CRAC) channels like those of the immune system. The CRAC channel blocker, S66, was a potent inhibitor of the VSMC signals, IC<sub>50</sub> 26 nM, which was almost two orders of magnitude greater than with leucocytes. S66 had no effect on PDGF- and ATP-evoked calcium release, overexpressed transient receptor potential canonical (TRPC)5 channels, native TRPC1/5-containing channels, stromal interaction molecule 1 clustering, non-selective cationic current evoked by store depletion and phenylephrine-evoked aortic contraction. S66 reduced PDGF-evoked VSMC migration while having only modest effects on cell proliferation and no effect on cell viability.

### CONCLUSIONS AND IMPLICATIONS

The data suggest that Orai1 has a role in human VSMC migration, and that a CRAC channel inhibitor has high potency and selectivity for the associated calcium entry, suggesting a distinct characteristic of vascular CRAC channels and the potential for selective chemical suppression of vascular remodelling.

### Abbreviations

CRAC channel, Ca<sup>2+</sup> release activated Ca<sup>2+</sup> channel; PDGF, platelet-derived growth factor; PGPC, 1-palmitoyl-2-glutaryl-phosphatidylcholine; siRNA, short interfering RNA; STIM1, stromal interaction molecule 1; S66, 3-fluoropyridine-4-carboxylic acid (2',5'-dimethoxybiphenyl-4-yl)amide; TRPC, transient receptor potential canonical; VSMC, vascular smooth muscle cell

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### Keywords

vascular smooth muscle; migration; calcium channel; store-operated calcium entry; platelet-derived growth factor; chemical blockade

### Received

4 October 2010

### Revised

16 February 2011

### Accepted

28 February 2011

## Introduction

Contractile smooth muscle cells regulate the diameters of most blood vessels. Although they are relatively quiescent, throughout life they retain capacity for profound but controlled plasticity that enables them to change reversibly to non-contractile and mobile phenotypes that play critical roles in vascular development, tissue remodelling and responses to injury (Owens *et al.*, 2004). This property of these cells contributes not only to physiology but to major disease phenomena and unwanted clinical responses, which include restenosis, atherosclerosis and neointimal hyperplasia of the saphenous vein (Angelini and Jeremy, 2002; Kumar *et al.*, 2006). The molecular events controlling phenotypic switching in smooth muscle cells and the properties of the non-contractile cells, such as mobility, are starting to emerge (Owens *et al.*, 2004).

Important events determining smooth muscle cell functions are plasma membrane ion fluxes such as  $\text{Ca}^{2+}$  entry (Beech, 2007). Voltage-gated  $\text{Ca}^{2+}$  entry channels (especially the dihydropyridine-sensitive L-type channels) have established roles in the regulation of blood pressure and the treatment of hypertension. However, other non-voltage-gated  $\text{Ca}^{2+}$  entry mechanisms exist and appear to make greater contributions in the non-contractile smooth muscle cells (Landsberg and Yuan, 2004; Beech, 2007). One of these mechanisms is store-operated  $\text{Ca}^{2+}$  entry, which was identified in vascular smooth muscle 30 years ago (Casteels and Droogmans, 1981). Various studies have suggested roles of transient receptor potential canonical (TRPC) channels (Alexander *et al.*, 2008) in the store-operated mechanism and shown the channels to have an important role in non-contractile properties of smooth muscle cells (Sweeney *et al.*, 2002; Kumar *et al.*, 2006; Takahashi *et al.*, 2007; Li *et al.*, 2008). Nevertheless, experiments using TRPC channel blockers or molecular approaches for TRPC disruption suggest that TRPC channels explain only part of the store-operated  $\text{Ca}^{2+}$  entry signal of vascular smooth muscles (VSMCs) (Li *et al.*, 2008) or even none of it according to some studies (Potier *et al.*, 2009). Therefore, an additional mechanism is inferred. A candidate for the additional mechanism is the store-operated  $\text{Ca}^{2+}$  release-activated  $\text{Ca}^{2+}$  (CRAC) channel that has been associated primarily with immune cells (Hogan *et al.*, 2010).

Determinants of CRAC channels are stromal interaction molecule 1 (STIM1) and Orai1 (Feske *et al.*, 2006; Cahalan, 2009; Hogan *et al.*, 2010). Orai1 is one of three related mammalian proteins encoded by distinct genes. The proteins are thought to contain four membrane-spanning segments and resemble, in some regards, the extensively-studied tetraspanin proteins, which have roles in mammalian cell morphology and motility. Tetraspanins had not been considered to be ion channel subunits, but various studies now suggest that Orai1 is the ion pore-forming subunit of CRAC channels (Hogan *et al.*, 2010). Despite the focus on immune cells and absence of a vascular phenotype in patients with disrupted Orai1, reports have suggested that Orai proteins are expressed, and that Orai1 is functional in VSMCs (Berra-Romani *et al.*, 2008; Baryshnikov *et al.*, 2009; Potier *et al.*, 2009; Bisailon *et al.*, 2010; Ng

*et al.*, 2010). One report has importantly described a whole-cell CRAC channel-like current in rat cultured VSMCs (Potier *et al.*, 2009), but the very small size of the current and failure to detect the current in the presence of physiological  $\text{Ca}^{2+}$  have generated challenges for establishing its relevance to VSMC biology. Here we describe an investigation of the relevance of Orai proteins and CRAC channels to proliferating human saphenous vein VSMCs with particular focus on identification of a chemical agent that could facilitate studies of the mechanism in the cardiovascular field.

## Methods

### Cell culture

Freshly discarded human saphenous vein segments were obtained anonymously and with informed consent from patients undergoing open heart surgery in the General Infirmary at Leeds. Approval was granted by the Leeds Teaching Hospitals Local Research Ethics Committee. Proliferating VSMCs were prepared using an explant technique (Porter *et al.*, 2001) and grown in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal calf serum (FCS), penicillin/streptomycin and L-glutamine at 37°C in a 5%  $\text{CO}_2$  incubator. Experiments were performed on VSMCs passaged two to five times; all of the cells stained positively for smooth muscle  $\alpha$ -actin and smooth muscle myosin heavy chain and were non-contractile. Tetracycline-induced expression of exogenous human TRPC5 in HEK 293 cells has been described previously (Zeng *et al.*, 2004). HL-60 (human promyelocytic leukaemia) cells were from ATTC (Teddington, Middlesex, UK) and were cultured in RPMI 1640 with 10% FCS and penicillin/streptomycin.

### VSMC transfections and knock-down validation

Cells ( $0.5\text{--}2 \times 10^6$ ) were centrifuged ( $100\times g$ ) for 10 min, resuspended in Basic Nucleofector solution (Amaxa GmbH, Lonza GmbH, Cologne, Germany), mixed with 1  $\mu\text{M}$  short interfering (si) RNA (Ambion Europe Ltd, Applied Biosystems, Warrington, UK; Table S1) and transferred into a cuvette for electroporation (Amaxa). With this method, the siRNA concentration is higher than used with lipid transfection methods, but the exposure time to the siRNA is considerably shorter. The scrambled control siRNA was Silencer Negative Control #1, which is a 19 bp scrambled sequence with no significant homology to human gene sequences (Ambion). Cells were transferred from cuvettes to pre-warmed culture medium and incubated in a 5%  $\text{CO}_2$  incubator at 37°C. Culture medium was changed after 24 h, and measurements were made after a further 24 h. When two siRNA probes were used as a cocktail, each probe was used at one-half of its usual concentration. Full-length cDNA for Orai1 (BC013386) was purchased from Geneservice (Source Bioscience LifeSciences, UK) and subcloned into pcDNA6/V5-His (Invitrogen, Paisley, UK) at *EcoRI/XhoI* sites. cDNA encoding eYFP-STIM1 was a gift from T Meyer. Clones were sequenced to confirm identities. Transfections with 1–1.5  $\mu\text{g}$  of DNA also occurred by electroporation.

### RNA isolation and quantitative reverse transcription PCR

Total RNA was extracted from saphenous vein smooth muscle cells using a Tri-reagent protocol followed by DNase I (Ambion) treatment (Fountain *et al.*, 2004). One microgram of total RNA was used for RT based on oligo-dT primers and AMV RT enzyme. The specificity of PCR was verified by reactions without RT (-RT) and by melt curve analysis of PCR products. Sequences of PCR primers are shown in Table S1. PCR products were electrophoresed on 2% agarose gels containing ethidium bromide. No PCR products occurred in the absence of RT. With RT, there was only a single product of the correct size, giving a single peak in the melt curve. All PCR products were sequenced to confirm identity (Lark Technologies, Takeley, Essex, UK). Real-time PCR was carried out using a Lightcycler (Roche, Welwyn Garden City, UK) largely as described previously (Fountain *et al.*, 2004). Relative abundance of target RNA was normalized to  $\beta$ -actin RNA, which showed no difference between samples. PCR efficiency ( $E$ ) was  $10^{(-1/\text{slope})}$ . Relative abundance of target RNA was calculated from  $(E_{\beta\text{-actin}}^{C_p}) / (E_{\text{target}}^{C_p})$ , where PCR cycle crossing points ( $C_p$ ) were determined by fit-points methodology. PCR reactions were at least in duplicate.

### Western blotting

Cells were collected using 1 mL of ice-cold phosphate-buffered saline (PBS) and pelleted at 4°C and then lysed in Laemmli sample buffer supplemented with complete protease inhibitor cocktail (Roche). The lysate was centrifuged to remove particulate matter, and equal amounts of protein were separated by 8% sodium dodecyl sulphate polyacrylamide gel electrophoresis and transferred onto nitrocellulose membrane. Membranes were incubated with blocking buffer (containing 0.1% Tween 20 and 5% nonfat dry milk) for 2 h at room temperature, followed by incubation in dialysed rabbit anti-Orai1 antibody (Alomone Labs Ltd., Jerusalem, Israel, #ACC-060) overnight at 4°C. Anti- $\beta$ -actin antibody (1:1000) was from Santa Cruz. Membranes (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA) were then washed and incubated with secondary antibody. Labelling was detected using Super-Signal West Pico chemiluminescent substrates (Pierce, Thermo Fisher Scientific Inc., Rockford, IL, USA).

### Intracellular calcium measurements

Cells were pre-incubated with fura-2AM for 1 h at 37°C followed by a 0.5 h wash at room temperature. HL-60 cells were centrifuged in the plate at  $2 \times 10^5$  cells per well. Measurements were made at room temperature on a 96-well plate reader (FlexStation, Molecular Devices Inc., Sunnyvale, CA, USA). The change ( $\Delta$ ) in intracellular calcium ( $\text{Ca}^{2+}_i$ ) concentration is indicated as the ratio of fura-2 emission intensities for 340 and 380 nm excitation ( $F$  ratio). Wells within columns of the 96-well plate were loaded alternately for test and control conditions. The recording solution contained (mM): 130 NaCl, 5 KCl, 8 D-glucose, 10 HEPES and 1.2  $\text{MgCl}_2$ , titrated to pH 7.4 with NaOH. When indicated, 0.2 or 1.5 mM  $\text{CaCl}_2$  was added. EGTA was not included in the  $\text{Ca}^{2+}$ -free recording solution because of the solution addition, rather than replacement, format of the FlexStation; therefore, contaminating  $\text{Ca}^{2+}$  was present (estimated to be 1–10  $\mu\text{M}$ ).

### Deconvolution microscopy

VSMCs, 48 h post-transfection, were detached and transferred to glass coverslips with fresh culture medium and allowed to spread for 24 h. After treatments with reagents, the cells were fixed in methanol for 10 min at  $-20^\circ\text{C}$ , washed and rehydrated in PBS, and mounted onto glass slides using ProLong Gold anti-fade (Molecular Probes®, Invitrogen, Paisley, UK). Cells were visualized on an Olympus IX-70 inverted microscope using a  $\times 100$  UPLAN objective (NA 1.35) supported by a DeltaVision deconvolution system (Applied Precision LLC Issaquah, WA, USA) with SoftWoRx image acquisition and analysis software. Images were captured on a Roper Coolsnap HQ CCD camera, and epifluorescence was recorded using filter sets for FITC. Wide-field optical sections of 0.2  $\mu\text{m}$  were acquired throughout the  $z$  plane of the cells and deconvolved using a constrained iterative algorithm assigned by Delta Vision.

### Cell assays

Cell migration assays were performed in duplicate in modified Boyden chambers with polycarbonate membranes containing 8  $\mu\text{m}$  pores (BD Biosciences, Oxford Science Park, Oxford, UK). Cells in suspension ( $1 \times 10^5$ ) were loaded in the upper chamber in DMEM supplemented with 0.4% FCS. The lower chamber contained DMEM (0.4% FCS) supplemented with the chemoattractant 10 ng  $\text{mL}^{-1}$  platelet-derived growth factor (PDGF)-BB (Invitrogen). After 8 h at 37°C in a 5%  $\text{CO}_2$  incubator, cells that had attached but not migrated were scraped from the upper surface, membranes were fixed in 70% ethanol at  $-20^\circ\text{C}$  and the migrated cells were stained with haematoxylin and eosin and evaluated by counting cell nuclei in 10 randomly chosen fields under light microscopy. For Orai1 knock-down cell proliferation experiments, equal numbers of cells from the same patient were transfected and seeded in parallel into six-well tissue culture plates in DMEM culture medium plus 10% FCS. Medium was changed after 24 h, and cells were incubated for a further 24 h. Cells were collected after trypsinization, stained with trypan blue, and the cell number was determined in duplicate wells and counted at least twice with a haemocytometer. Trypsinized wells were observed microscopically to confirm that all cells had been released. For S66 experiments, cells at 40–50% confluence were seeded in 96-well plates for the indicated time periods in at least triplicate wells. At the end of the specified times, cells were fixed and stained with haematoxylin and eosin for counting in four random fields. The medium was replaced every 24 h to provide fresh S66.

### Patch-clamp recordings

Recordings were made using the Patchliner planar patch-clamp system (Nanion Technologies, Munich, Germany) in whole-cell mode (Milligan *et al.*, 2009). Prior to recordings, cells were detached from culture flasks with Detachin (Gelantis Inc., AMS Biotechnology Europe Ltd., Abingdon, UK, cat. no. T100110) or 0.05% Trypsin/EDTA and resuspended at a density of  $1 \times 10^6$ – $5 \times 10^7$   $\text{mL}^{-1}$  in extracellular solution that was the same as the recording solution for  $\text{Ca}^{2+}$  measurements except it contained 0.2 mM  $\text{CaCl}_2$  and 100  $\mu\text{M}$  niflumic acid. The intracellular recording solution contained (mM): 40 EGTA, 17  $\text{CaCl}_2$ , 2  $\text{MgCl}_2$ , 8 NaCl, 1  $\text{Na}_2\text{ATP}$ , 10 HEPES, 66 L

glutamic acid, 66 CsOH, titrated to pH 7.2 with CsOH (calculated unbound  $\text{Ca}^{2+}$ , 100 nM). Corrections were not made for liquid–liquid junction potentials. Voltage ramps were applied from  $-100$  to  $+100$  mV for 1 s every 10 s from a holding potential of 0 mV. Currents were filtered at 1 kHz and sampled at 3 kHz.

### Isometric tension recording

For isometric tension recordings, 8 week old male C57/BL6 mice were killed by  $\text{CO}_2$  asphyxiation and cervical dissociation in accordance with Schedule 1 Code of Practice, UK Animals Scientific Procedures Act 1986. The thoracic aorta of the mouse was removed and placed in ice-cold Hank's solution. Fat was removed by dissection, and blood was flushed from the lumen with Hank's solution. Hanks solution contained (mM): NaCl, 137; KCl, 5.4;  $\text{CaCl}_2$ , 0.01;  $\text{NaH}_2\text{PO}_4$ , 0.34;  $\text{K}_2\text{HPO}_4$ , 0.44; D-glucose, 8; HEPES, 5. Vessels were mounted on two 40  $\mu\text{m}$  diameter wires for isometric tension recording in a 410A dual wire myograph system (Danish Myo Technology, Aarhus, Denmark). The wires were separated by 1 mm to provide basal tension, and the bath solution was at  $37^\circ\text{C}$  gassed continuously with 95%  $\text{O}_2$  and 5%  $\text{CO}_2$ . The bath solution contained (mM): 125 NaCl, 3.8 KCl, 25  $\text{NaHCO}_3$ , 1.5  $\text{MgSO}_4$ , 1.2  $\text{KH}_2\text{PO}_4$ , 8 D-glucose, 1.2  $\text{CaCl}_2$ , 0.02 EDTA. For  $\text{Ca}^{2+}$ -free solution (0  $\text{Ca}^{2+}$ ),  $\text{Ca}^{2+}$  was omitted.

### Reagents

S66 [3-fluoropyridine-4-carboxylic acid (2',5'-dimethoxybiphenyl-4-yl)amide] was a gift from GSK (Oxbridge, Middlesex, UK) (GSK1349571A); its chemical identity was confirmed independently by liquid chromatography mass spectrometry analysis (Leeds). S66 is an abbreviation of 'Synta 66' from patent WO 2005/009954 (Ng *et al.*, 2008; Di Sabatino *et al.*, 2009). All other reagents were from Sigma (Sigma-Aldrich, Gillingham, Dorset, UK) unless indicated otherwise.

### Data analysis

Mean data are presented as mean  $\pm$  SEM, where  $n$  represents the number of independent experiments and  $N$  represents the number of wells of a 96-well plate used in a single experiment. For patch-clamp experiments,  $n$  was the number of recordings from individual cells. In all experiments, comparisons were made independently with cells from at least three patient samples. Experiments were all performed in test and control pairs, and so  $t$ -tests were used for comparisons. Each experiment had its own paired control (ctrl), even though only single control columns (white) are shown in the figures. \* $P$ -values  $<0.05$ , which were taken to indicate significant difference; no significant difference is indicated by NS. Data were analysed and presented, and curves were fitted using Origin software (OriginLab Corporation, Northampton, MA, USA).

## Results

### Orai1 function in $\text{Ca}^{2+}$ entry evoked by store depletion in proliferating human saphenous VSMCs

Messenger RNAs encoded by the *Orai* genes were readily detected in proliferating VSMCs and had relatively high

abundance (Figure S1). To investigate if the gene expression is functionally relevant, we measured the  $\text{Ca}^{2+}$  entry signal occurring when  $\text{Ca}^{2+}$  was added back to cells that had been store-depleted by the pharmacological agent thapsigargin (Figure S2). Specific knock down of expression was achieved using small interfering (si) RNAs (Figure S3 and S4, Table S1). Knock down of Orai1 by two different Orai1 siRNAs suppressed store-operated  $\text{Ca}^{2+}$  entry, while knock down of Orai2 and Orai3 had no effect (Figure 1A, Figure S5). A third Orai1 siRNA was investigated, but it failed to have any effect on the expression of Orai1 (Figure S3), and so it acted as a negative control (Orai1 si.negative). Because the two active Orai1 siRNAs may have had off-target effects, rescue experiments were performed using wild-type Orai1 cDNA, which had no effect on its own but recovered the  $\text{Ca}^{2+}$  entry that was suppressed by Orai1 siRNA (Figure 1B).  $\text{Ca}^{2+}$  entry was also suppressed by the R91W dominant-negative mutant form of Orai1 (Derler *et al.*, 2009) (Figure 1A, Figure S5). The data support the hypothesis that Orai1 is functionally important for  $\text{Ca}^{2+}$  entry evoked by thapsigargin and suggest that CRAC channels are present.

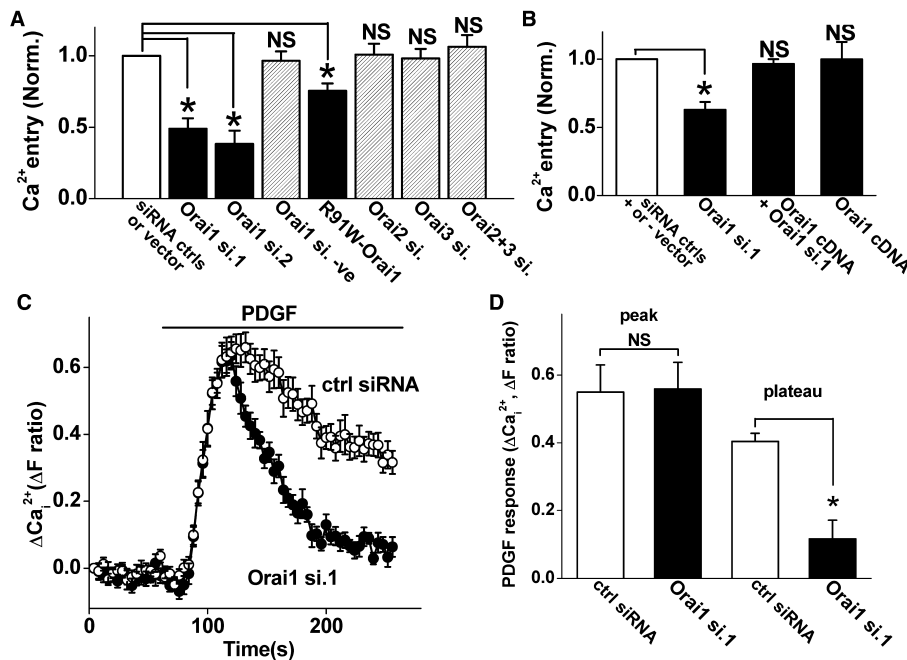
### Orai1 function in $\text{Ca}^{2+}$ entry evoked by PDGF

PDGF is an important physiological stimulant of VSMCs that causes  $\text{Ca}^{2+}$  release. Direct application of PDGF in  $\text{Ca}^{2+}$ -containing extracellular solution evoked a rise in intracellular  $\text{Ca}^{2+}$  that had rapid onset followed by decay towards a plateau phase of  $\text{Ca}^{2+}$  entry (Figure 1C). Knock down of Orai1 had no effect on the initial rise in  $\text{Ca}^{2+}$  but suppressed the  $\text{Ca}^{2+}$  entry (Figure 1C, D). The data suggest that PDGF-evoked  $\text{Ca}^{2+}$  entry depends substantially on Orai1.

### Potent and specific chemical inhibition of $\text{Ca}^{2+}$ entry

With studies of ion channels, it is helpful to have a potent and specific blocker that is easy to use and applicable to a range of assays. Therefore, we investigated if a CRAC channel blocker has effect on  $\text{Ca}^{2+}$  entry in VSMCs. The blocker tested was S66, which has previously been reported to specifically inhibit CRAC-related signals in immune cells with an  $\text{IC}_{50}$  of 1.4–3.0  $\mu\text{M}$  (Ng *et al.*, 2008; Di Sabatino *et al.*, 2009) whether applied before or after store depletion (Ng *et al.*, 2008). It was observed that the  $\text{Ca}^{2+}$  entry occurring upon returning  $\text{Ca}^{2+}$  to the medium (the add-back signal) in store-depleted VSMCs was strongly suppressed by S66 (Figure 2A). Unexpectedly, S66 had high potency, inhibiting the  $\text{Ca}^{2+}$  entry signal with an  $\text{IC}_{50}$  of 26 or 43 nM depending on whether the maximum or rate-of-rise  $\text{Ca}^{2+}$  signal was used (Figure 2B). We therefore made a direct comparison with human leucocytes using the same assay conditions. Consistent with previous reports, the leucocyte  $\text{Ca}^{2+}$  entry was much less sensitive, giving an  $\text{IC}_{50}$  of 1.76  $\mu\text{M}$  (Figure 2B). The effect of S66 on VSMCs occurred relatively rapidly, with a time constant of  $\sim 7$  s (Figure 2C, D). S66 had no effect on the initial rise in  $\text{Ca}^{2+}$  evoked by PDGF but suppressed the plateau phase of  $\text{Ca}^{2+}$  entry (Figure 3A, B). Consistent with the lack of effect on  $\text{Ca}^{2+}$  release, in the absence of extracellular  $\text{Ca}^{2+}$ , PDGF evoked a transient rise in intracellular  $\text{Ca}^{2+}$  that was unaffected by S66 (Figure 3C, D). Similar observations were made when ATP was the receptor agonist used to evoke  $\text{Ca}^{2+}$  release and  $\text{Ca}^{2+}$  entry (Figure 3E,





**Figure 1**

Effect of Orai1 knock down on Ca<sup>2+</sup> entry of human saphenous vein vascular smooth muscle cells (VSMCs). (A, B) Prior to recording Ca<sup>2+</sup> signals, VSMCs were divided into paired groups and transfected with control (siRNA ctrls, scrambled siRNA) or test siRNA (e.g. Orai1 si.1), or control DNA (vector) or vector expressing R91W-Orai1 or Orai1 (wild-type) cDNA. Although only one control column is shown in each graph, each test group had its own control, and the data were normalized (Norm.) to that control. NS indicates no significant difference compared with the matched control data. The 'Orai1 si.negative' was generated for the purpose of suppressing Orai1 expression but it failed to have any effect on the expression and so was used as a negative control. For Ca<sup>2+</sup> recording, VSMCs were pretreated with 1 μM thapsigargin in Ca<sup>2+</sup>-free bath solution for 30 min and exposed to 0.2 mM extracellular Ca<sup>2+</sup> as indicated. (A) Mean normalized (Norm.) data for experiments of the type illustrated in Figure S5 (*n/N* = 3–4/48–64). Raw mean control fura-2 ratio values for each of the seven groups were 0.55, 0.90, 0.54, 0.70, 0.56 and 0.57 (left to right). (B) Rescue of Ca<sup>2+</sup> entry in Orai1 knock down cells by Orai1 cDNA (*n/N* = 3/48). Raw mean control fura-2 ratio values for each of the three groups were 0.50, 0.48 and 0.45 (left to right). (C) Typical Ca<sup>2+</sup> responses to 100 ng·mL<sup>-1</sup> PDGF in the presence of extracellular 1.5 mM Ca<sup>2+</sup> after transfection with control (ctrl) or Orai1 siRNA 1 (*N* = 12 each). (D) Mean data for the type of experiment illustrated in (C) (*n/N* = 3/30).

F). The data suggest that S66 has high potency and specificity for store-operated Ca<sup>2+</sup> entry in human VSMCs, and that the potency is almost two orders of magnitude greater compared with immune cells.

### Lack of effect of S66 on endogenous or overexpressed TRPC channels

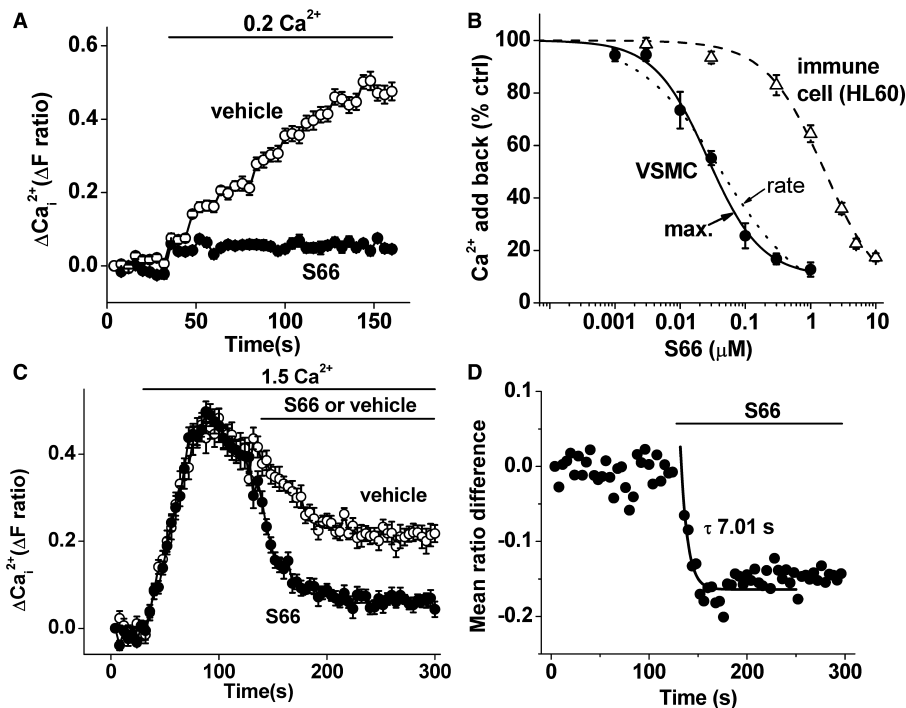
The above data suggest that S66 has specificity because it lacked effect on the PDGF receptor or Ca<sup>2+</sup> release mechanism. To further investigate the specificity, we tested S66 against the Ca<sup>2+</sup> signal evoked by the endogenous oxidized phospholipid PGPC (1-palmitoyl-2-glutaryl-phosphatidylcholine) (Al-Shawaf *et al.*, 2010). Although PGPC evokes a transient rise in intracellular Ca<sup>2+</sup>, this effect is not accounted for by Ca<sup>2+</sup> release but by Ca<sup>2+</sup> entry mediated by TRPC1/5-containing channels (Al-Shawaf *et al.*, 2010). The PGPC-evoked Ca<sup>2+</sup> signal was completely unaffected by S66 at the high concentration of 5 μM (Figure 4A, B). We also investigated human TRPC5 conditionally overexpressed in HEK 293 cells and stimulated by the lanthanide ion, gadolinium (Zeng *et al.*, 2004); this signal was also unaffected by S66 (Figure 4C). The data suggest that S66 does not inhibit endogenous TRPC1/5 or overexpressed TRPC5 channels.

### Lack of effect of S66 on STIM1 clustering

A widely accepted mechanism for coupling of depleted Ca<sup>2+</sup> stores to plasma membrane store-operated Ca<sup>2+</sup> channels is the clustering of STIM1 proteins in sarcoendoplasmic reticulum membranes and the subsequent interaction with plasma membrane channels (Cahalan, 2009). Therefore, the effects of S66 on the clustering of STIM1 were investigated. YFP-tagged STIM1 was expressed in VSMCs and observed by fluorescence microscopy. In the absence of store depletion, STIM1 showed the expected microtubule-like localization pattern (Figure 5A), consistent with it binding a plus-end microtubule tracking protein (Cahalan, 2009). Also, as expected, application of thapsigargin caused striking re-arrangement of STIM1 into clusters (Figure 5B). S66, however, had no effect on the clustering process (Figure 5C). These data suggest that S66 does not act by inhibiting store depletion, its sensing by STIM1 or the subsequent re-arrangements of STIM1 that are considered to be necessary for activation of store-operated channels.

### Lack of effect of S66 on store-operated non-selective cationic current

In human saphenous vein VSMCs, there is also a large non-selective cationic current that is evoked by thapsigargin and



**Figure 2**

S66 inhibition of  $\text{Ca}^{2+}$  add-back responses in store-depleted VSMCs. (A) Example of VSMC paired experiment showing inhibition by  $5 \mu\text{M}$  S66 of the  $\text{Ca}^{2+}$  entry signal observed when adding  $0.2 \text{ mM}$   $\text{Ca}^{2+}$  to thapsigargin-treated cells ( $N = 10$  each). (B) As for (A) but mean data for the maximum (max.)  $\text{Ca}^{2+}$  entry signal ( $n/N = 4/40$ ) with a fitted Hill equation ( $\text{IC}_{50}$   $26 \text{ nM}$ , slope  $1.05$ ). The rate of rise of the same  $\text{Ca}^{2+}$  entry events (rate) was also analysed, and the mean data (not shown but see Figure S6) were fitted with a Hill equation (dotted curve,  $\text{IC}_{50}$   $43 \text{ nM}$ , slope  $0.69$ ). Comparative maximum  $\text{Ca}^{2+}$  entry data are shown for HL-60 cells, also with a fitted Hill equation ( $\text{IC}_{50}$   $1.76 \mu\text{M}$ , slope  $1.0$ ). (C) As for (A) but  $5 \mu\text{M}$  S66 or its vehicle were applied after add-back of  $1.5 \text{ mM}$   $\text{Ca}^{2+}$  ( $N = 10$  each). (D) As for (C) but the mean difference between S66 and vehicle data points ( $n = 3$ ) with a fitted single exponential function (smooth curve,  $\tau = 7.01 \text{ s}$ ).

can be considered to be store-operated (Li *et al.*, 2008). Other investigators have observed a similar ionic current in response to store depletion (Trepakova *et al.*, 2001; Sweeney *et al.*, 2002). This ionic current in human saphenous vein VSMCs was completely unaffected by S66 at  $5 \mu\text{M}$  (Figure 6A–C). These data suggest that the action of S66 is not general for all types of store-operated channel.

#### Lack of effect of S66 on phenylephrine-evoked contraction

The possible effect of S66 on the contractile function of VSMCs was investigated by making isometric tension recordings from freshly-isolated aortic rings of the mouse. Phenylephrine was used as an  $\alpha_1$ -adrenoceptor agonist to evoke contraction (Figure 7A). S66 ( $5 \mu\text{M}$ ) had no effect on the evoked contraction whether applied before (Figure 7A, B) or acutely after phenylephrine (Figure 7B). Phenylephrine-evoked contraction did, nevertheless, depend substantially on  $\text{Ca}^{2+}$  entry (Figure 7B). These data further suggest the specificity of S66, as it had no effect on the  $\alpha_1$ -adrenoceptor-mediated signalling mechanisms,  $\text{Ca}^{2+}$  handling in contractile cells or the contractile machinery.

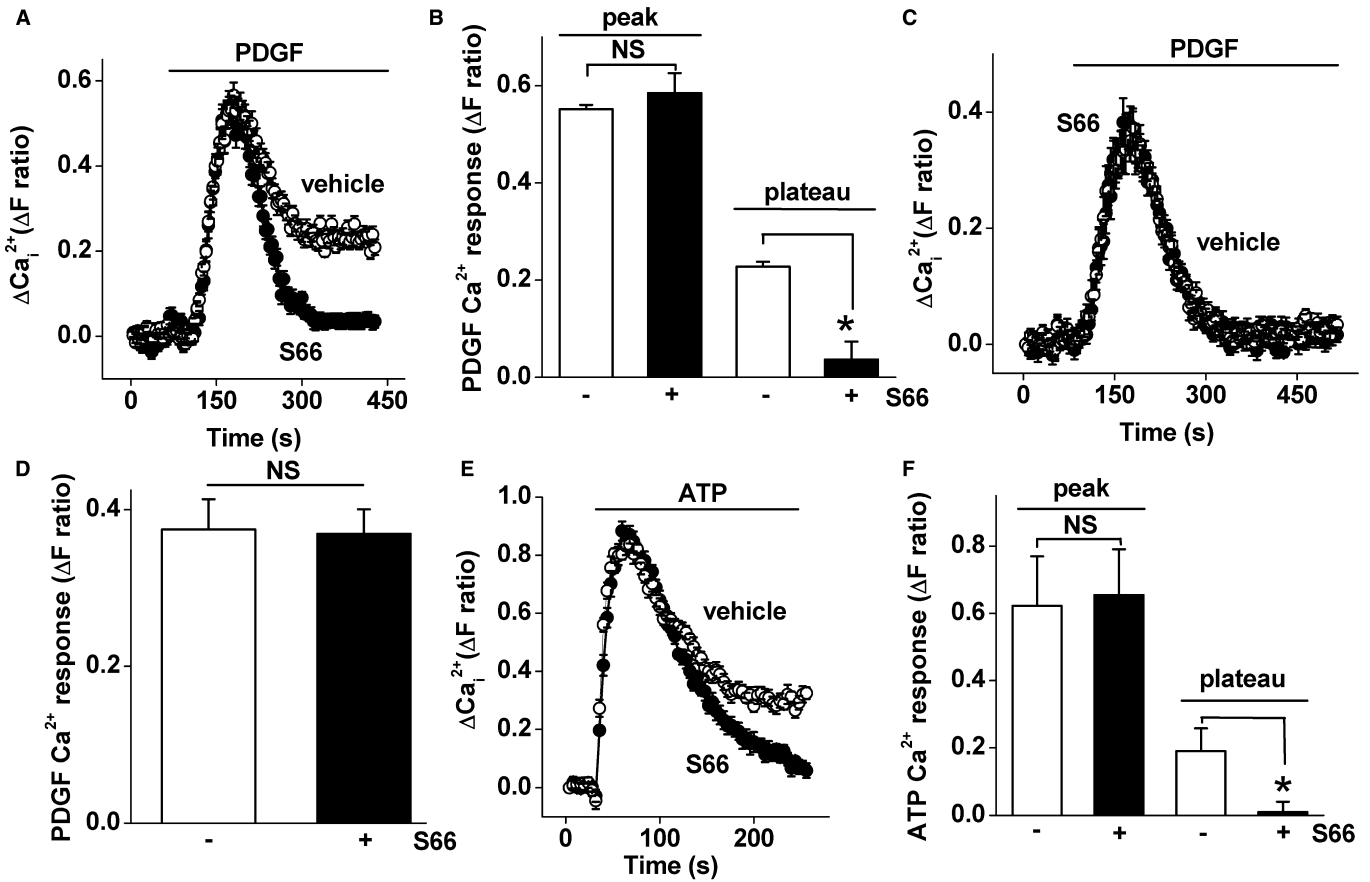
#### Potent inhibition of VSMC migration by S66

Because of the high potency and selectivity of S66, we used it to investigate the relevance of the  $\text{Ca}^{2+}$  entry mechanism to

VSMC migration and proliferation. Cell migration was equally suppressed by  $0.1$  and  $5 \mu\text{M}$  S66 (Figure 8A, Figure S7), suggesting high potency against this cellular parameter that is similar to that observed against  $\text{Ca}^{2+}$  entry.

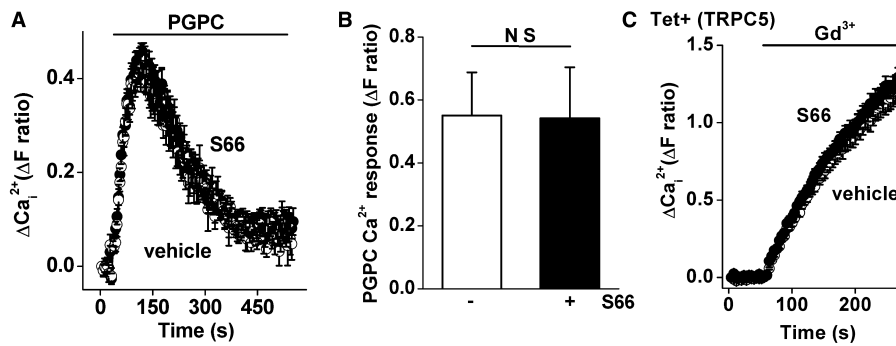
#### Modest effect of S66 on VSMC proliferation and lack of effect on cell viability

The saphenous vein VSMCs doubled in number over a period of  $\sim 7$  days in the presence of serum (data not shown), whereas in the absence of serum the cell number declined relative to the serum group (Figure 8B). The cell count in the presence of serum was quantified over 4 days in the presence of the S66 solvent (vehicle control) or different concentrations of S66. The cell counts were normalized to the vehicle controls at the specified time points (Figure 8B). At days 1 and 2, the cell count was unaffected by S66 and, similarly, concentrations of S66 below  $1 \mu\text{M}$  lacked effects on the cell counts at days 3 and 4 (Figure 8B). There were, however, inhibitory effects of  $1$  and  $5 \mu\text{M}$  S66 at days 3 and 4 (Figure 8B). Cell death, as indicated by trypan blue uptake, was small over these time periods in the presence of serum and was not affected by  $5 \mu\text{M}$  S66 (Figure 8C). The data suggest that S66-sensitive  $\text{Ca}^{2+}$  entry has a positive but relatively modest effect on cell proliferation and no effect on cell viability.



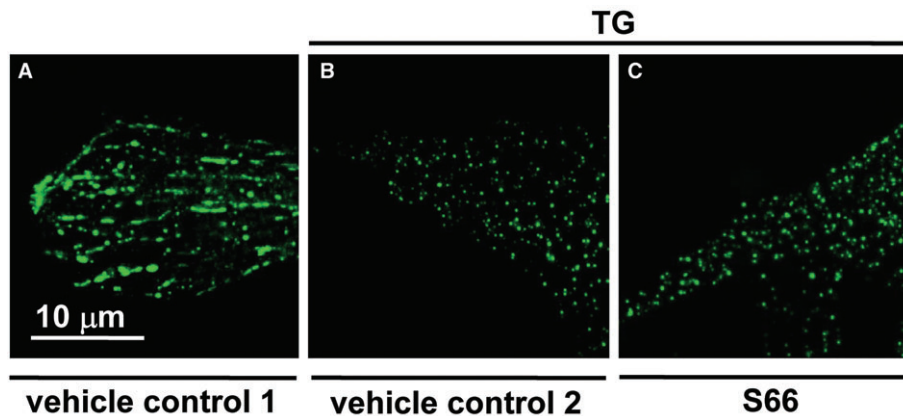
**Figure 3**

S66 inhibition of Ca<sup>2+</sup> entry evoked by physiological agonists. Recordings were made from VSMCs. (A) Intracellular Ca<sup>2+</sup> measurement showing responses to 100 ng·mL<sup>-1</sup> PDGF in 1.5 mM extracellular Ca<sup>2+</sup> and vehicle or 5  $\mu$ M S66 (*N* = 32 each). (B) Mean data for the type of experiment illustrated in (A) comparing the vehicle control and S66 (*n/N* = 3/72). (C, D) As for (A, B) but in the absence of extracellular Ca<sup>2+</sup> (*n/N* = 5/64) and therefore showing Ca<sup>2+</sup> release only. (E, F) As for (A, B) but using 100  $\mu$ M ATP instead of PDGF (*n/N* = 3/48).



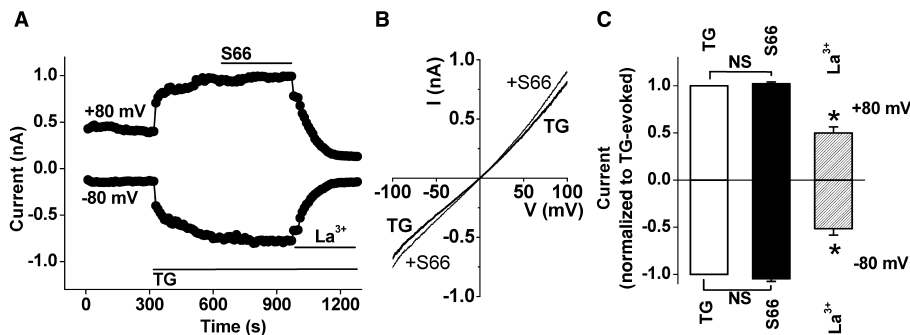
**Figure 4**

Resistance of TRPC1/5 channels to S66. Recordings were made from VSMCs (A, B) or HEK 293 cells (C). (A) Intracellular Ca<sup>2+</sup> measurement showing responses to 5  $\mu$ M 1-palmitoyl-2-glutaryl-phosphatidylcholine (PGPC) in 1.5 mM extracellular Ca<sup>2+</sup> and vehicle or 5  $\mu$ M S66 (black circles) (*N* = 16 for each). (B) As for (A) but mean data for the vehicle control and S66 (*n/N* = 3/64). (C) As for (A) but using HEK 293 cells with overexpression of TRPC5 showing responses to 50  $\mu$ M gadolinium (Gd<sup>3+</sup>) in the presence of vehicle or 5  $\mu$ M S66 (*N* = 12, typical of *n* = 3).



**Figure 5**

Resistance of STIM1 clustering to S66. The images all show VSMCs from the same batch transfected with plasmid expressing eYFP-STIM1 (green colour). VSMCs were then exposed for 5 min to: (A) the solvent for thapsigargin (vehicle control 1); (B) 1  $\mu$ M thapsigargin (TG) and the solvent for S66 (vehicle control 2); or (C) 1  $\mu$ M thapsigargin and 10  $\mu$ M S66. The scale bar in (A) applies to all of the images. Data are representative of two independent experiments on VSMCs from different patients.



**Figure 6**

Resistance of non-selective store-operated cationic current to S66. Recordings were made from VSMCs. (A) Example of ionic current evoked by 2  $\mu$ M thapsigargin (TG), showing effects of 5  $\mu$ M S66 and 50  $\mu$ M lanthanum ( $\text{La}^{3+}$ ). (B) As for (A) but example current–voltage relationships (I–V) for currents evoked by TG and then following the addition of S66. (C) As for (A) but mean data normalized to the current in the presence of TG alone, showing the effect of S66 and  $\text{La}^{3+}$  ( $n = 14$  cells from 3 patients).

### Strong inhibition of VSMC migration by Orai1 siRNA

The data obtained with S66 suggested a relatively specific role of the  $\text{Ca}^{2+}$  entry channels in cell migration. It was therefore investigated if knock-down of Orai1 with siRNA had a similar effect (Figure 8D). Orai1 knock down had a strong inhibitory effect on VSMC migration and no effect on cell proliferation over 2 days (Figure 8D). The inhibition of cell migration was quantitatively greater than that observed with S66 (Figure 8D; cf. Figure 8A). The data suggest that CRAC channels and Orai1 impact positively on VSMC migration, and that Orai1 may not only contribute through the CRAC channel mechanism.

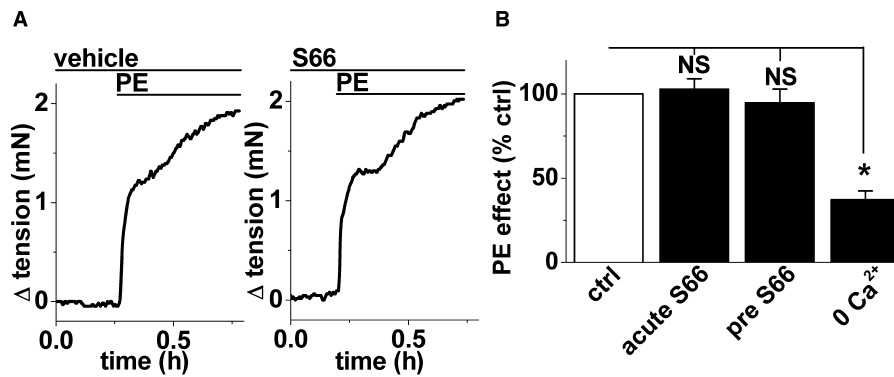
## Discussion and conclusions

These data support the hypothesis that Orai1 has a role in  $\text{Ca}^{2+}$  influx evoked by store depletion in VSMCs and reveal a

chemical inhibitor of the CRAC channel that is a promising tool for further investigation of the mechanism, showing ~100-fold selectivity for VSMCs over leucocytes. The chemical showed excellent specificity, lacking effects on PDGF receptors, ATP receptors,  $\text{Ca}^{2+}$  release, TRPC1/5 channel-dependent  $\text{Ca}^{2+}$  entry, STIM1 clustering, non-selective cationic current evoked by store depletion and the entire mechanisms of  $\alpha$ -adrenoceptor-mediated vascular contraction. In line with results from Orai1 knock-down experiments, S66 suppressed VSMC migration, although not as effectively as Orai1 knock down.

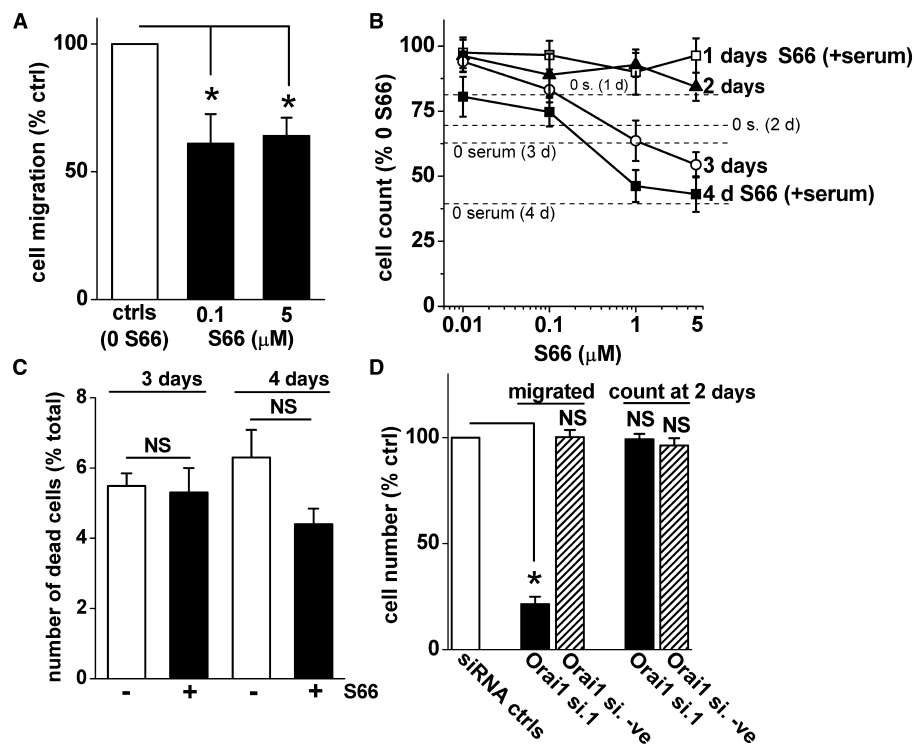
In addition to patent information (WO 2005/009954), effects of S66 on CRAC channel activity in rat basophilic leukaemia and Jurkat T cells have been reported in the scientific literature (Ng *et al.*, 2008; Di Sabatino *et al.*, 2009). S66 was found to have specificity because it did not affect plasma membrane  $\text{Ca}^{2+}$  ATPase, inward rectifier  $\text{K}^{+}$  current, hippocampal synaptic transmission or radioligand binding to a range of receptors and ion channels including the L-type  $\text{Ca}^{2+}$





**Figure 7**

Resistance of aortic contraction to S66. Recordings were made from mouse aorta. (A) Sequential isometric tension recordings from a ring of mouse aorta showing responses to 1 μM phenylephrine (PE) in the presence of vehicle (DMSO, left panel) or 5 μM S66 (right panel). (B) Mean phenylephrine-evoked contractions normalized to the vehicle control data (ctrl) for the acute (15 min) effect of 5 μM S66 (acute S66) and effect of pretreatment and continuous application of 5 μM S66 (pre S66) (as in A). The effect of removing extracellular Ca<sup>2+</sup> is shown for comparison (0 Ca<sup>2+</sup>).



**Figure 8**

Effects of S66 on VSMC migration, proliferation and viability. (A, B) Mean data for S66 effects on cell migration (A) and cell count (B) as percentages of their own vehicle controls (ctrls). (B) Cell count was measured in the presence of serum and various concentrations of S66 at 1, 2, 3 and 4 day time points (1–4 days) (symbols with error bars and connecting lines). The dashed horizontal lines are comparative mean data for cell counts in serum-free conditions as percentages of their serum controls. (C) Mean numbers of cells that failed to exclude trypan blue (i.e. non-viable cells) as a percentage of the total number of cells for the vehicle control (white bar) and S66 (black bar) groups at 3 and 4 days. (D) Mean data for number of cells migrating in Boyden chambers ('migrated') and with trypan blue exclusion in the presence of serum over 2 days ('count at 2 days'). Data obtained after transfection with Orai1 siRNA (Orai1 si.1) or Orai1 negative siRNA (Orai1 si.-ve) were normalized to their own scrambled siRNA controls (siRNA ctrls), even though only one control column is shown. (A, D) Raw mean migrated cell numbers in the controls (ctrls) were 28.9 and 32.5 (for the 0.1 and 5 S66 groups) and 30.4 and 31.8 (for the Orai1 si.1 and Orai1 si.-ve groups). (A–D) Data are from experiments performed at least twice with cells from at least four patients.

channel (Ng *et al.*, 2008; Di Sabatino *et al.*, 2009). Our data not only expand the evidence for specificity but also show that S66 has two orders of magnitude greater potency at the CRAC mechanism in VSMCs. In confirmation of prior observations, and under identical conditions to those used for the VSMC recordings, we showed that the CRAC mechanism in leucocytes is less sensitive to S66. The data suggest pharmacological opportunity and a fundamental difference between the CRAC mechanisms of the vascular and immune cells, even though Orai1 and STIM1 are involved in both cases.

The ionic current of Figure 6 has been previously described as a non-selective cationic current evoked by thapsigargin in saphenous vein smooth muscle cells and inhibited partially by extracellular antibody targeted to STIM1, suggesting the importance of plasma membrane STIM1 (Li *et al.*, 2008). The lack of cationic selectivity of this current, its large amplitude and relatively linear current-voltage relationship suggest that it is not mediated by a CRAC channel. Other investigators have observed similar currents (Trepakova *et al.*, 2001; Sweeney *et al.*, 2002; Stiber *et al.*, 2008). We show here that the current in saphenous vein smooth muscle cells is not sensitive to acute inhibition by S66, further distinguishing it from the CRAC mechanism. It might be expected that a CRAC channel current would have been evident in our electrophysiology experiments, but it was not observed, possibly because of the extremely small size of the current, which was not detectable in rat smooth muscle cells in the presence of extracellular calcium (Potier *et al.*, 2009).

Conflicting conclusions are reported about the contributions of TRPC channels to store-operated  $\text{Ca}^{2+}$  entry in VSMCs. Nevertheless, specifically in relation to human saphenous vein VSMCs, our data consistently point to a role of TRPC channels in the  $\text{Ca}^{2+}$  add-back signal of thapsigargin-treated cells, based on anti-TRPC1 blocking antibody, TRPC1/4/5 RNA interference and dominant-negative mutant TRPC experiments (Kumar *et al.*, 2006; Li *et al.*, 2008). The effects have, nevertheless, been only partial (up to 40%). Therefore, at least 60% of the  $\text{Ca}^{2+}$  add-back signal would seem to have been mediated by other mechanisms (i.e. non-TRPC1/4/5 mechanisms). These findings should be considered in the context of the present study where we observed that S66 had no effect on TRPC activity evoked by non-store depletion mechanisms but inhibited store-operated  $\text{Ca}^{2+}$  entry by about 90%. It seems therefore that there is a quantitative discrepancy with the previous data based on disruption of TRPC subunits. A reason for the discrepancy (i.e. of ~10% vs. ~40% TRPC contribution) could be that the TRPC contribution to the store-operated  $\text{Ca}^{2+}$  add-back signal depends on CRAC channel activity. If this is the case, the TRPC channels would contribute to the  $\text{Ca}^{2+}$  add-back signal only if the CRAC channels were first activated by store depletion and then the CRAC channels stimulated the TRPC channels, perhaps via  $\text{Ca}^{2+}$  facilitation. This may be an over-simplification, however, because of compelling evidence that TRPC channels are activated directly by store depletion due to physical coupling with STIM1 (Zeng *et al.*, 2008). Indeed, our immunoprecipitation and immune-staining results have supported the conclusion that a pool of TRPC1 interacts with STIM1 in saphenous vein smooth muscle cells (Li *et al.*, 2008). Although the relationship of TRPC channels to store deple-

tion remains puzzling, it is clear that such channels do not require store depletion in order to be activated in VSMCs (Al-Shawaf *et al.*, 2010).

The mechanism of action of S66 on  $\text{Ca}^{2+}$  entry is unknown. It acts relatively rapidly and so may function as an ion pore blocker of CRAC channels. Most simply, it would have such an effect by binding directly to Orai1. However, evidence to support such a hypothesis is lacking and the common role of Orai1 in  $\text{Ca}^{2+}$  entry of VSMCs and immune cells, and yet differential potencies of S66 on these cell types, argues against Orai1 as the target protein. However, if Orai1 is not the target protein, we do not have another clear candidate. Our data argue against S66 acting through disruption of the STIM1 mechanism for store-to-channel coupling (Figure 5) or as a general blocker of all types of store-operated channel (Figure 6). It may interfere with the physical coupling between STIM1 and Orai1, but it is unclear why the VSMC  $\text{Ca}^{2+}$  entry would be more potently affected.

One of the curious results of the study was that Orai1 knock down inhibited VSMC migration by about 80% whereas S66 inhibited it by only 40%. If Orai1 acted only as a subunit of CRAC channels, the percentage inhibition would be predicted to be similar or the effect of S66 may even be anticipated to be greater because it is such a strong inhibitor of the channels and siRNA knocks down the gene expression rather than deleting it. One explanation could be that Orai1 has a role in cell migration in addition to those that occur via the CRAC channel. However, additional evidence for such a hypothesis is needed because the possibility that S66 becomes less effective progressively in culture, either because of chemical modification or desensitization at its receptor, was not excluded.

In summary, the results of this study suggest the importance of Orai1 and CRAC channels as mediators of thapsigargin-, PDGF- and ATP-evoked  $\text{Ca}^{2+}$  entry in human saphenous vein VSMCs and that this mechanism potentiates migration of the cells but is not essential for migration. These findings support the hypothesis that CRAC channels are important in VSMCs, even though it has not, so far, been possible to detect an associated ionic current (i.e. I-CRAC) in physiological  $\text{Ca}^{2+}$  conditions. The study also reveals an easy-to-use small molecule inhibitor of the  $\text{Ca}^{2+}$  entry mechanism that has promising specificity and should be useful in further studies of the mechanism in a wide range of experiments. Perhaps most striking is the nanomolar potency of the inhibitor and approximate 100-fold selectivity for VSMCs over immune cells. This suggests that the CRAC channels of VSMCs are fundamentally different from those of immune cells, even though Orai1 and STIM1 are involved in both. From a pharmacological perspective, it would seem worthwhile to consider chemicals such as S66 as potential therapeutic inhibitors of vascular remodelling without negative implications for vascular tone or immune response.

## Acknowledgements

The study was supported by the British Heart Foundation and the Wellcome Trust. A. Sedo provided technical assistance.

## Conflict of interest

The authors state no conflict of interest.

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## Supporting information

Additional Supporting Information may be found in the online version of this article:

**Figure S1** Relevant gene expressions in human saphenous vein VSMCs. **(a)** Typical gel showing PCR products for Orai1, 2 and 3 mRNAs with (+) but not without (-) reverse transcriptase (RT) reaction. A DNA marker ladder is on the left. **(b)** Using real-time RT-PCR quantification, relative abundances of the mRNAs encoding the indicated proteins ( $n = 3-4$ ).

**Figure S2** Example  $\text{Ca}^{2+}$  add-back signals in human saphenous vein VSMCs with and without store depletion and showing comparison of effects of two chemically different store depletion agents, thapsigargin (TG) and cyclopiazonic acid (CPA). Intracellular  $\text{Ca}^{2+}$  was measured in cells pretreated (as indicated below) in  $\text{Ca}^{2+}$ -free bath solution for 30 min and then exposed to 0.2 mM extracellular  $\text{Ca}^{2+}$  as indicated by the horizontal bar labelled 'add-back', except for one group of cells where the  $\text{Ca}^{2+}$  was not added back (0  $\text{Ca}^{2+}$ ). The pretreatments in addition to the  $\text{Ca}^{2+}$ -free solution were 1  $\mu\text{M}$  TG, 10  $\mu\text{M}$  CPA, 1  $\mu\text{M}$  TG plus 10  $\mu\text{M}$  CPA (TG + CPA), 0.2% dimethylsulphoxide (DMSO) as the vehicle control for TG and CPA, or no TG, CPA or DMSO (none).

**Figure S3** Validation and specificity of siRNA knock down of *Orai* expression. See Table S1 for annotations. **(a)** Validation of knock down, showing relative abundances of Orai1, 2 and 3 mRNAs after transfection of vascular smooth muscle cells with Orai1, 2 and 3 siRNAs as indicated ( $n/N = 3/6$ ). The control was scrambled siRNA. **(b)** Validation of specificity, showing lack of effect on the non-target mRNAs (as indicated) after transfection of vascular smooth muscle cells with the indicated siRNAs ( $n/N = 2-3/4-6$ ).

**Figure S4** Endogenous Orai1 protein detection and knock down in human saphenous vein VSMCs. Western blot for lysates from cells transfected with control (scrambled) siRNA (siRNA ctrl) or Orai1 siRNA 1 (Orai1 si.1). The blot was probed

with anti-Orai1 antibody (upper panel) and anti- $\beta$ -actin antibody (lower panel). The predicted protein mass of Orai1 is 33 kDa (arrow).

**Figure S5** Typical original traces for the mean data shown in Figure 1A. For  $N = 16$  per individual experiment, intracellular  $\text{Ca}^{2+}$  measurements from VSMCs pretreated with 1  $\mu\text{M}$  thapsigargin in  $\text{Ca}^{2+}$ -free bath solution for 30 min and then exposed to 0.2 mM extracellular  $\text{Ca}^{2+}$  as indicated by the horizontal bars: comparing control (scrambled) siRNA with Orai1 siRNA 1 **(a)**, Orai2 siRNA **(b)**, Orai3 siRNA **(c)**, Orai2 and Orai3 siRNAs **(d)** and Orai1 siRNA 2 **(e)**; and comparing DNA vector with Orai1 R91W mutant (dominant negative) **(f)**.

**Figure S6** In support of the dotted curve in Figure 2B, the graph shows the mean rate of rise data points without normalization and with the fitted Hill equation. The  $y$ -axis is the linear slope of the initial rate of rise of the fura-2 fluorescence after extracellular  $\text{Ca}^{2+}$  was added to the medium.

**Figure S7** Original data for VSMC migration assays. Typical bright-field images of human saphenous vein VSMCs that had moved through pores in the polycarbonate membrane for vehicle control (ctrl) and 1  $\mu\text{M}$  S66. A cell is indicated by a white arrow and a pore by a black arrow.

**Table S1** PCR primer pairs (F, forward direction; R, reverse direction), PCR amplicon sizes, and siRNA probes sequences. The 'Orai1 si.negative' was generated for the purpose of suppressing Orai1 expression, but it failed to have any effect on the expression and so was used as a negative control

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