



ANOVLATORY INFERTILITY

Exogenous hCG activity, but not endogenous LH activity, is positively associated with live birth rates in anovulatory infertility

JOAN-CARLES ARCE¹ & JOHAN SMITZ²

¹Reproductive Health, Global Clinical & Non-Clinical R&D, Ferring Pharmaceuticals A/S, Copenhagen, Denmark²Follicle Biology Laboratory and Center for Reproductive Medicine, Vrije Universiteit Brussel (VUB), Brussels, Belgium

Objective. To evaluate, retrospectively, the roles of endogenous and exogenous luteinising hormone (LH) activity on live birth rate in ovulation induction cycles.

Methods. Associations between LH activity at baseline, end of stimulation and live birth rate were analysed in relation to patient characteristics, baseline and end of stimulation variables in WHO group II anovulatory women (n=155) stimulated with recombinant follicle-stimulating hormone (rFSH) or highly purified human menopausal gonadotrophin (HP-hMG). HP-hMG provides FSH and exogenous LH activity mainly in the form of human chorionic gonadotrophin (hCG).

Results. Serum LH concentrations at baseline or end of stimulation were not predictive of live birth rate in the rFSH group (n=79) or HP-hMG group (n=76). Serum hCG concentration at end of stimulation was a significant positive predictor in HP-hMG-treated women. Other variables were not independently predictive of live birth in either of the groups, except for a negative association between serum FSH concentrations at the start of stimulation and live birth in the rFSH-treated group.

Conclusions. Endogenous LH concentrations are not predictive of live birth in anovulatory WHO group II patients undergoing ovulation induction with rFSH or HP-hMG. On the other hand, exogenous hCG activity during HP-hMG stimulation is positively associated with treatment outcome.

Keywords: *Highly purified menotrophin, human chorionic gonadotrophin, infertility, live birth, luteinising hormone, ovulation induction*

Introduction

WHO group II anovulatory infertility is the most frequent anovulatory disorder, with the majority of such patients being diagnosed with polycystic ovary syndrome (PCOS) (van Santbrink et al., 1997; Laven et al., 2002). Management includes first-line treatment with clomiphene citrate (CC), and those who fail to ovulate or conceive during CC treatment respond well to go-

nadotrophin treatment (Hamilton-Fairley et al., 1991; Balen et al., 1994; Homburg & Howles, 1999).

Despite a normal follicle stimulating hormone (FSH) concentration, an elevated luteinising hormone (LH) concentration is not unusual in PCOS patients (Laven et al., 2002). Both the absolute concentration of circulating endogenous LH and the LH to FSH ratio are elevated in about 40% of these patients (Conway et al., 1989; Franks, 1989; Fauser et al., 1991; Balen et al., 1995; Taylor et al., 1997). The pathophysiology of the elevated LH concentration is not fully understood (Laven et al., 2002).

A number of studies are available on the impact of endogenous LH on fertility as well as treatment outcome during stimulated cycles in anovulatory patients. In women with regular spontaneous menstrual cycles, the endogenous LH concentration has been found to be higher in women with primary or secondary infertility than in fertile women (Regan et al., 1990). The association between elevated endogenous LH concentrations and miscarriage rates are inconsistent, both in women with normal ovarian morphology and in women with PCOS (Regan et al., 1990; Rai et al., 2000).

In PCOS women undergoing CC treatment, a high endogenous LH concentration *prior* to treatment has been associated with a high probability of conceiving (Kousta et al., 1997; Imani et al., 1999), whereas elevated endogenous LH concentrations *during* the follicular phase of CC treatment have been associated with poor treatment outcome (Shoham et al., 1990). An association between a high serum LH concentration in the follicular phase *during* gonadotrophin-releasing hormone (GnRH) pulsatile treatment and failure to conceive has been reported in PCOS patients (Jacobs & Homburg, 1990). A retrospective analysis in normogonadotrophic anovulatory patients (n=154) suggested that the endogenous LH concentration prior to the start of ovulation

induction with gonadotrophin preparations containing only FSH activity does not affect the chances of achieving a pregnancy (Mulders et al., 2003a). A few studies have measured LH concentrations during ovulation induction with FSH preparations (Coelingh Bennink et al., 1998; Balasch et al., 2003), but the impact of endogenous LH concentrations *during* or *at the end* of gonadotrophin stimulation on treatment outcome in PCOS patients has not been specifically addressed.

Due to the hypersecretion of LH in some patients with PCOS, gonadotrophin preparations containing only FSH activity were considered to have theoretical advantages over human menopausal gonadotrophin (hMG) preparations which contain both FSH and LH activity (Hillier, 1994; Shoham, 2002; Balasch, 2004). However, the existing data do not support these theoretical concerns, and there have been no studies powered to detect differences in clinical outcome. The available individual studies and meta-analyses have not been able to document significant differences in ongoing pregnancy and live birth rates between preparations containing only FSH activity and hMG preparations (Nugent et al., 2000; Platteau et al., 2006).

The major aim of the present retrospective study was to evaluate the roles of endogenous and exogenous LH activity on live birth rate in WHO group II anovulatory women stimulated with rFSH or highly purified menotrophins (HP-hMG), which provide FSH and exogenous LH activity mainly in the form of human chorionic gonadotrophin (hCG). Thus, serum concentrations of LH represent mainly the endogenous LH activity in both groups, and serum hCG represents the exogenous LH activity, which is present only in the HP-hMG group. Serum LH and hCG were compared between those women who achieved a live birth and those who did not, for both treatment regimens. To evaluate potential confounding predictors of live birth, we subsequently performed a full logistic regression analysis of the associations between live birth rate and an extensive range of patient characteristics, as well as endocrinological variables at baseline and end of stimulation.

Material and methods

Study population

This retrospective study was based on 184 women with normogonadotropic anovulatory infertility (WHO Group II) who participated in a prospective randomised controlled multicentre trial, which had ovulation rate as the primary outcome measure after ovulation induction with different gonadotrophin preparations [HP-hMG (Menopur, Ferring Pharmaceuticals A/S, Copenhagen, Denmark) and rFSH (Gonal-F, Merck Serono, Geneva, Switzerland)] using a low-dose step-up protocol (Platteau et al., 2006). HP-hMG contributes both FSH and LH activity; the LH-activity being almost completely (>95% immunoreactivity) derived from hCG molecules (Wolfenson et al., 2005). As reported (Platteau et al.,

2006), the ovulation rates in response to stimulation with the two gonadotrophin preparations are similar: 84.9% (79/93) for the rFSH group and 83.5% (76/91) for the HP-hMG group. In order to investigate the impact of a number of baseline and treatment-associated variables on live birth rate *per se* (i.e. ruling out the no live birth response due to lack of ovulation), only the women who ovulated after stimulation were included in the study cohort (n=155). Main inclusion criteria were chronic anovulation (amenorrhoea, oligomenorrhoea, or low progesterone concentrations in menstrual cycles of duration of 21–35 days), failure to ovulate with CC doses of at least 100 mg/day for at least 5 days or failure to conceive after three cycles of ovulation induction with CC, an age of at least 18 but not more than 39 years, a body mass index (BMI) of 19–35 kg/m², and early follicular serum FSH concentrations between 1 and 12 IU/l. The study was carried out in accordance with the Declaration of Helsinki on good clinical practice, and ethical committee approval was obtained in all participating centres. Written informed consent was obtained from all trial subjects.

Study protocol

Stimulation treatment was started 2–5 days after a spontaneous or progesterone-induced menstrual bleed. The starting dose of gonadotrophin was 75 IU daily, which was maintained for 7 days. After the first 7 days, the dose was either maintained or increased by 37.5 IU increments according to individual responses. All subjects were maintained on their specific dose concentration for at least 7 days. The maximum permitted daily dose was 225 IU, and the women were treated with the gonadotrophin for a maximum of 6 weeks. A single dose of 5,000 IU of hCG (Profasi, Merck Serono, Geneva, Switzerland) was given to trigger ovulation when one follicle of ≥ 17 mm or two to three follicles of ≥ 15 mm were observed by vaginal ultrasound. Any medication for luteal support (e.g. progesterone or hCG) was prohibited.

Blood samples were taken on stimulation day 1 (prior to the start dosing with gonadotrophins) and at the end of stimulation (at least 8 h after the last gonadotrophin dose). Serum was analysed for endocrine variables by a central laboratory using an electrochemiluminescence immunoassay (LH, FSH, hCG, testosterone, prolactin, sex hormone-binding globulin (SHBG)), a radioimmunoassay (estradiol, androstenedione), a chemoluminescence assay (insulin), and an enzymatic method (glucose). The lower detection limits of the validated analytical methods were as follows: LH 0.10 IU/l, FSH 0.10 IU/l, hCG 0.10 IU/l, total testosterone 0.17 nmol/l, prolactin 0.3 μ g/l, SHBG 2 nmol/l, estradiol 55 pmol/l, androstenedione 0.10 nmol/l, insulin 14.4 pmol/l and glucose 1.1 mmol/l.

Ovulation was defined as a serum progesterone concentration of ≥ 7.9 ng/ml (≥ 25 nmol/l) in the mid-luteal phase (6–9 days after hCG). Clinical pregnancy was

defined as a transvaginal ultrasound showing at least one intrauterine gestation sac with foetal heart beat 7–9 weeks after hCG administration. Live birth was defined as a cycle that resulted in at least one live born neonate, regardless of the number of other neonates (live born or still born).

Statistical analysis

Student's *t*-test was used for comparison of continuous variables between those women who did, and those who did not, achieve a live birth. The demographic and pre-stimulation variables as well as the variables obtained at end of ovarian stimulation for each treatment group were included in a primary univariate logistic regression analysis with the dependent variable live birth. The predictive value of each variable was summarised as an odds ratio with 95% confidence interval. The hCG values were multiplied by 10 to have a more interpretable OR estimate. All variables with a *p*-value <0.05 in the univariate regression analysis were then included in a secondary multivariate regression analysis for each treatment group. The multivariate model was reduced stepwise and the variables that were significant predictors of live birth at the <5% level are presented. Model fit was checked by the Hosmer and Lemeshow Goodness-of-Fit test. Tests for differences between variables in subgroups stratified into the 25th and 75th percentiles of exogenous serum hCG concentrations at end of stimulation were performed using the Kruskal–

Wallis test for continuous data and the Chi-square test for categorical data.

Results

Table I shows the demographics and baseline characteristics of the women who ovulated in response to stimulation. All prestimulation variables were comparable in the two treatment groups.

Seventeen women in the rFSH group had a positive clinical pregnancy test and 16 of them achieved a live birth (14 singletons and 2 multiples). In the HP-hMG group, 14 women had a positive clinical pregnancy test and 13 of them achieved a live birth (all singletons). The endogenous LH concentrations prior to treatment did not differ significantly between the live birth and non-live birth subgroups in rFSH- or HP-hMG-treated women (Table II). Neither were the endogenous LH concentrations at end of stimulation significantly different between the two subgroups of each treatment regimen. However, the mean exogenous serum concentration of hCG at end of HP-hMG treatment was found to be 30% higher (*p*=0.010) in the subgroup of women who achieved a live birth compared with the non-live birth subgroup.

In the group of women who were stimulated with rFSH, the univariate logistic regression analysis of the associations between demographic and prestimulation variables as well as the variables obtained at end

Table I. Demographics and baseline characteristics.

| Variables | rFSH (n=79) | HP-hMG (n=76) | All (n=155) |
|--|-------------|---------------|-------------|
| Female age (years) | 29.4±3.8 | 29.4±4.0 | 29.4±3.9 |
| BMI (kg/m ²) | 25.2±4.3 | 25.8±5.1 | 25.5±4.7 |
| Waist-to-hip ratio | 0.82±0.09 | 0.83±0.12 | 0.83±0.10 |
| Primary infertility (%) | 65 | 55 | 60 |
| Duration of infertility (years) | 2.9±1.9 | 2.9±1.8 | 2.9±1.9 |
| Previous ovulation induction cycles | 4.8±2.5 | 4.7±2.5 | 4.7±2.5 |
| Clomiphene citrate non-responders | | | |
| Failure to ovulate* (%) | 38 | 53 | 45 |
| Failure to conceive† (%) | 62 | 47 | 55 |
| Mean ovarian volume (cm ³) | 8.2±4.3 | 8.1±4.1 | 8.1±4.2 |
| Antral follicle count | 22±14 | 23±17 | 23±16 |
| LH (IU/l) | 7.6±4.5 | 7.2±4.8 | 7.4±4.6 |
| LH > 10 IU/l (%) | 23 | 22 | 23 |
| FSH (IU/l) | 5.4±2.7 | 5.1±1.3 | 5.2±2.1 |
| LH:FSH ratio | 1.6±1.2 | 1.5±1.1 | 1.5±1.1 |
| Prolactin (µg/l) | 12±7 | 13±15 | 12±11 |
| Androstenedione (nmol/l) | 7.2±3.5 | 8.0±5.0 | 7.6±4.3 |
| Total testosterone (nmol/l) | 1.6±0.6 | 1.8±0.7 | 1.7±0.6 |
| SHBG (nmol/l) | 62±43 | 57±38 | 59±40 |
| Free androgen index | 4.1±3.3 | 4.9±4.6 | 4.5±4.0 |
| Estradiol (pmol/l) | 158±63 | 156±91 | 157±77 |
| Glucose (mmol/l) | 5.1±0.7 | 5.2±0.7 | 5.1±0.7 |
| Insulin (pmol/l) | 98±101 | 109±117 | 103±109 |
| Insulin:glucose ratio | 2.8±2.6 | 2.9±2.7 | 2.8±2.7 |

Variables expressed as mean±SD, or %.

*At least 100 mg/day for at least 5 days.

†After three cycles.

Table II. Serum concentrations of LH at baseline, and of LH and hCG at end of gonadotrophin stimulation.

| LH activity | rFSH | | | HP-hMG | | |
|----------------------|----------------------|-------------------|-------|----------------------|-------------------|-------|
| | No live birth (n=63) | Live birth (n=16) | p | No live birth (n=63) | Live birth (n=13) | p |
| Baseline | | | | | | |
| Endogenous LH (IU/l) | 7.56±4.42 | 8.00±4.93 | 0.454 | 6.97±4.04 | 8.54±7.63 | 0.287 |
| End of stimulation | | | | | | |
| Endogenous LH (IU/l) | 14.5±17.8* | 9.41±6.54 | 0.289 | 11.2±9.91* | 9.45±9.11 | 0.666 |
| Exogenous hCG (IU/l) | – | – | – | 0.98±0.35* | 1.27±0.39 | 0.010 |

The data are expressed as mean±SD.

*57 women attended the end-of-stimulation visit.

Table III. Univariate logistic regression analysis of the association between live birth and clinical, sonographic and endocrinological parameters prior to start of ovarian stimulation and at end of stimulation.

| Variables | rFSH (n=79) | | | HP-hMG (n=76) | | |
|--|-------------|------------|---------|---------------|-----------|---------|
| | OR | 95% CI | p-value | OR | 95% CI | p-value |
| Baseline | | | | | | |
| Female age (years) | 0.89 | 0.76–1.04 | 0.139 | 0.94 | 0.80–1.09 | 0.395 |
| BMI (kg/m ²) | 1.07 | 0.94–1.21 | 0.318 | 0.96 | 0.85–1.08 | 0.508 |
| Waist-to-hip ratio | 1.02 | 0.55–1.88 | 0.944 | 1.32 | 0.84–2.05 | 0.226 |
| Menstrual cycle pattern* | 3.75 | 0.78–18.03 | 0.098 | 6.00 | 0.73–49.3 | 0.095 |
| Duration of infertility (years) | 0.90 | 0.64–1.27 | 0.551 | 0.70 | 0.41–1.19 | 0.188 |
| Failure to ovulate on cc | 0.54 | 0.18–1.63 | 0.271 | 0.65 | 0.19–2.19 | 0.482 |
| Failure to conceive on cc | 1.86 | 0.62–5.65 | 0.271 | 1.55 | 0.46–5.26 | 0.482 |
| Antral follicle count | 1.02 | 0.99–1.06 | 0.192 | 1.03 | 1.00–1.07 | 0.070 |
| Mean ovarian volume (cm ³) | 1.02 | 0.90–1.15 | 0.761 | 0.99 | 0.85–1.16 | 0.948 |
| LH (IU/l) | 1.02 | 0.91–1.15 | 0.723 | 1.06 | 0.95–1.19 | 0.289 |
| LH (≤10 IU/l vs. >10 IU/l) | 0.59 | 0.17–2.02 | 0.397 | 0.48 | 0.12–1.87 | 0.289 |
| FSH (IU/l) | 0.58 | 0.38–0.90 | 0.015 | 1.08 | 0.69–1.70 | 0.733 |
| Estradiol (pmol/l) | 1.00 | 0.99–1.01 | 0.993 | 0.99 | 0.97–1.00 | 0.041 |
| Prolactin (µg/l) | 0.97 | 0.88–1.07 | 0.516 | 0.92 | 0.80–1.05 | 0.207 |
| Androstenedione (nmol/l) | 1.07 | 0.92–1.25 | 0.357 | 0.94 | 0.80–1.10 | 0.443 |
| Total testosterone (nmol/l) | 1.96 | 0.77–4.95 | 0.155 | 0.56 | 0.20–1.53 | 0.255 |
| SHBG (nmol/l) | 1.00 | 0.99–1.01 | 0.863 | 1.01 | 0.99–1.02 | 0.481 |
| Free androgen index | 1.14 | 0.98–1.33 | 0.090 | 0.86 | 0.69–1.09 | 0.208 |
| Glucose (mmol/l) | 0.67 | 0.28–1.60 | 0.361 | 0.60 | 0.24–1.52 | 0.282 |
| Insulin (pmol/l) | 1.00 | 1.00–1.01 | 0.811 | 0.99 | 0.98–1.00 | 0.205 |
| Insulin: glucose ratio | 1.05 | 0.87–1.27 | 0.629 | 0.77 | 0.50–1.16 | 0.210 |
| End of stimulation | | | | | | |
| Follicular development (multiple vs. mono) | 1.81 | 0.58–5.63 | 0.306 | 0.63 | 0.16–2.54 | 0.516 |
| Duration of gonadotrophin (days) | 1.04 | 0.93–1.17 | 0.507 | 1.02 | 0.93–1.12 | 0.675 |
| Total dose of gonadotrophin (IU) | 1.00 | 1.00–1.00 | 0.641 | 1.00 | 1.00–1.00 | 0.735 |
| Endometrial thickness (mm) | 1.25 | 0.98–1.61 | 0.078 | 0.90 | 0.67–1.20 | 0.461 |
| Estradiol (pmol/l) | 1.00 | 1.00–1.00 | 0.349 | 1.00 | 1.00–1.00 | 0.442 |
| FSH (IU/l) | 0.80 | 0.59–1.08 | 0.147 | 1.18 | 0.87–1.61 | 0.282 |
| hCG (IU/l) | – | – | – | 1.25 | 1.04–1.50 | 0.015 |
| LH (IU/l) | 0.97 | 0.92–1.02 | 0.273 | 0.98 | 0.91–1.05 | 0.568 |
| LH (≤10 IU/l vs. >10 IU/l) | 0.87 | 0.28–2.67 | 0.807 | 1.01 | 0.29–3.47 | 0.993 |

OR, odds ratio; CI, confidence interval.

*Amenorrhea and oligomenorrhea vs. anovulatory cycles.

of ovarian stimulation and live birth showed that there was no significant association between the endogenous LH concentration and live birth rate; neither at baseline [OR=1.02 (95% CI: 0.91–1.15), $p=0.723$] nor at end of stimulation [OR=0.97 (95% CI: 0.92–1.02), $p=0.273$] (Table III). The only variable significantly associated with live birth rate in this treatment group was the serum concentration of FSH at baseline, which was negatively associated with live birth rate [OR=0.58 (95% CI: 0.38–0.90), $p=0.015$]. The variables yielding a p -value ≤ 0.1 in the univariate tests entered into a multivariate logistic regression analysis. As in the univariate tests, only FSH at baseline was a significant predictor of live birth rate at the 5% level.

In the univariate logistic regression analysis of the group of women stimulated with HP-hMG, two variables were found to be significantly associated with the live birth rate; the serum concentrations of estradiol at start of stimulation [OR=0.99 (95% CI: 0.97–1.00), $p=0.041$] and exogenous hCG at end of stimulation [OR=1.25 (95% CI: 1.04–1.50), $p=0.015$]. Similar to the rFSH group, neither the endogenous LH concentration at start of stimulation [OR=1.06 (95% CI: 0.95–1.19), $p=0.289$] nor the LH concentration at end of stimulation [OR=0.98 (95% CI: 0.91–1.05), $p=0.568$] was significantly associated with live birth rate. Furthermore, there was no correlation ($p=0.340$) between the concentrations of LH and hCG at end of stimulation in the group of women stimulated with HP-hMG (data not shown). There was a trend towards an association between live birth rate and antral follicle count [OR=1.03 (95% CI: 1.00–1.07), $p=0.070$], while no other demographic, baseline, clinical, sonographic or endocrine variable was associated with live birth rate. When eliminating the non-significant variables stepwise in a multivariate logistic regression analysis, the exogenous hCG concentration at the end of stimulation was the only significant predictor of live birth rate at the 5% level in the HP-hMG group.

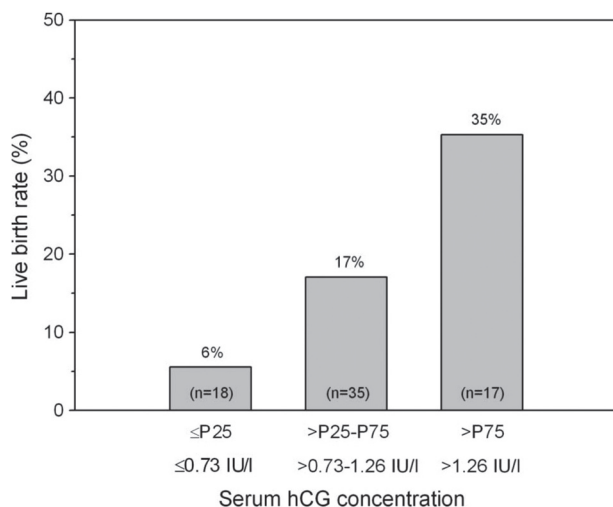


Figure 1. Live birth rate according to the $\leq P25$, $>P25$ – $P75$ and $>P75$ percentiles of serum hCG concentrations at end of HP-hMG stimulation. Seventy women attended the end-of-stimulation visit.

Model fit was checked by the Hosmer and Lemeshow Goodness-of-Fit test, which gave an acceptable fit ($p=0.518$).

The HP-hMG treated women were stratified into the 25th and 75th percentiles (P) of exogenous serum hCG concentrations at end of stimulation, resulting in subgroups of $\leq P25$, $>P25$ – $P75$, and $>P75$. The live birth rate was lowest (6%) in the $\leq P25$ quartile and highest (35%) in the $>P75$ quartile (Figure 1). No significant differences between the percentile subgroups of hCG were observed regarding the demographic and baseline characteristics and the endocrine and sonographic variables at the start of stimulation, except for a significantly ($p=0.028$) higher antral follicle count in the $>P25$ – $P75$ percentile compared to the two extreme quartiles and a trend ($p=0.099$) towards higher BMI in those women with lower hCG concentrations. Concerning stimulation characteristics, the duration of stimulation, the total dose of gonadotrophin and the dose on the last stimulation day as well as the levels of FSH and estradiol at end of stimulation were found to be highest in the $>P75$ quartile (Table IV).

Discussion

The present study did not find a significant association between serum concentrations of LH at baseline and live birth rate, neither in the rFSH group nor in the HP-hMG group. In line with this observation, the endogenous LH levels were similar in women who achieved a live birth and in those who did not. The lack of association between the LH level and outcome in the current study is in agreement with the results of a retrospective study in women with normogonadotropic anovulatory infertility resistant to CC-therapy treated with FSH activity gonadotrophin preparations (Mulders et al., 2003a), but in contrast to the meta-analysis of four studies in this patient category undergoing ovulation induction with FSH (Mulders et al., 2003b). The meta-analysis found a small, but significant, positive association between elevated basal LH levels and pregnancy rates. The present study, however, used a single sensitive LH assay applied to all study samples in a central laboratory, which may partly explain the different results.

The present study shows for the first time that increased serum concentrations of exogenous hCG at the end of stimulation were associated with higher live birth rates in normogonadotropic anovulatory infertile women treated with HP-hMG. Actually, among all the variables obtained at baseline or end of stimulation, the hCG concentration was the only variable predictive of treatment outcome in the multivariate logistic regression analysis. This is an interesting, novel finding in a PCOS population with already normal or high endogenous LH concentrations prior to starting the ovulation induction treatment. In this context, it is important to note that any interpretation of the relation between endogenous LH concentrations and outcome in HP-hMG-treated women is confounded by the fact

that there is an intervention with exogenous hCG, supplementing additional LH activity.

The higher serum hCG concentrations (as well as FSH concentrations) in patients in the highest quartile group at the end of stimulation could be explained by their higher final daily dose of HP-hMG. These patients had a longer duration of stimulation in order to achieve the desired follicular response, but also had significantly higher estradiol levels than the patients in the other quartiles, despite an apparently comparable degree of follicular response. The finding of higher estradiol levels despite similar follicular response could reflect the effect of higher hCG levels on follicular differentiation. Also, the number of antral follicles was different between the percentile subgroups, but was not higher for the >75th percentile. A trend towards a lower BMI in the highest quartile group is in line with a previous report (Chan et al., 2003), which showed an inverse association between BMI and bioavailability of hCG. Despite

all these observations, the concentration of hCG at the end of HP-hMG stimulation was identified as the sole significant positive predictor of live birth rate, while demographic variables, including BMI, or other variables at baseline or at end of stimulation did not add further to the prediction.

The interpretation of this investigation is influenced by certain methodological aspects of the hormone measurements. Immunoassay determination of LH concentrations based on blood sampling every 15 minutes leads to documented major oscillations within short intervals, which are attributed to the pulsatile release of LH and potentially to the exogenous pharmacological interventions (Griesinger & Diedrich, 2006). In the present study, the analysis of endogenous LH concentrations at baseline and the end of stimulation was based on a single blood sample on each occasion which may not truly reflect the mean daily endogenous LH concentration or the overall exposure of reproductive tissues to

Table IV. Demographics, baseline and stimulation characteristics by hCG concentration at end of HP-hMG stimulation.

| Variables | hCG (IU/l) | | | p-value* |
|--|--------------|------------------|--------------|----------|
| | ≤P25 n=18 | >P25–P75 n=35 | >P75 n=17 | |
| Baseline | | | | |
| Female age (years) | 30.1±3.6 | 29.3±3.9 | 28.6±4.7 | 0.561 |
| BMI (kg/m ²) | 27.8±4.1 | 26.0±5.5 | 24.3±5.3 | 0.099 |
| Waist-to-hip ratio | 0.83±0.06 | 0.83±0.12 | 0.86±0.17 | 0.759 |
| Duration of infertility (years) | 3.0±1.5 | 2.7±2.1 | 2.7±1.3 | 0.432 |
| Primary infertility | 39% | 61% | 53% | 0.251 |
| Antral follicle count | 17±11 | 27±15 | 21±24 | 0.028 |
| Mean ovarian volume (cm ³) | 8.2±3.5 | 8.4±4.2 | 7.3±4.5 | 0.468 |
| LH (IU/l) | 5.3±2.0 | 6.9±4.0 | 9.1±7.4 | 0.452 |
| FSH (IU/l) | 5.4±1.4 | 4.7±1.3 | 5.4±1.3 | 0.174 |
| Estradiol (pmol/l) | 132±40 | 176±114 | 151±78 | 0.237 |
| Progesterone (nmol/l) | 3.1±1.3 | 4.7±10.0 | 2.9±2.3 | 0.545 |
| Prolactin (µg/l) | 16.6±27.3 | 11.9±6.6 | 10.5±6.3 | 0.533 |
| Androstenedione (nmol/l) | 7.7±3.7 | 7.9±4.7 | 9.2±7.2 | 0.974 |
| Total testosterone (nmol/l) | 1.7±0.5 | 1.8±0.6 | 1.8±1.0 | 0.583 |
| SHBG (nmol/l) | 43.3±25.8 | 60.2±38.1 | 61.5±47.0 | 0.200 |
| Glucose (mmol/l) | 5.3±0.7 | 5.2±0.8 | 5.0±0.6 | 0.714 |
| Insulin (pmol/l) | 135±132 | 120±136 | 74±57 | 0.099 |
| Insulin:glucose ratio | 3.6±3.0 | 3.2±3.2 | 2.1±1.5 | 0.133 |
| End of stimulation | | | | |
| Subjects with monofollicular development (%) | 67 | 69 | 59 | 0.466 |
| Number of follicles ≥12 mm | 2±1 | 2±2 | 2±2 | 0.780 |
| Duration of gonadotrophin (days) | 9.9±2.6 | 13.6±5.3 | 19.9±7.7 | <0.001 |
| Total dose of gonadotrophin (IU) | 746±198 | 1225±669 | 2014±1102 | <0.001 |
| Last gonadotrophin dose (IU) | 75±0 | 97.5±27.6 | 121.3±33.9 | <0.001 |
| Endometrial thickness (mm) | 9.6±1.8 | 9.5±2.2 | 9.5±2.6 | 0.964 |
| Estradiol (pmol/l) | 831±247 | 1455±1524 | 1644±1552 | 0.017 |
| FSH (IU/l) | 8.4±2.5 | 9.1±1.2 | 11.6±0.90 | <0.001 |
| hCG (IU/l) | 0.6±0.1 | 1.0±0.2 | 1.5±0.2 | <0.001 |
| LH (IU/l) | 14.8±10.9 | 8.8±6.6 | 10.9±12.6 | 0.053 |
| Subjects with LH > 10 IU/l (%) | 56 | 29 | 41 | 0.156 |

Variables expressed as mean±SD, or %.

*Kruskal–Wallis test for continuous and chi-square test for categorical data.

LH. Furthermore, the biological activity of the LH and hCG concentrations may not readily be extrapolated from the results of an immunoassay. Given the longer terminal half-life of hCG, the bioactivity contribution *in vivo* of hCG may be greater than could actually be extrapolated from immunoassay concentrations (de Leeuw et al., 1996).

As WHO type II anovulatory patients generally have normal or elevated LH concentrations, supplementation of LH activity is not considered to be required in this patient category. However, the finding in the present study that the live birth rate in the >75th percentile of hCG in HP-hMG stimulated women was 35% compared to 20% in the rFSH group may suggest that exogenous hCG exerts effects other than pure LH substitution. To date, no clinical studies have been adequately designed to investigate the effect of LH supplementation on live birth rates in anovulatory infertility WHO type II patients. Most of the data on potential favourable effects of LH activity on oocyte/embryo quality and on the endometrium are based on superovulating patients undergoing *in vitro* fertilisation/intracytoplasmic sperm injection (IVF/ICSI) cycles or oocyte donors, which usually exclude patients with PCOS (Acevedo et al., 2004; Platteau et al., 2004; Lisi et al., 2005; Smitz et al., 2007; Ziebe et al., 2007; Weghofer et al., 2008; Hudleston et al., 2009).

In summary, in anovulatory patients with normal or elevated baseline endogenous LH concentrations undergoing ovulation induction with gonadotrophins, the LH concentration at baseline or end of stimulation is not significantly associated with live birth rate. The exogenous LH activity, provided mainly in the form of hCG by using the HP-hMG gonadotrophin preparation, does not appear to be detrimental to live birth rates. Patients with higher serum hCG levels at the end of stimulation actually had the higher live birth rates in this treatment regimen.

Acknowledgements

The study was sponsored by Ferring Pharmaceuticals A/S, Copenhagen, Denmark. The authors thank Klaus Juel Olsen, PhD, Larix Aps, Denmark, for statistical assistance, and Göran Pettersson, PhD, Ferring Pharmaceuticals A/S, Copenhagen, Denmark for support with the preparation of this manuscript. We also thank all the centres from which the study cohort was derived: Belgium: Virga Jesse Ziekenhuis, Hasselt; AZ-VUB, Brussels; AZ Groeninge, Kortrijk; Hôpital Saint Vincent, Rocourt; Hôpital Erasme, Brussels; Centre Hospitalier Notre Dame, Charleroi; AZ St. Lucas, Gent; Univerzitair Ziekenhuis, Gent. United Kingdom: Leeds Hospital; Ninewells Hospital, Dundee; Glasgow Royal Infirmary; The Jessop Wing, Sheffield; Liverpool Women's Hospital; Princess Anne Hospital, Southampton; Guy's Hospital, London. Sweden: Uppsala University Hospital; Sahlgrenska University Hospital, Gothenburg; Lund

University Hospital; Karlstad Hospital; Helsingborg Hospital. Denmark: Copenhagen University Hospital; Brødstrup Hospital; Hvidovre Hospital; Randers Hospital; Skive Hospital; Holbæk Hospital; Herlev Hospital; Odense University Hospital; Skejby Hospital.

Declaration of interest: Joan-Carles Arce is employee of Ferring Pharmaceuticals. Johan Smitz has nothing to disclose.

References

- Acevedo, B., Sanchez, M., Gomez, J. L., Cuadros, J., Ricciarelli, E., & Hernández, E. R. (2004). Luteinizing hormone supplementation increases pregnancy rates in gonadotropin-releasing hormone antagonist donor cycles. *Fertility & Sterility*, 82, 343–347.
- Balasz, J. (2004). *The role of FSH and LH in ovulation induction: current concepts and the contribution of recombinant gonadotropins. Textbook of assisted reproductive techniques: laboratory and clinical perspectives*, In: Gardner, D. K., Weissman, A., Howles, C. M., Shoham, Z., (Eds), 2nd ed London Taylor and Francis, p 541–565
- Balasz, J., Fábregues, F., Casamitjana, R., Peñarrubia, J., & Vanrell, J. A. (2003). A pharmacokinetic and endocrine comparison of recombinant follicle-stimulating hormone and human menopausal gonadotrophin in polycystic ovary syndrome. *Reproductive Biomedicine Online*, 6, 296–301.
- Balen, A. H., Braat, D. D., West, C., Patel, A., & Jacobs, H. S. (1994). Cumulative conception and live birth rates after the treatment of anovulatory infertility: safety and efficacy of ovulation induction in 200 patients. *Human Reproduction*, 9, 1563–1570.
- Balen, A. H., Conway, G. S., Kaltsas, G., Techatrasak, K., Manning, P. J., West, C., & Jacobs, H. S. (1995). Polycystic ovary syndrome: the spectrum of the disorder in 1741 patients. *Human Reproduction*, 10, 2107–2111.
- Chan, C. C., Ng, E. H., Chan, M. M., Tang, O. S., Lau, E. Y., Yeung, W. S., et al. (2003). Bioavailability of hCG after intramuscular or subcutaneous injection in obese and non-obese women. *Human Reproduction*, 18, 2294–2297.
- Coelingh Bennink, H. J., Fauser, B. C., & Out, H. J. (1998). Recombinant follicle-stimulating hormone (FSH; Puregon) is more efficient than urinary FSH (Metrodin) in women with clomiphene citrate-resistant, normogonadotropic, chronic anovulation: a prospective, multicenter, assessor-blind, randomized, clinical trial. European Puregon Collaborative Anovulation Study Group. *Fertility & Sterility*, 69, 19–25.
- Conway, G. S., Honour, J. W., & Jacobs, H. S. (1989). Heterogeneity of the polycystic ovary syndrome: clinical, endocrine and ultrasound features in 556 patients. *Clinical Endocrinology*, 30, 459–470.
- de Leeuw, R., Mulders, J., Voortman, G., Rombout, F., Damm, J., & Kloosterboer, L. (1996). Structure-function relationship of recombinant follicle stimulating hormone (Puregon). *Molecular Human Reproduction*, 2, 361–369.
- Fauser, B. C., Pache, T. D., Lamberts, S. W., Hop, W. C., de Jong, F. H., & Dahl, K. D. (1991). Serum bioactive and immunoreactive luteinizing hormone and follicle-stimulating hormone levels in women with cycle abnormalities, with or without polycystic ovarian disease. *Journal of Clinical Endocrinology and Metabolism*, 73, 811–817.
- Franks, S. (1989). Polycystic ovary syndrome. *Trends in Endocrinology and Metabolism*, 1, 60–63.
- Griesinger, G. & Diedrich, K. (2006). Role of LH in ovarian stimulation: considerations. *Reproductive Biomedicine Online*, 12, 404–406.
- Hamilton-Fairley, D., Kiddy, D., Watson, H., Sagle, M., & Franks, S. (1991). Low-dose gonadotrophin therapy for induction of

- ovulation in 100 women with polycystic ovary syndrome. *Human Reproduction*, 6, 1095–1099.
- Hillier, S. G. (1994). Current concepts of the roles of follicle stimulating hormone and luteinizing hormone in folliculogenesis. *Human Reproduction*, 9, 188–191.
- Homburg, R. & Howles, C. M. (1999). Low-dose FSH therapy for anovulatory infertility associated with polycystic ovary syndrome: rationale, results, reflections and refinements. *Human Reproduction Update*, 5, 493–499.
- Huddleston, H. G., Jackson, K. V., Doyle, J. O., & Racowsky, C. (2009). hMG increases the yield of mature oocytes and excellent-quality embryos in patients with a previous cycle having a high incidence of oocyte immaturity. *Fertility & Sterility*, 92, 946–949.
- Imani, B., Eijkemans, M. J., te Velde, E. R., Habbema, J. D., & Fauser, B. C. (1999). Predictors of chances to conceive in ovulatory patients during clomiphene citrate induction of ovulation in normogonadotropic oligoamenorrhic infertility. *Journal of Clinical Endocrinology and Metabolism*, 84, 1617–1622.
- Jacobs, H. S. & Homburg, R. R. (1990). The endocrinology of conception. *Baillieres Clinical Endocrinology and Metabolism*, 4, 195–205.
- Kousta, E., White, D. M., & Franks, S. (1997). Modern use of clomiphene citrate in induction of ovulation. *Human Reproduction Update*, 3, 359–365.
- Laven, J. S., Imani, B., Eijkemans, M. J., & Fauser, B. C. (2002). New approach to polycystic ovary syndrome and other forms of anovulatory infertility. *Obstetrical & Gynecological Survey*, 57, 755–767.
- Lisi, F., Rinaldi, L., Fishel, S., Caserta, D., Lisi, R., & Campbell, A. (2005). Evaluation of two doses of recombinant luteinizing hormone supplementation in an unselected group of women undergoing follicular stimulation for in vitro fertilization. *Fertility & Sterility*, 83, 309–315.
- Mulders, A. G., Eijkemans, M. J., Imani, B., & Fauser, B. C. (2003a). Prediction of chances for success or complications in gonadotrophin ovulation induction in normogonadotrophic anovulatory infertility. *Reproductive Biomedicine Online*, 7, 170–178.
- Mulders, A. G., Laven, J. S., Eijkemans, M. J. C., Hughes, E. G., & Fauser, B. C. (2003b). Patient predictors for outcome with gonadotrophin ovulation induction in women with normogonadotrophic anovulatory infertility: a meta-analysis. *Human Reproduction Update*, 9, 429–449.
- Nugent, D., Vandekerckhove, P., Hughes, E., Arnot, M., & Lilford, R. (2000). Gonadotrophin therapy for ovulation induction in subfertility associated with polycystic ovary syndrome. *Cochrane Database of Systematic Reviews 2000*, Issue 3. Art. No.: CD000410. DOI: 10.1002/14651858.CD000410.
- Platteau, P., Nyboe Andersen, A., Balen, A., Devroey, P., Sørensen, P. Helmggaard, L., et al. (2006). Similar ovulation rates, but different follicular development with highly purified menotropin compared with recombinant FSH in WHO Group II anovulatory infertility: a randomized controlled study. *Human Reproduction*, 21, 1798–1804.
- Platteau, P., Smits, J., Albano, C., Sørensen, P., Arce, J.-C., & Devroey, P. (2004). Exogenous luteinizing hormone activity may influence the treatment outcome in *in vitro* fertilization but not in intracytoplasmic sperm injection cycles. *Fertility & Sterility*, 81, 1401–1404.
- Rai, R., Backos, M., Rushworth, F., & Regan, L. (2000). Polycystic ovaries and recurrent miscarriage—a reappraisal. *Human Reproduction*, 15, 612–615.
- Regan, L., Owen, E. J., & Jacobs, H. S. (1990). Hypersecretion of luteinising hormone, infertility, and miscarriage. *Lancet*, 336, 1141–1144.
- Shoham, Z. (2002). The clinical therapeutic window for luteinizing hormone in controlled ovarian stimulation. *Fertility & Sterility*, 77, 1170–1177.
- Shoham, Z., Borenstein, R., Lunenfeld, B., & Pariente, C. (1990). Hormonal profiles following clomiphene citrate therapy in conception and nonconception cycles. *Clinical Endocrinology*, 33, 271–278.
- Smits, J., Andersen, A. N., Devroey, P., Arce, J. C., & MERIT Group. (2007). Endocrine profile in serum and follicular fluid differs after ovarian stimulation with HP-hMG or recombinant FSH in IVF patients. *Human Reproduction*, 22, 676–687.
- Taylor, A. E., McCourt, B., Martin, K. A., Anderson, E. J., Adams, J. M., & Schoenfeld, D. (1997). Determinants of abnormal gonadotropin secretion in clinically defined women with polycystic ovary syndrome. *Journal of Clinical Endocrinology and Metabolism*, 82, 2248–2256.
- van Santbrink, E. J., Hop, W. C., & Fauser, B. C. (1997). Classification of normogonadotrophic infertility: polycystic ovaries diagnosed by ultrasound versus endocrine characteristics of polycystic ovary syndrome. *Fertility & Sterility*, 67, 452–458.
- Weghofer, A., Munné, S., Brannath, W., Chen, S., Tomkin, G., Cekleniak, N., et al. (2008). The impact of LH-containing gonadotropins on diploidy rates in preimplantation embryos: long protocol stimulation. *Human Reproduction*, 23, 499–503.
- Wolfenson, C., Groisman, J., Couto, A. S., Hedenfalk, M., Cortvrint, R. G., Smits, J. E., et al. (2005). Batch-to-batch consistency of human-derived gonadotrophin preparations compared with recombinant preparations. *Reproductive Biomedicine Online*, 10, 442–454.
- Ziebe, S., Lundin, K., Janssens, R., Helmggaard, L., Arce, J. C., & MERIT (Menotropin vs Recombinant FSH in vitro Fertilisation Trial) Group. (2007). Influence of ovarian stimulation with HP-hMG or recombinant FSH on embryo quality parameters in patients undergoing IVF. *Human Reproduction*, 22, 2404–2413.