# Reproducibility of Post-Amphetamine [<sup>11</sup>C]FLB 457 Binding to Cortical D<sub>2/3</sub> Receptors

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### Abstract

In a recent positron emission tomography (PET) study, we demonstrated the ability to measure amphetamineinduced dopamine (DA) release in the human cortex with the relatively high affinity dopamine  $D_{2/3}$  radioligand [<sup>11</sup>C]FLB 457. Herein we report on reproducibility and reliability of [<sup>11</sup>C]FLB 457 binding potential relative to nondisplaceable uptake (BP<sub>ND</sub>) following an acute amphetamine challenge. Ten healthy human subjects were studied twice with [<sup>11</sup>C]FLB 457 following an acute amphetamine (oral, 0.5 mg kg<sup>-1</sup> dose) challenge on two-separate days approximately one week apart.  $D_{2/3}$  receptor binding parameters were estimated using a two-tissue compartment kinetic analysis in the cortical regions of interest and cerebellum (reference region). The test-retest variability and intraclass correlation coefficient were assessed for distribution volume (V<sub>T</sub>), binding potential relative to plasma concentration (BP<sub>P</sub>), and BP<sub>ND</sub> of [<sup>11</sup>C]FLB 457. The test-retest variability of [<sup>11</sup>C]FLB 457 V<sub>T</sub>, BP<sub>P</sub> and BP<sub>ND</sub> were  $\leq$ 17%, 22% and 11% respectively. These results, which are consistent with the published test-retest variability for this ligand measured under baseline conditions demonstrate that the post-amphetamine [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> is reproducible. These data further support the use [<sup>11</sup>C]FLB 457 and amphetamine to characterize cortical dopamine transmission in neuropsychiatric disorders.

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## Introduction

The competition between dopamine (DA) and D<sub>2/3</sub> radiotracer binding (such as [11C]raclopride and [123I]IBZM) following an amphetamine challenge is a noninvasive measure of the change in extracellular DA induced by the challenge [1-7] and is thought to provide information as to the status of DA transmission in the brain. Previous investigations have used this technique to report abnormal DA release in the striatum in patients with schizophrenia [4,8], alcoholism [9] and drug abuse [10-12]. A limitation of these previous studies is that the relatively low binding potential for [<sup>11</sup>C]raclopride and [<sup>123</sup>I]IBZM in the extrastriatal regions precluded the investigation of DA in the cortical regions that have been implicated in these disorders. Thus, it is of interest to develop an imaging paradigm to measure cortical DA in schizophrenia and addiction. In a recent PET study, we demonstrated the ability to detect amphetamine-induced DA release in the human cortex with the dopamine  $D_{2/3}$  radioligand [<sup>11</sup>C]FLB 457 [13]. The results of this study, which showed a significant reduction in the in vivo binding of [11C]FLB 457 following oral amphetamine (0.5 mg kg<sup>-1</sup>) led to further characterization of this imaging paradigm as a tool to measure cortical DA release. In a series of validation studies following this report we have shown: good reproducibility for [11C]FLB 457 BP<sub>ND</sub> under baseline conditions [14]; no carryover mass induced decrease in BP<sub>ND</sub> [14]; a relatively small fraction of D<sub>2/3</sub> receptor specific binding for [11C]FLB 457 in the cerebellar reference region [15]; and a linear relationship between the amphetamine-induced decrease in [11C]FLB 457  $BP_{ND}$  and increase in extracellular DA using combined PET and microdialysis [16]. In addition, we have replicated our initial report of amphetamine-induced displacement of [11C]FLB 457 BPND in an independent cohort of subjects [17]. In a previous study, we demonstrated that the test-retest variability for [11C]FLB 457 BP<sub>ND</sub> measured under control conditions (i.e., at baseline) is an acceptable  $\leq$  15% in the cortical regions of interest -- a result which is also consistent with other published [11C]FLB 457 reproducibility studies [14,18,19]. Here, we were interested in evaluating the test-retest variability of the post-amphetamine [11C]FLB 457  $BP_{ND}$  to ensure that it is reproducible. To evaluate this issue we conducted test and retest amphetamine studies to measure the **Table 1.** Reproducibility of [<sup>11</sup>C]FLB 457 and amphetamine in plasma.

			BSSD		WSSD		
Parameter	Mean	BSSD	cv	WSSE	cv	VAR + SD	ICC
[ <sup>11</sup> C]FLB 457 fp (%)	0.38	0.05	0.14	0.06	0.15	23.0% ± 18.6%	-0.05
[ <sup>11</sup> C]FLB 457 Clearance (L/h)	69.72	14.33	0.21	8.85	0.13	23.7% ± 15.3%	0.45
Amphetamine level 0 min (ng/mL)	87.34	11.83	0.14	4.11	0.05	8.1% ± 7.2%	0.78
Amphetamine level 45 min (ng/mL)	76.60	11.22	0.15	3.45	0.05	7.1% ± 6.6%	0.83
Amphetamine level 90 min (ng/mL)	73.82	11.08	0.15	4.16	0.06	9.8% ± 8.8%	0.75

Values are the mean of 10 subjects with each value measured twice. BSSD CV = between subject standard deviation coefficient of variation, WSSD CV = within subject standard deviation coefficient of variation, VAR = test/retest variability, ICC = intraclass correlation coefficient.

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reproducibility of the post-amphetamine  $V_T$  (regional distribution volume), BP<sub>P</sub> and BP<sub>ND</sub> in ten healthy human subjects.

#### **Materials and Methods**

#### **Ethics statement**

The Institutional Review Board and Radioactive Drug Research Committee of the University of Pittsburgh approved the study. All subjects provided written informed consent.

## Study design

A total of 20 PET scans were acquired for this study in ten healthy control (4 females/6 males; 2 Asian/8 Caucasian; age  $23 \pm 4$ , weight 70  $\pm$  12 kg) subjects. Each subject was scanned with [<sup>11</sup>C]FLB 457 following amphetamine (oral, 0.5 mg kg<sup>-1</sup>) in a test and retest condition separated by one week.

## **PET Protocol**

Radiolabeling of [<sup>11</sup>C]FLB 457 was performed as outlined in previously published procedures [20]. Imaging experiments with amphetamine were conducted on the Siemens ECAT EXACT HR+ scanner consistent with previously described image acquisition protocols [13]. [<sup>11</sup>C]FLB 457 was administered as a bolus intravenous injection three-hours following the administration of 0.5 mg kg<sup>-1</sup> of d-amphetamine (Dexedrine, oral formulation) and emission data were collected for 90 minutes. An oral amphetamine dose of 0.5 mg kg<sup>-1</sup> is consistent with what has been used in previous PET investigations to measured dopamine release in the striatal and extra striatal regions [21-25]. Previous microdialysis studies in primates have shown that 0.5 mg kg<sup>-1</sup> of amphetamine increases extracellular dopamine concentrations by 1320  $\pm$ 432% [16]. **Table 2.** Reproducibility of post-amphetamine [ $^{11}$ C]FLB 457 total distribution volume (V<sub>T</sub>, mL cm<sup>-3</sup>).

			BSSD		WSSD		
Region	Mean	BSSD	cv	WSSD	CV	VAR + SD	ICC
Cerebellum	3.90	0.51	0.13	0.37	0.09	14.7% ± 11.1%	0.32
Medial Temporal Lobe	8.45	1.55	0.18	0.79	0.09	14.9% ± 11.1%	0.59
Anterior Cingulate Cortex	6.97	1.15	0.16	0.68	0.10	14.7% ± 13.0%	0.48
Dorsolateral prefrontal Cortex	5.96	1.32	0.22	0.54	0.09	14.4% ± 10.7%	0.72
Orbital Frontal Cortex	7.09	1.49	0.21	0.72	0.10	16.8% ± 11.7%	0.62
Medial Prefrontal Cortex	6.36	1.06	0.17	0.54	0.09	13.5% ± 11.1%	0.59
Temporal Cortex	9.64	2.30	0.24	0.90	0.09	14.8% ± 11.3%	0.73
Parietal Cortex	6.13	1.61	0.26	0.54	0.09	14.0% ± 10.8%	0.80
Occipital Cortex	5.81	1.59	0.27	0.53	0.09	14.7% ± 10.7%	0.80

Values are the mean of 10 subjects with each value measured twice. BSSD CV = between subject standard deviation coefficient of variation, WSSD CV = within subject standard deviation coefficient of variation, VAR = test/retest variability, ICC = intraclass correlation coefficient.

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Following radiotracer injection, arterial samples were collected manually approximately every 6 seconds for the first 2 minutes and thereafter at longer intervals. A total of 35 samples were obtained per scan. Following centrifugation, plasma was collected in 200 µL aliquots and activities were counted in a gamma well counter. To determine the plasma activity representing unmetabolized [11C]FLB 457 parent compound, six samples (collected at 4, 10, 20, 40, 60 and 80 min) were further processed using high-performance liquid chromatography methods [26] and fitted using a Hill model [27,28]. The input function was then calculated as the product of total counts and interpolated parent fraction at each time point. The measured input function values were fitted to a sum of three exponentials from the time of peak plasma activity and the fitted values were used as the input to the kinetic analysis. The clearance of the parent compound (C<sub>1</sub>, L/h) was calculated as the ratio of the injected dose to the area under the curve of the input function [29]. In addition, measurement of plasma free fraction (f<sub>P</sub>) for [<sup>11</sup>C]FLB 457 was performed [30]. Amphetamine plasma levels were measured in three arterial samples obtained at time 0 min, 45 min and 90 relative to the PET scan as previously described [31]. These data ensured that differences in plasma amphetamine concentration did not bias the test and retest comparison.

**Table 3.** Reproducibility of post-amphetamine [ $^{11}$ C]FLB 457 binding potential relative to plasma concentrations (BP<sub>P</sub>, mL cm<sup>-3</sup>).

			BSSD		WSSD		
Region	Mean	BSSD	cv	WSSD	CV	VAR + SD	ICC
Medial Temporal Lobe	4.55	1.20	0.26	0.45	0.10	16.1% ± 11.1%	0.75
Anterior Cingulate Cortex	3.07	0.74	0.24	0.32	0.11	16.9% ± 14.5%	0.68
Dorsolateral prefrontal Cortex	2.06	0.91	0.44	0.19	0.09	14.8% ± 9.3%	0.92
Orbital Frontal Cortex	3.19	1.08	0.34	0.38	0.12	21.7% ± 13.4%	0.78
Medial Prefrontal Cortex	2.46	0.69	0.28	0.18	0.08	12.4% ± 11.1%	0.87
Temporal Cortex	5.74	1.90	0.33	0.56	0.10	15.3% ± 11.3%	0.84
Parietal Cortex	2.23	1.21	0.54	0.18	0.08	13.9% ± 9.8%	0.96
Occipital Cortex	1.91	1.17	0.61	0.19	0.10	17.2% ± 9.8%	0.95

Values are the mean of 10 subjects with each value measured twice. BSSD CV = between subject standard deviation coefficient of variation, WSSD CV = within subject standard deviation coefficient of variation, VAR = test/retest variability, ICC = intraclass correlation coefficient.

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#### **MRI Protocol**

Prior to PET imaging, a magnetization prepared rapid gradient echo structural MRI scan was obtained using a Siemens 3 Tesla Trio scanner for determination of regions of interest. MRI segmentation was performed using the automated segmentation tool [32] implemented in the FMRIB Software Library v4.0 [33].

#### Analysis of PET data

PET data were reconstructed and processed with the image analysis software MEDx (Sensor Systems, Inc., Sterling, Virginia) and SPM2 (www.fil.ion.ucl.ac.uk/spm) as described in [13]. Frame-to-frame motion correction for head movement and MR-PET image alignment were performed using a mutual information algorithm implemented in SPM2. Time activity curves were generated for the eight cortical regions of interest and cerebellum (reference region) using the criteria and methods outlined in [13,14]. Sampled cortical regions (n = 8) included the medial temporal lobe (MTL), anterior cingulate cortex (ACC), dorsolateral prefrontal cortex (DLPFC), orbital frontal cortex (OFC, defined using criteria outlined in Lacerda 2003), medial prefrontal cortex (MPFC), temporal cortex (TC), parietal cortex (PC), and occipital cortex (OC). The three outcome measures provided are regional tissue distribution volume (VT, mL cm-3), binding potential relative to plasma concentration (BPP, mL cm-3) and binding potential relative to non-displaceable uptake (BPND, unitless) [34]. Derivation of [11C]FLB 457 VT in the regions of interest and cerebellum were performed using a two-tissue compartment kinetic analysis using the arterial input function as described in [13].

#### Statistical analysis

The reproducibility of the plasma ( $f_p$ ,  $C_L$  and amphetamine levels) and brain ( $V_T$ ,  $BP_P$  and  $BP_{ND}$ ) outcome measures were evaluated for their *variability* and *reliability*.

The test-retest variability (VAR) was calculated as the absolute value of the difference between the test and retest, divided by the mean of the test and retest values.

To evaluate the within-subject variability relative to the between-subject variability, both within-subject standard deviation (WSSD) and between-subject standard deviation (BSSD) were calculated and expressed as fraction of mean value (WS CV and BS CV). The reliability of the measurements was assessed by the intraclass correlation coefficient (ICC) calculated as [35]:

BSMS	S-V	VSMSS	
	(	1) WOMO	ļ

BSMSS + (n-1)WSMSS

where BSMSS is the mean sum of square between subjects, WSMSS is the mean sum of square within subjects and n is the number of repeated observations (n = 2 in this study). This statistic estimates the relative contributions of between and within subject variability and assumes values from -1 (i.e. BSMSS = 0) to 1 (identity between test and retest, i.e. WSMSS = 0).

### Results

## **Baseline scan parameters**

The mean injected dose for [<sup>11</sup>C]FLB 457 in the test and retest conditions were  $8.4 \pm 0.3$  mCi and  $7.5 \pm 1.4$  mCi. The mean injected specific activity in the test and retest conditions were 10186  $\pm$  3638 Ci/mmol and 7805  $\pm$  3762 Ci/mmol. The mean injected mass for [<sup>11</sup>C]FLB 457 in the test and retest conditions were  $0.3 \pm 0.1 \mu g$  and  $0.4 \pm 0.1 \mu g$ . There were no significant differences in injected dose or mass between the test and the retest conditions (paired t test, p > 0.05).

### Plasma analysis

The mean [<sup>11</sup>C]FLB 457 fp,  $C_L$ , and amphetamine plasma levels (measured at 0, 45 and 90 min following the [<sup>11</sup>C]FLB 457 injection) and their corresponding VAR and ICC are provided in Table 1.

#### Brain analysis

The mean VT,  $BP_{P}$ ,  $BP_{ND}$  and their corresponding VAR and ICC for the regions of interest are provided in Tables 2, 3 and 4.

## Discussion

The results of this study show that the post-amphetamine [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> is reproducible. The test-retest variability of  $\leq$  15% for [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> in the cortical regions of interest measured in the post-amphetamine condition is comparable to

**Table 4.** Reproducibility of post-amphetamine [ $^{11}C$ ]FLB 457 binding potential relative to non specific uptake (BP<sub>ND</sub>, unitless).

			BSSD		WSSD		
Region	Mean	BSSD	CV	WSSD	cv	VAR + SD	ICC
Medial Temporal Lobe	1.16	0.26	0.22	0.06	0.05	6.8% ± 5.8%	0.91
Anterior Cingulate Cortex	0.78	0.15	0.19	0.03	0.04	6.9% ± 5.2%	0.91
Dorsolateral prefrontal Cortex	0.52	0.19	0.37	0.02	0.03	5.4% ± 4.9%	0.98
Orbital Frontal Cortex	0.81	0.21	0.27	0.05	0.06	11.1% ± 7.9%	0.90
Medial Prefrontal Cortex	0.63	0.15	0.24	0.02	0.03	4.3% ± 4.3%	0.97
Temporal Cortex	1.45	0.36	0.25	0.05	0.03	3.9% ± 3.9%	0.97
Parietal Cortex	0.55	0.26	0.47	0.01	0.03	4.2% ± 2.8%	0.99
Occipital Cortex	0.47	0.26	0.56	0.02	0.05	6.7% ± 4.9%	0.99

Values are the mean of 10 subjects with each value measured twice. BSSD CV = between subject standard deviation coefficient of variation, WSSD CV = within subject standard deviation coefficient of variation, VAR = test/retest variability, ICC = intraclass correlation coefficient.

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that reported in the baseline condition in our previous report [14]. It was necessary to evaluate the test-retest variability for the post-amphetamine PET measurements because [11C]FLB 457 BP<sub>ND</sub> is lower following amphetamine compared to baseline [13]. The good reproducibility of [11C]FLB 457 BP<sub>ND</sub> in the baseline and post-amphetamine conditions suggest that the relatively low cortical binding potential in itself does not pose a problem to the use of this tool to measure cortical DA release. This point is further illustrated in Table 5 which shows an effect size (d) of 0.5 to 2.2 to measure amphetamine-induced change ( $\Delta$ ) of [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> in the cortical regions. These d values are comparable to that observed with [11C]raclopride to detect of an effect for amphetamine in the striatal subdivisions (d=0.8 to 1.9, derived as ABPND/test-retest variability using data in [36,37]). This suggests that the [11C]FLB 457 BP<sub>ND</sub> measured under baseline and post-amphetamine conditions will be distinguishable.

**Table 5.** Effect size to measure amphetamine-induced displacement of [ $^{11}C$ ]FLB 457 BP<sub>ND</sub>.

			Post-AMPH T-RT Effe			
Region	Δ BP <sub>ND</sub> (%	%) BASE T-RT (%)	) (%)	size (d)		
Medial Temporal Lobe	-7 ± 6	11 ± 5	7 ± 6	0.76		
Anterior Cingulate Cortex	-8 ± 8	15 ± 8	7 ± 5	0.68		
Dorsolateral prefrontal Cortex	-13 ± 15	8 ± 6	5 ± 5	1.95		
Orbital Frontal Cortex	-8 ± 15	7 ± 6	11 ± 8	0.87		
Medial Prefrontal Cortex	-11 ± 14	6 ± 4	4 ± 4	2.16		
Temporal Cortex	-4 ± 9	10 ± 6	4 ± 4	0.53		
Parietal Cortex	-12 ± 13	8 ± 4	4 ± 3	1.90		
Occipital Cortex	-5 ± 20	10 ± 4	7 ± 5	0.58		

% values shown are mean ± standard deviation (SD);  $\Delta$  BP<sub>ND</sub> is amphetamine-induced displacement of [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> [13,15];

BASE T-RT is test-retest variability [<sup>11</sup>C]FLB 457 BP<sub>ND</sub> under baseline conditions [14];

Post-AMPH T-RT is test-retest variability of [<sup>11</sup>C]FLB 457 BPND under postamphetamine conditions (this study, Table 4);

Effect size (d) is computed as mean  $\Delta$  BP <sub>ND</sub>/ mean pooled variability; Pooled variability was calculated as the square root of (BASE T-RT<sup>2</sup> + POST-AMPH T-RT<sup>2</sup>)/2 to incorporate both the baseline and post-amphetamine test-retest data. doi: 10.1371/journal.pone.0076905.t005

The measured test-retest variability for  $V_T$  (14-17%) and  $BP_P$ (12-22%) was higher than BP<sub>ND</sub> (4-11%) in the postamphetamine condition. This is consistent with what has been reported for [11C]FLB 457 in the baseline condition, and other DA D<sub>2/3</sub> PET radioligands such as [<sup>11</sup>C]raclopride and [<sup>11</sup>C]NPA [38-40]. BP\_{ND} as opposed to BP\_P and V\_T is associated with lower test-retest variability because it is less vulnerable to the experimental errors associated with the measurement of the plasma input function. Therefore, it is the preferred outcome measure in amphetamine challenge studies that measure a relatively small decrease in radiotracer binding (~10-15%) [41]. An important assumption in the use of  $\Delta BP_{ND}$  to quantify dopamine release is that amphetamine does not affect the nonspecific binding in the brain ( $V_{\mbox{\tiny ND}}$ ). This assumption is tested in amphetamine-PET studies by documenting  $V_{\mbox{\tiny ND}}$  in the baseline and post-amphetamine condition. The use of  $\Delta BP_P$  and  $\Delta V_T$  to quantify dopamine release is necessary when this assumption fails because these outcome measures are somewhat less influenced by amphetamine-induced changes in  $V_{ND}$  [42]. Thus, it was necessary to document the test-retest variability for all outcome measures --  $BP_{\mbox{\tiny ND}},\ BP_{\mbox{\tiny P}}$  and  $V_{\mbox{\tiny T}}$  in the postamphetamine condition. These results suggest that the use of  $\Delta BP_P$  and  $\Delta V_T$  to quantify dopamine release in amphetamine challenge studies might be limited by its relatively higher testretest variability.

In summary, we evaluated the reproducibility of the postamphetamine [<sup>11</sup>C]FLB 457 BP<sub>ND</sub>, and found it to be consistent with that measured under baseline conditions. The results of this reproducibility study support the use of [<sup>11</sup>C]FLB 457 to measure cortical dopamine release despite its relatively low binding potential (BP<sub>ND</sub>).

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## References

- Laruelle M, Iyer RN, AI-Tikriti MS, Zea-Ponce Y, Malison R et al. (1997) Microdialysis and SPECT measurements of amphetamine-induced dopamine release in nonhuman primates. Synapse 25: 1-14. doi: 10.1002/(SICI)1098-2396(199701)25:1. PubMed: 8987142.
- Laruelle M, Abi-Dargham A, van Dyck CH, Rosenblatt W, Zea-Ponce Y et al. (1995) SPECT imaging of striatal dopamine release after amphetamine challenge. J Nucl Med 36: 1182-1190. PubMed: 7790942.
- Kegeles LS, Zea-Ponce Y, Abi-Dargham A, Rodenhiser J, Wang T et al. (1999) Stability of [123I]IBZM SPECT measurement of amphetamine-induced striatal dopamine release in humans. Synapse 31: 302-308. doi:10.1002/(SICI)1098-2396(19990315)31:4. PubMed: 10051112.
- Breier A, Su TP, Saunders R, Carson RE, Kolachana BS et al. (1997) Schizophrenia is associated with elevated amphetamine-induced synaptic dopamine concentrations: Evidence from a novel positron emission tomography method. Proc Natl Acad Sci U S A 94: 2569-2574. doi:10.1073/pnas.94.6.2569. PubMed: 9122236.
- Carson RE, Breier A, deBartolomeis A, Saunders RC, Su TP et al. (1997) Quantification of amphetamine-induced changes in [C-11]raclopride binding with continuous infusion. J Cereb Blood Flow Metab 17: 437-447. PubMed: 9143226.
- Endres CJ, Kolachana BS, Saunders RC, Su T, Weinberger D et al. (1997) Kinetic modeling of [C-11]raclopride: Combined PETmicrodialysis studies. J Cereb Blood Flow Metab 17: 932-942. PubMed: 9307606.
- Villemagne VL, Wong DF, Yokoi F, Stephane M, Rice KC et al. (1999) GBR12909 attenuates amphetamine-induced striatal dopamine release as measured by [(11)C]raclopride continuous infusion PET scans. Synapse 33: 268-273. doi:10.1002/(SICI)1098-2396(19990915)33:4. PubMed: 10421707.
- Laruelle M, Abi-Dargham A, van Dyck CH, Gil R, De Souza CD et al. (1996) Single photon emission computerized tomography imaging of amphetamine-induced dopamine release in drug free schizophrenic subjects. Proc Natl Acad Sci U S A 93: 9235-9240. doi:10.1073/pnas. 93.17.9235. PubMed: 8799184.
- Martinez D, Gil R, Slifstein M, Hwang DR, Huang Y et al. (2005) Alcohol dependence is associated with blunted dopamine transmission in the ventral striatum. Biol Psychiatry 58: 779-786. doi:10.1016/ j.biopsych.2005.04.044. PubMed: 16018986.
- Volkow ND, Wang GJ, Fowler JS, Logan J, Gatley SJ et al. (1997) Decreased striatal dopaminergic responsiveness in detoxified cocainedependent subjects. Nature 386: 830-833. doi:10.1038/386830a0. PubMed: 9126741.
- Martinez D, Narendran R, Foltin RW, Slifstein M, Hwang DR et al. (2007) Amphetamine-induced dopamine release is markedly blunted in cocaine dependent subjects and predictive of the choice to self administer cocaine. Am J Psychiatry 164: 622-629. doi:10.1176/ appi.ajp.164.4.622. PubMed: 17403976.
- Martinez D, Saccone PA, Liu F, Slifstein M, Orlowska D et al. (2012) Deficits in dopamine D(2) receptors and presynaptic dopamine in heroin dependence: commonalities and differences with other types of addiction. Biol Psychiatry 71: 192-198. doi:10.1016/j.biopsych. 2011.08.024. PubMed: 22015315.
- Narendran R, Frankle WG, Mason NS, Rabiner EA, Gunn RN et al. (2009) Positron emission tomography imaging of amphetamine-induced dopamine release in the human cortex: a comparative evaluation of the high affinity dopamine D2/3 radiotracers [11C]FLB 457 and [11C]fallypride. Synapse 63: 447-461. doi:10.1002/syn.20628. PubMed: 19217025.

acquisition of PET data and assisted with the care of all subjects during PET procedures.

#### **Author Contributions**

Conceived and designed the experiments: RN. Performed the experiments: RN NSM. Analyzed the data: RN MH. Contributed reagents/materials/analysis tools: NSM. Wrote the manuscript: RN MH NSM.

- Narendran R, Mason NS, May MA, Chen CM, Kendro S et al. (2011) Positron emission tomography imaging of dopamine D/ receptors in the human cortex with [(1) (1)C]FLB 457: reproducibility studies. Synapse 65: 35-40. doi:10.1002/syn.20813. PubMed: 20506186.
- Narendran R, Mason NS, Chen CM, Himes M, Keating P et al. (2011) Evaluation of dopamine D/ specific binding in the cerebellum for the positron emission tomography radiotracer [(1) (1)C]FLB 457: implications for measuring cortical dopamine release. Synapse 65: 991-997. doi:10.1002/syn.20926. PubMed: 21360596.
- Narendran R, Jedema HP, Lopresti B, Mason NS, Gurnsey K et al. (2013) Imaging dopamine transmission in the frontal cortex: a simultaneous microdialysis and [11C]FLB 457 PET study. Mol Psychiatry (In press).
- Deuitch L, Gurnsey K, Ruskiewicz J, Himes M, Griswold K et al. (2012). Imaging prefrontal cortical dopamine transmission with [11C]FLB 457 and amphetamine. New Orleans, LA.
- Sudo Y, Suhara T, Inoue M, Ito H, Suzuki K et al. (2001) Reproducibility of [11 C]FLB 457 binding in extrastriatal regions. Nucl Med Commun 22: 1215-1221. doi: 10.1097/00006231-200111000-00008. PubMed: 11606887.
- Vilkman H, Kajander J, Någren K, Oikonen V, Syvälahti E et al. (2000) Measurement of extrastriatal D2-like receptor binding with [11C]FLB 457--a test-retest analysis. Eur J Nucl Med 27: 1666-1673. doi:10.1007/ s002590000342. PubMed: 11105823.
- Halldin C, Farde L, Högberg T, Mohell N, Hall H et al. (1995) Carbon-11-FLB 457: a radioligand for extrastriatal D2 dopamine receptors. J Nucl Med 36: 1275-1281. PubMed: 7790956.
- Narendran R, Frankle WG, Mason NS, Rabiner EA, Gunn RN et al. (2009) Positron Emission Tomography Imaging of Amphetamine-Induced Dopamine Release in the Human Cortex: A comparative evaluation of the high affinity dopamine D2/3 radiotracers [11C]FLB 457 and [11C]fallypride. Synapse 63: 447-461. doi:10.1002/syn.20628. PubMed: 19217025.
- Narendran R, Mason NS, Laymon CM, Lopresti BJ, Velasquez ND et al. (2010) A comparative evaluation of the dopamine D(2/3) agonist radiotracer [11C](-)-N-propyl-norapomorphine and antagonist [11C]raclopride to measure amphetamine-induced dopamine release in the human striatum. J Pharmacol Exp Ther 333: 533-539. doi:10.1124/ jpet.109.163501. PubMed: 20103586.
- Bailer UF, Narendran R, Frankle WG, Himes ML, Duvvuri V et al. (2012) Amphetamine induced dopamine release increases anxiety in individuals recovered from anorexia nervosa. Int J Eat Disord 45: 263-271. doi:10.1002/eat.20937. PubMed: 21541980.
- Riccardi P, Li R, Ansari MS, Zald D, Park S et al. (2005) Amphetamine-Induced Displacement of [(18)F] Fallypride in Striatum and Extrastriatal Regions in Humans. Neuropsychopharmacology 31: 1016-1026.
  Cropley VL, Innis RB, Nathan PJ, Brown AK, Sangare JL et al. (2008)
- Cropley VL, Innis RB, Nathan PJ, Brown AK, Sangare JL et al. (2008) Small effect of dopamine release and no effect of dopamine depletion on [(18)F]fallypride binding in healthy humans. Synapse 62: 399-408. doi:10.1002/syn.20506. PubMed: 18361438.
- Olsson H, Halldin C, Swahn CG, Farde L (1999) Quantification of [11C]FLB 457 binding to extrastriatal dopamine receptors in the human brain. J Cereb Blood Flow Metab 19: 1164-1173. PubMed: 10532641.
- Hill AV (1910) The possible effects of the aggregation of the molecules of haemoglobin on its disassociation curves. Phys Med Biol 1991: 749-761.
- Gunn RN, Sargent PA, Bench CJ, Rabiner EA, Osman S et al. (1998) Tracer kinetic modeling of the 5-HT1A receptor ligand [carbonyl-11C]WAY- 100635 for PET. Neuroimage 8: 426-440. doi: 10.1006/nimg.1998.0379. PubMed: 9811559.

- Abi-Dargham A, Laruelle M, Seibyl J, Rattner Z, Baldwin RM et al. (1994) SPECT measurement of benzodiazepine receptors in human brain with [123-I]iomazenil: kinetic and equilibrium paradigms. J Nucl Med 35: 228-238. PubMed: 8294990.
- Gandelman MS, Baldwin RM, Zoghbi SS, Zea-Ponce Y, Innis RB (1994) Evaluation of ultrafiltration for the free fraction determination of single photon emission computerized tomography (SPECT) radiotracers: β-CIT, IBF and iomazenil. J Pharm Sci 83: 1014-1019. doi:10.1002/jps.2600830718. PubMed: 7965658.
- Reimer ML, Mamer OA, Zavitsanos AP, Siddiqui AW, Dadgar D (1993) Determination of amphetamine, methamphetamine and desmethyldeprenyl in human plasma by gas chromatography/negative ion chemical ionization mass spectrometry. Biol Mass Spectrom 22: 235-242. doi:10.1002/bms.1200220404. PubMed: 8481411.
- Zhang Y, Brady M, Smith S (2001) Segmentation of brain MR images through a hidden Markov random field model and the expectationmaximization algorithm. IEEE Trans Med Imaging 20: 45-57. doi: 10.1109/42.906424. PubMed: 11293691.
- Smith SM, Jenkinson M, Woolrich MW, Beckmann CF, Behrens TE et al. (2004) Advances in functional and structural MR image analysis and implementation as FSL. NeuroImage 23 Suppl 1: S208-S219. doi: 10.1016/j.neuroimage.2004.07.051. PubMed: 15501092.
- Innis RB, Cunningham VJ, Delforge J, Fujita M, Gjedde A et al. (2007) Consensus nomenclature for in vivo imaging of reversibly binding radioligands. J Cereb Blood Flow Metab 27: 1533-1539. doi:10.1038/ sj.jcbfm.9600493. PubMed: 17519979.
- Kirk RE (1982) Experimental design: procedures for the behavioral sciences. Pacific Grove, California: Brooks/Cole Publishing Company.
- 36. Martinez D, Slifstein M, Broft A, Mawlawi O, Hwang DR et al. (2003) Imaging human mesolimbic dopamine transmission with positron emission tomography. Part II: amphetamine-induced dopamine release in the functional subdivisions of the striatum. J Cereb Blood Flow Metab

Off J International Society Of Cerebral Blood Flow Metabolism 23: 285-300.

- 37. Mawlawi O, Martinez D, Slifstein M, Broft A, Chatterjee R et al. (2001) Imaging human mesolimbic dopamine transmission with positron emission tomography: I. Accuracy and precision of D(2) receptor parameter measurements in ventral striatum. J Cereb Blood Flow Metab Off J International Society Of Cerebral Blood Flow Metabolism 21: 1034-1057.
- Mavlawi O, Martinez D, Slifstein M, Broft A, Chatterjee R et al. (2001) Imaging human mesolimbic dopamine transmission with positron emission tomography: I. Accuracy and precision of D2 receptor parameter measurements in ventral striatum. J Cereb Blood Flow Metab 21: 1034-1057. PubMed: 11524609.
- Narendran R, Frankle WG, Mason NS, Laymon CM, Lopresti BJ et al. (2009) PET Imaging of D2/3 agonist binding in healthy human subjects with the radiotracer [11C]-N-propyl-nor-apomorphine (NPA): preliminary evaluation and reproducibility studies. Synapse 63: 574-584. doi: 10.1002/syn.20633. PubMed: 19301416.
- Narendran R, Mason NS, May MA, Chen CM, Kendro S et al. (2011) Positron emission tomography imaging of dopamine D(2)/(3) receptors in the human cortex with [(1) (1)C]FLB 457: reproducibility studies. Synapse 65: 35-40. doi:10.1002/syn.20813. PubMed: 20506186.
- Laruelle M (2000) Imaging synaptic neurotransmission with in vivo binding competition techniques: a critical review. J Cereb Blood Flow Metab Off J International Society Of Cerebral Blood Flow Metabolism 20: 423-451. PubMed: 10724107.
- 42. Narendran R, Mason NS, Laymon C, Lopresti B, Velasquez N et al. (2010) A comparative evaluation of the dopamine D2/3 agonist radiotracer [11C]NPA and antagonist [11C]raclopride to measure amphetamine-induced dopamine release in the human striatum. J Pharmacol Exp Ther 63: 574-584.