

RESEARCH ARTICLE

Open Access



# Suppression of miR-181a attenuates H<sub>2</sub>O<sub>2</sub>-induced death of mesenchymal stem cells by maintaining hexokinase II expression

Seahyoung Lee<sup>1†</sup>, Ina Yun<sup>2†</sup>, Onju Ham<sup>2</sup>, Se-Yeon Lee<sup>2</sup>, Chang Yeon Lee<sup>3</sup>, Jun-Hee Park<sup>3</sup>, Jiyun Lee<sup>2</sup>, Hyang-Hee Seo<sup>2</sup>, Eunhyun Choi<sup>1</sup> and Ki-Chul Hwang<sup>1\*</sup>

## Abstract

**Background:** Low survival rate of transplanted cells compromises the efficacy of cell therapy. Hexokinase II (HKII) is known to have anti-apoptotic activity through its interaction with mitochondria. The objective was to identify miRNAs targeting HKII and investigate whether miRNA-mediated modulation of HKII could improve the survival of mesenchymal stem cells (MSCs) exposed to H<sub>2</sub>O<sub>2</sub>. The expression of HKII in MSCs exposed to H<sub>2</sub>O<sub>2</sub> was evaluated, and HKII-targeting miRNA was screened based on miRNA-target prediction databases. The effect of H<sub>2</sub>O<sub>2</sub> on the expression of the selected HKII-targeting miRNA was examined and the effect of modulation of the selected HKII-targeting miRNA using anti-miRNA on H<sub>2</sub>O<sub>2</sub>-induced apoptosis of MSC was evaluated.

**Results:** H<sub>2</sub>O<sub>2</sub> (600 μM) induced cell death of MSCs and decreased mitochondrial HKII expression. We have identified miR-181a as a HKII-targeting miRNA and H<sub>2</sub>O<sub>2</sub> increased the expression of miR-181a in MSCs. Delivery of anti-miR-181a, which neutralizes endogenous miR-181a, significantly attenuated H<sub>2</sub>O<sub>2</sub>-induced decrease of HKII expression and disruption of mitochondrial membrane potential, improving the survival of MSCs exposed to H<sub>2</sub>O<sub>2</sub>.

**Conclusions:** These findings suggest that H<sub>2</sub>O<sub>2</sub>-induced up-regulation of miR-181a contributes to the cell death of MSCs by down-regulating HKII. Neutralizing miR-181a can be an effective way to prime MSCs for transplantation into ischemic tissues.

**Keywords:** Hexokinase II, miRNA-181a, Cell death, Mesenchymal stem cell

## Background

For many decades, ischemic heart diseases such as myocardial infarction (MI) have been a leading cause of death worldwide [1]. Cell therapy using various stem cells is one of the therapeutic approaches to manage such heart disease [2]. However, high death rate of the transplanted cells has hampered the efficacy of stem cell-based cell therapy [3]. One of the major reasons for such low survival of transplanted cells is that the transplanted cells are also immediately exposed to a harsh microenvironment

(i.e., low tissue oxygen and nutrients, and increased reactive oxygen species (ROS) in case of ischemic heart disease), which can induce various types of cell death, including apoptosis [4]. Consequently, discovery of alternative means to attenuate or prevent the onset of apoptosis of transplanted cells is of utmost importance for developing effective therapeutic strategy.

One of the molecules known to play an important role in preventing apoptosis is hexokinase II (HKII) [5]. HK family is composed of four different isoforms (1 to 4, or I to IV) and they can initiate the glycolysis pathway where glucose is converted to glucose-6-phosphate [6]. Additionally, HKs are known to have non-enzymatic activities such as anti-apoptotic activity [7]. A previous study reported that myocardial ischemia/reperfusion (I/R) injury decreased the cardiac expression of HKII as well

\*Correspondence: kchwang@cku.ac.kr

†Seahyoung Lee and Ina Yun contributed equally to this work

<sup>1</sup>Institute for Bio-Medical Convergence, College of Medicine, Catholic Kwandong University, Gangneung, Gangwon-do 210-701, Republic of Korea

Full list of author information is available at the end of the article

as the binding to mitochondria, and it was associated with decreased cardiac function [8]. Thus, assuming the cell death mechanisms of transplanted cells are similar to those of host cells, maintaining the expression of HKII may improve the survival of transplanted cells. Regarding the regulation of HKII expression, we speculate that microRNAs (miRNA or miR) may be involved. MicroRNAs are approximately 22-nucleotide-long RNAs that attenuate certain gene expression by either inhibiting mRNA translation or degrading mRNAs [9], and studies have demonstrated that miRNAs, such as miR-143 and miR-125a-5p, negatively regulated HKII expression in cancer cells [10–12]. This prompted us to examine possible involvement of HKII-targeting microRNAs (miRNAs) in the death of transplanted cells.

In the present study, we examined the possibility of miRNA-dependent HKII regulation during ROS-induced death of mesenchymal stem cells (MSCs). We identified miR-181a as a HKII-targeting miRNA and demonstrated that its expression increased during ROS-induced apoptosis of MSCs. We further examined the potential of anti-miR-181a as an anti-apoptotic agent for MSCs exposed to ROS in vitro.

## Results

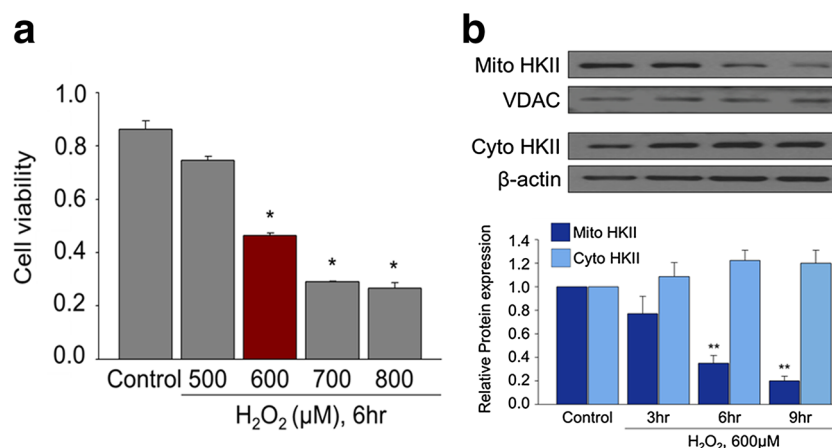
### H<sub>2</sub>O<sub>2</sub> induced cell death of MSCs and decrease of mitochondrial HKII expression

H<sub>2</sub>O<sub>2</sub>, which has been used to induce apoptosis of MSCs [13], was used to simulate oxidative stress after transplantation. H<sub>2</sub>O<sub>2</sub> concentration-dependently decreased cell viability of MSCs, and 600 μM or higher concentrations of H<sub>2</sub>O<sub>2</sub> significantly decreased cell viability of MSCs after 6 h of treatment (Fig. 1a). Since 6 h of

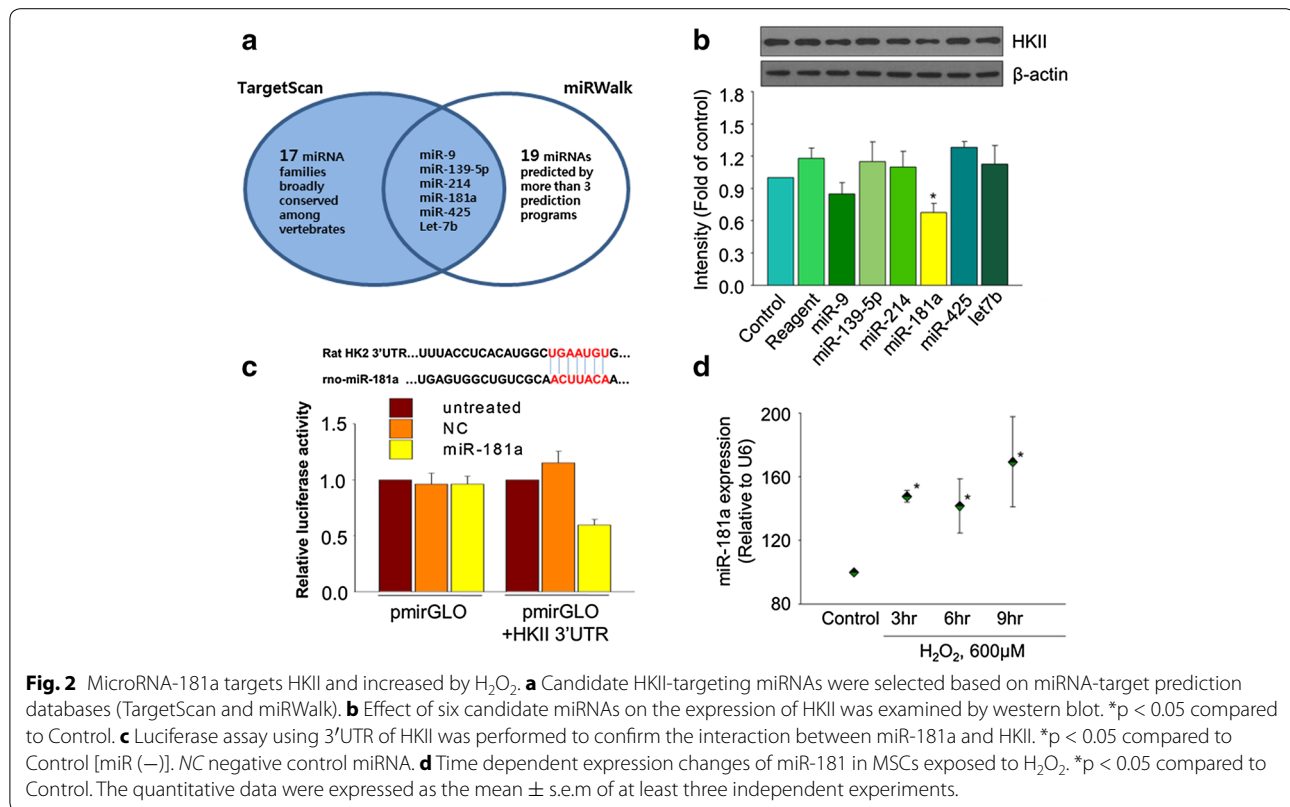
treatment with 600 μM of H<sub>2</sub>O<sub>2</sub> significantly induced cell death of MSCs, this particular condition was used for further experiments. For the expression of HKII, H<sub>2</sub>O<sub>2</sub> time-dependently decreased HKII in mitochondrial fraction, while it had no significant effect on HKII in cytosolic fraction (Fig. 1b), suggesting the decrease of mitochondrial HKII contributed to cell death of MSCs exposed to H<sub>2</sub>O<sub>2</sub>.

### Identification of HKII targeting miRNA and its expression during H<sub>2</sub>O<sub>2</sub>-induced cell death

To screen miRNAs targeting HKII, miRNA-target protein prediction databases (<http://www.targetscan.org> and <http://www.umm.uni-heidelberg.de/apps/zmf/mirwalk/>) were utilized. The TargetScan database predicted 23 miRNAs families broadly conserved among vertebrates. In case of the miRWalk database, it provided comparative analysis by 8 prediction programs and the number of miRNAs predicted by more than 3 prediction programs was 25. Among these miRNAs, 6 miRNAs were shared by the two groups (Fig. 2a). We transfected MSCs with those 6 miRNAs and examined the expression of HKII. Among these 6 miRNAs, miR-181a significantly decreased the expression of HKII in MSCs (Fig. 2b). To examine whether miR-181a directly targets HKII, we also conducted a luciferase assay. The luciferase assay used in the present study is to evaluate the effect of miRNA-dependent post-transcriptional regulation of target genes [14]. In this luciferase assay, 3'UTR of HKII was cloned into a plasmid containing the gene codes for luciferase. Since the 3'UTR of HKII was positioned immediately after the luciferase gene in the constructed plasmids, if miRNA effectively recognize



**Fig. 1** Decrease of mitochondrial HKII during H<sub>2</sub>O<sub>2</sub>-induced MSC cell death. MSCs were exposed to increasing concentrations of H<sub>2</sub>O<sub>2</sub> simulating ROS for 6 h. **a** Cell viability of MSCs exposed to varying concentrations of H<sub>2</sub>O<sub>2</sub>. \*p < 0.05 compared to Control. **b** Time dependent changes of mitochondrial and cytosolic expressions of HKII in MSCs exposed to H<sub>2</sub>O<sub>2</sub>. \*\*p < 0.05 compared to Control. The quantitative data were expressed as the mean ± s.e.m of at least three independent experiments.



and target the 3'UTR of HKII, the translation of luciferase is also affected and the expression of luciferase is suppressed. The result of the luciferase assay indicated that miR-181a transfection decreased the expression of luciferase linked to the 3'UTR of HKII (pmirGLO-HKII 3'UTR), while the luciferase expression of the cells transfected with control luciferase vector lacking the 3'UTR of HKII (pmirGLO) was not affected by miR-181a transfection. This indicated that miR-181a directly targeted the 3'UTR of HKII (Fig. 2c). To examine the possible relationship between oxidative stress and miR-181a, the expression of miR-181a in MSCs treated with H<sub>2</sub>O<sub>2</sub> was evaluated. According to the real time PCR data, H<sub>2</sub>O<sub>2</sub> significantly up-regulated the expression of miR-181a in a time-dependent manner (Fig. 2d).

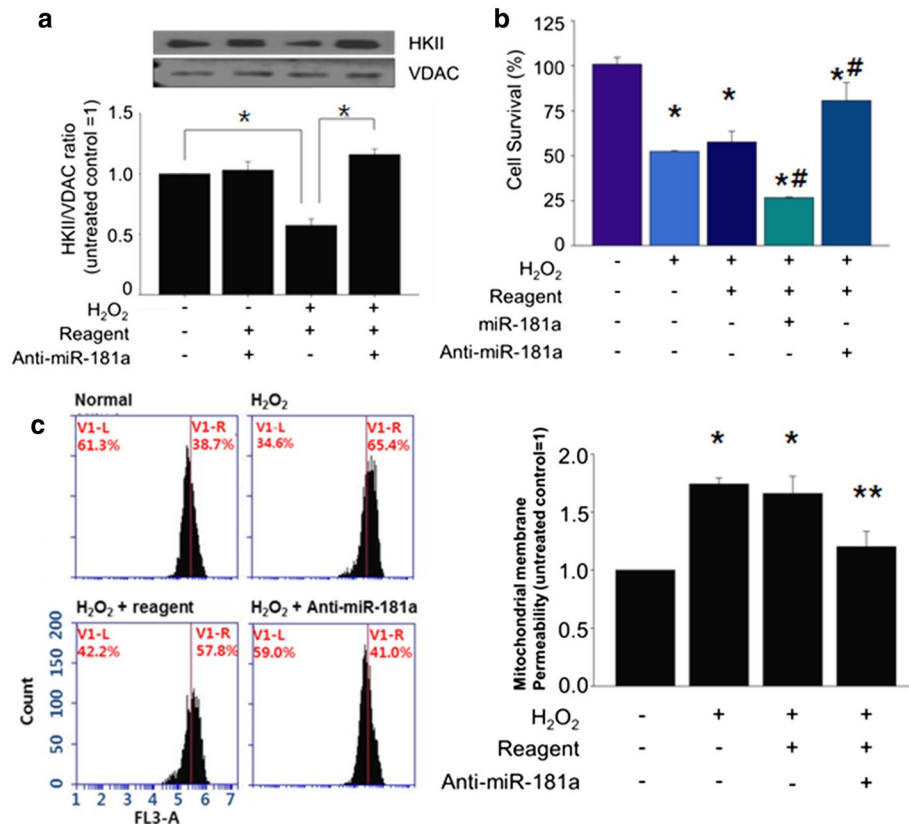
#### Neutralization of miR-181a attenuates cell death of MSCs induced by H<sub>2</sub>O<sub>2</sub>

Since we speculate that H<sub>2</sub>O<sub>2</sub>-induced up-regulation of miR-181a contributed to the down-regulation of mitochondrial HKII expression, we neutralized endogenous miR-181a using anti-miR-181a and examined its effect on mitochondrial HKII expression. Without H<sub>2</sub>O<sub>2</sub> treatment, anti-miR-1861a transfection had no significant effect on mitochondrial HKII expression. However, with H<sub>2</sub>O<sub>2</sub> treatment, anti-miR-181a attenuated mitochondrial

HKII expression compared to H<sub>2</sub>O<sub>2</sub> only treated group (Fig. 3a). This suggested that miR-181a-dependent regulation of HKII may be negligible under normal (without H<sub>2</sub>O<sub>2</sub>) condition. For the effect of anti-miR-181a on the survival of MSCs exposed to H<sub>2</sub>O<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>-induced decrease of MSC survival was further decreased by miR-181a treatment, while anti-miR-181a treatment reversed such decrease (Fig. 3b), indicating suppression of H<sub>2</sub>O<sub>2</sub>-induced increase of miR-181a may be an effective way to enhance cell survival. Furthermore, H<sub>2</sub>O<sub>2</sub> induced significant accumulation of tetramethylrhodamine methyl esters (TMRM) in the inner membrane of mitochondria indicating loss of mitochondrial membrane potential. However, such increase of TMRM accumulation was attenuated by anti-miR-181a transfection prior to H<sub>2</sub>O<sub>2</sub> treatment suggesting that anti-miR-181a suppressed H<sub>2</sub>O<sub>2</sub>-induced mitochondrial membrane potential loss (Fig. 3c).

#### Discussion

One of the major problems of cell therapy is low survival rate of transplanted cells. Thus developing an effective strategy to enhance the survival of the transplanted cells is of importance. Here, we report that miRNA-dependent modulation of HKII improves the survival of MSCs exposed to ROS.



**Fig. 3** Anti-miR-181a attenuates HKII expression and H<sub>2</sub>O<sub>2</sub>-induced cell death of MSCs. **a** Effect of anti-miR-181a on HKII expression was examined by western blot. \*p < 0.05. **b** Effect of anti-miR-181a on the survival of MSCs exposed to H<sub>2</sub>O<sub>2</sub> was evaluated. \*p < 0.05 compared to untreated control. #p < 0.05 compared to H<sub>2</sub>O<sub>2</sub> only treated group. **c** Effect of anti-miR-181a on H<sub>2</sub>O<sub>2</sub>-induced mitochondrial membrane potential disturbance was examined by using cytometry with TMRM. \*p < 0.05 compared to untreated control. \*\*p < 0.05 compared to H<sub>2</sub>O<sub>2</sub> only treated group. The quantitative data were expressed as the mean ± s.e.m of at least three independent experiments.

The anti-apoptotic activity of HKII is based on their interaction with mitochondria [15, 16]. HKII, having the hydrophobic mitochondrial binding domain in their N-terminus [16, 17], can bind to mitochondria in association with the voltage-dependent anion channel (VDAC) [18], constituting the mitochondria permeability transition pore (PT-pore) [19]. This mitochondrial localization of HKII maintains mitochondrial integrity and prevent bcl-2 homologous antagonist killer (BAK) and bcl-2 associated protein X (BAX) mediated mitochondrial cytochrome c release and subsequent apoptosis [20, 21]. Our data also suggested that ROS-induced cell death via dissociation of HKII from mitochondria in our experimental settings.

H<sub>2</sub>O<sub>2</sub>-induced up-regulation of miR-181a has been also reported in rat cardiomyocytes [22]. That particular study clearly demonstrated that H<sub>2</sub>O<sub>2</sub>-induced up-regulation of miR-181a significantly contributed to apoptosis of cardiomyocytes. Thus, our premise that the cell death mechanisms of transplanted cells may be similar to those

of host cells was supported. Although the above mentioned study did not examined the relationship between the up-regulation of miR-181a and HKII expression, it is possible that miR-181a-mediated down-regulation of HKII also contributed to the observed cell death of cardiomyocytes in that previous study.

To date, only few miRNAs that regulate HKII have been identified in mostly cancer cells. For example, miR-143 has been reported to down-regulate HKII in colon cancer cells, reducing cell proliferation [11]. Other studies also have demonstrated that miR-143 significantly increased the number of apoptotic cells compared to control in cancer cells [10, 12], suggesting down-regulation of HKII significantly contributes to progression of apoptosis. On the other hand, miR-143 repressive miR-155 led to up-regulation of HKII and promoted breast cancer cell proliferation [23]. These observations together suggest that miRNA-mediated regulation of HKII does occur, and modulation of HKII-targeting miRNAs to control cell survival is a feasible therapeutic approach.

The approach used in the present study was not intended to increase the mitochondrial binding efficacy of HKII, but to attenuate H<sub>2</sub>O<sub>2</sub>-induced increase of miR-181a and subsequent decrease of HKII expression. Although our data indicate that this approach works in vitro, it should be further validated in vivo as well with optimized delivery method of anti-miR-181a. Although MSCs were primed by anti-miR-181a treatment before exposed to H<sub>2</sub>O<sub>2</sub> in this study, simultaneous delivery of primed MSCs and anti-miR-181a to damaged heart can be therapeutically more effective because anti-miR-181a delivery to damaged heart may enhance the survival of host cardiomyocytes on the verge of activation of apoptotic signaling as well. Another limitation of this study is lacking information about the effect of miR-181a modulation on the differentiation potential of MSCs. Since this study was focused on testing the feasibility of enhancing MSC survival by modulating key miRNA and its target, we have not examined the effect of anti-miR-181a on the differentiation of MSCs. Since previous studies have indicated that miR-181a accelerated endothelial cell [24] and adipocyte [25] differentiation of embryonic stem cells and pre-adipocyte, respectively, it can be speculated that suppressing miR-181a may alter the differentiation potential of MSCs as well. Nevertheless, it should be empirically proved, and examining such possibility in further studies will help us to develop improved regenerative therapeutic strategies involving MSC-based cell therapy.

## Conclusions

In summary, we demonstrate that anti-miR-181a attenuates cell death of MSCs exposed to H<sub>2</sub>O<sub>2</sub> through suppression of miR-181a and subsequent down-regulation of HKII. With further optimization of delivery strategy and in vivo validation, anti-miR-181a can be an effective therapeutic agent for improving the survival of transplanted cells in cell therapy, and the result of this study warrants further studies to investigate more detailed underlying mechanisms.

## Methods

### Animals

Four-week-old Sprague–Dawley rats were used for MSC isolation. The rats were anesthetized with zoletil (20 mg/kg) and xylazine (5 mg/kg). All animal experimental procedures were approved by the Institutional Animal Care and Use Committee (IACUC) of Yonsei University College of Medicine and performed in accordance with the Guidelines and Regulations for Animal Care.

### Isolation and culture of rat MSCs

Bone marrow-derived MSCs were isolated and collected from aspirates of rat femurs and tibiae with 10 mL of

MSC medium consisting of Dulbecco's modified Eagle's medium (DMEM)-low glucose supplemented with 10 % fetal bovine serum (FBS) (Invitrogen) and 1 % antibiotic-penicillin and streptomycin. Mononuclear cells recovered from the interface of Ficoll-Paque PLUS (GE healthcare)-separated bone marrow were washed twice and resuspended in DMEM with 10 % FBS, then plated at a density of  $1 \times 10^6$  cells/100 mm dish. The cultures were maintained at 37 °C in a humidified atmosphere containing 5 % CO<sub>2</sub>. After 72 h, the non-adherent cells were discarded, and the adherent cells were thoroughly washed twice with phosphate-buffered saline (PBS). Fresh MesenPRO RS™ Medium (Invitrogen) was added and then replaced every 3 days for approximately 10 days to achieve stable multi-potentiality.

### H<sub>2</sub>O<sub>2</sub> treatment

The cells were cultured for 24 h before the experiments. For inducing apoptosis, H<sub>2</sub>O<sub>2</sub> was added to the culture media at a final concentration of 500–800 μM for 6 h.

### Cell viability assay

To measure cell viability, CCK reagent (cell counting kit-8, Dojindo) was added to each well for a final concentration of 0.5 mg/mL and the cells were incubated at 37 °C for 2 h. The absorbance of the medium was measured at 450 nm using a microplate reader.

### Transfection of miRNA and anti-miRNA

Transfections of miRNA mimics and anti-miRNAs were performed using siLentFect™ Lipid reagent (Life Science Research). Mature specific miRNAs (Genolution Pharmaceuticals, Inc., Korea) were used at a final concentration of 100 nM. Anti-miRNAs was used at a final concentration of 50 nM. After 4 h of incubation in a CO<sub>2</sub> incubator at 37 °C, the medium was changed to 10 % FBS containing DMEM.

### Western blot

Cells were washed once in PBS and lysed in lysis buffer (Cell Signaling Technology) with protease and phosphatase inhibitor cocktail. Protein concentrations were determined using the BCA protein assay kit (Thermo Science). Proteins were separated in a SDS–polyacrylamide gel and transferred to the PVDF membrane (Millipore). After blocking the membrane with 0.1 % Tris-buffered saline–Tween 20 (TBS-T, 0.1 % Tween 20) containing 10 % skim milk for 1 h at room temperature, the membrane was washed twice with TBS and incubated with primary antibodies for overnight at 4 °C. The membrane was washed three times with 0.1 %TBS-T for 5 min and then incubated for 1 h at room temperature with horseradish peroxidase (HRP)-conjugated secondary

antibodies. After extensive washing, the bands were detected by enhanced chemiluminescence reagent (ECL, Santa Cruz Biotechnology). The band intensities were quantified using NIH Image J version 1.34e software. The primary antibodies for HKII and  $\beta$ -actin were from Cell Signaling (28675) and Sigma (A1978), respectively.

#### Luciferase assay using the 3'UTR of HKII

The 3'UTR sequences of HKII was amplified using primers with XbaI (forward) and EcoRI (reverse) endonuclease sites. The 3'UTR fragment was then cloned into the pmirGLO vector. HeLa cells were plated at a density of  $1 \times 10^5$  cell/well in a 12 well plate, and then transfected with either pmirGLO control vector or pmirGLO vector with HKII 3'UTR using Lipofectamine 2000. After 48 h, relative luciferase activity was measured by using Dual Luciferase assay kit (Promega) according to the manufacturer's instructions. The Renilla luciferase was used for normalization.

#### Real-time PCR

Purified total RNA (100 ng) was used for reverse transcription (TaqMan<sup>®</sup> MicroRNA Reverse Transcriptase Kit, Applied Biosystems) in combination with TaqMan<sup>®</sup> MicroRNA assays to quantify the levels of each specific miRNA according to the manufacturer's instructions. The amplification and detection of specific products were performed using a TaqMan Small RNA Assay kit (AB Applied Biosystems) at 95 °C for 10 min, followed by 40 cycles of 95 °C for 15 s and 60 °C for 60 s. The threshold cycle (Ct) of each target gene was defined automatically; the Ct value was located in the linear amplification phase of the PCR and was normalized to the Ct value of the control transcript U6 ( $\Delta$ Ct value). The relative expression level of each miRNA was calculated using the  $\Delta\Delta$ Ct method and is reported as the fold-change in expression ( $2^{-\Delta\Delta$ Ct).

#### Tetramethylrhodamine methyl ester (TMRM) staining

Mitochondrial membrane potential was determined by using tetramethylrhodamine methyl esters (TMRM) fluorescent dye (Invitrogen) [26]. Loss of mitochondrial membrane potential triggers the accumulation of cationic dye (TMRM) in the mitochondria. To evaluate mitochondrial membrane permeability, the cells were loaded with 200 nM of TMRM for 30 min at 37 °C in medium. Then, the cell pellets were washed with pre-warmed with PBS and re-suspended with PBS to analyze. The data were performed by BD ACCURI C6 cytometer (BD Biosciences) and analyzed with BD Accuri C6 Software.

#### Statistical analysis

Quantitative data were expressed as the mean  $\pm$  S.E.M (standard error of measurement) of at least 3 independent

experiments. For statistical analysis, one-way ANOVA with Bonferroni correction was performed using the OriginPro 8 SR4 software (ver. 8.0951, OriginLab Corporation, Northampton, MA, USA). A *p* value of less than 0.05 was considered to be statistically significant.

#### Authors' contributions

SL participated in the study design and drafted the manuscript. IY conceived the study and carried out cell culture including cell preparation. OH, SYL, CL participated in cell preparation and cell culture. JP, JL, and HS carried out miRNA-related testing. EC provided technical support. KH conceived the study, participated in the study design. All authors read and approved the final manuscript.

#### Author details

<sup>1</sup> Institute for Bio-Medical Convergence, College of Medicine, Catholic Kwangdong University, Gangneung, Gangwon-do 210-701, Republic of Korea. <sup>2</sup> Brain Korea 21 PLUS Project for Medical Science, Yonsei University, Seoul 120-752, Republic of Korea. <sup>3</sup> Department of Integrated Omics for Biomedical Sciences, Yonsei University, Seoul 120-752, Republic of Korea.

#### Acknowledgements

This study was supported by a Korea Science and Engineering Foundation grant funded by the Korean government (MEST) (NRF-2011-0019243, NRF-2011-0019254, NRF-2015M3A9E6029519, and NRF-2015M3A9E6029407) and a Grant from the Korea Health 21 R&D Project, Ministry of Health and Welfare, Republic of Korea (A120478).

#### Compliance with ethical guidelines

#### Competing interests

The authors declare that they have no competing interests.

Received: 25 March 2015 Accepted: 5 August 2015

Published online: 18 August 2015

#### References

- Mathers C, Fat DM, Boerma JT, World Health Organization. The global burden of disease: 2004 update. Geneva: World Health Organization; 2008.
- Garbern JC, Lee RT. Cardiac stem cell therapy and the promise of heart regeneration. *Cell Stem Cell*. 2013;12(6):689–98.
- Herberts CA, Kwa MS, Hermens HP. Risk factors in the development of stem cell therapy. *J Transl Med*. 2011;9:29.
- Mughal W, Kirshenbaum LA. Cell death signalling mechanisms in heart failure. *Exp Clin Cardiol*. 2011;16(4):102–8.
- Miyamoto S, Murphy AN, Brown JH. Akt mediates mitochondrial protection in cardiomyocytes through phosphorylation of mitochondrial hexokinase-II. *Cell Death Differ*. 2008;15(3):521–9.
- Wolf A, Agnihotri S, Micallef J, Mukherjee J, Sabha N, Cairns R, et al. Hexokinase 2 is a key mediator of aerobic glycolysis and promotes tumor growth in human glioblastoma multiforme. *J Exp Med*. 2011;208(2):313–26.
- Dai W, Wang F, Lu J, Xia Y, He L, Chen K et al. By reducing hexokinase 2, resveratrol induces apoptosis in HCC cells addicted to aerobic glycolysis and inhibits tumor growth in mice. *Oncotarget*. 2015;6(15):13703–17
- Wu R, Smeele KM, Wyatt E, Ichikawa Y, Eerbeek O, Sun L, et al. Reduction in hexokinase II levels results in decreased cardiac function and altered remodeling after ischemia/reperfusion injury. *Circ Res*. 2011;108(1):60–9.
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*. 2007;131(4):11–29.
- Jiang S, Zhang LF, Zhang HW, Hu S, Lu MH, Liang S, et al. A novel miR-155/miR-143 cascade controls glycolysis by regulating hexokinase 2 in breast cancer cells. *EMBO J*. 2012;31(8):1985–98.
- Gregersen LH, Jacobsen A, Frankel LB, Wen JY, Krogh A, Lund AH. MicroRNA-143 down-regulates Hexokinase 2 in colon cancer cells. *BMC Cancer*. 2012;12:232–41.

12. Fang R, Xiao T, Fang ZY, Sun YH, Li F, Gao YJ, et al. MicroRNA-143 (miR-143) Regulates cancer glycolysis via targeting hexokinase 2 gene. *J Biol Chem*. 2012;287(27):23227–35.
13. Wei H, Li Z, Hu S, Chen X, Cong X. Apoptosis of mesenchymal stem cells induced by hydrogen peroxide concerns both endoplasmic reticulum stress and mitochondrial death pathway through regulation of caspases, p38 and JNK. *J Cell Biochem*. 2010;111(4):967–78.
14. Jin Y, Chen Z, Liu X, Zhou X. Evaluating the microRNA targeting sites by luciferase reporter gene assay. *Methods Mol Biol*. 2013;936:117–27.
15. Majewski N, Nogueira V, Bhaskar P, Coy PE, Skeen JE, Gottlob K, et al. Hexokinase-mitochondria interaction mediated by Akt is required to inhibit apoptosis in the presence or absence of Bax and Bak. *Mol Cell*. 2004;16(5):819–30.
16. Sun L, Shukair S, Naik TJ, Moazed F, Ardehali H. Glucose phosphorylation and mitochondrial binding are required for the protective effects of hexokinases I and II. *Mol Cell Biol*. 2008;28(3):1007–17.
17. Sui DX, Wilson JE. Structural determinants for the intracellular localization of the isozymes of mammalian hexokinase: intracellular localization of fusion constructs incorporating structural elements from the hexokinase isozymes and the green fluorescent protein. *Arch Biochem Biophys*. 1997;345(1):111–25.
18. McCommis KS, Baines CP. The role of VDAC in cell death: friend or foe? *Biochim Biophys Acta*. 2012;1818(6):1444–50.
19. Zheng Y, Shi Y, Tian C, Jiang C, Jin H, Chen J, et al. Essential role of the voltage-dependent anion channel (VDAC) in mitochondrial permeability transition pore opening and cytochrome c release induced by arsenic trioxide. *Oncogene*. 2004;23(6):1239–47.
20. Gottlob K, Majewski N, Kennedy S, Kandel E, Robey RB, Hay N. Inhibition of early apoptotic events by Akt/PKB is dependent on the first committed step of glycolysis and mitochondrial hexokinase. *Gene Dev*. 2001;15(11):1406–18.
21. Majewski N, Nogueira V, Robey RB, Hay N. Akt inhibits apoptosis downstream of BID cleavage via a glucose-dependent mechanism involving mitochondrial hexokinases. *Mol Cell Biol*. 2004;24(2):730–40.
22. Wang L, Huang H, Fan Y, Kong B, Hu H, Hu K, et al. Effects of down-regulation of microRNA-181a on H<sub>2</sub>O<sub>2</sub>-induced H9c2 cell apoptosis via the mitochondrial apoptotic pathway. *Oxid Med Cell Longev*. 2014;2014:960362.
23. Higgs G, Slack F. The multiple roles of microRNA-155 in oncogenesis. *J Clin Bioinform*. 2013;3(1):17.
24. Kane NM, Howard L, Descamps B, Meloni M, McClure J, Lu R, et al. Role of microRNAs 99b, 181a, and 181b in the differentiation of human embryonic stem cells to vascular endothelial cells. *Stem Cells*. 2012;30(4):643–54.
25. Li H, Chen X, Guan L, Qi Q, Shu G, Jiang Q, et al. MiRNA-181a regulates adipogenesis by targeting tumor necrosis factor-alpha (TNF-alpha) in the porcine model. *PLoS One*. 2013;8(10):e71568.
26. Scaduto RC Jr, Grotyohann LW. Measurement of mitochondrial membrane potential using fluorescent rhodamine derivatives. *Biophys J*. 1999;76(1 Pt 1):469–77.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

