

Meta-fibrosis links positive energy balance and mitochondrial metabolism to insulin resistance [version 1; referees: 3 approved]

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V1 First published: 27 Sep 2017, 6(F1000 Faculty Rev):1758 (doi: 10.12688/f1000research.11653.1)

Latest published: 27 Sep 2017, 6(F1000 Faculty Rev):1758 (doi: 10.12688/f1000research.11653.1)

Abstract

Obesity and insulin resistance often emerge from positive energy balance and generally are linked to low-grade inflammation. This low-grade inflammation has been called "meta-inflammation" because it is a consequence of the metabolic dysregulation that can accompany overnutrition. One means by which meta-inflammation is linked to insulin resistance is extracellular matrix expansion secondary to meta-inflammation, which we define here as "meta-fibrosis". The significance of meta-fibrosis is that it reflects a situation in which the extracellular matrix functions as a multi-level integrator of local (for example, mitochondrial reactive oxygen species production) and systemic (for example, inflammation) inputs that couple to cellular processes creating insulin resistance. While adipose tissue extracellular matrix remodeling has received considerable attention, it is becoming increasingly apparent that liver and skeletal muscle extracellular matrix remodeling also contributes to insulin resistance. In this review, we address recent advances in our understanding of energy balance, mitochondrial energetics, meta-inflammation, and meta-fibrosis in the development of insulin resistance.



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Competing interests: The authors declare that they have no competing interests.

How to cite this article: Lark DS and Wasserman DH. Meta-fibrosis links positive energy balance and mitochondrial metabolism to insulin resistance [version 1; referees: 3 approved] *F1000Research* 2017, 6(F1000 Faculty Rev):1758 (doi: 10.12688/f1000research.11653.1)

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Grant information: The author(s) declared that this work was funded by National Institutes of Health (grants DK054902, DK050277, DK059637), NIDDK Mouse Metabolic Phenotyping Centers MICROMouse Program (grant 15GRU2558) and American Heart Association (grant 16POST29910001).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

First published: 27 Sep 2017, 6(F1000 Faculty Rev):1758 (doi: 10.12688/f1000research.11653.1)

Introduction

Advances in industrial and agricultural technology combined with lower rates of energy expenditure through physical activity have had the unintended consequence of creating a dramatic rise in the prevalence of obesity, insulin resistance (IR), hypertension, and dyslipidemia. These comorbidities are principal components of the metabolic syndrome as well as risk factors for type 2 diabetes mellitus and cardiovascular disease. The public health impact of these altered metabolic states is clear when considering that, in 2012, approximately 33% of United States citizens (over 100 million people) were projected to have at least one component of the metabolic syndrome¹.

Positive energy balance at the whole-body level and altered oxidative metabolism at the cellular level are central to the development of IR. However, the conduit linking nutrient status and cellular energetics to pathophysiological states like IR is incompletely defined. In this commentary, we provide a framework for how mitochondrial energetics along with metabolically driven inflammation (meta-inflammation) and extracellular matrix (ECM) remodeling leading to fibrosis (meta-fibrosis) link overnutrition to IR (Figure 1). As several recent discoveries suggest, there is a



Figure 1. Positive energy balance promotes insulin resistance via metabolism-driven inflammation and fibrosis. Energy balance is defined as the difference between absorbed dietary macronutrients (Supply) and energy expenditure (Demand). Energy supply is determined by the quantity and composition of macronutrients consumed, whereas energy demand is determined by exercise, non-exercise activity thermogenesis, and resting metabolic rate. A net positive energy balance (Supply > Demand) leads to obesity and a cascade of events that includes mitochondrial carbon stress (that is, an oversupply of macronutrients to mitochondria). This metabolic stress on mitochondria can promote meta-inflammation and meta-fibrosis that ultimately contribute to cellular and systemic insulin resistance.

great deal to be learned regarding the etiology of IR by studying organ-level physiological events in the context of the extracellular milieu. The focus here will be on metabolism, molecular organization, and cell signaling in the pathogenesis of IR. The important roles of gene transcription and epigenetics in the development of IR are beyond the scope of this commentary. Readers are directed to recent reviews on these topics^{2,3}.

Energy balance and the metabolic syndrome

Energy balance is defined as the gastrointestinal absorption of dietary macronutrients minus whole-body energy expenditure. Human evolution has selected for traits that facilitate the efficient mobilization, metabolism, and storage of macronutrients. The biological significance of these adaptations lies in the need to store nutrients during times of nutrient excess and the ability to mobilize fuel in situations of nutrient deficiency. Nutrient storage is important for acute bouts of elevated energy expenditure or prolonged periods during which food is not readily available. Indeed, mechanisms for storing excess glucose (glycogen), lipids (triglyceride), and amino acids (protein) obtained from the diet are exquisitely sensitive. While these adaptations have been critical for survival and species propagation, people living in industrialized societies now have easy access to high-calorie foods and do not need to expend considerable energy to obtain their food. This has led to a sustained positive energy balance. Since this is a situation rarely encountered during the course of human evolution, the body is poorly equipped to adapt to dietary excess. As such, the chronic energy surplus incurred by overnutrition and sedentary behavior has become a persistent metabolic burden that leads to adipose tissue expansion and obesity in many individuals⁴. Obesity, in turn, is central to the development of IR.

The evolutionarily conserved mechanisms that make survival possible during periods of famine also make humans refractory to weight loss. Resistance to weight loss and weight maintenance is recognized as a primary barrier to improving metabolic health⁵. This is most clearly demonstrated when considering the effects of caloric restriction and physical activity on energy balance and body weight. In both obese and non-obese humans⁶⁻⁸, prolonged caloric restriction results in significant weight loss, but it is accompanied by reductions in resting metabolic rate (RMR) beyond that which can be accounted for by weight loss alone. Since RMR is a primary contributor to the daily energy budget⁹, this represents a significant barrier to long-term weight loss. It is notable that exercise alone is only marginally effective as a therapy for weight loss^{10–12}. This is likely due to both metabolic and behavioral obstacles. Exercise training fails to increase RMR in obese individuals with diabetes¹³, and this is potentially due to increased metabolic efficiency¹⁴. Exercise training has had mixed results in eliciting weight loss in both rodents¹⁵ and humans¹⁰ and is explained in part by increased food intake. In addition to RMR, "non-exercise activity thermogenesis" (NEAT) is a major contributor to energy expenditure in mice and humans¹⁶. Mice given access to a running wheel increase their physical activity and energy expenditure over a four-week period, but the metabolic cost of activity progressively decreases concurrently with decreased NEAT¹⁷. This is significant because fat gain with overnutrition in humans is positively correlated with an increase in NEAT¹⁸. Whether the bidirectional modulation of NEAT

based on whole-body energy balance is modifiable therapeutically remains to be seen but may be a viable strategy for combating obesity. Complicating therapeutic strategies further is a growing body of literature demonstrating that the metabolic adaptations that occur with weight loss predispose an individual to accelerated weight regain and increased adiposity upon cessation of a supervised diet or exercise regimen or both¹⁹. Notably, recent work suggests that glucocorticoid antagonism mitigates the weight regain and IR that occur following cessation of voluntary exercise in rats²⁰. A better understanding of how humans resist weight loss, even in the setting of obesity, is critically important in that it may reveal novel therapeutic strategies for treating obesity and IR.

Mitochondrial energetics and the pathogenesis of insulin resistance

As the demand-driven terminus of oxidative metabolism, mitochondria are intricately involved in the maintenance of energy balance, and several recent reviews have highlighted the importance of mitochondrial energetics to the etiology of IR²¹⁻²³. At the level of the mitochondrion, energy balance is established by a dynamic rate of carbon flux through the tricarboxylic acid (TCA) cycle that supports ATP production via oxidative phosphorylation. In the setting of overnutrition, there is a supply/demand mismatch that results in excess anaplerotic flux of carbon from fatty acids entering the TCA cycle relative to the ATP demand leading to IR²⁴. Excessive anaplerotic flux creates a mitochondrial "carbon stress" that has been well documented in both skeletal muscle (SkM) and liver. This carbon stress promotes IR through incompletely defined mechanisms that likely involve post-translational protein modifications that alter insulin signaling or protein trafficking (that is, GLUT4 translocation). The teleological explanation for limiting SkM glucose uptake in the face of excess dietary lipids may be that SkM is unable to efficiently convert excess intracellular glucose to an inert metabolite (that is, fatty acids).

In the liver, greater fatty acid availability accelerates anaplerotic flux contributing to IR that correlates with the severity of nonalcoholic fatty liver disease (NAFLD) in humans²⁵. This appears to be linked, at least in part, to incomplete β -oxidation in the setting of overnutrition²⁶. This hypothesis is supported by findings that acyl-carnitine, the carbon chain intermediate of β -oxidation, is increased in human plasma²⁷ as well as rodent SkM²⁶. Free carnitine in SkM is also reduced in the setting of obesity or high-fat feeding or both²⁸, suggesting a reduced capacity to handle excess fatty acids. Collectively, excess dietary fatty acids entering metabolically active tissues overload the mitochondria, leading to IR. A teleological explanation for why mitochondria induce IR may be to mitigate oxidative damage induced by overnutrition²⁹.

Mitochondria can also engage in cataplerosis, which is removal of carbons from the TCA cycle. In SkM, one proposed role for cataplerosis is as a buffering system to avoid mitochondrial carbon excess that can lead to increased reactive oxygen species (ROS) production during overnutrition²⁴. SkM cataplerosis occurs in large part via carnitine acetyltransferase (CrAT), an enzyme that is responsible for exporting acetyl and acyl groups bound to carnitine from the mitochondrial matrix into the cytosol. Mice with SkM-specific deletion of CrAT have impaired glucose

tolerance and increased oxidative stress³⁰, illustrating a need for mitochondrial carbon efflux (that is, cataplerosis) to preserve SkM metabolic homeostasis in the setting of overnutrition. In the liver, cataplerosis is essential for the production of both glucose (gluconeogenesis) and ketones (ketogenesis). Predominantly expressed in gluconeogenic organs (liver and kidney), phosphoenolpyruvate carboxykinase (PEPCK) converts oxaloacetate to pyruvate and is a key enzyme for gluconeogenesis. Loss of PEPCK in mice reduces hyperglycemia in leptin receptordeficient (db/db) diabetic mice³¹. Similarly, ketogenesis exerts partial protection against high-fat diet (60% calories from fat)induced hyperglycemia and fatty liver, primary complications linked to obesity and overnutrition³². Notably, however, mice fed a ketogenic diet (more than 90% calories from fat) are lean and hypoinsulinemic but also display fatty liver^{33,34}. This may be due to the impaired liver mitochondrial respiratory capacity observed in mice fed a short-term (14 days) ketogenic diet³⁵. Strategies to increase cataplerosis in a tissue- and product-specific fashion could yield valuable strategies for preserving glucose homeostasis and insulin sensitivity but should be considered in the context of also preventing the development of fatty liver.

How does mitochondrial carbon excess promote IR? Carbon turnover that exceeds metabolic demand leads to accumulation of reducing equivalents (NADH and FADH,) that exert greater "reducing pressure" (that is, more electrons) on the electron transport system³⁶. This buildup of reducing equivalents in the matrix and electrons within the electron transport system promotes the formation of ROS that modulate a wide variety of normal and pathophysiological cellular processes³⁷. For example, acute or chronic high-fat feeding increases mitochondrial ROS production that has been shown in some^{29,38–40}, but not all⁴¹, reports to be causal for the development of IR. Notably, fatty acids can also "uncouple" oxidative phosphorylation⁴², raising the possibility that mitochondrial oxidative efficiency may be an additional mechanism to manage carbon excess in obesity. Targeting this mechanism may be feasible in light of recent work demonstrating that mitochondrial oxidative efficiency is a dynamic process that is acutely sensitive to energetic demand⁴³. Historically, the use of mitochondrial uncouplers as therapeutic agents has been met with skepticism following a string of deaths linked to the protonophore 2-dinitrophenol in the 1930s. However, recent efforts have provided new lead compounds that may be promising in the treatment of obesity⁴⁴⁻⁴⁶. While mitochondria-targeted therapies are being studied intensively and hold great promise, an alternative approach may be to address downstream effectors of mitochondrial oxidants. The downstream processes affected by mitochondrial oxidants are incompletely defined but include inflammation and expansion of the ECM. The remainder of this article will be spent discussing these processes in the context of their individual, and collective, contributions to the etiology of IR.

Inflammation and extracellular matrix expansion in the etiology of insulin resistance

Low-grade metabolically driven "meta-inflammation"⁴⁷ contributes to IR in obesity⁴⁸. There are numerous intersecting mechanisms linking inflammation and ROS⁴⁹, including a critical role for the innate immune system that is coupled to macrophage infiltration^{50,51}. Macrophages recruited with chronic overnutrition are pro-inflammatory (M1; CD11b⁺) and secrete tumor necrosis factor alpha (TNF α) that has been shown to contribute to IR in adipose, SkM, and liver⁵²⁻⁵⁴. M1 macrophages also play a critical role in wound healing. It has been observed that the metainflammatory response to obesity that includes M1 macrophage infiltration is responsible for the accumulation of ECM proteins in insulin-sensitive tissues⁵⁵. The evidence linking these processes in adipose, SkM, and liver is outlined below.

Adipose tissue function is reliant upon, and in certain situations compromised by, the ECM surrounding adipocytes⁵⁶. Healthy adipose tissue expansion involves a balance between enzymatic degradation and subsequent synthesis of ECM proteins⁵⁷. Pathogenic obesity in humans is characterized by adipose tissue fibrosis due to excessive ECM deposition and reduced ECM degradation that is associated with IR⁵⁸⁻⁶¹. Paradoxically, recent work by Muir et al.62 showed that diabetics have reduced adipose tissue fibrosis and greater adipose tissue hypertrophy. Genetically obese (ob/ob) mice have increased expression of genes encoding collagens⁶³ that is exacerbated by high-fat feeding⁶⁴. Genetic loss of the adipose tissue-abundant collagen VI in mice mitigates adipocyte inflammation, diet-induced obesity (DIO), and glucose intolerance while permitting greater adipocyte hypertrophy63. Beyond collagen, various other ECM componentsincluding osteopontin^{65,66}, hyaluronan⁶⁷, thrombospondins^{68,69}, and microfibril-associated glycoprotein 1 (MAGP1)⁷⁰—accumulate in adipose tissue with obesity and contribute to IR. Adipose tissue ECM expansion is attenuated by the anti-diabetic drug metformin⁷¹, a drug that is also known to reduce mitochondrial ROS production³⁹. Whether metformin improves metabolic health by mitigating mitochondrial ROS production or ECM accumulation or both remains to be addressed directly.

Obesity induces SkM ECM expansion^{55,72,73} that would be expected to increase the resistance to glucose delivery, an essential controller of glucose uptake⁷⁴. Even short-term (28 days) high-fat feeding⁷⁵ is sufficient to induce SkM ECM expansion. This appears to be reversible as SkM collagen accumulation is ameliorated in obese mice following exercise training⁷³ and preventable in mice with genetic enhancement of SkM mitochondrial ROS scavenging⁵⁵. A genetic knockout of matrix metalloprotease-9 (MMP-9), a key ECM-degrading enzyme, in obese mice causes increased collagen and a further deterioration of SkM insulin action⁷⁶. Treatment with pegylated hyaluronidase causes degradation of hyaluronan and rescues IR in obese mice⁷⁷. These studies demonstrate a direct link between ECM accumulation and insulin action in SkM.

In the setting of obesity, circulating lipids are incompletely sequestered in adipose tissue and consequently accumulate in SkM and liver and lead to IR. NAFLD is a primary risk factor for the development of IR and diabetes via liver fibrosis^{78,79}. Mice fed a high-fat high-fructose diet exhibit liver fibrosis that accompanies lipid accumulation and IR^{80,81}. The extent and scope to which overnutrition alters liver ECM are not completely known, highlighting a need for future studies.

ECM accumulation is recognized as a structural barrier between cells and the vascular space that restricts molecular transport⁸². More recently, a body of evidence has emerged indicating that cellular changes that accompany ECM accumulation are receptor-mediated. As such, the ECM is a biomolecular "motherboard" that determines the physical and metabolic properties of the tissue and the cells that they envelope. A greater understanding of how the ECM changes in obesity and the contribution of individual ECM proteins will be necessary in defining extracellular processes impacting metabolic health.

Extracellular matrix expansion and integrins in the setting of obesity

Integrins are a class of receptors that bind ECM proteins and have numerous overlapping functions, including cell adhesion, mechanotransduction, and differentiation^{83,84}. ECM receptors are involved in a myriad of receptor signaling events through physical and functional interactions with growth factor receptors, including the insulin receptor⁸⁵. In this way, ECM receptors orchestrate dynamic and specific signaling responses to diverse physiological and pathophysiological conditions. Integrins functionally link ECM changes to a multitude of conditions, including IR⁸⁶ (summarized in Figure 2).

Integrins are heterodimers consisting of α and β subunits with varying ligand specificities and expression in different tissues. Differentiated insulin-sensitive cells from SkM, adipose tissue, and liver express a variety of α subunit isoforms but express only a single β integrin isoform (β 1)^{87–89}. Whole-body loss of the integrin α 1 subunit, a pro-fibrotic integrin receptor subunit that exclusively binds to β1, fails to protect against diet-induced SkM IR in mice; however, loss of the anti-fibrotic $\alpha 2$ isoform that also binds to $\beta 1$ is protective⁵⁵. It is interesting to note that combined SkM and myocardial loss of the integrin $\beta 1$ subunit results in IR in lean mice⁹⁰. Integrin-linked kinase (ILK) is a protein that physically associates with the intracellular tail of the β integrin subunit⁹⁰. In contrast to the IR caused by knockout of the integrin $\beta 1$ subunit in both SkM and myocardium of lean mice90, SkM-specific loss of ILK (mILK-KO mice) results in improved SkM insulin action in DIO mice91. Liver-specific deletion of ILK also protects against IR in DIO mice92. Whether adipocyte ILK deletion has effects on nutrient metabolism remains to be determined.

Despite its name, ILK lacks a functional kinase domain but rather functions as a scaffold for at least 26 high-fidelity binding partners⁹³. Most notable among these binding proteins are PINCH and parvin, which, together with ILK, form an ILK/PINCH/Parvin (IPP) complex. PINCH consists of two isoforms (PINCH1 and PINCH2) that have both distinct and overlapping cellular functions⁹⁴. In the context of glucose homeostasis, PINCH can bind to Nck2, which in turn interacts with insulin receptor substrate-1 (IRS-1)⁹⁵, a requisite for insulin signaling. Nck2 is highly expressed in epididymal adipose tissue and its genetic deletion in mice causes IR and increased lipolysis⁹⁶. PINCH has also been implicated in the phosphorylation of Akt via interactions with ILK⁹⁷. Three ubiquitously expressed isoforms of parvin exist (α , β , and γ). α - and β -parvin both can bind directly to f-actin and in this way



Figure 2. Putative mechanisms for the role of integrins in the development of insulin resistance. Extracellular matrix (ECM) proteins are ligands for integrins, a family of cell surface receptors. Integrins are linked to the regulation of glucose metabolism through numerous mechanisms. Integrins can co-localize with transforming growth factor beta (TGFβ) and insulin receptors that are key regulators of glucose uptake into tissues. Integrins are also involved in intracellular signaling through the integrin-linked kinase (ILK)/PINCH/Parvin (IPP) complex. PINCH is characterized as a modulator of kinase signaling pathways as it regulates Nck2 and Akt, requisite proteins for insulin signaling. Parvin is involved in the regulation of cytoskeletal dynamics that permit remodeling and translocation of mitochondria and various intracellular proteins (that is, glucose transporters). The integration of integrins with regulatory nodes for glucose metabolism highlights the potential significance of ECM-integrin signaling in the etiology of insulin resistance.

regulate cytoskeletal dynamics98. Parvin-mediated regulation of actin cytoskeletal dynamics is thought to occur, at least in part, via interactions between parvin and the Rho GTPase Rac199,100 and actin depolymerizing factor protein cofilin¹⁰¹. Rac1 is required for insulin-stimulated glucose uptake and is impaired during IR¹⁰², representing a potential link between integrins and insulin action. A potential role for γ -parvin in the context of insulin action has not been elucidated. Rac1 and cofilin are also involved in the regulation of numerous mitochondrial processes, including fission¹⁰³, apoptosis¹⁰⁴, and translocation¹⁰⁵, demonstrating a link between integrins and the regulation of oxidative metabolism. Whether integrins and the IPP complex directly regulate Rac1 or cofilin in obesity is not yet known, nor is it known what role the IPP complex may play in obesity through its other binding partners. A recent report shows that focal adhesion kinase (FAK), an alternative downstream target of integrin activation, can modulate insulin sensitivity through regulation of adipocyte survival¹⁰⁶. In light of the complexities of the ECM, integrins, and intracellular signaling pathways, much remains to be learned about ECM-integrin interactions in IR.

Summary and future directions

The etiology of IR involves both cell-intrinsic regulation of nutrient metabolism and integrated systems pathophysiology. The established paradigm of meta-inflammation coupled with the emerging concept of meta-fibrosis illustrates the complex nature of IR; however, several major questions remain to be addressed. For example, the composition and organization of the ECM must be elucidated so that the contribution of individual proteins or complexes or both can be mechanistically understood. Additionally, the role of downstream intracellular substrates of integrin signaling must be defined in the context of IR and cellular metabolism. The complex nature and broad importance of ECM/integrin function will be better understood through interdisciplinary studies that draw expertise from numerous fields (such as mechanobiology, biophysics, endocrinology, and molecular metabolism).

It is anticipated that future studies will provide a more complete understanding of how the ECM functions as a biophysical regulator of whole-body function and support the development of novel therapeutics aimed at treating IR by mitigating meta-fibrosis.

Competing interests

The authors declare that they have no competing interests.

Grant information

The author(s) declared that this work was funded by National Institutes of Health (grants DK054902, DK050277, DK059637), NIDDK Mouse Metabolic Phenotyping Centers MICROMouse Program (grant 15GRU2558) and American Heart Association (grant 16POST29910001).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

- F Aguilar M, Bhuket T, Torres S, et al.: Prevalence of the metabolic syndrome in the United States, 2003–2012. JAMA. 2015; 313(19): 1973–4.
 PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Kang S, Tsai LT, Rosen ED: Nuclear Mechanisms of Insulin Resistance. Trends Cell Biol. 2016; 26(5): 341–51.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Gross B, Pawlak M, Lefebvre P, et al.: PPARs in obesity-induced T2DM, dyslipidaemia and NAFLD. Nat Rev Endocrinol. 2017; 13(1): 36–49. PubMed Abstract | Publisher Full Text
- Endorsed by The Obesity Society, Young DR, Hivert MF, et al.: Sedentary Behavior and Cardiovascular Morbidity and Mortality: A Science Advisory From the American Heart Association. Circulation. 2016; 134(13): e262–79. PubMed Abstract | Publisher Full Text
- Donnelly JE, Blair SN, Jakicic JM, *et al.*: American College of Sports Medicine Position Stand. Appropriate physical activity intervention strategies for weight loss and prevention of weight regain for adults. *Med Sci Sports Exerc.* 2009; 41(2): 459–71.
 - PubMed Abstract | Publisher Full Text
- Tremblay A, Chaput JP: Adaptive reduction in thermogenesis and resistance to lose fat in obese men. Br J Nutr. 2009; 102(4): 488–92.
 PubMed Abstract | Publisher Full Text
- Doucet E, St-Pierre S, Alméras N, et al.: Evidence for the existence of adaptive thermogenesis during weight loss. Br J Nutr. 2001; 85(6): 715–23.
 PubMed Abstract | Publisher Full Text
- Redman LM, Heilbronn LK, Martin CK, et al.: Metabolic and behavioral compensations in response to caloric restriction: implications for the maintenance of weight loss. PLoS One. 2009; 4(2): e4377.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Speakman JR, Selman C: Physical activity and resting metabolic rate. Proc Nutr Soc. 2003; 62(3): 621–34.
 PubMed Abstract | Publisher Full Text
- Thomas DM, Bouchard C, Church T, et al.: Why do individuals not lose more weight from an exercise intervention at a defined dose? An energy balance analysis. Obes Rev. 2012; 13(10): 835–47.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Byrne NM, Wood RE, Schutz Y, et al.: Does metabolic compensation explain the majority of less-than-expected weight loss in obese adults during a short-term severe diet and exercise intervention? Int J Obes (Lond). 2012; 36(11): 1472–8. PubMed Abstract | Publisher Full Text
- Shaw K, Gennat H, O'Rourke P, et al.: Exercise for overweight or obesity. Cochrane Database Syst Rev. 2006; (4): CD003817.
 PubMed Abstract | Publisher Full Text
- Jennings AE, Alberga A, Sigal RJ, et al.: The effect of exercise training on resting metabolic rate in type 2 diabetes mellitus. *Med Sci Sports Exerc.* 2009; 41(8): 1558–65.
 - PubMed Abstract | Publisher Full Text
- Amati F, Dubé JJ, Shay C, et al.: Separate and combined effects of exercise training and weight loss on exercise efficiency and substrate oxidation. J Appl Physiol (1985). 2008; 105(3): 825–31.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Maclean PS, Bergouignan A, Cornier MA, et al.: Biology's response to dieting: the impetus for weight regain. Am J Physiol Regul Integr Comp Physiol. 2011; 301(3): FIS81–600.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- Garland T Jr, Schutz H, Chappell MA, et al.: The biological control of voluntary exercise, spontaneous physical activity and daily energy expenditure in relation to obesity: human and rodent perspectives. J Exp Biol. 2011; 214(Pt 2):

206–29. PubMed Abstract | Publisher Full Text | Free Full Text

 F O'Neal TJ, Friend DM, Guo J, et al.: Increases in Physical Activity Result in Diminishing Increments in Daily Energy Expenditure in Mice. Curr Biol. 2017; 27(3): 423–30.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

F1000 recommended

- Levine JA, Eberhardt NL, Jensen MD: Role of nonexercise activity
 the second second
- thermogenesis in resistance to fat gain in humans. Science. 1999; 283(5399): 212–4. PubMed Abstract | Publisher Full Text
- F MacLean PS, Higgins JA, Giles ED, et al.: The role for adipose tissue in weight regain after weight loss. Obes Rev. 2015; 16 Suppl 1: 45–54.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- F Teich T, Dunford EC, Porras DP, et al.: Glucocorticoid antagonism limits adiposity rebound and glucose intolerance in young male rats following the cessation of daily exercise and caloric restriction. Am J Physiol Endocrinol Metab. 2016; 311(1): E56–68.
- PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

 21.
 Lark DS, Fisher-Wellman KH, Neufer PD: High-fat load: mechanism(s) of insulin
- resistance in skeletal muscle. Int J Obes Suppl. 2012; 2(Suppl 2): S31–S36.
 PubMed Abstract | Publisher Full Text | Free Full Text
 Hesselink MK, Schrauwen-Hinderling V, Schrauwen P: Skeletal muscle
- mitochondria as a target to prevent or treat type 2 diabetes mellitus. Nat Rev Endocrinol. 2016; 12(11): 633–45. PubMed Abstract | Publisher Full Text
- Theurey P, Rieusset J: Mitochondria-Associated Membranes Response to Nutrient Availability and Role in Metabolic Diseases. Trends Endocrinol Metab. 2017; 28(1): 32–45.
 PubMed Abstract | Publisher Full Text
- Muoio DM, Neufer PD: Lipid-induced mitochondrial stress and insulin action in muscle. Cell Metab. 2012; 15(5): 595–605.
 PubMed Abstract | Publisher Full Text | Free Full Text
- E Satapati S, Kucejova B, Duarte JA, et al.: Mitochondrial metabolism mediates oxidative stress and inflammation in fatty liver. J Clin Invest. 2015;
- 125(12): 4447–62.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 Koves TR. Ussher JR. Noland RC. *et al.*: Mitochondrial overload and incomplete
- fatty acid oxidation contribute to skeletal muscle insulin resistance. Cell Metab. 2008; 7(1): 45–56. PubMed Abstract | Publisher Full Text
- Adams SH, Hoppel CL, Lok KH, et al.: Plasma acylcarnitine profiles suggest incomplete long-chain fatty acid beta-oxidation and altered tricarboxylic acid cycle activity in type 2 diabetic African-American women. J Nutr. 2009; 139(6): 1073–81.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- Noland RC, Koves TR, Seiler SE, et al.: Carnitine insufficiency caused by aging and overnutrition compromises mitochondrial performance and metabolic control. J Biol Chem. 2009; 284(34): 22840–52.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Hoehn KL, Salmon AB, Hohnen-Behrens C, et al.: Insulin resistance is a cellular antioxidant defense mechanism. Proc Natl Acad Sci U S A. 2009; 106(42): 17787–92.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Muoio DM, Noland RC, Kovalik JP, et al.: Muscle-specific deletion of carnitine acetyltransferase compromises glucose tolerance and metabolic flexibility. *Cell Metab.* 2012; 15(5): 764–77.
 PubMed Abstract | Publisher Full Text | Free Full Text

- Gómez-Valadés AG, Méndez-Lucas A, Vidal-Alabró A, et al.: Pck1 gene silencing in the liver improves glycemia control, insulin sensitivity, and dyslipidemia in db/db mice. Diabetes. 2008; 57(8): 2199–210.
 PubMed Abstract | Publisher Full Text | Free Full Text
- F Cotter DG, Ercal B, Huang X, et al.: Ketogenesis prevents diet-induced fatty liver injury and hyperglycemia. J Clin Invest. 2014; 124(12): 5175–90.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Garbow JR, Doherty JM, Schugar RC, et al.: Hepatic steatosis, inflammation, and ER stress in mice maintained long term on a very low-carbohydrate ketogenic diet. Am J Physiol Gastrointest Liver Physiol. 2011; 300(6): G956–67. PubMed Abstract | Publisher Full Text | Free Full Text
- F Klein MS, Newell C, Bomhof MR, et al.: Metabolomic Modeling To Monitor Host Responsiveness to Gut Microbiota Manipulation in the BTBR^{T-MD} Mouse. J Proteome Res. 2016; 15(4): 1143–50.
 PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Newell C, Shutt TE, Ahn Y, et al.: Tissue Specific Impacts of a Ketogenic Diet on Mitochondrial Dynamics in the BTBR^{T+th} Mouse. Front Physiol. 2016; 7: 654.
- PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation 36. Fisher-Wellman KH, Neufer PD: Linking mitochondrial bioenergetics to insulin
- resistance via redox biology. Trends Endocrinol Metab. 2012; 23(3): 142–53. PubMed Abstract | Publisher Full Text | Free Full Text
- Jones DP: Radical-free biology of oxidative stress. Am J Physiol Cell Physiol. 2008; 295(4): C849–68.
 PubMed Abstract | Publisher Full Text | Free Full Text
- F Anderson EJ, Lustig ME, Boyle KE, et al.: Mitochondrial H2O2 emission and cellular redox state link excess fat intake to insulin resistance in both rodents and humans. J Clin Invest. 2009; 119(3): 573–81.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Kane DA, Anderson EJ, Price JW 3rd, *et al.*: Metformin selectively attenuates mitochondrial H2O2 emission without affecting respiratory capacity in skeletal muscle of obese rats. *Free Radic Biol Med.* 2010; 49(6): 1082–7.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Lark DS, Kang L, Lustig ME, et al.: Enhanced mitochondrial superoxide scavenging does not improve muscle insulin action in the high fat-fed mouse. PLoS One. 2015; 10(5): e0126732.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Paglialunga S, van Bree B, Bosma M, et al.: Targeting of mitochondrial reactive oxygen species production does not avert lipid-induced insulin resistance in muscle tissue from mice. *Diabetologia*. 2012; 55(10): 2759–68.
 PubMed Abstract | Publisher Full Text
- Anderson EJ, Yamazaki H, Neufer PD: Induction of endogenous uncoupling protein 3 suppresses mitochondrial oxidant emission during fatty acidsupported respiration. J Biol Chem. 2007; 282(43): 31257–66.
 PubMed Abstract | Publisher Full Text
- Lark DS, Torres MJ, Lin CT, et al.: Direct real-time quantification of mitochondrial oxidative phosphorylation efficiency in permeabilized skeletal muscle myofibers. Am J Physiol Cell Physiol. 2016; 311(2): C239–45. PubMed Abstract | Publisher Full Text | Free Full Text
- Kenwood BM, Weaver JL, Bajwa A, et al.: Identification of a novel mitochondrial uncoupler that does not depolarize the plasma membrane. Mol Metab. 2014; 3(2): 114–23.
- PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 45. Ost M, Keipert S, Klaus S: Targeted mitochondrial uncoupling beyond UCP1
- The fine line between death and metabolic health. *Biochimie*. 2017; 134: 77–85. PubMed Abstract | Publisher Full Text
- Lou P, Hansen BS, Olsen PH, et al.: Mitochondrial uncouplers with an extraordinary dynamic range. Biochem J. 2007; 407(1): 129–40.
 PubMed Abstract | Publisher Full Text | Free Full Text
- F Hotamisligii GS: Inflammation and metabolic disorders. Nature. 2006; 444(7121): 860–7.
 PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 48. F Xu H, Barnes GT, Yang Q, et al.: Chronic inflammation in fat plays a crucial
- Tole in the development of obesity-related insulin resistance. J Clin Invest. 2003; 112(12): 1821–30. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Mittal M, Siddiqui MR, Tran K, *et al.*: Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal.* 2014; 20(7): 1126–67.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Man K, Kutyavin VI, Chawla A: Tissue Immunometabolism: Development, Physiology, and Pathobiology. Cell Metab. 2017; 25(1): 11–26.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Lackey DE, Olefsky JM: Regulation of metabolism by the innate immune system. Nat Rev Endocrinol. 2016; 12(1): 15–28.
 PubMed Abstract | Publisher Full Text
- Hotamisligil GS, Shargill NS, Spiegelman BM: Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. Science. 1993; 259(5091): 87–91.
 PubMed Abstract | Publisher Full Text
- 53. Plomgaard P, Bouzakri K, Krogh-Madsen R, et al.: Tumor necrosis factor-alpha induces skeletal muscle insulin resistance in healthy human subjects via

inhibition of Akt substrate 160 phosphorylation. *Diabetes*. 2005; 54(10): 2939–45. PubMed Abstract | Publisher Full Text

- Lang CH, Dobrescu C, Bagby GJ: Tumor necrosis factor impairs insulin action on peripheral glucose disposal and hepatic glucose output. *Endocrinology*. 1992; 130(1): 43–52.
 PubMed Abstract | Publisher Full Text
- Kang L, Ayala JE, Lee-Young RS, et al.: Diet-induced muscle insulin resistance is associated with extracellular matrix remodeling and interaction with integrin alpha2beta1 in mice. Diabetes. 2011; 60(2): 416–26.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Rutkowski JM, Stern JH, Scherer PE: The cell biology of fat expansion. J Cell Biol. 2015; 208(5): 501–12.
- PubMed Abstract | Publisher Full Text | Free Full Text

 57.
 Crewe C, An YA, Scherer PE: The ominous triad of adipose tissue dysfunction:
- inflammation, fibrosis, and impaired angiogenesis. J Clin Invest. 2017; 127(1): 74–82. PubMed Abstract | Publisher Full Text | Free Full Text
- F Vila IK, Badin PM, Marques MA, et al.: Immune cell Toll-like receptor 4 mediates the development of obesity- and endotoxemia-associated adipose tissue fibrosis. Cell Rep. 2014; 7(4): 1116–29.
 PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Spencer M, Yao-Borengasser A, Unal R, et al.: Adipose tissue macrophages in insulin-resistant subjects are associated with collagen VI and fibrosis and demonstrate alternative activation. Am J Physiol Endocrinol Metab. 2010; 299(6): E1016–27.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- 60. F Dankel SN, Svärd J, Matthä S, et al.: COL6A3 expression in adipocytes associates with insulin resistance and depends on PPARγ and adipocyte size. Obesity (Silver Spring). 2014; 22(8): 1807–13. PubMed Abstract | Publisher Full Text | F1000 Recommendation
- F Guglielmi V, Cardellini M, Cinti F, et al.: Omental adipose tissue fibrosis and insulin resistance in severe obesity. Nutr Diabetes. 2015; 5: e175.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- F Muir LA, Neeley CK, Meyer KA, et al.: Adipose tissue fibrosis, hypertrophy, and hyperplasia: Correlations with diabetes in human obesity. Obesity (Silver Spring). 2016; 24(3): 597–605.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Khan T, Muise ES, Iyengar P, et al.: Metabolic dysregulation and adipose tissue fibrosis: role of collagen VI. Mol Cell Biol. 2009; 29(6): 1575–91.
- PubMed Abstract | Publisher Full Text | Free Full Text
 Huber J, Löffler M, Bilban M, et al.: Prevention of high-fat diet-induced adipose
- tissue remodeling in obese diabetic mice by n-3 polyunsaturated fatty acids. Int J Obes (Lond). 2007; 31(6): 1004–13.
 PubMed Abstract | Publisher Full Text
- Kiefer FW, Zeyda M, Todoric J, et al.: Osteopontin expression in human and murine obesity: extensive local up-regulation in adipose tissue but minimal systemic alterations. Endocrinology. 2008; 149(3): 1350–7. PubMed Abstract | Publisher Full Text
- Nomiyama T, Perez-Tilve D, Ogawa D, et al.: Osteopontin mediates obesityinduced adipose tissue macrophage infiltration and insulin resistance in mice. J Clin Invest. 2007; 117(10): 2877–88.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Han CY, Subramanian S, Chan CK, et al.: Adipocyte-derived serum amyloid A3 and hyaluronan play a role in monocyte recruitment and adhesion. *Diabetes*. 2007; 56(9): 2260–73.
 PubMed Abstract | Publisher Full Text
- Varma V, Yao-Borengasser A, Bodles AM, et al.: Thrombospondin-1 is an adipokine associated with obesity, adipose inflammation, and insulin resistance. Diabetes. 2008; 57(2): 432–9.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Inoue M, Jiang Y, Barnes RH 2nd, et al.: Thrombospondin 1 mediates highfat diet-induced muscle fibrosis and insulin resistance in male mice. Endocrinology. 2013; 154(12): 4548–59.
 PubMed Abstract | Publisher Full Text | Free Full Text
- F Craft CS, Pietka TA, Schappe T, et al.: The extracellular matrix protein MAGP1 supports thermogenesis and protects against obesity and diabetes through regulation of TGF-B. Diabetes. 2014; 63(6): 1920–32.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 71. F Luo T, Nocon A, Fry J, et al.: AMPK Activation by Metformin Suppresses Abnormal Extracellular Matrix Remodeling in Adipose Tissue and Ameliorates Insulin Resistance in Obesity. Diabetes. 2016; 65(8): 2295–310. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Berria R, Wang L, Richardson DK, et al.: Increased collagen content in insulinresistant skeletal muscle. Am J Physiol Endocrinol Metab. 2006; 290(3): E560–5. PubMed Abstract | Publisher Full Text
- F Pincu Y, Linden MA, Zou K, et al.: The effects of high fat diet and moderate exercise on TGFβ1 and collagen deposition in mouse skeletal muscle. Cytokine. 2015; 73(1): 23–9.
- PubMed Abstract | Publisher Full Text | F1000 Recommendation
 74. Wasserman DH: Four grams of glucose. Am J Physiol Endocrinol Metab. 2009; 296(1): E11–21.
 - PubMed Abstract | Publisher Full Text | Free Full Text

- 75. F Tam CS, Chaudhuri R, Hutchison AT, et al.: Skeletal muscle extracellular matrix remodeling after short-term overfeeding in healthy humans. *Metab Clin Exp.* 2017; 67: 26–30. PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Kang L, Mayes WH, James FD, et al.: Matrix metalloproteinase 9 opposes dietinduced muscle insulin resistance in mice. Diabetologia. 2014; 57(3): 603–13. PubMed Abstract | Publisher Full Text | Free Full Text
- 77. F Kang L, Lantier L, Kennedy A, et al.: Hyaluronan accumulates with high-fat feeding and contributes to insulin resistance. Diabetes. 2013; 62(6): 1888–96. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Ekstedt M, Franzén LE, Mathiesen UL, et al.: Long-term follow-up of patients with NAFLD and elevated liver enzymes. *Hepatology*. 2006; 44(4): 865–73. PubMed Abstract | Publisher Full Text
- McCullough AJ: The clinical features, diagnosis and natural history of nonalcoholic fatty liver disease. *Clin Liver Dis.* 2004; 8(3): 521–33, viii. PubMed Abstract | Publisher Full Text
- Kohli R, Kirby M, Xanthakos SA, *et al.*: High-fructose, medium chain trans fat diet induces liver fibrosis and elevates plasma coenzyme Q9 in a novel murine model of obesity and nonalcoholic steatohepatitis. *Hepatology*. 2010; 52(3): 934–44.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- F Luo Y, Burrington CM, Graff EC, et al.: Metabolic phenotype and adipose and liver features in a high-fat Western diet-induced mouse model of obesitylinked NAFLD. Am J Physiol Endocrinol Metab. 2016; 310(6): E418–39. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Sun K, Tordjman J, Clément K, *et al.*: Fibrosis and adipose tissue dysfunction. *Cell Metab.* 2013; 18(4): 470–7.
- PubMed Abstract | Publisher Full Text | Free Full Text
- Gattazzo F, Urciuolo A, Bonaldo P: Extracellular matrix: a dynamic microenvironment for stem cell niche. *Biochim Biophys Acta*. 2014; 1840(8): 2506–19.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- Ringer P, Colo G, Fässler R, et al.: Sensing the mechano-chemical properties of the extracellular matrix. Matrix Biol. 2017; pii: S0945-053X(17)30018-5.
 PubMed Abstract | Publisher Full Text
- Kim SH, Turnbull J, Guimond S: Extracellular matrix and cell signalling: the dynamic cooperation of integrin, proteoglycan and growth factor receptor. *J Endocrinol.* 2011; 209: 139–51.
 PubMed Abstract | Publisher Full Text
- 86. Williams AS, Kang L, Wasserman DH: The extracellular matrix and insulin resistance. Trends Endocrinol Metab. 2015; 26(7): 357–66. PubMed Abstract | Publisher Full Text | Free Full Text
- Schwander M, Shirasaki R, Pfaff SL, *et al.*: Beta1 integrins in muscle, but not in motor neurons, are required for skeletal muscle innervation. *J Neurosci.* 2004; 24(37): 8181–91.

PubMed Abstract | Publisher Full Text

- Schwander M, Leu M, Stumm M, et al.: Beta1 integrins regulate myoblast fusion and sarcomere assembly. Dev Cell. 2003; 4(5): 673–85.
 PubMed Abstract | Publisher Full Text
- Liadaki K, Casar JC, Wessen M, et al.: β4 integrin marks interstitial myogenic progenitor cells in adult murine skeletal muscle. J Histochem Cytochem. 2012; 60(1): 31–44.
 - PubMed Abstract | Publisher Full Text | Free Full Text
- Zong H, Bastie CC, Xu J, et al.: Insulin resistance in striated muscle-specific integrin receptor beta1-deficient mice. J Biol Chem. 2009; 284(7): 4679–88.
 PubMed Abstract | Publisher Full Text | Free Full Text
- 91. Kang L, Mokshagundam S, Reuter B, et al.: Integrin-Linked Kinase in Muscle Is

Necessary for the Development of Insulin Resistance in Diet-Induced Obese Mice. Diabetes. 2016; 65(6): 1590–600. PubMed Abstract | Publisher Full Text | Free Full Text

- Williams AS, Trefts E, Lantier L, *et al.*: Integrin-Linked Kinase Is Necessary for the Development of Diet-Induced Hepatic Insulin Resistance. *Diabetes.* 2017; 66(2): 325–34.
- PubMed Abstract | Publisher Full Text | Free Full Text
- Dobreva I, Fielding A, Foster LJ, et al.: Mapping the integrin-linked kinase interactome using SILAC. J Proteome Res. 2008; 7(4): 1740–9. PubMed Abstract | Publisher Full Text
- Kovalevich J, Tracy B, Langford D: PINCH: More than just an adaptor protein in cellular response. J Cell Physiol. 2011; 226(4): 940–7.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Tu Y, Liang L, Frank SJ, et al.: Src homology 3 domain-dependent interaction of Nck-2 with insulin receptor substrate-1. Biochem J. 2001; 354(Pt 2): 315–22.
 PubMed Abstract | Publisher Full Text | Free Full Text
- 96. F Dusseault J, Li B, Haider N, et al.: Nck2 Deficiency in Mice Results in Increased Adiposity Associated With Adipocyte Hypertrophy and Enhanced Adipogenesis. Diabetes. 2016; 65(9): 2652–66. PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Xu Z, Fukuda T, Li Y, *et al.*: Molecular dissection of PINCH-1 reveals a mechanism of coupling and uncoupling of cell shape modulation and survival. *J Biol Chem*. 2005; 280(30): 27631–7.
 PubMed Abstract | Publisher Full Text
- Nikolopoulos SN, Turner CE: Actopaxin, a new focal adhesion protein that binds paxillin LD motifs and actin and regulates cell adhesion. J Cell Biol. 2000; 151(7): 1435–48.
 PubMed Abstract | Publisher Full Text | Free Full Text
- Berrier AL, Martinez R, Bokoch GM, *et al.*: The integrin beta tail is required and sufficient to regulate adhesion signaling to Rac1. *J Cell Sci.* 2002; 115(Pt 22): 4285–91.
- PubMed Abstract | Publisher Full Text | F1000 Recommendation

 100.
 Pignatelli J, LaLonde SE, LaLonde DP, et al.: Actopaxin (α-parvin)

 phosphorylation is required for matrix degradation and cancer cell invasion.
 J Biol Chem. 2012; 287(44): 37309–20.

 PubMed Abstract | Publisher Full Text | Free Full Text
 Preserver Full Text
- 101. E Shibue T, Brooks MW, Weinberg RA: An integrin-linked machinery of cytoskeletal regulation that enables experimental tumor initiation and metastatic colonization. *Cancer Cell.* 2013; 24(4): 481–98. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 102. Sylow L, Jensen TE, Kleinert M, et al.: Rac1 signaling is required for insulinstimulated glucose uptake and is dysregulated in insulin-resistant murine and human skeletal muscle. Diabetes. 2013; 62(6): 1865–75. PubMed Abstract | Publisher Full Text | Free Full Text
- 103. F Li S, Xu S, Roelofs BA, et al.: Transient assembly of F-actin on the outer mitochondrial membrane contributes to mitochondrial fission. J Cell Biol. 2015; 208(1): 109–23. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Chua BT, Volbracht C, Tan KO, *et al.*: Mitochondrial translocation of cofilin is an early step in apoptosis induction. *Nat Cell Biol.* 2003; 5(12): 1083–9.
- PubMed Abstract | Publisher Full Text

 105.
 F

 Matveeva EA, Venkova LS, Chernoivanenko IS, et al.: Vimentin is involved
- in regulation of mitochondrial motility and membrane potential by Rac1. Biol Open. 2015; 4(10): 1290–7. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- F Luk CT, Shi SY, Cai EP, et al.: FAK signalling controls insulin sensitivity through regulation of adipocyte survival. Nat Commun. 2017; 8: 14360.
 PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

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Version 1

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Competing Interests: No competing interests were disclosed.

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Competing Interests: No competing interests were disclosed.

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