

Reduction of blood pressure by store-operated calcium channel blockers

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Abstract

The voltage-operated Ca^{2+} channels (VOCC), which allow Ca^{2+} influx from the extracellular space, are inhibited by anti-hypertensive agents such as verapamil and nifedipine. The Ca^{2+} entering from outside into the cell triggers Ca^{2+} release from the sarcoplasmic reticulum (SR) stores. To refill the depleted Ca^{2+} stores in the SR, another type of Ca^{2+} channels in the cell membrane, known as store-operated Ca^{2+} channels (SOCC), are activated. These SOCCs are verapamil and nifedipine resistant, but are SKF 96465 (SK) and gadolinium (Gd^{3+}) sensitive. Both SK and Gd^{3+} have been shown to reduce $[\text{Ca}^{2+}]_i$ in the smooth muscle, but their effects on blood pressure have not been reported. Our results demonstrated that both SK and Gd^{3+} produced a dose-dependent reduction in blood pressure in rat. The combination of SK and verapamil produced an additive action in lowering the blood pressure. Furthermore, SK, but not Gd^{3+} suppressed proliferation of vascular smooth muscle cells in the absence or presence of lysophosphatidic acid (LPA). SK decreased the elevation of $[\text{Ca}^{2+}]_i$ induced by LPA, endothelin-1 (ET-1) and angiotensin II (Ang II), but did not affect the norepinephrine (NE)-evoked increase in $[\text{Ca}^{2+}]_i$. On the other hand, Gd^{3+} inhibited the LPA and Ang II induced change in $[\text{Ca}^{2+}]_i$, but had no effect on the ET-1 and NE induced increase in $[\text{Ca}^{2+}]_i$. The combination of verapamil and SK abolished the LPA- or adenosine-5'-triphosphate (ATP)-induced $[\text{Ca}^{2+}]_i$ augmentation. These results suggest that SOCC inhibitors, like VOCC blocker, may serve as promising drugs for the treatment of hypertension.

Keywords: Ca^{2+} antagonists • antihypertensive agents • intracellular Ca^{2+} • cell proliferation • hypertension

Introduction

In view of the effect of different vasoactive agents such as Ang II, NE, ET-1 and LPA on blood pressure [1–3], a wide variety of receptor-blocking drugs are being used for the treatment of hypertension. As the vasoactive hormones and agents are known to promote Ca^{2+} entry into the smooth muscle cells [4, 5], several VOCC antagonists or L-type Ca^{2+} channel antagonists including verapamil are also known to produce beneficial effect in hypertensive cases [6]. The increase in intracellular Ca^{2+} concentration ($[\text{Ca}^{2+}]_i$) in smooth muscle cells is not only dependent upon the entry of extracellular Ca^{2+} but Ca^{2+} released from the intracellular store has also been demonstrated to play a critical role [7]. The release of Ca^{2+} from the intracellular Ca^{2+} stores not only contributes to the increase of $[\text{Ca}^{2+}]_i$ directly, but also causes Ca^{2+} influx through voltage-independent Ca^{2+} channels, which are called Ca^{2+} release-activated currents or SOCC [8]. Although SOCC blockers such as SKF 96365 (SK) and Gd^{3+} have been shown to reduce $[\text{Ca}^{2+}]_i$ in vascular smooth muscle cells (VSMC) [9], the

effects of these agents on blood pressure as well as their actions on LPA-induced elevation of blood pressure have not been reported previously. Although SK has been shown to reduce the ET-1-induced vasoconstriction of rat cerebral arteries [10], the effects of this agent on VSMC with respect to cell proliferation and changes in $[\text{Ca}^{2+}]_i$ because of different agonists have not been studied. In view of the fact that hypertension is associated with an increase in smooth muscle cell proliferation and $[\text{Ca}^{2+}]_i$ [11], this study was undertaken to test the action of SK and Gd^{3+} on blood pressure, cell proliferation and $[\text{Ca}^{2+}]_i$. The specificity of the effect of SK was also examined by using various agents such as Ang II, ET-1, NE and LPA, which are known to increase the $[\text{Ca}^{2+}]_i$ and blood pressure. Some experiments were carried out to investigate the effect of SOCC blocker, SK in combination with VOCC antagonist, verapamil on blood pressure and $[\text{Ca}^{2+}]_i$.

Materials and methods

The use of animals and experimental protocol were according to the guidelines of the Canadian Council of Animal Care as approved by the

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Animal Care Committee of the University of Manitoba. SKF 96365 was purchased from Biomol Research Laboratory (Plymouth Meeting, PA, USA). Gadolinium, L- α -LPA and verapamil were obtained from Sigma Chemicals (Oakville, ON, Canada); Fura-2 acetoxymenthyl/ester (Fura 2-AM) was purchased from Molecular Probes (Eugene, OR, USA); [^3H] - thymidine was purchased from Amersham (Oakville, ON, Canada). DMEM and foetal bovine serum (FBS) were obtained from Invitrogen (Burlington, ON, Canada), whereas the A10 VSMC line was from the American Type Culture Collection (Manassas, VA, USA).

Measurement of blood pressure

Male Sprague Dawley rats weighing 250–300 g were anaesthetized by an intraperitoneal injection of ketamine (90 mg/kg) and xylazine (10 mg/kg) mixture. The right carotid artery was exposed and cannulated with a microtip pressure transducer (model SPR-249; Millar Instruments, Houston, TX, USA). The catheter was inserted carefully into the lumen of the carotid artery, then catheter was secured with a silk ligature around the artery and blood pressure values were recorded using the computer programme AcqKnowledge for Windows 3.5 (Biopac Systems Inc., Goleta, CA, USA). Arterial systolic and diastolic pressures were measured simultaneously [12].

Cell number count

The cultured A10 VSMC were treated with 0.25% trypsin-1 mM ethylenediaminetetraacetic acid (EDTA) for 2 min. and were then collected and centrifuged at $240 \times g$ for 5 min. at room temperature. The supernatant was removed and the cells were suspended in *N*-2-hydroxyethylpiperazine-*N'*-2-ethane sulphonic acid (HEPES) buffer; 0.1 ml of 0.4% Trypan blue was added to 0.5 ml cell suspension and kept for 5 min. at room temperature. About 20 μl of cell suspension was filled in the haemocytometer and the non-stained cells were counted under microscope.

Measurement of DNA synthesis

A10 VSMC were cultured in DMEM containing 10% FBS and 0.01 mg/ml gentamicin (Gibco, Burlington, ON, Canada) at 37°C with 95% air and 5% CO₂. DNA synthesis in A10 VSMC was measured by [^3H]-thymidine incorporation into the DNA of the cells [13]; cells were cultured in 12-well plates. Before the experiments, cells were incubated in serum-free DMEM for 20 hrs. Lysophosphatidic acid was added 10 min. after the addition of different inhibitors and after incubation for 4 hrs, 1 μCi [^3H]-thymidine was added. The reaction was terminated after 22 hrs by keeping the cell culture plates on the ice and removing the culture medium. The cells were washed three times with 1 ml HEPES buffer (mM/l: NaCl 145, KCl 4.5, CaCl₂ 1.0, MgSO₄·7H₂O 1.0, HEPES 10, glucose 5, bovine serum albumin 0.1%, KH₂PO₄ 1.0, pH 7.4). These cells were then incubated for 1 hr in cold 5% trichloroacetic acid on the ice, washed two more times with 0.5 ml HEPES buffer, and incubated with 0.2 ml NaOH (0.5 N) for 1 hr. The aliquots were transferred to scintillation vial. The radioactivity was counted in a Beckman LS 6500 scintillation counter after the addition of 10 ml Cytoscint-ES (MP Biomedicals, Santa Ana, CA, USA).

Measurement of [Ca^{2+}]_i

The cultured A10 VSMC were incubated with 0.25% trypsin-1 mM EDTA for 2 min. and then the cells were harvested and centrifuged at $240 \times g$ for 5 min. at room temperature. The supernatant was discarded and the cells were incubated with 10 μM Fura 2-AM in HEPES buffer for 40 min. at 37°C. The cells were then washed twice with HEPES buffer and the cell number was adjusted to 0.3×10^6 cells/ml by adding HEPES buffer. The fluorescence intensity of Fura-2 was determined by a SLM DMX-1100 dual-wavelength spectrofluorometer (SLM Instruments, Inc, Urbana, IL, USA); the ratio (R) of fluorescence signal at 340/380 (nm) was calculated automatically. The R_{max} and R_{min} values were determined by the addition of 40 μl Triton X-100 (10%) and 20 μl EGTA (40 mM) to a cuvette with 2 ml cell suspension respectively. The [Ca^{2+}]_i was calculated according to the following formula: [Ca^{2+}]_i = $224 \times [(R - R_{\min}) / (R_{\max} - R)] \times \text{Sf}_2 / \text{Sb}_2$, where Sf₂ and Sb₂ are the fluorescence proportionality coefficients obtained at 380 nm under R_{min} and R_{max} conditions respectively [14].

Statistical analysis

The data are expressed as mean \pm S.E.M. Statistical analysis was performed with the Microcal Origin Version 6 (Microcal Software Inc., Northampton, MA, USA). The data analysis was carried out by one-way ANOVA analysis, the comparison of mean values of the two groups was performed by Student's *t*-test. *P* values less than 0.05 were considered to be significantly different.

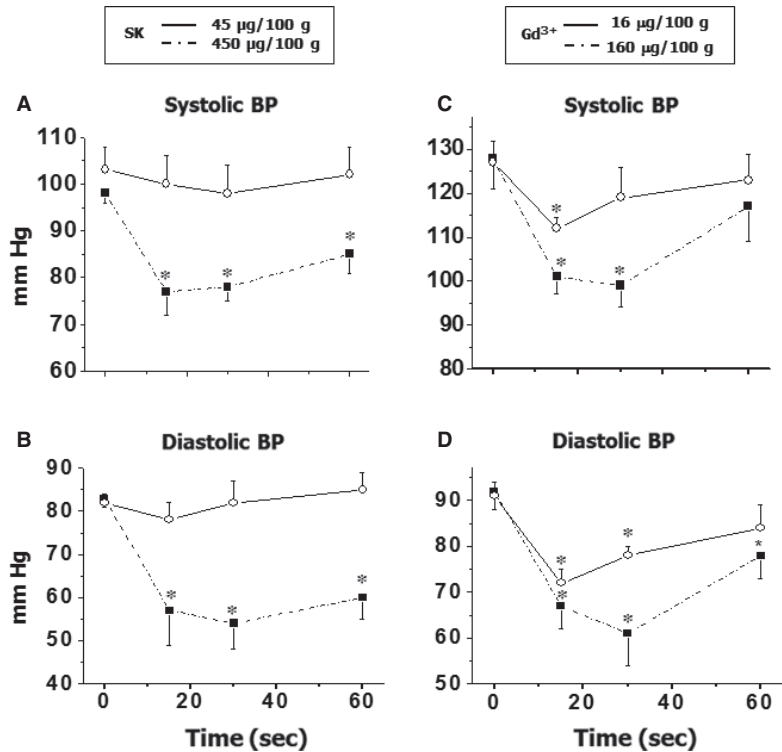
Results

SOCC blockers on blood pressure

The effects of SOCC blockers on blood pressure in rats were tested by injecting different doses of SK or Gd³⁺ intravenously. The blood pressure was monitored before and after treatment. In preliminary experiments, both SK (45–450 $\mu\text{g}/100$ g b.w.) and Gd³⁺ (16–160 $\mu\text{g}/100$ g b.w.) were found to lower blood pressure in a dose-dependent manner. As shown in Figure 1, both agents induced dose- and time-dependent reductions in systolic (25% by SK and 23% by Gd³⁺, Figures 1A and 1C, respectively) and diastolic (35% by SK and 33% by Gd³⁺, Figures 1B and 1C, respectively) blood pressures. The maximum effects were achieved within 30 sec. of injection. Both systolic blood pressure and diastolic blood pressure were still significantly lower at 60 sec. of the injection of SK; however, there is no significant difference in the systolic blood pressure at 60 sec. following Gd³⁺ treatment.

In view of the importance of LPA in the development of hypertension [3], the effect of SOCC blockers on the LPA-induced elevation of blood pressure were tested by giving different doses of SK (4.5–45 $\mu\text{g}/100$ g b.w.; i.v.) 30 sec. prior to the treatment with LPA (5.6 $\mu\text{g}/100$ g b.w.). It was observed that SK pre-treatment caused a dose-dependent inhibition of the LPA-induced change in blood pressure; higher dose of SK (45 $\mu\text{g}/100$ g b.w.) abolished the effect of LPA (Fig. 2). As VOCC antagonists are commonly used for the control

Fig. 1 The effect of SK and Gd³⁺ on blood pressure. The blood pressure was recorded from the carotid artery before and after treatment by a microtip pressure transducer and computer programme Acq-knowledge for Windows 3.5. **P* < 0.05 compared with basal value; *n* = 6.



of blood pressure, it was planned to test if the combination use of VOCC and SOCC blockers can produce an additive effect. The blood pressure was monitored before and after injection of SK (45 µg/100 g b.w.), verapamil (15 µg/100 g b.w.) or combination of these two agents. It is pointed out that the use of verapamil in combination with SK was based on the fact that verapamil is well known to block VOCC in both the VSMC and the heart [6, 15] and such a combination could be expected to reduce the side effects of this agent. As shown in Table 1, the combination of SK and verapamil produced a stronger effect on the diastolic blood pressure in comparison to the treatments with SK or verapamil alone.

SOCC blockers on cell proliferation

The ratio of arterial lumen and wall thickness is an essential factor for the regulation of blood pressure [11]. As cell proliferation plays critical role in the thickness of blood vessel, the effect of SOCC blockers on cell proliferation was tested in cultured A10 VSMC. As shown in Figure 3, SK caused a dose-dependent inhibition of cell proliferation in the absence or presence of LPA as reflected by the change in cell numbers; however, Gd³⁺ treatment had no significant effect on cell number. To confirm this finding, [³H] thymidine incorporation, an index of DNA synthesis and cell proliferation, was examined in cultured A10 VSMC. As shown in Figure 4, [³H] thymidine incorporation with or without LPA was significantly inhibited by SK treatment, meanwhile Gd³⁺ had no significant effect on [³H] thymidine incorporation.

Store-operated Ca²⁺ channel blocker on intracellular Ca²⁺

Intracellular Ca²⁺ concentration is a key factor for the control of vascular tone as well as cell proliferation [16]. To examine the effects of SOCC blockers on intracellular calcium mobilization, the effects of SK and Gd³⁺ on the various vasoactive agonists (such as LPA, ATP, NE, Ang II and ET-1)-evoked changes in [Ca²⁺]_i were examined in cultured A10 VSMC. Figure 5 demonstrated that SK and Gd³⁺ had no significant effect on basal [Ca²⁺]_i; however, LPA-induced increase in [Ca²⁺]_i was inhibited by both SK and Gd³⁺ in a concentration-dependent manner. But these antagonists had no significant effect on NE-induced increase in [Ca²⁺]_i (Fig. 6). For ET-1-induced elevation of [Ca²⁺]_i, SK demonstrated a concentration-dependent suppressive effect; whereas, Gd³⁺ had no significant action (Fig. 7). On the other hand, for Ang II-induced increase in [Ca²⁺]_i, both SK and Gd³⁺ exerted inhibitory effects.

Combination of store-operated Ca²⁺ channel blocker and voltage-dependent Ca²⁺ channel blocker on [Ca²⁺]_i

The mobilization of intracellular Ca²⁺ is elicited by Ca²⁺ influx through VOCC, SOCC, receptor operated Ca²⁺ channel (ROCC) and Ca²⁺ release from intracellular store [17]. To examine if the combination of SOCC and VOCC blockers is more effective than the individual agent,

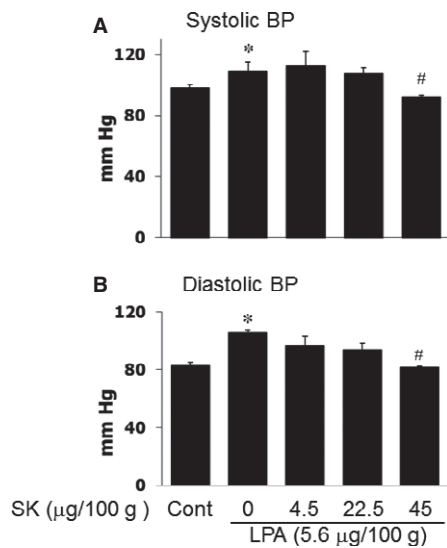


Fig. 2 The effect of SK on LPA-induced blood pressure elevation. The blood pressure was recorded from carotid artery prior to and after treatment by a microtip pressure transducer and computer programme Acq-knowledge for Windows 3.5. * $P < 0.05$ compared to control (Cont) values (basal line), # $P < 0.05$ compared to LPA injection group but without SK; $n = 6$.

the actions of SK and verapamil alone or in combination were examined on the LPA- or ATP-induced increase in $[Ca^{2+}]_i$ (Fig. 8). Both SK and verapamil suppressed the LPA- or ATP-induced elevation of $[Ca^{2+}]_i$ in A10 VSMC significantly; the combination of SK and verapamil demonstrated a stronger inhibitory effect; the LPA-induced response was inhibited by 85%, 51% and 95% following the treatment with SK, verapamil and combination of SK and verapamil respectively. For the ATP-induced response, the inhibitory actions were: 49% by SK, 62% by verapamil and 91% by SK plus verapamil. The reason for choosing LPA and ATP as agonists in this experiment is based on the fact that these agents are broadly used for Ca^{2+} mobilization in VSMC [14].

Discussion

Hypertension is a major cause of many cardiovascular diseases such as stroke, heart attack, atherosclerosis, chronic heart failure and kidney failure. These cardiovascular diseases affect more than 65 million Americans [18]. Voltage-operated Ca^{2+} channels blockers, including verapamil, nifedipine and diltiazem are useful agents for the reduction of blood pressure in the majority of population; however, some patients do not respond to these drugs. In addition, the major side effect of VOCC blockers is the increase of heart failure incidence as the heart beat is dependent on the Ca^{2+} influx from extracellular space through VOCC. Blocking these channels would reduce cardiac contractility and eventually lead to heart failure [15] and this effect may limit their clinical use in hypertension. On the other hand, Ca^{2+} influx through SOCC is triggered by hormones, growth factors and neurotransmitters. The route of Ca^{2+} influx through SOCC provides a minimal contribution to the beat-to-beat Ca^{2+} transients in the heart, but plays a major role in the increase of $[Ca^{2+}]_i$ in VSMC [19, 20]. Thus, SOCC blockers may have less side effects on cardiac contractility when used for the treatment of hypertensive patients.

It is known that Ang II, ET-1 and LPA are involved in the development of hypertension and atherosclerosis [3]. Voltage-operated Ca^{2+} channels blockers have only a partial inhibitory effect on ET-1-induced increase in $[Ca^{2+}]_i$ and cell contraction [21]. Arun *et al.* [22] have reported that verapamil and diltiazem do not inhibit the Ang II-induced contraction of vascular smooth muscle, indicating that L-type Ca^{2+} channels are not involved in this response. In an experiment using rat tail artery, the Ang II- and ET-1-induced contraction was found to be insensitive to L-type Ca^{2+} channel antagonist, nifedipine, but blocked by SK, indicating that SK may be useful for the treatment of hypertension in patients insensitive to L-type calcium channel blocker [23]. Bova *et al.* [24] demonstrated that norbormide, a vasoconstrictor agent for rat small artery, caused a concentration-dependent contraction, which was not sensitive to verapamil, but was almost completely inhibited by 30 μ M SK. These reports suggest that SOCC blockers may be superior to VOCC antagonists in certain cases.

In this study, we have observed for the first time that SOCC blocker SK reduced blood pressure significantly, and SK suppressed

Table 1 Effects of SK (SK&F 96365), verapamil and SK+verapamil combination on blood pressure in rats

Time	SK ($N = 4$)		Verapamil ($N = 5$)		Combination ($N = 8$)	
	SBP	DBP	SBP	DBP	SBP	DBP
0	129 \pm 11	85 \pm 6	124 \pm 7	87 \pm 4	134 \pm 4	89 \pm 2
10 sec.	113 \pm 4	74 \pm 10	98 \pm 3*	67 \pm 3*	96 \pm 2*	54 \pm 2* [†]
30 sec.	120 \pm 9	77 \pm 7	97 \pm 3*	66 \pm 4*	92 \pm 2*	47 \pm 2* [†]
1 min.	139 \pm 12	90 \pm 7	99 \pm 2*	72 \pm 1*	99 \pm 4*	61 \pm 4* [†]
5 min.	142 \pm 12	91 \pm 13	113 \pm 10	78 \pm 4	128 \pm 6	83 \pm 4

* $P < 0.05$ compared with the respective control value (before the treatment).

[†] $P < 0.05$ compared with respective values of verapamil (15 μ g/100 g) group and SK (45 μ g/100 g) group.

SBP: systolic blood pressure (mm Hg); DBP: diastolic blood pressure (mm Hg).

Fig. 3 The effect of SK or Gd³⁺ on cell proliferation in the presence or absence of LPA. Prior to treatment, the cells were cultured in serum free medium for 20 hrs, then different concentrations of SK were added to different wells and 10 min. later, LPA was added to all the wells except for control (Con) group. After continuing culture for 24 hrs, the cell number was counted. **P* < 0.05 compared to control value; #*P* < 0.05 compared to the group with LPA but no SK or Gd³⁺; *n* = 6.

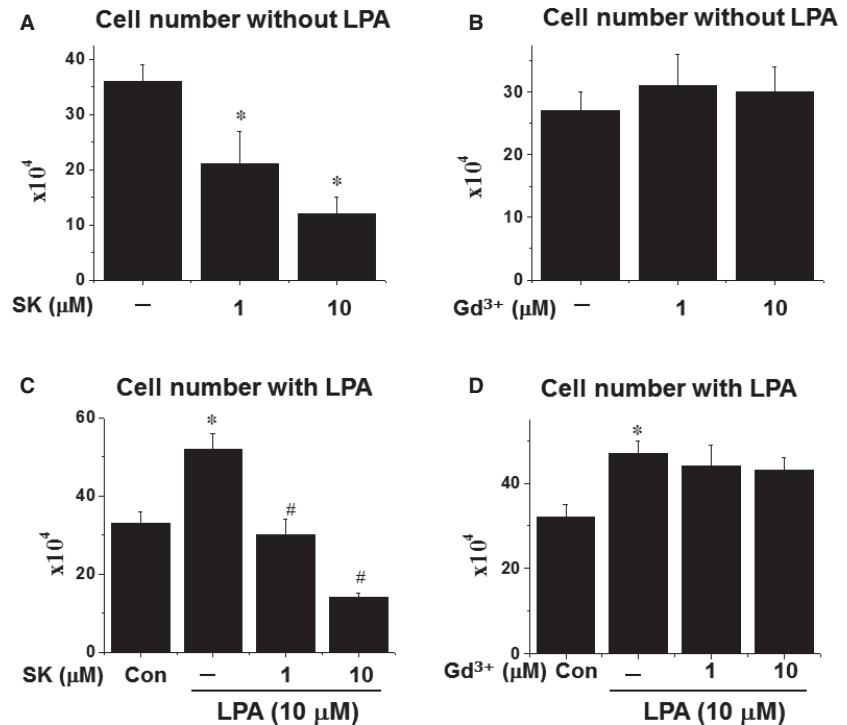
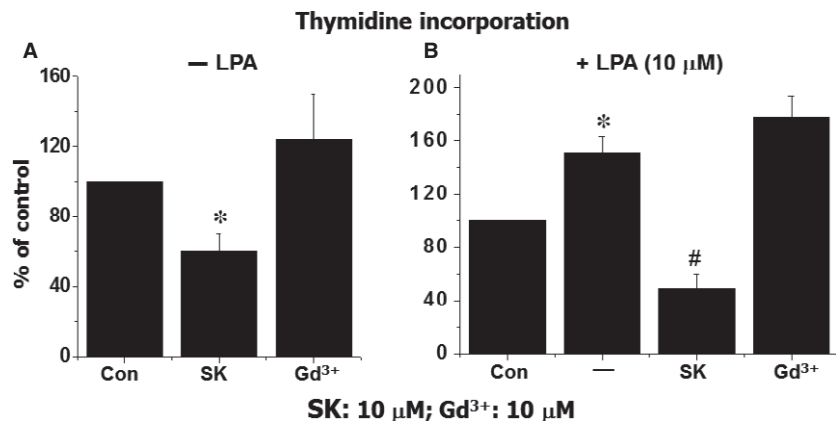


Fig. 4 The effect of SK or Gd³⁺ on DNA synthesis in the presence or absence of LPA. Prior to treatment, the cells were cultured in serum free medium for 20 hrs, then different concentration of SK or Gd³⁺ were added to different wells and 10 min. later, LPA was added to all the wells except for control (Con) group. After incubation for 4 hrs, ³H-thymidine was added, then reaction was terminated 20 hrs later. **P* < 0.05 compared to control value; #*P* < 0.05 compared to the group with LPA but no SK or Gd³⁺; *n* = 6.



the LPA-induced elevation of blood pressure. We have also shown the reduction of blood pressure by Gd³⁺, another SOCC blocker [9]. These results indicate that SOCC blockers may be used for the control of blood pressure. It should be noted that because the ET-1-induced pulmonary contraction was partially blocked by SK and Gd³⁺, different investigators [25, 26] have reported the role of SOCCs in the pathogenesis of pulmonary hypertension. However, some caution should be exercised about this viewpoint regarding the use of SOCC blockers in the treatment of pulmonary hypertension because Gd³⁺, unlike SK, was unable to block the ET-1-induced increase in [Ca²⁺]_i in VSMC. It is also pointed out that SK and Gd³⁺ produced toxic effects in the body [27], thus SOCC blockers with low toxic and less side effects need to be developed. Zuo *et al.* [28] have demonstrated that tyrosine

kinase inhibitors depressed the SOCC activity and it is possible to use tyrosine kinase inhibitors to reduce blood pressure. Accordingly, it would be interesting to investigate the effects of SK and Gd³⁺ on the tyrosine kinase activity in VSMC.

It has been shown that SOCCs play an important role in mitogenic response to growth factor in VSMC [20], and the effect of L-type Ca²⁺ channel blocker in this regard are controversial. Xiao *et al.* [29] reported that ET-1 caused a concentration-dependent increase in cell count and [³H] thymidine incorporation in VSMC; both nifedipine and SK inhibited these responses. However, Kawanabe *et al.* [30] reported that nifedipine had no effect on ET-1-induced augmentation of [Ca²⁺]_i in sustained phase and ET-1-stimulated cell proliferation. As SK significantly inhibited both changes in [Ca²⁺]_i and cell growth,

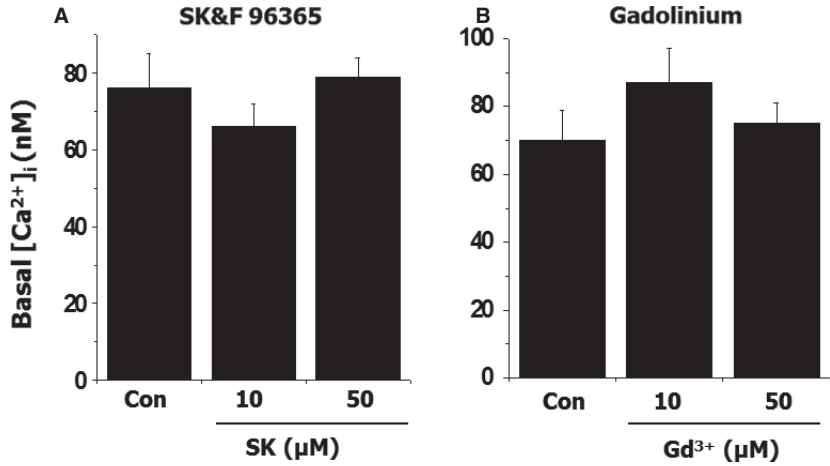


Fig. 5 The effect of SK or Gd³⁺ on intracellular free Ca²⁺ concentration in vascular smooth muscle cells. The cells were incubated with 10 μM Fura 2-AM in HEPES buffer for 30 min. The intensity of fluorescence before and after treatment was recorded by a SLM DMX -1100 dual wavelength spectrofluorometer; *n* = 6.

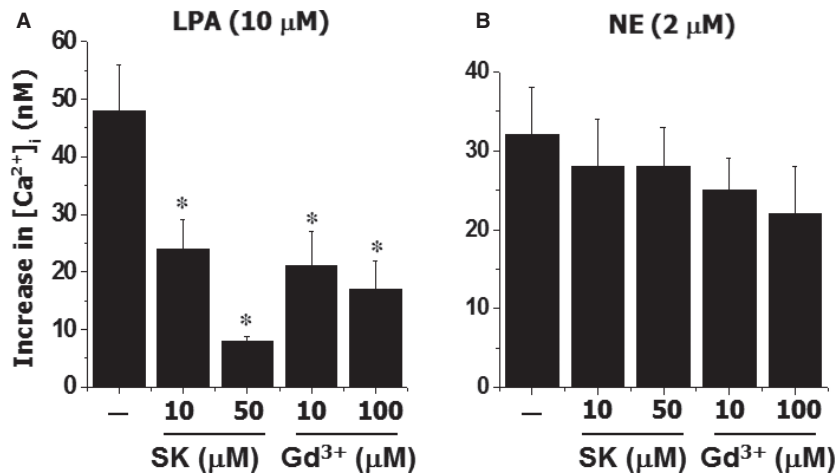


Fig. 6 The effect of SK or Gd³⁺ on LPA or NE-induced change in intracellular free Ca²⁺ concentration in vascular smooth muscle cells. The cells labelled with Fura 2-AM were incubated with different concentrations of SK or Gd³⁺ for 30 sec. prior to the challenging with LPA or NE. **P* < 0.05 compared to control value with LPA or NE alone; *n* = 6.

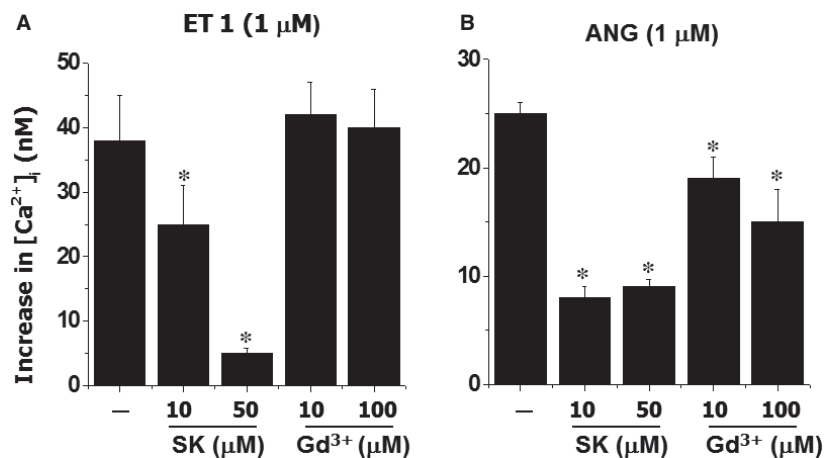
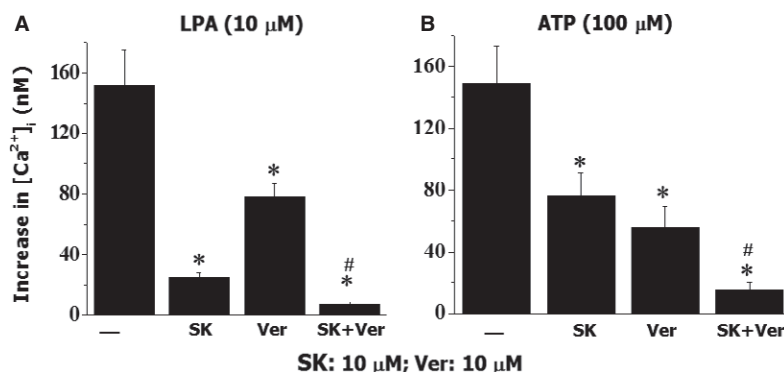


Fig. 7 The effect of SK or Gd³⁺ on ET 1 or ANG-induced change in intracellular free Ca²⁺ concentration in vascular smooth muscle cells. The cells labelled with Fura 2-AM were incubated with different concentrations of SK or Gd³⁺ for 30 sec. prior to the challenging with ET 1 or ANG. **P* < 0.05 compared to control value with ET 1 or ANG alone; *n* = 6.

it appears that SOCC inhibitors may prevent remodelling of VSMC. This view is consistent with the observations of Leung *et al.* [20], who have shown the role of SOCC in VSMC proliferation. Although

both SK and Gd³⁺ produce a reduction in blood pressure, these agents were found to exert different actions with respect to VSMC proliferation and [Ca²⁺]_i changes. In this regard, it should be noted

Fig. 8 The effect of SK, verapamil (Ver) or SK + Ver on LPA- or ATP-induced changes in intracellular free Ca^{2+} concentration in vascular smooth muscle cells. The cells labelled with Fura 2-AM were incubated with SK, Ver or SK+Ver for 30 sec. prior to the challenging with LPA or ATP. * $P < 0.05$ compared to control value with LPA or ATP alone without inhibitors, # $P < 0.05$ compared to other groups; $n = 6$.



that there are three important components in SOCC, namely, a stromal interaction molecule (which is Ca^{2+} sensor), a pore forming protein (which allows Ca^{2+} influx) and a transient receptor (which regulates the function of SOCC) [31–33]. As all three components of SOCC are important in the control of Ca^{2+} influx in VSMC [32, 33], it is likely that the difference in the actions of SK and Gd^{3+} on cell proliferation and $[\text{Ca}^{2+}]_i$ observed in this study may be as a result of their effects on different targeting sites in SOCC. Because the expression of the transient receptor component of SOCC is up-regulated in the vasculature of hypertensive rats [33], future studies should be carried out to investigate the effects of different SOCC blockers on this component.

It is noteworthy that both SK and Gd^{3+} have been shown to block Ca^{2+} influx through SOCC in VSMC and their inhibitory effects are agonist and cell-type dependent. For instance, in arteriolar smooth muscle cells [34, 35], Gd^{3+} produced a concentration-dependent inhibition of SOCC-mediated Ca^{2+} entry; however, the effects of SK on cyclopiazonic acid and thapsigargin depletion-induced SOCC activation were different. In our study, both SK and Gd^{3+} inhibited LPA-induced alteration of $[\text{Ca}^{2+}]_i$, as observed previously [7]; however, it had no significant effect on the increase in $[\text{Ca}^{2+}]_i$ induced by NE. This is because NE evoked the increase in $[\text{Ca}^{2+}]_i$ mainly by cell membrane depolarization [36] and thus VOCC were activated [37]. On the other hand, ET-1 has been shown to increase $[\text{Ca}^{2+}]_i$ through SOCC which was blocked by SK [38]; the results in this study are consistent with this observation. Furthermore, Gd^{3+} produced no effect on ET-1-induced $[\text{Ca}^{2+}]_i$ change in our experiments, indicating the distinct effect of these two blockers on SOCC in VSMC. Taken together, LPA, Ang II, NE and ET-1, were all observed to increase in $[\text{Ca}^{2+}]_i$, but the mechanisms seemed to be different.

We have shown that both SK and verapamil, when used in combination, produced a synergistic effect on LPA- and ATP-induced $[\text{Ca}^{2+}]_i$ increase in VSMC. Other investigators have reported an additive inhibitory effect on ET-1-induced vasoconstriction in cerebral arteries by using SK and nifedipine (VOCC blocker) [10]. These observations suggest that the use of SOCC blockers in combination with VOCC inhibitor may be more effective in lowering the blood pressure. However, extensive studies by using other agonists including ET-1, Ang II and NE are required for establishing the beneficial effects of the combination of SOCC and VOCC blockers. Another limitation of this study is concerned with the specificity of SK and Gd^{3+} on SOCC. In this regard, it has been reported that these two agents in higher concentrations may also inhibit ROCC and other Ca^{2+} channels [39–41]. Thus more specific chemical inhibitors or specific antibodies are needed for further study. Nevertheless, on the basis of the results described in this study, it is suggested that SOCC blockers may be considered as a new category of antihypertensive agents and SOCC may be a molecular target for drug development.

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Conflicts of interest

The authors declare no conflict of interest with any grant funding agency.

References

- Kohan DE, Rossi NF, Inscho EW, *et al*. Regulation of blood pressure and salt homeostasis by endothelin. *Physiol Rev*. 2011; 91: 1–77.
- Unger T, Paulis L, Sica DA. Therapeutic perspectives in hypertension: novel means for rennin-angiotensin-aldosterone system modulation and emerging device-based approaches. *Eur Heart J*. 2011; 32: 2739–47.
- Xu YJ, Aziz OA, Bhugra P, *et al*. Potential role of lysophosphatidic acid in hypertension and atherosclerosis. *Can J Cardiol*. 2003; 19: 1525–36.
- Schiffirin EL. Vascular protection with newer antihypertensive agents. *J Hypertens Suppl*. 1998; 16: S25–9.
- Schiffirin EL. Intracellular signal transduction for vasoactive peptides in hypertension. *Can J Physiol Pharmacol*. 1994; 72: 954–62.

6. Elliott WJ, Ram CVS. Calcium channel blockers. *J Clin Hyperten*. 2011; 13: 687–9.
7. Hill-Eubanks DC, Werner ME, Heppner TJ, et al. Calcium signaling in smooth muscle. *Cold Spring Harb Perspect Biol*. 2011; 3: a004549.
8. Nauli SM, Kawanabe Y. Involvement of extracellular Ca²⁺ influx through voltage-independent Ca²⁺ channels in endothelin-1 function. *Cell Signal*. 2005; 17: 911–6.
9. Poburko D, Lhote P, Szado T, et al. Basal calcium entry in vascular smooth muscle. *Eur J Pharmacol*. 2004; 28: 19–29.
10. Mamo YA, Angus JA, Ziogas J, et al. The role of voltage-operated and non-voltage-operated calcium channels in endothelin-induced vasoconstriction of rat cerebral arteries. *Eur J Pharmacol*. 2014; 742: 65–73.
11. Houssaini A, Abid S, Mouraret N, et al. Rapamycin reverses pulmonary artery smooth muscle cell proliferation in pulmonary hypertension. *Am J Respir Cell Mol Biol*. 2013; 48: 568–77.
12. Xu YJ, Rathi SS, Zhang M, et al. Mechanism of the positive inotropic effect of lysophosphatidic acid in rat heart. *Cardiovasc Pharmacol Ther*. 2002; 7: 109–15.
13. Xu YJ, Rathi SS, Chapman DC, et al. Mechanisms of lysophosphatidic acid-induced DNA synthesis in vascular smooth muscle cells. *J Cardiovasc Pharmacol*. 2003; 41: 381–7.
14. Xu YJ, Saini HK, Cheema SK, et al. Mechanisms of lysophosphatidic acid-induced increase in intracellular calcium in vascular smooth muscle cells. *Cell Calcium*. 2005; 38: 569–79.
15. Colovina VA. Cell proliferation in associated with enhanced capacitative Ca²⁺ entry in human arterial myocytes. *Am J Physiol (Cell Physiol)*. 1999; 277: C343–9.
16. Jackson WF. Ion channels and vascular tone. *Hypertension*. 2000; 35: 173–8.
17. DeSimone ME, Crowe A. Nonpharmacological approaches in the management of hypertension. *J Am Acad Nurse Pract*. 2009; 21: 189–96.
18. Colucci WS, Fifer MA, Lorell BH, et al. Calcium channel blockers in congestive heart failure: theoretic consideration and clinical experience. *Am J Med*. 1985; 78: 9–17.
19. Guibert C, Ducret T, Savineau JP. Voltage-independent calcium influx in smooth muscle. *Prog Biophys Mol Biol*. 2008; 98: 10–23.
20. Leung FP, Yung LM, Yao X, et al. Store-operated calcium entry in vascular smooth muscle. *Br J Pharmacol*. 2008; 153: 846–57.
21. McNair LL, Salamanca DA, Khalil RA. Endothelin-1 promotes Ca²⁺ antagonist-insensitive coronary smooth muscle contraction via activation of epsilon-protein kinase C. *Hypertension*. 2004; 43: 897–904.
22. Arun KHS, Kaul CL, Ramarao P. AT1 receptors and L-type calcium channels: functional coupling in supersensitivity to angiotensin II in diabetic rats. *Cardiovas Res*. 2005; 65: 374–86.
23. Jiang Y, Triggler CR. Lack of involvement of endothelin-1 in angiotensin II-induced contraction of the isolated rat tail artery. *Brit J Pharmacol*. 2000; 131: 1055–64.
24. Bova S, Trevisi L, Cima S, et al. Signaling mechanisms for selective vasoconstrictor effect of norbormide on the rat small arteries. *J Pharmacol Experim Therapeu*. 2000; 296: 458–63.
25. Firth AL, Won JY, Park WS. Regulation of Ca²⁺ signaling in pulmonary hypertension. *Korean J Physiol Pharmacol*. 2013; 17: 1–8.
26. Yang K, Lu W, Jiang Q, et al. PPAP_γ inhibits hypoxia-induced store-operated calcium entry by suppressing caveolin-1. *Am J Respir Cell Mol Biol*. 2015; Doi:10.1165/rcmb.2015-00020C.
27. Bernstein EJ, Schmidt-Lauber C, Kay J. Nephrogenic system fibrosis: a system fibrosis resulting from gadolinium exposure. *Best Pract Res Lin Rheumatol*. 2012; 26: 489–503.
28. Zuo WL, Du JY, Huang JH, et al. Tyrosine phosphorylation modulates store-operated calcium entry in cultured rat epididymal basal cells. *J Cell Physiol*. 2011; 226: 1069–73.
29. Xiao G-N, Guan Y-Y, He H. Effect of Cl⁻ channels on endothelin-1-induced proliferation of rat vascular smooth muscle cells. *Life Sci*. 2002; 70: 2233–41.
30. Kawanabe Y, Hashimoto N, Masaki T. Characterization of Ca²⁺ channels involved in endothelin-1-induced contraction of rabbit basilar artery. *J Cardiovas Pharmacol*. 2002; 40: 438–47.
31. Giachini FRC, Chiao C-W, Carneiro FS, et al. Increased activation of stromal inter-action molecular-1/Orai-1 in aorta from hypertensive rats: a novel insight into vascular dysfunction. *Hypertension*. 2009; 53: 409–16.
32. Eder P, Poteser M, Groschner K. TRPC3: a multifunctional, pore-forming signaling molecule. *Handb Exp Pharmacol*. 2007; 179: 77–92.
33. Liu D, Yang D, He H, et al. Increased transient receptor potential canonical type 3 channels in vasculature from hypertensive rats. *Hypertension*. 2009; 53: 70–6.
34. Flemming R, Xu SZ, Beech DJ. Pharmacological profile of store-operated channels in cerebral arteriolar smooth muscle cells. *Br J Pharmacol*. 2003; 139: 955–65.
35. Nilsson H, Videbaek LM, Toma C, et al. Role of intracellular calcium for norepinephrine-induced depolarization in rat mesenteric small arteries. *J Vasc Res*. 1998; 35: 36–44.
36. Yousof MH, Williams KI, Oriowo MA. Source(s) of activator calcium for norepinephrine-induced vasoconstriction in the perfused rabbit isolated ovarian vascular bed: a role for tyrosine kinase. *Gen Pharmacol*. 1999; 32: 563–79.
37. Liskova S, Petrova M, Karen P, et al. Contribution of Ca²⁺-dependent Cl⁻ channels to norepinephrine-induced contraction of femoral artery is replaced by increasing EDCF contribution during ageing. *Biomed Res Int*. 2014; 2014: 289361.
38. McFadzean I, Gibson A. The developing relationship between receptor-operated and store-operated calcium channels in smooth muscle. *Br J Pharmacol*. 2002; 135: 1–13.
39. Singh A, Hildebrand ME, Garcia E, et al. The transient receptor potential channel antagonist SKF96365 is a potent blocker of low-voltage-activated T-type calcium channels. *Bt J Pharmacol*. 2010; 160: 1464–75.
40. Song M, Chen D, Yu SP. The TRPC channel blocker SKF96365 inhibits glioblastoma cell growth by enhancing reverse mode of the Na⁺/Ca²⁺ exchanger and increasing intracellular Ca²⁺. *Br J Pharmacol*. 2014; 171: 3432–47.
41. Chen KH, Liu H, Yang L, et al. SKF-96365 strongly inhibits voltage-gated sodium current in rat ventricular myocytes. *Pflugers Arch*. 2014; 467: 1227–36.