

Amine derivatives of furocoumarin induce melanogenesis by activating Akt/GSK-3 β / β -catenin signal pathway

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Background: Melanogenesis, or the biosynthesis of melanin, plays a critical role in the pigmentation of skin, hair, and eyes. Reduced melanogenesis may lead to depigmentation conditions such as vitiligo. Psoralen, a furocoumarin derivative, is closely associated with melanogenesis, and its derivative 8-methoxypsoralen is used in psoralen and ultraviolet A therapy for pigmentation disorders. In a previous study, we synthesized a new series of amine derivatives of furocoumarin, of which 5-(morpholinomethyl)-3-phenyl-7*H*-furo[3,2-*g*]chromen-7-one (encoded as D206008) showed a remarkable melanogenic effect in B16 murine cells.

Methods: In this study, we examined the effects of D206008 on the melanogenesis-related pathways in B16 cells. D206008 increased melanin production and tyrosinase (TYR) activity, as well as the mRNA and protein expression levels of the melanogenic enzymes TYR, TRP-1 and TRP-2, and the melanogenesis-related transcription factor microphthalmia-associated transcription factor (MITF) in a dose-dependent (0–100 μ M) and time-dependent (0–48 hours) manner.

Results: Mechanistically, D206008 inhibited β -catenin degradation by enhancing the phosphorylation of Akt and glycogen synthase kinase-3 β (GSK-3 β), which increased the accumulation of β -catenin in the cytoplasm. Nuclear translocation of β -catenin also increased in response to D206008 treatment.

Conclusion: Taken together, these data indicate that D206008 promotes melanin synthesis by stimulating the nuclear translocation of β -catenin, which activates MITF transcription and eventually melanogenesis.

Keywords: amine derivatives of furocoumarin, melanogenesis, Akt/GSK-3 β / β -catenin

Introduction

Pigmentation of skin, hair, and eyes is regulated by melanin synthesis.^{1,2} Abnormal pigmentation, either due to dysregulated melanin synthesis or melanocyte development, results in esthetic imperfections. Any impairment that decreases melanin biosynthesis either targets the melanocyte number or function and may lead to depigmentation conditions such as vitiligo.^{3,4} Melanogenesis occurs in specialized vesicles called the melanosomes within the melanocytes.^{5,6} Three structurally related enzymes, tyrosinase (TYR) and the tyrosinase-related proteins 1 and 2 (TRP-1 and TRP-2), are involved in melanin synthesis. These proteins are transcriptionally regulated by the microphthalmia-associated transcription factor (MITF), which harbors a basic helix–loop–helix leucine zipper structure⁷ and is a key regulator of melanocyte development and melanogenesis.⁸ Several signal transduction pathways are involved in melanogenesis via the regulation of MITF and TYR expression, such as the key Akt/glycogen synthase kinase-3 β (GSK-3 β)/ β -catenin pathway.⁹ Activation by the Frizzled receptor represses GSK-3 β ,

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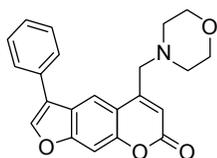
which leads to the accumulation of β -catenin in cytoplasm¹⁰ and eventual nuclear translocation, finally resulting in the transcriptional activation of MITF.^{11,12} Activation of the protein kinase Akt mediates phosphorylation of GSK-3 β , which induces melanin synthesis via upregulation of MITF.

Natural products or plant extracts have been used for centuries for the treatment of pigmentation disorders. *Psoralea corylifolia* L. is used for repigmentation in traditional Uyghur medicine,^{13–15} and several psoralen compounds that have been isolated from this plant, for instance, 8-methoxypsoralen (8-MOP) and 5-methoxypsoralen (5-MOP),¹⁶ are used with ultraviolet radiation in repigmentation therapies.¹⁷ Although these compounds have good melanogenesis potential, they have several side effects such as increased risk of skin cancer and hepatic steatosis.^{18,19} It is important therefore to isolate/synthesize novel plant-derived compounds with better melanogenesis activity and lower toxicity. Recently, we synthesized a new series of amine derivatives of furocoumarin and evaluated their melanogenic effects in B16 murine cells, relative to 8-MOP. One of derivatives 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one (D206008) (Figure 1) resulted in higher melanogenesis and TYR activity than 8-MOP in the B16 cells without being cytotoxic. The aim of the present study was to investigate the molecular mechanisms underlying the melanogenic activity of D206008.

Materials and methods

Materials

D206008 was synthesized by The Key Laboratory of Plant Resource Sand Chemistry of Arid Zone, Xinjiang Technical Institute of Physics and Chemistry, Chinese Academy of Sciences:²⁰ Yield 42%, light yellow solid, m.p. 166°C–167°C; purity for HPLC =98.3%; ¹H NMR (400 MHz, CDCl₃) δ 8.39 (s, 1H), 7.85 (s, 1H), 7.66 (d, $J=7.1$ Hz, 2H), 7.57–7.50 (m, 3H), 7.45 (t, $J=7.3$ Hz, 1H), 6.53 (s, 1H), 3.80–3.73 (m, 4H), 3.71 (s, 2H), 2.63–2.56 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 161.24, 157.18, 152.21, 142.99, 131.22, 129.83, 129.34, 128.23, 127.64, 123.95, 122.42, 116.68, 115.53, 113.81, 100.31, 67.14, 60.15, 53.87; IR (KBr) ν : 2,926, 1,725, 1,633, 1,575, 1,455, 1,384, 1,320, 1,263, 1,112, 986, 866 cm⁻¹;



**5-(morpholinomethyl)-3-phenyl-7H-furo
[3,2-g]chromen-7-one**

Figure 1 The structure of D206008.

HRMS (ESI) calcd for C₂₂H₂₀NO₄ [M+H]⁺ 362.1387, found 362.1390. D206008 was dissolved in DMSO and stored at –20°C as a stock solution (50 mM). L-3-(3,4-dihydroxyphenyl) alanine (L-DOPA) (CAS No 59-92-7) was obtained from Generay Biotech (Shanghai, China), 8-MOP (CAS:298-81-7) was obtained from Sigma Aldrich (Milan, Italy), and Akt inhibitor IV was obtained from EMD Biosciences (La Jolla, CA, USA). 6-Bromindirubin-3'-oxime (BIO) was purchased from AMQUAR Biology (Shanghai, China). Phosphor-Akt (Thr308, #5106), Akt (#5373), phosphor-GSK-3 β (Ser9, #9323), GSK-3 β (#9832), phosphor β -catenin (Ser 33,37,41#9561), phospho- β -catenin (Ser675, #4176), β -catenin (#8480), and NUP98 (#2598) were obtained from Cell Signaling Technology (Beverly, MA, USA), and the antibodies against β -actin and tyrosinase (C-19), TRP-1 (H-90), and TRP-2 (H-150) were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). Anti-MITF was obtained from Chemicon (Temecula, CA, USA). Goat anti-rabbit (BA1054), goat anti-mouse (BA1050), rabbit anti-goat (BA1060), and anti-GADPH were obtained from BOSTER Biological Technology Co Ltd. (Wuhan, China).

B16 cell culture

The B16 murine melanoma cells (Xiangf Bio, Shanghai, China) were maintained in DMEM (Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% FBS (Thermo Fisher Scientific), penicillin G (100 U/mL), and streptomycin (100 μ g/mL) (Gibco-BRL, Grand Island, NY, USA) and incubated in 5% CO₂ at 37°C.

Cell viability assay

Viability of B16 cells was measured by CCK-8 solution (Promega, Madison, MA, USA) assay. After D206008 treatments (0–100 μ M), the B16 cells were incubated for 48 hours at 37°C and the cell viability of each treated sample was measured in triplicate using CCK-8 assay kit according to the manufacturer's instructions. Then culture medium was removed, and 10 μ L of CCK solution was added into each well of the 96-well plates. After incubation for 2 hours at 37°C, the absorbance was read at a wavelength of 450 nm using a Spectra Max M5 (Molecular Devices Company, San Diego, CA, USA). The same volume of cells without treatment was used as a blank control. All treatments were performed in triplicate, and all experiments were repeated three times.

TYR activity

The cellular TYR activity proceeded as mentioned before method with slight modifications.⁹ B16 cells were incubated

with various concentrations (0–100 μM) of D206008 in DMEM containing 10% FBS for 24 hours; the cells were washed twice with cold PBS, and then lysed in PBS buffer containing 1% sodium deoxycholate + 1% TritonX-100 solution. Next, 3 μL supernatants of cell extracts were used to measure the total protein content by BCA kit assay; the remaining part was used to measure TYR activity 10 μL , 10 mM L-DOPA and 90 μL test supernatants plated in 96-wellplates, then incubated at 37°C for 30 minutes. The OD of supernatants was measured at 490 nm using a microplate reader.

Melanin content measurement

The melanin release of B16 cells was measured using a previously described method with slight modifications.²¹ Briefly, the cells were treated with various concentrations of D206008 in DMEM containing 10% FBS for 48 hours, washed with ice-cold PBS twice, and then lysed with RIPA lysis buffer containing 1 M PNPP, 1 M sodium NaF, 10 mM PMSF, 100 mM benzamide, 100 mM DTT, and 200 mM sodium orthovanadate. Three microliters of supernatants of cell extracts were used to measure the total protein content by BCA kit assay and then dissolved in 1 mM NaOH at 80°C for 1 hour, which supernatants was measured at 405 nm.

Western blot analysis

B16 cells were treated with or without D206008. The protein was prepared from B16 cells according to the method mentioned above. The concentration of protein was measured by BCA assay kit. Proteins per lane were separated by 10% SDS-PAGE. Proteins were transferred to a polyvinylidene fluoride membrane. Membranes were blocked with 5% milk or 5% BSA and incubated overnight at 4°C with appropriate antibodies. And then, the membranes were further incubated for 2 hours with the corresponding secondary antibody. Following incubation, the proteins of membranes were detected using enhanced chemiluminescence (ECL) Western blotting detection reagents and photographed with the ChemiDoc MP Imaging System (Bio-Rad Laboratories Inc., Hercules, CA, USA). All determinations were performed three times.

Primer sequence in quantitative real-time PCR

Total cellular RNA was prepared from B16 cells treated with D206008 and isolated with TRIzol reagent in accordance with the manufacturer's instructions. Quantitative PCR was performed to determine the expression

of target genes. The primers were as follow: forward 5'-GTCGTCACCCTGAAAATCCTAACT-3' and reverse 5'-CATCGCATAAAACCTGATGGC-3' for Tyr (111 bp); forward 5'-ACCCATTTGTCTCCCAATGA-3' and reverse 5'-GTCCAATAGGTGCGTTTTCC-3' for TRP-1 (130 bp); forward 5'-TACCATCTGTTGTGGCTGGA-3' and reverse 5'-TGGGTCATCTTGCTG-3' for TRP-2 (147 bp); forward 5'-AGTACAGGAGCTGGAGATG-3' and reverse 5'-GTGAGATCCAGAGTTGTCGT-3' for MITF (181 bp). β -Actin was used as an internal control in all cases, and its primer sequence was as follow: forward 5'-TCAAGATCATTGCTCCTCCTG-3' and reverse 5'-CTGCTTGCTGATCCA-CATCTG-3' (59 bp). All determinations were performed three times. The reaction parameters were 95°C for 10 minutes, followed by 40 cycles of 15 seconds at 95°C for melting and 1 minute at 60°C for annealing. The real-time PCR was performed using Applied Biosystems 7300 PCR machine (Applied Bioscience, Foster City, CA, USA). The results were normalized to the controls.

Statistics

All results were expressed as mean \pm SD and standard error. Statistical analysis was performed with one-way ANOVA followed by Tukey's multiple comparison test. Statistical analysis was performed using GraphPad Prism 6 (La Jolla, CA, USA). *P*-values <0.05 were considered statistically significant.

Results

Effect of D206008 on B16 cell viability

To determine the cytotoxic effects of D206008 on cell viability, B16 melanoma cells were incubated with different concentrations of D206008 (0–100 μM) for 48 hours, and their viability was measured using the CCK-8 assay. As shown in Figure 2, D206008 had no obvious cytotoxic effects on B16 cells.

Effects of D206008 on TYR activity and melanin content in B16 cells

To determine the effects of D206008 on pigmentation, we examined the melanin content of B16 cells treated with different concentrations (0–100 μM) of D206008 for 48 hours, and with 100 μM for varying durations (0–48 hours). As shown in Figure 3, D206008 increased melanin content in B16 cells in a dose-dependent (Figure 3A) and time-dependent manner (Figure 3C). Similarly, the activity of cellular TYR also increased significantly in a concentration-dependent manner following incubation with D206008 (Figure 3B).

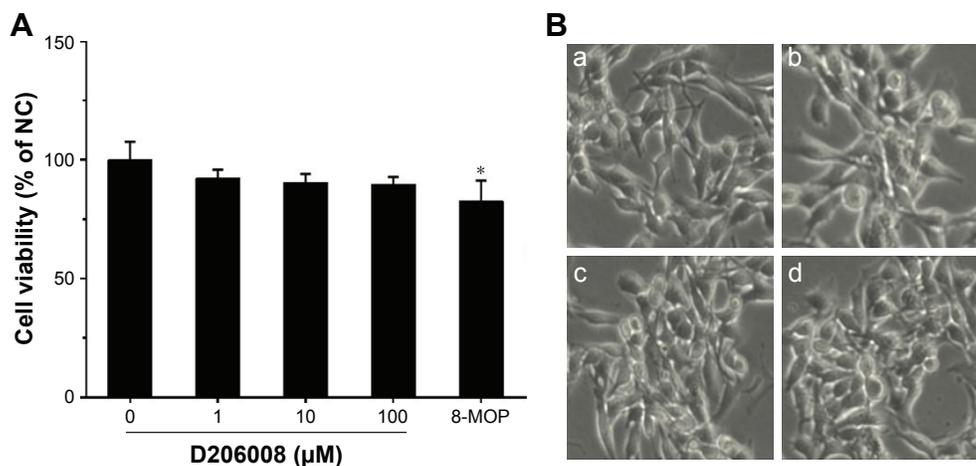


Figure 2 Effect of D206008 on cell viability and morphology.

Notes: (A) The effect of different concentrations (0–100 μM) of D206008 on B16 melanoma cell viability was measured by CCK-8 assay. The data are expressed as the mean ± SD (* $P < 0.05$ compared to DMSO vehicle group). (B) Morphological changes in D206008-treated B16 cells at 200× magnification. (a) 0.1% DMSO, (b) 1 μM, (c) 10 μM, and (d) 100 μM D206008.

Abbreviations: D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; 8-MOP, 8-methoxypsoralen.

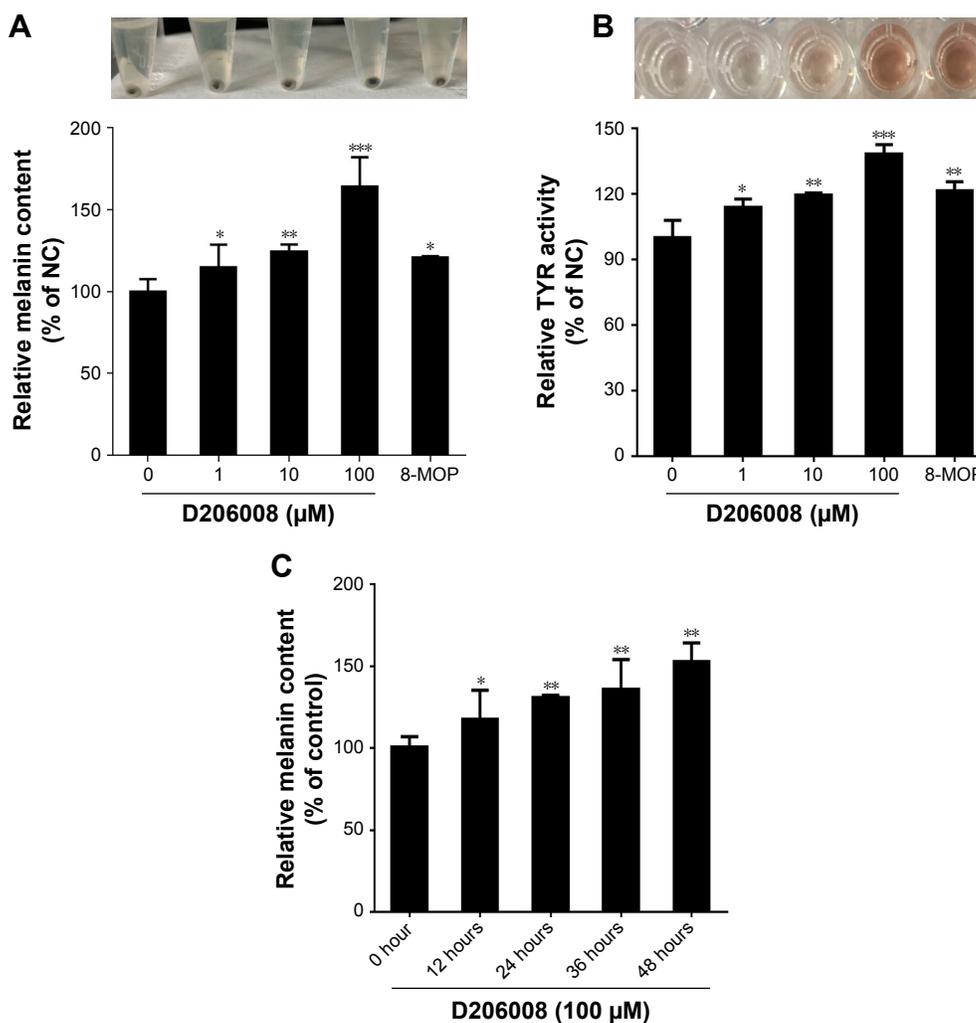


Figure 3 Effect of D206008 on melanin content and TYR activity in B16 melanoma cells.

Notes: (A) Melanin content of cells treated with different concentrations of D206008 for 48 hours. (B) TYR activity in the similarly treated B16 melanoma cells. (C) Effect of 100 μM D206008 treatment for varying durations (0–48 hours) on melanin content. DMSO (0.1%) was used as the vehicle control and 8-MOP (100 μM) as the positive control. Results are expressed as mean ± SD. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared to DMSO (0.1%) group.

Abbreviations: D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; 8-MOP, 8-methoxypsoralen; TYR, tyrosinase.

Effects of D206008 on the expression level of MITF and TYR family

The effect of D206008 on the expression of melanogenic genes was evaluated by real-time PCR and Western blotting. As shown in Figure 4C, the mRNA levels of MITF and its

downstream TYR family genes such as TYR, TRP-1, and TRP-2 were increased upon D206008 treatment. The levels of MITF, TYR, TRP-1, and TRP-2 proteins were also significantly increased by D206008 treatment in a dose-dependent (Figure 4A) and time-dependent (Figure 4B) manner in

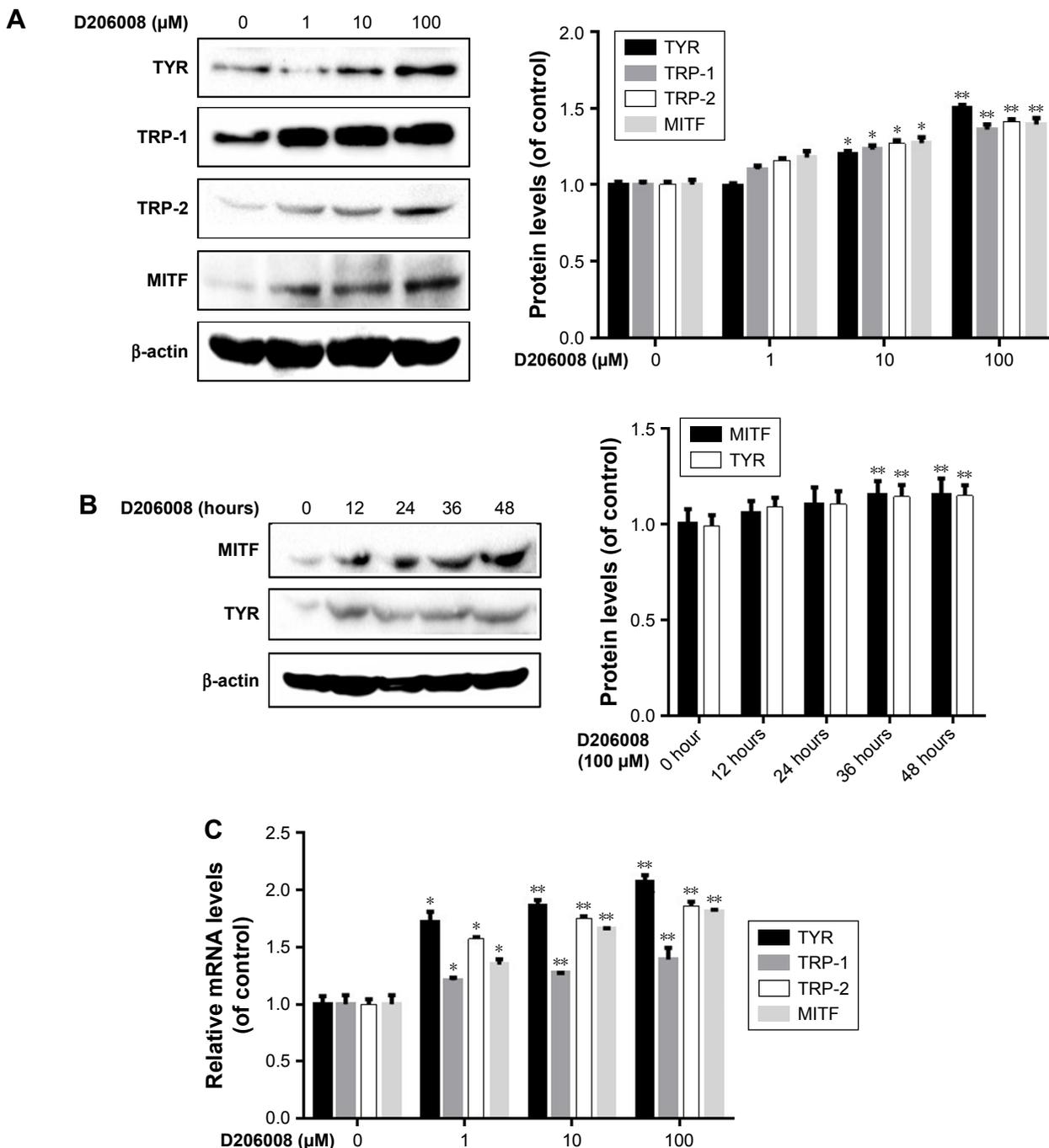


Figure 4 Effect of D206008 on the expression of melanogenic genes.

Notes: (A) The cells were treated with different concentrations (0–100 μM) of D206008 for 48 hours and the levels of MITF, TYR, TRP-1, and TRP-2 proteins were evaluated by Western blotting. (B) Cells were treated with 100 μM D206008 for varying durations (0–48 hours) and the levels of melanogenic factors were evaluated by Western blotting. The band densities of proteins were measured by the Quantity One program, and the expression levels of each protein was normalized against β-actin expression level. (C) Cells were treated with different concentrations (0–100 μM) of D206008 for 48 hours and expression of melanogenic genes was analyzed by RT-PCR. The data were expressed as mean ± SD and analyzed by one-way ANOVA followed by Tukey's test. * $P < 0.05$, ** $P < 0.01$, compared to control group. All experiments were performed three times.

Abbreviations: D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; MITF, microphthalmia-associated transcription factor; TYR, tyrosinase; RT-PCR, real-time PCR.

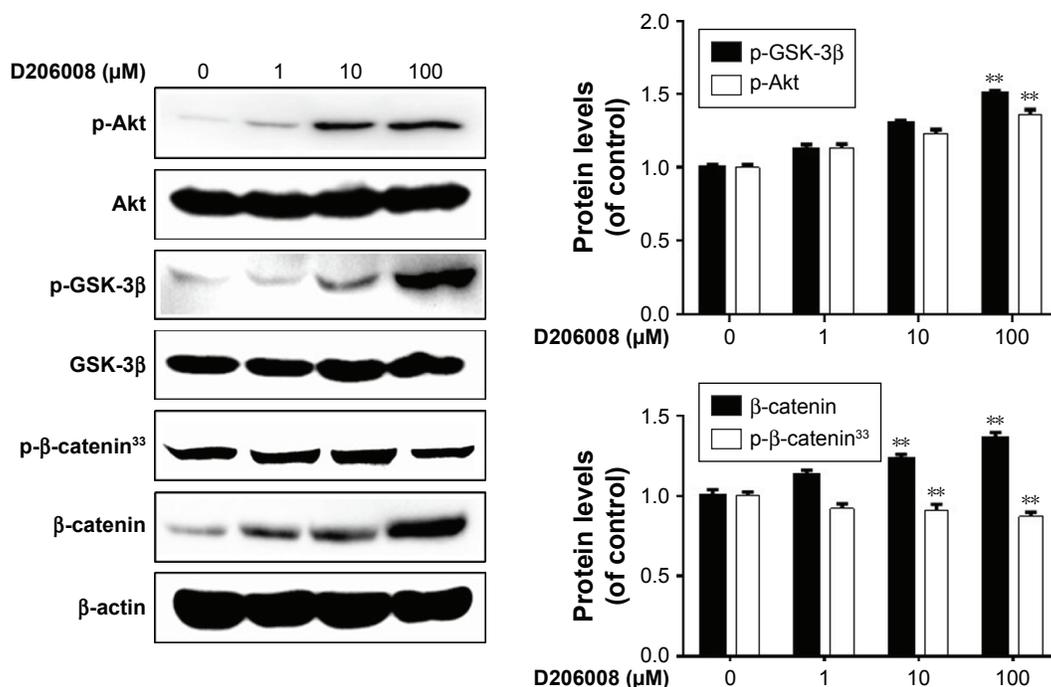
B16 cells. These results indicated that D206008-induced pigmentation occurred via the upregulation of MITF and TYR family genes.

D206008 induces melanogenesis by activating the Akt/GSK-3 β / β -catenin pathway

To determine the underlying mechanism of D206008-induced melanogenesis, we investigated the possible role of

the Akt/GSK-3 β / β -catenin pathway. B16 cells treated with different concentrations of D206008 showed higher levels of p-Akt and p-GSK-3 β in a concentration-dependent manner. In contrast, p- β -catenin-ser33 levels decreased and the intracellular β -catenin content increased in a dose-dependent manner (Figure 5A). Furthermore, the nuclear β -catenin content was significantly increased in the D206008-treated cells relative to the untreated cells, while the cytoplasmic protein expression did not change (Figure 5B).

A



B

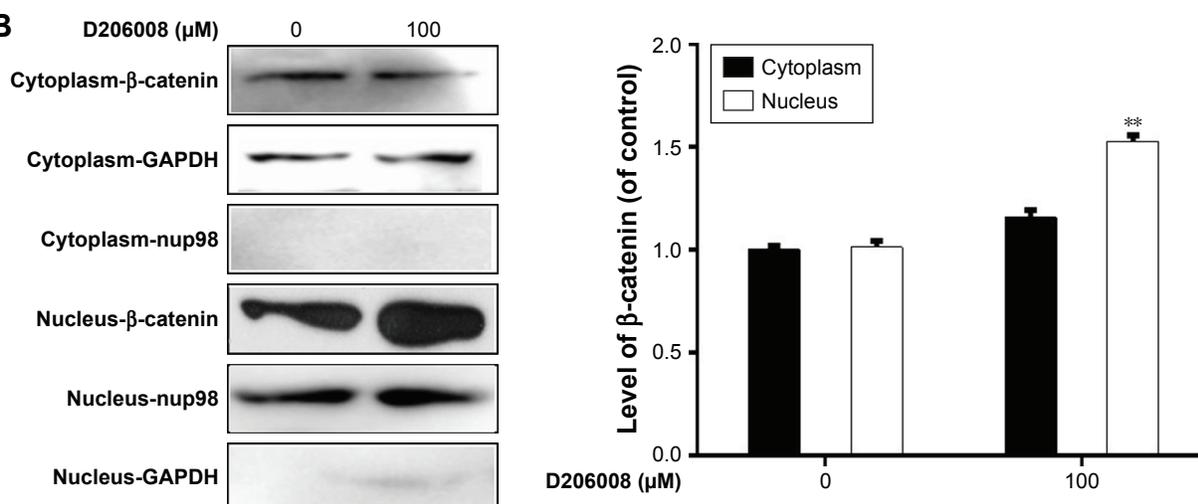


Figure 5 D206008 activates β -catenin signaling pathway via the phosphorylation of GSK-3 β and Akt.

Notes: (A) Cells were treated with different concentrations (0–100 μ M) of D206008 for 48 hours, and the levels of p-Akt, p-GSK-3 β , p- β -catenin, and β -catenin proteins were analyzed relative to β -actin expression. (B) B16 cells treated with either vehicle (0.1% DMSO) or D206008 (100 μ M) for 48 hours were analyzed for the levels of nuclear and cytoplasmic β -catenin. Cytoplasmic protein levels were normalized against GAPDH and nuclear protein against nup98. The band densities of proteins were measured by the Quantity One program. The data are shown as mean \pm SD and analyzed by one-way ANOVA followed by Tukey's test. ** P < 0.01, compared to control group. All experiments were performed three times.

Abbreviations: D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; GSK-3 β , glycogen synthase kinase-3 β .

The effect of GSK-3 β and Akt inhibitor on D206008-induced melanogenesis

To further confirm the involvement of the Akt/GSK-3 β / β -catenin signaling pathway in D206008-induced melanogenesis, the B16 cells treated with D206008 were exposed to either the specific p-GSK-3 β inhibitor BIO or Akt inhibitor IV. D206008-induced increase in melanin content and TYR activity were further augmented by BIO and inhibited by Akt inhibitor IV (Figure 6). GSK-3 β , which is a negative regulator of Wnt signaling, when GSK-3 β was inhibited, could increase β -catenin accumulation in the cytoplasm and promotes β -catenin nuclear translocation, ultimately transcriptionally upregulates MITF and thus activates the melanogenic genes expression. These results clearly indicated that the Akt/GSK-3 β / β -catenin signal pathway is involved in D206008-induced melanogenesis (Figure 7).

Discussion

An aberrant decrease in skin melanin biosynthesis may result in depigmentation disorders such as vitiligo,²² which in addition to seriously affecting an individual's appearance

also cause psychological distress and affect quality of life.²³ Although a number of melanogenic agents have been developed,²⁴⁻²⁶ most have serious side effects such as allergies, contact dermatitis, eczema, and cytotoxicity. Therefore, novel, safe, and effective melanogenic drugs have to be developed.

Some plant-derived compounds can promote melanocyte regeneration and have been used in various medicinal systems. Psoralens are furocoumarins isolated from *P. corylifolia* L, and several natural and synthetic photosensitive derivatives of psoralen have been tested in depigmentation disorders.²⁷ *P. corylifolia* grows in the high-altitude regions of southern Xin Jiang, and its dried ripe fruits are used to treat vitiligo in Uygur medicine.²⁸ Psoralen compounds isolated from this plant have been used with ultraviolet radiation for the treatment of irregular pigmentation problems such as vitiligo as they activate TYR and promote the synthesis of melanin.^{29,30} The main psoralen used in psoralen and ultraviolet A is 8-MOP, which unfortunately has secondary gastrointestinal effects and increases the risk of severe complications.^{31,32} Therefore, it is vital to isolate or synthesize compounds with better activity and low toxicity

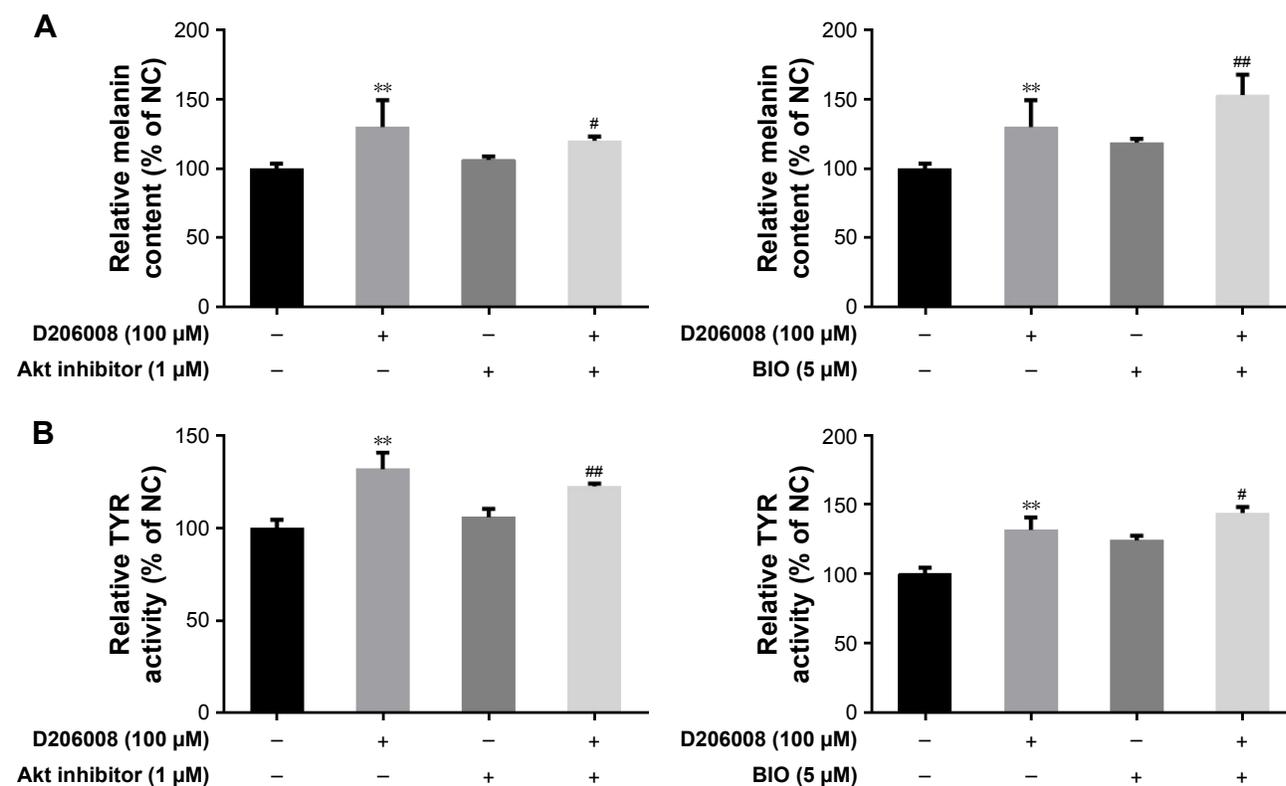


Figure 6 Effect of Akt and GSK-3 β inhibitors on D206008-induced melanogenesis.

Notes: B16 cells were pretreated with 6-bromindirubin-3'-oxime (BIO) (5 μ M) or Akt inhibitor IV (1 μ M) for 2 hours and then incubated with D206008 (100 μ M) for (A) 48 hours to measure the melanin content or (B) 24 hours to measure TYR activity. Result shows that pretreating the cells with the specific GSK-3 β inhibitor BIO increased the tyrosinase activity and melanin content. ## P <0.01, * P <0.05 compared to only D206008-treated group. D206008-induced increase in melanin content and TYR activity was inhibited by Akt inhibitor IV. ## P <0.01, * P <0.05 compared to only D206008-treated group. Data are shown as mean \pm SD and analyzed by one-way ANOVA followed by Tukey's test. ** P <0.01, compared to control group; all experiments were performed three times.

Abbreviations: D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; GSK-3 β , glycogen synthase kinase-3 β ; TYR, tyrosinase.

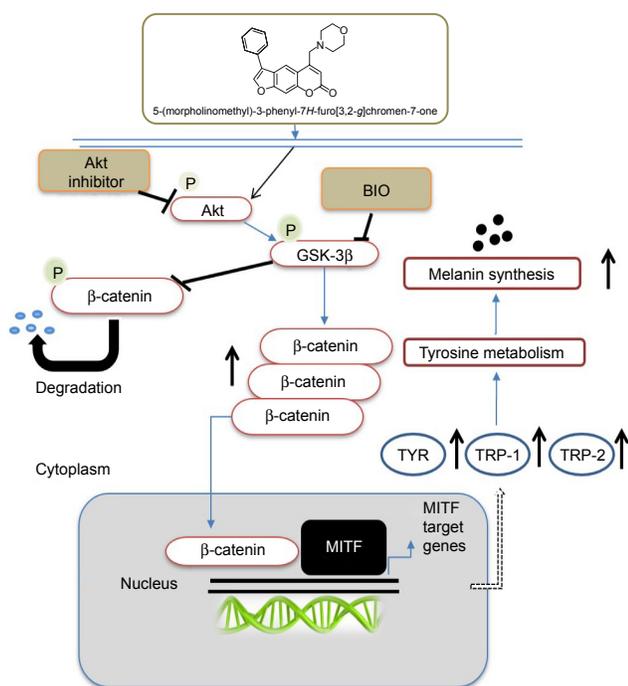


Figure 7 Potential mechanism by D206008-mediated melanogenesis in B16 cells. **Notes:** D206008 activates Akt and GSK-3 β via phosphorylation and decreases the phosphorylation level of β -catenin, which increases its accumulation in cytoplasm, and subsequent nuclear translocation. This leads to the upregulation of the melanogenic genes *TYR*, *TRP-1*, and *TRP-2* via MTF. D206008 promoted phosphorylation of Akt and led to increased intracellular p-GSK-3 β expression, resulting in the accumulation of the β -catenin in the cytoplasm by inhibiting the phosphorylation of β -catenin, which translocated into the nucleus and activated the transcription of melanogenic genes *TYR*, *TRP-1*, and *TRP-2* via MTF. D206008 activates Akt and GSK-3 β via phosphorylation and decreases the phosphorylation level of β -catenin, which increases its accumulation in cytoplasm, and subsequent nuclear translocation. This leads to the upregulation of the melanogenic genes *TYR*, *TRP-1*, and *TRP-2* via MTF. D206008-induced melanin synthesis was completely blocked by GSK-3 β and Akt inhibitors. D206008-induced melanin synthesis was completely blocked by GSK-3 β and Akt inhibitors. **Abbreviations:** BIO, 6-bromindirubin-3'-oxime; D206008, 5-(morpholinomethyl)-3-phenyl-7H-furo[3,2-g]chromen-7-one; GSK-3 β , glycogen synthase kinase-3 β ; MTF, microphthalmia-associated transcription factor; TYR, tyrosinase.

to treat pigmentation disorders. Pang et al synthesized coumarin derivatives bearing isoxazole moieties as melanogenic stimulator from 5-(bromomethyl) I soxazoles and 4-methylumbelliferone.³³ Niu et al synthesized ester coumarin derivatives that were potent on melanin synthesis and TYR stimulations.³⁴ Recently, we synthesized a new series of amine derivatives of furocoumarin compounds and evaluated their melanogenic effects in B16 murine cells. One of derivatives D206008 exhibited better melanogenesis compared to the positive control (8-MOP), in terms of both melanin synthesis and TYR activity, without any cytotoxicity.

Melanin is produced in melanocytes and involves three structurally related enzymes – TYR, TRP-1, and TRP-2 – of which TYR is the main rate-limiting enzyme. It hydroxylates L-tyrosine to L-DOPA and subsequently oxidizes L-DOPA to dopaquinone. TRP-1 and TRP-2 catalyze further

oxidation steps in melanin synthesis.³⁵ Therefore, studies have largely focused on enhancing TYR activity to restore melanin synthesis. D206008 promoted melanin synthesis by increasing TYR activity and therefore total melanin content in a dose- and time-dependent manner. Interestingly, even low doses of D206008 (1 μ M) had a positive effect on melanogenesis. Furthermore, D206008 treatment significantly increased the expression levels of TYR, TRP-1, and TRP-2. The melanogenic enzymes are regulated by the MTF, a key regulator of melanocyte development and melanogenesis.^{36,37} To determine the underlying molecular mechanism of the pro-melanogenic effect of D206008, we analyzed the expression of MTF and TYR genes, both of which were significantly upregulated by D206008 in a time- and dose-dependent manner, as was the MTF protein. Taken together, D206008 promotes melanogenesis in B16 melanoma cells by upregulating MTF and its downstream TYR family genes.

The Akt/GSK-3 β / β -catenin signaling pathway is closely related to melanocyte development and melanogenesis. Akt phosphorylation and activation lead to the phosphorylation of GSK-3 β ,³⁸ which prevents degradation of β -catenin and increases its stability. Accumulation of β -catenin in the cytoplasm promotes its nuclear translocation, where it transcriptionally upregulates MTF and thus activates the melanogenic genes.^{39,40} The key regulator of this pathway is the level of intracellular β -catenin.⁴¹ A previous study found that β -catenin regulates melanogenesis, which upregulated MTF transcription by activation of β -catenin signal pathway, then MTF enhance tyrosinase gene expression and promote melanogenesis.^{42,43} D206008 also enhanced Akt and GSK-3 β phosphorylation and decreased that of β -catenin, which increased its accumulation in the cytoplasm of B16 cells. D206008 promoted phosphorylation of Akt and led to increased intracellular p-GSK-3 β expression, resulting in the accumulation of β -catenin in the cytoplasm by inhibiting the phosphorylation of β -catenin, which translocated into the nucleus and activated the transcription of melanogenic genes *TYR*, *TRP-1*, and *TRP-2* via MTF. These results show that D206008 induced melanogenesis via phosphorylation of GSK-3 β and Akt, which increased the nuclear translocation of β -catenin and activated MTF. To further confirm the possible involvement of GSK-3 β and Akt in melanogenesis, we pretreated the cells with the specific GSK-3 β inhibitor BIO and Akt inhibitor IV. Previous research had revealed that BIO makes β -catenin accumulate in cells, which results in an increase in TYR activity and melanin content. The Akt inhibitor reversed the TYR

activity and melanin content.⁴⁴ In our study, BIO increased TYR activity and melanin content in B16 cells treated with D206008; opposite effects were seen with the Akt inhibitor IV. In conclusion, D206008 promotes melanogenesis via the Akt/GSK-3 β / β -catenin pathway. Although ester coumarin derivatives synthesized by Niu et al were potent on melanin synthesis and TYR stimulations by upregulation of MITF and TYR family via Akt/GSK-3 β / β -catenin signaling pathways, it was low water solubility, which may reduce its effect in vivo. Thus D206008 was designed and synthesized to improve this deficiency since the nitrogen atoms could easily form hydrogen bond in water. Further, our data indicated that D206008 increased melanin production and TYR activity by Akt/GSK-3 β / β -catenin signaling pathway, a finding that needs to be supported in animal models of hypopigmentation to comprehensively understand the repigmentation effect of this drug.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work.

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