



Topical Agents for Nonrestorative Management of Dental Erosion: A Narrative Review

Darren Dhananthat Chawhuaveang ^(D), Ollie Yiru Yu *^(D), Iris Xiaoxue Yin^(D), Walter Yu Hang Lam ^(D) and Chun Hung Chu ^(D)

> Department of Restorative Dental Sciences, Faculty of Dentistry, The University of Hong Kong, Hong Kong SAR, China; dhanant@connect.hku.hk (D.D.C.); irisxyin@hku.hk (I.X.Y.); retlaw@hku.hk (W.Y.H.L.); chchu@hku.hk (C.H.C.)

* Correspondence: ollieyu@hku.hk; Tel.: +852-2859-0310

Abstract: A nonrestorative approach to the management of dental erosion is the foremost option: controlling dental erosion. The objectives of this study are to provide an overview and to summarise the effects and properties of topical anti-erosive agents as a nonrestorative treatment of dental erosion. A literature search was conducted on five databases of peer-reviewed literature—Cochrane Library, EMBASE, PubMed, Scopus and Web of Science—to recruit articles published between 1 January 2000 and 31 December 2021. The literature search identified 812 studies; 95 studies were included. Topical anti-erosive agents can be broadly categorised as fluorides, calcium phosphate-based agents, organic compounds and other anti-erosive agents. In the presence of saliva, fluorides promote the formation of fluorapatite on teeth through remineralisation. Calcium phosphate-based agents supply the necessary minerals that are lost due to the acid challenge of erosion. Some organic compounds and other anti-erosive agents prevent or control dental erosion by forming a protective layer on the tooth surface, by modifying salivary pellicle or by inhibiting the proteolytic activity of dentine collagenases. Topical anti-erosive agents are promising in managing dental erosion. However, current evidence shows inconsistent or limited results for supporting the use of these agents in clinical settings.

Keywords: tooth erosion; fluorides; anti-erosive agents

1. Introduction

Dental erosion is the loss of minerals from the dental hard tissue caused by nonbacterial acids [1]. This chemical process often occurs with mechanical wear, such as abrasion and attrition, and is known as erosive tooth wear [1,2]. Erosive tooth wear damages the tooth structure, negatively affecting the aesthetics and the function of the natural dentition [1–3]. The loss of tooth structure with opening dentinal tubular often initiates dentine hypersensitivity, which can result in severe, persistent pain and discomfort [2,3]. With the increased consumption of acidic food and beverage, the prevalence of dental erosion has increased significantly over the past few decades [1,4,5]. The estimated global prevalence of dental erosion in primary dentition ranged from 30% to 50% and from 20% to 45% in permanent dentition [6]. Almost all middle-aged populations and more than half of children and adolescents are affected by dental erosion [7]. Therefore, dental erosion has a long-term impact on the oral and general health of the population [2,3].

Restorative treatment for dental erosion can be challenging, invasive and extensive [8,9]. Therefore, it is imperative to identify dental erosion at an early stage, and nonrestorative management aiming for preventive care should be provided [1,10]. Nonrestorative approaches to dental erosion commonly include dietary analysis and counselling, oral health education and topical use of anti-erosive agents [5,11,12]. Dietary analysis and counselling are important for preventing excessive consumption of acidic beverages and foodstuffs that potentially damage dentition [1,10]. Oral health education is essential to promoting the protective factors that effectively prevent dental erosion [10,13].



Citation: Chawhuaveang, D.D.; Yu, O.Y.; Yin, I.X.; Lam, W.Y.H.; Chu, C.H. Topical Agents for Nonrestorative Management of Dental Erosion: A Narrative Review. *Healthcare* 2022, 10, 1413. https://doi.org/10.3390/ healthcare10081413

Academic Editor: Massimo Corsalini

Received: 1 July 2022 Accepted: 25 July 2022 Published: 28 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Among the various nonrestorative approaches, topical anti-erosive agents are commonly used to manage dental erosion [11,12].

Although nonrestorative approaches should be prioritised in managing dental erosion, anti-erosive agents for preventing and managing dental erosion have never been elucidated in the literature. Therefore, this narrative review aims to provide an overview and to summarise the effects and properties of topical anti-erosive agents as a nonrestorative treatment of dental erosion.

2. Anti-Erosive Agents in the Literature

The literature search of this narrative review was conducted on five databases: Cochrane Library, EMBASE, PubMed, Scopus and Web of Science, to recruit articles published between 1 January 2000 and 31 December 2021. The keywords used in the search were ("Dental erosion" OR "Tooth erosion" OR "Enamel erosion" OR "Dentine erosion" OR "Dentine erosion" OR "Eroded" OR "Erosive lesion" OR "Erosive demineralisation*" OR "Erosive tooth wear") AND ("Prevention" OR "Protection" OR "Protective effect*" OR "Anti-erosive"). Figure 1 shows a flowchart of our systemic search of the literature. The inclusion criteria were (i) studies that assessed the anti-erosive effects of topical agents and (ii) studies published between 1 January 2010 and 30 September 2021. The exclusion criteria were (i) studies on dental caries, (ii) studies on the anti-erosive effects of salivary pellicle, (iii) studies on the restorative management of dental erosion/erosive tooth wear and (iv) studies on the remineralising effects of carbon dioxide laser. Topical anti-erosive agents can be broadly categorised as topical fluorides, calcium phosphate-based agents, organic compounds and other anti-erosive agents (Figure 2).

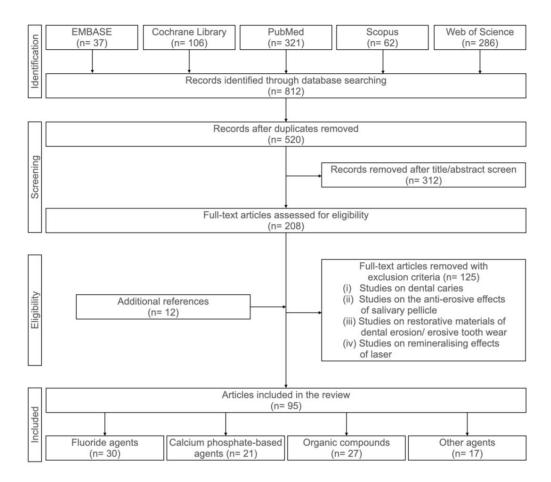


Figure 1. Flowchart summarising the reference-selection process.

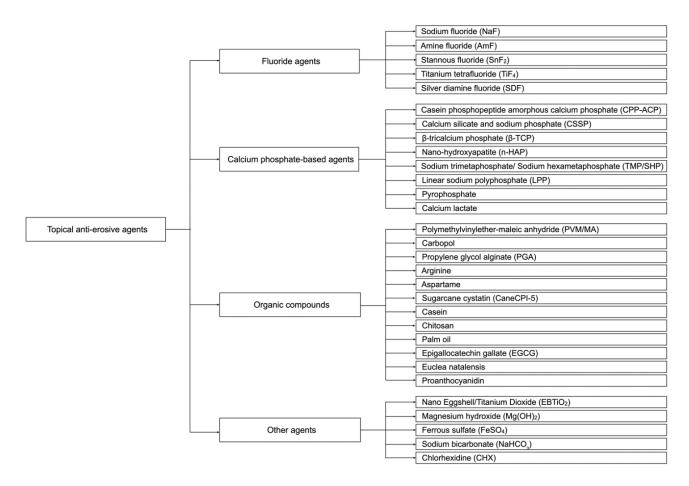


Figure 2. Topical anti-erosive agents.

2.1. Fluoride Agents

Fluoride agents are the most common topical anti-erosive agents [5] and include sodium fluoride (NaF), amine fluoride (AmF), stannous fluoride (SnF₂), titanium tetrafluoride (TiF₄) and silver diamine fluoride (SDF). They are divided into two groups: conventional monovalent fluoride, such as NaF and AmF [5], and polyvalent fluoride with metal cations, such as TiF₄, SnF₂ and SDF [5,11,14]. Topical fluoride facilitates the formation of a protective layer on the tooth surface and enhances the acid resistance of dental hard tissue [14,15]. Some fluoride agents can modify salivary pellicle and enhance the acid resistance of salivary pellicle [16,17]. Fluoride can also inhibit the activation of matrix metalloproteinases (MMPs) in the dentine matrix [14,15].

2.1.1. Sodium Fluoride (NaF)

NaF is a conventional monovalent fluoride [5]. There are plenty of commercially available NaF products [18–21]. Over the counter, NaF toothpaste contains up to 1450 ppm fluoride [5,20]. NaF toothpaste with 5000 ppmF is also available as a prescription [5,22]. In addition, 5% NaF varnish containing 22,600 ppmF is a commonly used professional fluoride agent [5,20]. NaF application leads to a calcium fluoride-like layer (CaF₂) forming on the tooth surface [5,22]. This layer is a protective barrier that blocks acid diffusion through the tooth's surface [23]. CaF acts as a reservoir of calcium and fluoride ions to enhance the remineralisation process of the hard tissue [5,22]. Sodium ions may interact with the salivary proteins in the acquired salivary pellicle [12,24]. It improves the re-adsorption of proteins on the eroded tooth surface and may enhance the erosive resistance of salivary pellicle [24]. Moreover, NaF is an MMP inhibitor that inhibits MMP activation [25] and

maintains the demineralised organic matrix (DOM), which acts as a barrier to acid diffusion on the dentine surface [25,26].

NaF reduced surface loss, mineral loss and microhardness change in enamel specimens after erosive challenges [20,27,28]. However, some studies reported that the anti-erosive effects of NaF are limited [5,12,23,29,30].

2.1.2. Amine Fluoride Containing Agents (AmF)

Products containing AmF are available in different forms such as toothpaste, mouthrinse and solution [31–35]. In collaboration with the University of Zurich, a company (GABA, Schweiz, Colgate-Palmolive, Therwil, Switzerland) developed and patented AmF with NaF and SnCl₂ [12,36]. Using organic molecules as carriers, they demonstrated that AmF significantly reduced the solubility of the enamel and thereby increased the resistance of enamel against acid attack [12,36]. AmF provide CaF₂ precipitates as a protective layer on the tooth surface [12,37]. Moreover, AmF have a pronounced affinity regarding enamel by raising the quantity of fluoride in the saliva [37].

AmF combined with SnCl₂ or SnF₂ reduced human enamel or dentine loss after erosive challenges [12,38]. AmF alone decreased calcium ion release in human dentine specimens in acid challenges [39], but the protective effect was limited [34,35,40].

2.1.3. Stannous Fluoride Containing Agents (SnF₂)

Anti-erosive products using SnF_2 or $SnCl_2$ with fluoride as active ingredients are available on the market [41–43]. The stannous ions worked synergistically with fluoride and enhanced the anti-erosive effects of fluoride [15,44,45]. Stannous ions in these products have a higher affinity for mineral contents than organic contents [15]. These stannous ions interact with hydroxyapatite and fluoride and form a layer of Sn_2OHPO_4 , $Sn_3F_3PO_4$, Ca (SnF_3)₂ and CaF_2 on the tooth surface [36,46–49]. This layer is more stable and acidresistant than the layer formed after NaF application alone [15,36,46]. Moreover, stannous ions interact with salivary proteins in acquired salivary pellicle [16,24] and enhance the acid resistance of salivary pellicle [16,17]. Stannous ions can also inhibit the activation of MMPs in the dentine matrix and prevent the organic matrix of the demineralised dentine surface from degrading [50].

 SnF_2 reduced surface loss, decreased calcium ions release and decreased the microhardness change in enamel specimens after erosive challenge [24,33,51,52]. Nevertheless, the protective layer of SnF_2 is not stable at neutral or alkaline conditions [15]. It should be noted that $SnF_2/SnCl_2$ may cause staining on the dental hard tissue with prolonged use [32].

2.1.4. Titanium Tetrafluoride (TiF_4)

No commercially available product of TiF₄ is currently available. TiF₄ reacts with the hydroxyapatite on the tooth surface and forms a glaze-like layer of titanium dioxide (TiO₂) and hydrated titanium phosphate (TiPO₄) [50,53,54]. The TiO₂ and TiPO₄ layer coats the tooth surface and acts as a protective layer against erosive challenges [53,54]. Furthermore, this layer promotes fluoride uptake to the tooth surface [50,53] and increases the CaF₂ deposits on the tooth surface [55]. The interaction of TiF₄ and fluorapatite was found in the subsurface area in the demineralised enamel [54,56]. This layer is more acid-resistant than the CaF₂ layer formed after NaF application [53].

TiF₄ reduced human dentine loss, decreased microhardness change in human enamel specimens and occluded human dentinal tubule in erosive attacks [50,57–59]. However, the extreme acidity of the TiF₄ solution may weaken its anti-erosive effect [60,61]. The interaction with saliva also reduces the protective effect of TiF₄, which is unavoidable in the oral cavity [54,62]. Another concern is that TiF₄ can cause reversible staining on the tooth surface [50,63,64].

2.1.5. Silver Diamine Fluoride (SDF)

SDF is an alkaline agent (pH \sim 10) [65]; 38% SDF is a solution that contains a high concentration of fluoride and silver ions [65,66]. Several SDF products are available on the market [66,67]. SDF application facilitates the deposition of silver compounds on the tooth surface and may act as a protective layer against dental erosion [16,67]. SDF reduces demineralisation and promotes remineralisation on the tooth surface by promoting the deposition of CaF₂ and the formation of fluorapatite crystals on the tooth surface [16,65,68]. SDF can also inhibit MMPs [69] and may decrease the erosive demineralisation of dentine [69].

SDF has been widely investigated in anti-caries activity but the evidence of SDF on managing dental erosion is insufficient [70]. SDF decreased bovine enamel and dentine loss after erosive challenges [16]. A main disadvantage of SDF is the permanent black staining on the tooth surface [66,69,70].

2.2. Calcium Phosphate-Based Agents

Calcium phosphate-based agents supply the necessary minerals that are lost due to the acid challenge of erosion [71]. Calcium phosphate-based agents include casein phosphopeptide amorphous calcium phosphate (CPP-ACP), calcium silicate sodium phosphate (CSSP), β -tricalcium phosphate (β -TCP), nano-hydroxyapatite (n-HAP), sodium trimetaphosphate/sodium hexametaphosphate (TMP/SHP), linear sodium polyphosphate (LPP), pyrophosphate and calcium lactate.

2.2.1. Casein Phosphopeptide Amorphous Calcium Phosphate (CPP-ACP)

CPP-ACP is used in oral health products such as varnish, cream, chewing gum and acidic drinks for the management of dental caries [11]. The addition of fluoride to CPP-ACP results in the formation of casein phosphopeptide amorphous calcium fluoride phosphate (CPP-ACFP). CPP-ACP comprises a high amount of calcium and phosphate ions [36,49]. These ions form a calcium hydrogen phosphate precipitation layer [49], which acts as a physical barrier between acids and tooth surfaces [72–74]. Notably, the precipitation layer showed non-homogeneous distribution and may be easily detached by acids [73,75]. Calcium and phosphate ions are maintained around the tooth surface even under acidic conditions and reduce demineralisation process [11,36,49]. It also promotes remineralisation by supplying the mineral ions on the eroded surface [11,36]. In addition, CPP-ACP can modify the acquired salivary pellicle by increasing the electron-dense layer of the pellicle [73,76].

In situ studies revealed that CPP-ACP reduced the surface loss, microhardness change and calcium ions release in human enamel specimens [73,77–79]. Similarly, in situ studies of CPP-ACFP reported that CPP-ACFP reduced bovine enamel loss [80] and decreased microhardness change in human enamel after erosive cycles [81]. The effectiveness of CPP-ACP depends on the concentration, the vehicles and time of application [73–75]. The optimal time of application of CPP-ACP is contradictory in the literature [73,82]. A previous study showed that CPP-ACP had a limited working time [73].

2.2.2. Calcium Silicate and Sodium Phosphate (CSSP)

CSSP has been used as an additive in oral hygiene products for many years [83]. CSSP provides calcium and phosphate ions and raises the concentration of the mineral ions surrounding the tooth surface to saturated levels [36,83]. Thus, it helps reduce the demineralisation on the tooth surface [32,36] and promotes the remineralisation process of erosive dental hard tissue [32,36,84]. The CSSP also forms a protective layer by depositing calcium silicate particles on the tooth surface [32,83].

No study has investigated the effect of CSSP alone. In situ studies of CSSP reported that CSSP reduced bovine enamel loss [83], decreased the human enamel hardness change [85] and decreased dentine permeability after erosive challenges when combined with fluoride [86]. However, the protective effect of CSSP has a relatively short effective time [32]. The protective layer is unstable and can be easily removed by a strong acid [84]. In addition, CSSP may induce demineralisation on the tooth surface by itself due to the high acidity of the agent [32].

2.2.3. β -Tricalcium Phosphate (β -TCP)

 β -TCP is a bioactive agent mainly comprising calcium and phosphate [71]. β -TCP provides calcium and phosphate ions to the tooth surface [71]. The calcium and phosphate ions maintain saturation levels around the tooth surface and induce mineral deposition on the tooth surface [71,87]. Moreover, β -TCP provides nucleation of the remineralisation process [71,87]. This nucleation facilitates the remineralisation of the eroded surface [71].

No study has investigated the anti-erosive effect of β -TCP alone. Previous in situ studies of β -TCP revealed that β -TCP reduced calcium and phosphate ion release, surface loss and hardness change in human enamel specimens when used with fluoride [71,87–89].

2.2.4. Nano-Hydroxyapatite (n-HAP)

n-HAP or nano-sized zinc-carbonate-hydroxyapatite is a synthetic hydroxyapatite, of which its size is approximately 20–100 nm [90–92]. The common concentration of n-HAP ranges between 1 and 10% [90]. n-HAP chemically binds to the natural apatite on the tooth structure and forms a crystalised apatite layer [90,93]. In addition, n-HAP can release calcium ions to maintain the mineral ions at a high level around the tooth surface and oral environment, which decrease demineralisation and increase remineralisation [90,94]. n-HAP can also be a template for crystal growth and remineralisation [93,95].

One in situ study reported that n-HAP decreased the human enamel hardness change after erosive challenge [90]. However, it was also reported that n-HAP had limited preventive effects on dental erosion [90].

2.2.5. Sodium Trimetaphosphate/Sodium Hexametaphosphate (TMP/SHP)

TMP and SHP are inorganic polyphosphate compounds [29,96]. TMP is usually used as an additive in fluoride varnish, toothpaste and mouthrinse, whereas SHP is mainly added to fluoride toothpaste [14,29,49]. TMP can provide an acid-resistant layer by adsorbing onto the hydroxyapatite structure of the tooth surface and the collagen of the dentine surface [29,97–99]. The phosphate structures in the protective layer can incorporate the CaF₂ layer due to fluoride application [98]. SHP has a similar action to TMP on the tooth surface [14]. Additionally, SHP may infiltrate into the demineralised organic contents at the dentine surface and facilitates the formation of a scaffold for remineralisation [14]. SHP also enhances the level of acid-resistant salivary proteins in the acquired salivary pellicle [49,97].

No study has investigated the anti-erosion effect of TMP or SHP alone. In situ studies demonstrated that TMP or SHP reduced surface loss and decreased surface microhardness change in bovine enamel when combined with fluoride [100,101]. A low concentration of TMP/SHP worked synergistically with fluoride [99], while a high concentration of TMP/SHP may reduce the anti-erosive effect of fluoride [100].

2.2.6. Linear Sodium Polyphosphate (LPP)

LPP is a long-chain polyphosphate agent commonly used as the additive of nonalcoholic drinks [102,103]. LPP has phosphate groups that can bind with positively charged particles on the tooth structure [102,103] and forms an acid-resistant layer [103,104]. The LPP also works synergistically with fluoride and stannous ions in controlling dental erosion [102,103].

LPP decreased bovine enamel loss in in situ erosive challenges when combined with fluoride [104]. Likewise, in vitro studies demonstrated that LPP decreased surface loss and hydroxyapatite dissolution in bovine enamel and dentine when combined with SnCl₂ and fluoride [102,103]. LPP is more effective on enamel than on dentine because dentine has fewer binding sites for LPP [102]. However, some studies showed that LPP might compete

with the anti-erosive protein in the salivary pellicle [104] or fluoride [105] for the binding site on the tooth surface and decrease their protective effect against dental erosion [104,105].

2.2.7. Pyrophosphate

Pyrophosphate or phytate is an organic polyphosphate, which is mainly found in cereals and seeds [106]. It is a cyclic structure with six phosphate groups without direct phosphate–phosphate bonds [105,106]. The pyrophosphate rapidly adsorbs and multi-point binds with hydroxyapatite of the tooth surface [105]. Moreover, it inhibits the diffusion of the ions between acids and the tooth surface [105,106]. Thus, pyrophosphate reduces erosive demineralisation [105,106].

One in situ study reported that pyrophosphate completely inhibited the remineralisation property of fluoride [105]. It may also compete with other mineral ions and inhibit them from binding to the tooth surface [105].

2.2.8. Calcium Lactate

Calcium lactate or calcium effervescent tablets have been used to prevent the softening of dental hard tissue caused by erosive drinks [28,48]. Calcium lactate mouthrinses or solutions provide extra calcium ions in the saliva [28,48] and reduce the demineralisation of the tooth during acid attacks [48]. Moreover, calcium ions may react with the fluoride in the oral cavity and increase CaF deposition on the tooth surface [28,107]. The CaF acts as a fluoride and calcium ion reservoir and promotes remineralisation [93,95].

An invitro study reported that calcium lactate with fluoride decreased bovine enamel loss after erosive challenges [107]. However, an in situ study reported that a calcium lactate-containing solution could not prevent enamel erosion [48].

2.3. Organic Compounds

Anti-erosive organic compounds include polymer agents, such as polymethylvinylethermaleic anhydride (PVM/MA), carbopol and propylene glycol alginate (PGA), and these agents are derived from animals or plants' carbohydrates; lipids; or proteins, such as arginine, aspartame, sugarcane cystatin (CaneCPI-5), casein, chitosan, palm oil, epigallocatechin gallate (EGCG), Euclea natalensis plants and proanthocyanidin.

2.3.1. Polymethylvinylether-Maleic Anhydride (PVM/MA)

PVM/MA is a film-forming polymer, which is commonly used as an additive in beverages [108]. It is added to oral health products to control dental erosion [108]. PVM/MA binds to the mineral ions in the enamel and forms a protective layer [108,109]. It also binds to mineral ions and type I collagen in the dentine [110]. However, the protective layer formed by PVM/MA could easily be removed by acids because of the weak bond of PVM/MA with the tooth [108]. PVM/MA also helps with fluoride retention on the tooth surface and sustains fluoride release for a longer period [109–111].

An in situ study reported that PVM/MA enhanced surface hardness on bovine enamel after erosion compared with the control group [111]. Previous in vitro studies showed that PVM/MA positively interacted with fluoride and reduced surface loss in human and bovine dentine specimens [108,110,111].

2.3.2. Carbopol

Carbopol is a high molecular weight polymer with a negatively charged centre, which allows it to chelate with calcium ions [40,112]. Carbopol strongly binds with calcium ions [17,40] and forms a protective layer that covers the tooth surface [40,112]. Some studies have demonstrated that carbopol enhanced fluoride adsorption and promoted fluoride retention in the oral cavity [40,112].

Studies reported that carbopol with fluoride reduced enamel loss, increased enamel hardness and decreased the hydroxyapatite dissolution in erosive challenges [40,112]. However, the acidity (pH = 2.7-3.3) of carbopol may intensify the dissolution of dental

hard tissue [40]. Carbopol also competes with salivary proteins for the binding sites on the tooth surface and may interfere with the anti-erosive potential of the acquired salivary pellicle [17].

2.3.3. Propylene Glycol Alginate (PGA)

PGA is a natural polymer derived from brown seaweeds that is commonly used in the food and biomedical industry, such as emulsifiers, stabilisers and thickening agents [108,113]. It has low toxicity, low cost, and high viscosity and biocompatibility [108,113]. The carboxylic groups of the PGA can bind to the calcium ions on the tooth surface and provides a protective layer [108,113]. The anti-erosive effect of the PGA is affected by the hydrogen ion level in acidic solutions because the hydrogen ions bind with the carboxylic groups of PGA and change the number of available carboxylic groups in PGA that can bind to calcium ions on the tooth surface [113]. PGA also competes with salivary proteins in salivary pellicle for binding sites on the tooth surface [113]. PGA with fluoride reduced bovine enamel and dentine loss after erosive challenge [108,113]. Notably, PGA alone cannot protect the tooth surface from erosion [108,113].

2.3.4. Arginine

Arginine is a positively charged amino acid that has a strong affinity to the dentine surface [56,114]. It improves the attachment of calcium to the tooth surface and promotes calcium carbonate precipitation [49,115,116]. This precipitation layer enhances the resistance of dental hard tissue to acids [115,116]. The precipitation layer helps to occlude dentinal tubules and alleviates dentine hypersensitivity [114,115].

The combined application of arginine with fluoride or calcium carbonate showed a positive result in controlling erosion [114,116,117]. The previous in situ studies demonstrated that arginine with fluoride decreased dentine permeability to acids by occluding human dentinal tubule [114,118]. In vitro studies reported that arginine reduced the hardness change and surface loss on bovine enamel [56,116].

2.3.5. Aspartame

Aspartame is a synthetic dipeptide and an artificial non-saccharide sweetener [119]. It is widely used as a sugar substitute in food and drink products [119]. Aspartame can be degraded to phenylalanine, aspartic acid and methanol [119]. Phenylalanine contains carboxylic and amino groups that can capture hydrogen ions in the erosive acids and can reduce the acidity of the acids [119]. Hence, it can reduce the demineralisation of the enamel surface [119,120].

A previous study showed that the anti-erosive effect of aspartame is limited [120]. All in situ and in vitro studies that investigated the anti-erosive property of aspartame revealed aspartame had no significant protective effect on the bovine enamel compared to no treatment [119,120].

2.3.6. Sugarcane Cystatin (CaneCPI-5)

CaneCPI-5 is a novel synthesised sugarcane cystatin [121]. It strongly binds to hydroxyapatite of the enamel surface and forms a protective layer [121,122]. The CaneCPI-5 can also improve the protective effect of the salivary pellicle by increasing the number of acid-resistant proteins such as cystatin B [121,123,124]. In addition, CaneCPI-5 inhibits MMPs and reduces the severity of dentine erosion [122,123].

One in situ study revealed that CaneCPI-5 reduced bovine enamel loss in erosive challenge [121]. In vitro studies reported that CaneCPI-5 reduced surface loss in bovine enamel and dentine and hardness change in bovine dentine [122,123].

2.3.7. Casein

Casein is a protein that is commonly found in colostrum or milk products [125,126]. Casein comprises three subfractions: α -, β - and κ -casein [125]. The casein can adsorb onto

the hydroxyapatite and forms a protective layer of milk protein on the tooth surface [125,126]. Casein has many amino acid sequences such as phosphoserine, histidine, glutamate and aspartate [126]. These amino acids could act as a buffer to increase pH and decrease demineralisation during acidic conditions [126]. Furthermore, casein can modify the pellicle's compositions and enhances the protective effects of salivary pellicle [125,126].

Casein reduced calcium and phosphate ion release in human enamel specimens after in situ erosive challenges [125]. Casein reduced surface loss and reduced microhardness change on bovine enamel in in vitro erosive challenges [126].

2.3.8. Chitosan

Chitosan has been used as an additive in some fluoridated oral health products [47]. It increases the viscosity and stability of fluoride-containing agents [55,104] and enhances the anti-erosive capacity of fluoride agents [49,104]. Chitosan alone provides a protective layer on the tooth surface by binding with mineral ions on the tooth surface [14,55]. It can be modified by phosphorus ions and forms phosphorylated chitosan, which promotes the remineralisation process by chelating with calcium and phosphate ions on the tooth surface [127]. The alkaline phosphorylated chitosan inhibits MMP degradation and preserves the integrity of collagen fibrils [127]. Chitosan also modifies the acquired salivary pellicle by increasing the attachment of acid-resistant salivary proteins [47,123,128].

Previous in situ studies supported that chitosan with fluoride could reduce human enamel surface loss in erosive challenges [128,129]. In vitro studies reported that chitosan with fluoride decreased bovine dentine loss [130,131]. Chitosan alone had a weak antierosive effect [55,127,131]. It should be noted that chitosan with different pH levels has different anti-erosion effects [127]. Alkaline phosphorylated chitosan is more acid-resistant than neutral or acidic ones [127].

2.3.9. Palm Oil

Palm oil is an edible, safe and low-cost natural product [132,133]. Palm oil has been widely used in foods, cosmetics and medical products [133]. It can modify the composition and ultrastructure of the outer layer of the salivary pellicle and form an acid-resistant hydrophobic lipid-enriched pellicle [132].

Palm oil reduced bovine enamel loss in erosive challenges in an in situ study [132]. An in vitro study reported palm oil decreased bovine enamel hardness change [133].

2.3.10. Epigallocatechin Gallate (EGCG)

EGCG is a type of polyphenol that is found in green tea [134,135]. EGCG forms an acid-resistant precipitation layer by binding with calcium ions on the tooth surface [134]. In addition, EGCG inhibits MMP activity in the dentine matrix and decreases erosive demineralisation in the dentine [134–136]. Moreover, EGCG modifies acquired salivary pellicle's compositions by increasing the number of acid-resistant proteins [137].

The in situ studies revealed that EGCG reduced bovine dentine loss in erosive challenges [26,138]. An in vitro study reported that EGCG decreased human enamel and dentine loss after acid challenges [134]. However, the effectiveness of EGCG gradually reduced after interacting with acids and could not protect the tooth from prolonged erosive attacks [134,135]. One in vitro study also stated that EGCG might have an antagonistic effect with fluoride [134].

2.3.11. Euclea Natalensis Plant Extracts

Euclea natalensis is a plant from Africa [139,140]. The active ingredients of Euclea natalensis extracts are taninus; naphthoquinone; flavonoids; and polyphenolic compounds that demonstrated fungicidal, antibacterial, insecticidal, phytotoxic, cytotoxic and anticarcinogenic properties [139,140]. Taninus interacts with salivary proteins in the salivary pellicle and forms protein-taninus complexes [140]. Therefore, Euclea natalensis compounds can modify salivary pellicle and form a protective layer on the tooth surface [139,140]. Naphthoquinone is an MMP inhibitor that preserves the organic matrix of the dentine surface in an acid attack [139].

An in situ study reported that Euclea natalensis plant extracts decreased human dentine loss in a 5-day erosive challenge [139]. Euclea natalensis plant extracts also reduced human dentine permeability in an in vitro study [140]. However, Euclea natalensis plant extracts cause temporary light yellow staining on the tooth surface and oral tissues [140]. It should be noted that the effects of the Euclea natalensis plant against erosive demineralisation are scarce in the literature, with only one in situ study and one in vitro study available [139,140].

2.3.12. Proanthocyanidin

Proanthocyanidin is a polyphenol that is commonly found in natural fruits and nuts [141,142]. It has been used in adhesive products in restorative dentistry [139,141]. Proanthocyanidin is effective at inhibiting MMPs 1, 2, 8 and 9 [141,142]. It reduces dentine loss by preserving collagen fibrils and delays acid diffusion [143]. In addition, proanthocyanidin enhances collagen strength by increasing collagen cross-links [144] and forms an insoluble complex that promotes precipitation of calcium ions from saliva [141].

Proanthocyanidin reduced bovine dentine loss in erosive challenges [141,142]. However, the by-products of proanthocyanidin are acidic and decrease the pH of the surrounding tissue [143]. Moreover, proanthocyanidin causes brownish staining in the dentine surface [143].

2.4. Other Anti-Erosive Agents

Other anti-erosive agents are agents that cannot be classified into the groups mentioned above, including nano eggshell/titanium dioxide (EBTiO₂), magnesium hydroxide (Mg(OH)₂), ferrous sulfate (FeSO₄) and sodium bicarbonate (NaHCO₃) and chlorhexidine.

2.4.1. Nano Eggshell/Titanium Dioxide (EBTiO₂)

EBTiO₂ is an eggshell-modified agent containing calcium, phosphorus, strontium, zinc, fluoride, copper and titanium ions [95,145]. EBTiO₂ facilitates the formation of a protective layer of calcium carbonate and TiO₂ on the tooth surface [145]. The calcium carbonate can also occlude the dentinal tubule and protects dentine from demineralisation [146]. EBTiO₂ also provides mineral ions on the tooth surface to decrease demineralisation and to promote remineralisation [146].

EBTiO₂ reduced bovine enamel loss in an in situ erosive challenge [145] and occluded bovine dentinal tubule in an in vitro setting [146]. It should be noted that the calcium carbonate in EBTiO₂ is unstable in the acidic environment, which limits its use in highly acidic conditions [147]. There are also concerns about the safety and biocompatibility of EBTiO₂ [146].

2.4.2. Magnesium Hydroxide $(Mg(OH)_2)$

 $Mg(OH)_2$ is an inorganic compound with low solubility and cytotoxicity [148,149]. It is an active component of antacid products for neutralising acidity from gastroesophageal reflux disease [148,149]. $Mg(OH)_2$ reacts with hydrogen ions and produces magnesium chloride (MgCl₂), insoluble magnesium salt and water [148,150]. $Mg(OH)_2$ also reacts with calcium ions or calcium carbonate in oral fluids and form a protective layer composed of magnesium carbonate (MgCO₃) and calcium hydroxide (Ca(OH)₂) on the tooth surface [148]. Magnesium ions in $Mg(OH)_2$ can incorporate into the surface layer of the enamel and modify the enamel crystallographic properties [148,151], which makes the enamel surface stronger [151].

Previous in situ studies reported that $Mg(OH)_2$ reduced human enamel loss [150] and human enamel microhardness change after erosive cycles [148,150]. However, $Mg(OH)_2$ could not prevent erosion in acidic solutions with a pH lower than 2 [148,150] because $Mg(OH)_2$ can easily dissolve in highly acidic conditions [149].

2.4.3. Ferrous Sulfate (FeSO₄)

FeSO₄ or iron ion is an inhibitor of MMP-2 and 9 [152,153], which inhibits the hydrolysing of the collagen fibrils in the dentine surface [152,154]. The application of FeSO₄ on the tooth surface facilitates the formation and precipitation of a protective layer composed of ferric phosphate (FePO₄) and ferric salts on the tooth surface [154,155].

FeSO₄ reduced both human and bovine dentine surface loss [152,154] and reduced the microhardness change in the human enamel and dentine after erosive challenges in in situ settings [154]. Nevertheless, FeSO₄ may cause stains on the tooth surface, particularly when it is used in high concentrations or prolonged periods [152,155].

2.4.4. Sodium Bicarbonate (NaHCO₃)

NaHCO₃ has high solubility and can be rapidly dissolved in oral fluid [156,157]. It acts as an acid neutraliser due to its alkalinity [156]. The bicarbonate ions from NaHCO₃ work synergistically with salivary bicarbonate and increase the buffering capacity of the oral fluid [156,158].

In situ studies demonstrated that NaHCO₃ reduced the surface loss of bovine enamel [156], but it had no effects on the microhardness change in bovine enamel after erosive challenges [156–158]. Hence, the anti-erosive of NaHCO₃ is limited [157,158].

2.4.5. Chlorhexidine (CHX)

CHX is used in controlling dental erosion because it is an inhibitor of MMPs [141,159,160]. It inhibits collagen degradation by inhibiting MMPs 2, 8 and 9 in acidic attacks [141,159]. In addition, CHX binds with calcium ions and forms a protective precipitation layer on the tooth surface [26].

CHX reduced the surface loss and hardness change in dentine specimens after erosive challenges in in situ settings [159–161]. CHX should not be used with fluoride in controlling dental erosion because it has an antagonistic effect with fluoride [161]. It should be noted that prolonged use of CHX may cause tooth discolouration, loss of taste and mucosal irritation [141].

3. Discussion

The included studies were conducted with various study protocols and different assessments such as surface loss, microhardness change or surface morphology of the specimens. Due to the heterogeneous research methodology and outcome measures, it was not possible to provide a quantitative assessment of the anti-erosive effects of the included agents in the current study. Therefore, the effects of the included topical anti-erosive agents were summarised without quantitative analysis.

This review introduced four categories of anti-erosive agents for the nonrestorative management of dental erosion, but not all of them are available on the market. The commercially available agents for anti-erosive are shown in Table 1. Fluoride agents revealed the most promising properties in the included studies, especially the combination agents between conventional monovalent and polyvalent fluoride. Unfortunately, more than 2/3 of the included agents are under development. Approximately half of the included studies investigated the remineralising effect of the anti-erosive agents. These studies used erosive challenge to simulate the clinical situation in patients who had been affected by dental erosion. The topical anti-erosive agents were applied on the teeth after erosive challenges. The other half of the studies investigated the preventive effects of the anti-erosive agents, where the anti-erosive agents were applied before the erosive challenge on the teeth. Therefore, the options of topical anti-erosive agents for clinical use are currently limited and the application time of topical anti-erosive is inconclusive.

Agents	Concentration of Active Ingredients	Delivery System	Example of Product Names and Companies	
NaF	1100 ppm F of NaF	Toothpaste	 Colgate[®] Total[®], Colgate-Palmolive [33,43] Crest[®] Cavity Protection, The Procter & Gamble [44] 	
	1450 ppm F of NaF	Toothpaste	 SensodyneTM ProNamel[®], GSK [18,19,21] Crest[®] Decay Protection, The Procter & Gamble [41] Colgate[®] Total[®], Colgate-Palmolive [46] 	
	5000 ppm F of NaF	Toothpaste	• Prevident 5000 [®] , Colgate–Palmolive [20]	
AmF + SnCl ₂	800 ppm Sn SnCl ₂ + 500 ppmF of NaF+ AmF	Mouthrinse	• Elmex [®] Erosion Protection dental rinse, GABA Schweiz, Colgate-Palmolive [12,50]	
$AmF + SnF_2$	250 ppm F of AmF + SnF ₂	Mouthrinse	Meridol [®] , Colgate-Palmolive [34]	
	1450 ppmF of AmF + SnF ₂	Toothpaste	• Meridol [®] , GABA Benelux [35]	
SnF ₂	0.4–0.454% SnF ₂ , 1100–1450 ppm F of SnF ₂	Toothpaste	 Crest[®] Pro HealthTM Advanced Gum Protection, The Procter & Gamble [33,42,43] Solidox, Lilleborg [51] 	
	1450 ppm F of NaF + SnF_2	Toothpaste	 Oral-B[®] Pro-Expert Sensitive; The Procter & Gamble [21,47] 	
CPP-ACP	10% CPP-ACP	Gum	• Trident Total [®] /Recaldent [®] , Trident [74]	
	10% CPP-ACP	Cream	 GC Tooth Mousse, GC Company [73,75,81] MI Paste, GC company [80] 	
	10% CPP-ACP with 900 ppm F of NaF	Cream	 GC Tooth Mousse Plus, GC Company [81] MI Paste Plus, GC company [75,80] 	
	2% CPP-ACP + 22,600 ppm F of NaF	Varnish	 MI varnish, GC Company [13] 	
CSSP	CSSP with 1450 ppm F of SMFP	Toothpaste	Regenerate TM Enamel Science, Unilever Oral Care [83,86]	
β-ΤСΡ	5% TCP + 22,600 ppm F of NaF	Varnish	• Clinpro white varnish, 3M ESPE [13]	
n-HAP	2.25–20% n-HAP	Toothpaste	 Biorepair Sensitive Repair, Coswell [95] PrevDent, PrevDent International [90] 	
	1% n-HAP + 1450 ppm F of NaF	Toothpaste	ApaCare, Cumdente [90]	
Arginine	1.5% Arginine + 1450 ppm F of SMFP 8% Arginine + 1450 ppm F of SMFP	Toothpaste	 Colgate[®] Maximum Cavity Protection, Colgate-Palmolive [115,117] 	
		Toothpaste	 Colgate Sensitive Pro-Relief[®], Colgate-Palmolive [114,118] 	
Chitosan	0.5% Chitosan + 1400 ppm F AmF/NaF + 3500 ppm Sn SnCl ₂	Toothpaste	• Elmex [®] Erosionschutz and Elmex [®] Erosion Protection, GABA Schweiz, Colgate-Palmolive [47]	
CHX	0.2% Chlorhexidine	Mouthrinse	 Periogard, Colgate-Palmolive [159] Klorhex, Drogsan Pharmaceutical [161] 	

 Table 1. Summary of commercially available anti-erosive agents in the literature.

NaF, sodium fluoride; SnF₂, stannous fluoride; AmF, amine fluoride; SnCl₂, stannous chloride; CPP-ACP, casein phosphopeptide amorphous calcium phosphate; CPP-ACFP, casein phosphopeptide amorphous calcium fluoride phosphate; CSP, calcium silicate and sodium phosphate; β -TCP, β -tricalcium phosphate; F, fluoride; SMFP, sodium monofluorophosphate; ppm, parts per million; n-HAP, nano hydroxyapatite; CHX, chlorhexidine.

Based on previous studies, we have summarised the anti-erosive properties of the topical anti-erosive agents (shown in Figure 3). Most anti-erosive agents can precipitate on the tooth surface and form a protective layer [21,47]. This protective layer reduces the mineral ion exchange between acids and the tooth [103,116]. Some agents such as fluoride and magnesium hydroxide can modify the hydroxyapatite crystal in the tooth and make it more acid-resistant [148,151]. Fluoride agents and most organic compounds can modify the salivary pellicle's thickness, compositions or ultrastructure and can enhance the anti-erosive capacity of salivary pellicle [16,17,33,76,124]. Some agents are alkaline and can neutralise the acidity of the erosive solutions on the tooth surface [65,149,156]. Calcium phosphate-based agents can supply calcium or phosphate ions that are lost from the tooth surface in acidic conditions [36,71,109–111]. Some agents can inhibit MMPs, preserve the demineralised organic matrix of the dentine structure and hamper acid diffusion [83,141,152,154,159,160]. The properties of each anti-erosive agent are shown in Table 2.

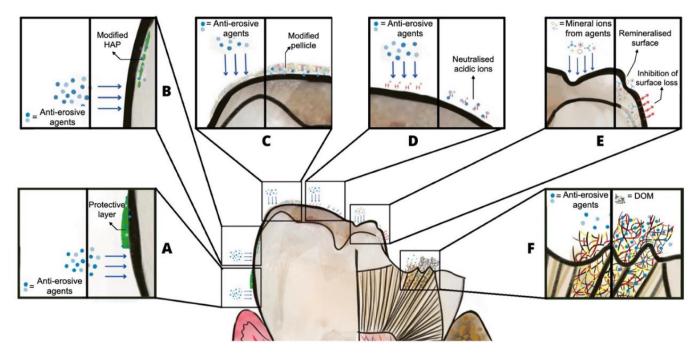


Figure 3. Anti-erosive properties of the topical anti-erosive agents: (**A**) form a protective layer on the tooth surface, (**B**) modify hydroxyapatite, (**C**) modify salivary pellicle, (**D**) neutralise acidic ions, (**E**) supply mineral ions and (**F**) inhibit MMP activation.

Future research should focus on enhancing the anti-erosive effects of topical antierosive agents. Most topical anti-erosive agents are used to prevent dental erosion or to control dental erosion in the early stages. The effectiveness of these agents on the management of advanced dental erosion or erosive tooth wear is not satisfactory. The effects of the topical anti-erosive agents may be increased by adding one or more anti-erosive properties. Favourable properties of anti-erosive agents includes forming a protective layer on the tooth surface, modifying hydroxyapatite crystal, modifying salivary pellicle, supplying calcium/phosphate ions, neutralising the acidity and inhibiting MMP activation (shown in Figure 3). No topical anti-erosive agents have all of these favourable properties. Therefore, developing novel agents with most or all the properties and enhanced antierosive effects are essential for the nonrestorative management of dental erosion.

	Form a Protective Layer	Modify Hydroxy- Apatite	Modify Salivary Pellicle	Neutralise Acid	Supply Mineral Ions	Inhibit MMPs
			Fluoride agents			
NaF	\checkmark	\checkmark		×	×	
AmF	V	V	$\frac{\sqrt{2}}{2}$	×	×	?
SnF ₂	V	v v	1	×	×	
TiF_4	v v	V	?	×	×	?
SDF	$\sqrt[n]{}$	$\sqrt[v]{}$?	\checkmark	×	\checkmark
	•	Calciun	n phosphate-base			
CPP-ACP	\checkmark	×	/	×	\checkmark	×
CSSP	v v	×	$\stackrel{}{?}$	×	V	×
β-ΤСΡ	v	×	?	×	v	×
n-HAP	v v	×	?	×	V	×
TMP/SHP	V	\checkmark	/*	×	V	×
LPP	V	V ×	V /*	×	V ./	×
Pyrophosphate	V	. /	?	×	V	×
Calcium lactate	v v	$\stackrel{\checkmark}{\times}$?	×	V V	×
	v	(Organic compound	ds	v	
PVM/MA	/	×	?	×	×	×
Carbopol	\checkmark	×	/*	×	×	×
PGA	V	×	V /*	×	×	×
Arginine		×	$\stackrel{\vee}{?}$	×	×	×
Aspartame	$\frac{\sqrt{2}}{2}$	×	?	×	×	×
CaneCPI-5		×	•	×	×	,
Casein				×	×	
Chitosan	\checkmark		\checkmark			×
Palm oil	$\frac{\sqrt{2}}{2}$	×	\checkmark	×	×	\checkmark
	<u>،</u>	×		×	×	×
EGCG	\checkmark	×	\checkmark	×	×	\checkmark
Euclea	\checkmark	×		×	×	./
natalensis	v		v ?			v
Proanthocyanidin	\checkmark	×	· · · · · · · · · · · · · · · · · · ·	×	×	\checkmark
EPT:O	,	N.	Other agents		,	
EBTiO ₂		×	?	×		×
Mg(OH) ₂	\checkmark	\checkmark	?	\checkmark	×	×
FeSO ₄	\checkmark	×	?	×	×	\checkmark
NaHCO ₃	?	×	?	\checkmark	×	×
CHX	\checkmark	×	?	?	×	\checkmark

Table 2. Summary of the properties of topical anti-erosive agents in the literature.

 $\sqrt{}$, positive impact; $\sqrt{*}$, negative impact; \times , no property; ?, unknown; NaF, sodium fluoride; SnF2, stannous fluoride; AmF, amine fluoride; TiF₄, titanium tetrafluoride; SDF, silver diamine fluoride; CPP-ACP, casein phosphopeptide amorphous calcium phosphate; CPP-ACFP, casein phosphopeptide amorphous calcium fluoride phosphate; CSSP, calcium silicate and sodium phosphate; β -TCP, β -tricalcium phosphate; TMP, sodium trimetaphosphate; SHP, sodium hexametaphosphate; LPP, linear sodium polyphosphate; n-HAP, nano-hydroxyapatite; PVM/MA, polymethylvinylether-maleic anhydride; PGA, propylene glycol alginate; EGCG, epigallocatechin gallate; CaneCPI-5, sugarcane cystatin; EBTiO₂, nano eggshell/titanium dioxide; Mg(OH)₂, magnesium hydroxide; FeSO₄, ferrous sulfate; NaHCO₃, sodium bicarbonate; CHX, chlorhexidine.

Clinical studies with higher evidence levels are needed to prove the effectiveness of these topical anti-erosive agents. Although several of the anti-erosive agents provided promising results in protecting the tooth from dental erosion, it should be noted that most studies of anti-erosive agents were in vitro studies or in situ studies that cannot simulate the complex oral environment [162]. The results are not comparable with those found in clinical situations due to the difference in the erosive challenge, the small number of subjects and the short follow-up period.

4. Conclusions

Dental erosion is a common oral health problem that requires early intervention. Topical anti-erosive agents are commonly used for the nonrestorative management of dental erosion. These agents can be categorised as topical fluorides, calcium phosphatebased agents, organic compounds and other anti-erosive agents. Most topical anti-erosive agents are used to prevent dental erosion or control dental erosion in the early stages. Future research should be conducted to validate the clinical effectiveness of the anti-erosive agents and to develop topical agents with enhanced anti-erosive effects.

Author Contributions: D.D.C. wrote the original draft and prepared all figures and tables; O.Y.Y. conceptualised the study, designed the figures and tables, and revised the manuscript; I.X.Y. and W.Y.H.L. critically commented on the manuscript; C.H.C. conceptualised and critically commented on the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the General Research Fund of Research Grants Council of Hong Kong SAR, China (No. 17100820).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors thank Steven Morrell for proofreading this manuscript and Suthasinee Inchai for contributing to the design of the figures.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Schlueter, N.; Amaechi, B.T.; Bartlett, D.; Buzalaf, M.A.R.; Carvalho, T.S.; Ganss, C.; Hara, A.T.; Huysmans, M.C.D.; Lussi, A.; Moazzez, R.; et al. Terminology of erosive tooth wear: Consensus report of a workshop organized by the ORCA and the cariology research group of the IADR. *Caries Res.* 2020, 54, 2–6. [CrossRef]
- 2. Seong, J.; Claydon, N.; Macdonald, E.; Garner, S.; Newcombe, R.G.; West, N. A randomised clinical trial to determine the abrasive effect of the tongue on human enamel loss with and without a prior erosive challenge. *J. Dent.* **2017**, *58*, 48–53. [CrossRef]
- Warreth, A.; Abuhijleh, E.; Almaghribi, M.A.; Mahwal, G.; Ashawish, A. Tooth surface loss: A review of literature. *Saudi Dent. J.* 2020, *32*, 53–60. [CrossRef]
- 4. Gupta, M.; Pandit, I.; Srivastava, N.; Gugnani, N. Dental erosion in children. J. Oral Health Comm. Dent. 2009, 3, 56–61.
- Lussi, A.; Buzalaf, M.A.R.; Duangthip, D.; Anttonen, V.; Ganss, C.; João-Souza, S.H.; Baumann, T.; Carvalho, T.S. The use of fluoride for the prevention of dental erosion and erosive tooth wear in children and adolescents. *Eur. Arch. Paediatr. Dent.* 2019, 20, 517–527. [CrossRef]
- 6. Schlueter, N.; Luka, B. Erosive tooth wear—A review on global prevalence and on its prevalence in risk groups. *Br. Dent. J.* **2018**, 224, 364–370. [CrossRef]
- 7. Okunseri, C.; Wong, M.C.M.; Yau, D.T.W.; McGrath, C.; Szabo, A. The relationship between consumption of beverages and tooth wear among adults in the United States. *J. Public Health Dent.* **2015**, *75*, 274–281. [CrossRef]
- 8. Peutzfeldt, A.; Jaeggi, T.; Lussi, A. Restorative therapy of erosive lesions. Monogr. Oral Sci. 2014, 25, 253–261. [CrossRef]
- 9. Belmar da Costa, M.; Delgado, A.H.S.; Pinheiro de Melo, T.; Amorim, T.; Mano Azul, A. Analysis of laboratory adhesion studies in eroded enamel and dentin: A scoping review. *Biomater. Investig. Dent.* **2021**, *8*, 24–38. [CrossRef]
- 10. Taji, S.; Seow, W.K. A literature review of dental erosion in children. Aust. Dent. J. 2010, 55, 358–367. [CrossRef]
- 11. Colombo, M.; Dagna, A.; Moroni, G.; Chiesa, M.; Poggio, C.; Pietrocola, G. Effect of different protective agents on enamel erosion: An in vitro investigation. *J. Clin. Exp. Dent.* **2019**, *11*, 113–118. [CrossRef] [PubMed]
- 12. Da Silva, C.V.; Ramos-Oliveira, T.M.; Mantilla, T.F.; de Freitas, P.M. Frequency of application of AmF/NaF/SnCl2 solution and its potential in controlling human enamel erosion progression: An in situ study. *Caries Res.* 2017, *51*, 141–148. [CrossRef] [PubMed]
- 13. Bayrak, S.; Tuloglu, N.; Bicer, H.; Sen Tunc, E. Effect of fluoride varnish containing CPP-ACP on preventing enamel erosion. *Scanning* **2017**, 2017, 1897825–1897827. [CrossRef]
- 14. Buzalaf, M.A.R.; Magalhães, A.C.; Wiegand, A. Alternatives to fluoride in the prevention and treatment of dental erosion. *Monogr. Oral Sci.* **2014**, *25*, 244–252. [CrossRef]
- 15. O'Toole, S.; Mistry, M.; Mutahar, M.; Moazzez, R.; Bartlett, D. Sequence of stannous and sodium fluoride solutions to prevent enamel erosion. *J. Dent.* **2015**, *43*, 1498–1503. [CrossRef]
- 16. Ainoosah, S.E.; Levon, J.; Eckert, G.J.; Hara, A.T.; Lippert, F. Effect of silver diamine fluoride on the prevention of erosive tooth wear in vitro. *J. Dent.* 2020, 103, 1–6. [CrossRef] [PubMed]

- 17. Da Silva Ávila, D.M.; Zanatta, R.F.; Scaramucci, T.; Aoki, I.V.; Torres, C.R.G.; Borges, A.B. Randomized in situ trial on the efficacy of Carbopol in enhancing fluoride/stannous anti-erosive properties. *J. Dent.* **2020**, *101*, 1–6. [CrossRef]
- Nehme, M.; Jeffery, P.; Mason, S.; Lippert, F.; Zero, D.T.; Hara, A.T. Erosion remineralization efficacy of gel-to-foam fluoride toothpastes in situ: A randomized clinical trial. *Caries Res.* 2016, *50*, 62–70. [CrossRef] [PubMed]
- 19. Nehme, M.; Parkinson, C.R.; Zero, D.T.; Hara, A.T. Randomised study of the effects of fluoride and time on in situ remineralisation of acid-softened enamel. *Clin. Oral Investig.* **2019**, *23*, 4455–4463. [CrossRef] [PubMed]
- Ren, Y.-F.; Liu, X.; Fadel, N.; Malmstrom, H.; Barnes, V.; Xu, T. Preventive effects of dentifrice containing 5000 ppm fluoride against dental erosion in situ. J. Dent. 2011, 39, 672–678. [CrossRef] [PubMed]
- 21. Hooper, S.; Seong, J.; Macdonald, E.; Claydon, N.; Hellin, N.; Barker, M.L.; He, T.; West, N.X. A randomised in situ trial, measuring the anti-erosive properties of a stannous-containing sodium fluoride dentifrice compared with a sodium fluoride/potassium nitrate dentifrice. *Int. Dent. J.* **2014**, *64*, 35–42. [CrossRef]
- 22. Lussi, A.; Jaeggi, T.; Zero, D. The role of diet in the aetiology of dental erosion. Caries Res. 2004, 38, 34-44. [CrossRef] [PubMed]
- Scaramucci, T.; Borges, A.B.; Lippert, F.; Frank, N.E.; Hara, A.T. Sodium fluoride effect on erosion–abrasion under hyposalivatory simulating conditions. Arch. Oral Biol. 2013, 58, 1457–1463. [CrossRef]
- Kensche, A.; Buschbeck, E.; König, B.; Koch, M.; Kirsch, J.; Hannig, C.; Hannig, M. Effect of fluoride mouthrinses and stannous ions on the erosion protective properties of the in situ pellicle. *Sci. Rep.* 2019, *9*, 1–10. [CrossRef]
- Kato, M.T.; Bolanho, A.; Zarella, B.L.; Salo, T.; Tjäderhane, L.; Buzalaf, M.A.R. Sodium fluoride inhibits MMP-2 and MMP-9. J. Dent. Res. 2014, 93, 74–77. [CrossRef]
- Kato, M.T.; Leite, A.L.; Hannas, A.R.; Buzalaf, M.A.R. Gels containing MMP inhibitors prevent dental erosion in situ. J. Dent. Res. 2010, 89, 468–472. [CrossRef]
- Creeth, J.E.; Kelly, S.A.; Martinez-Mier, E.A.; Hara, A.T.; Bosma, M.L.; Butler, A.; Lynch, R.J.M.; Zero, D.T. Dose–response effect of fluoride dentifrice on remineralisation and further demineralisation of erosive lesions: A randomised in situ clinical study. J. Dent. 2015, 43, 823–831. [CrossRef]
- Turssi, C.P.; Hara, A.T.; Amaral, F.L.B.; França, F.M.G.; Basting, R.T. Calcium lactate pre-rinse increased fluoride protection against enamel erosion in a randomized controlled in situ trial. *J. Dent.* 2014, 42, 534–539. [CrossRef]
- Danelon, M.; Pessan, J.P.; Prado, K.M.; Ramos, J.P.; Emerenciano, N.G.; Moretto, M.J.; Martinhon, C.C.R.; Delbem, A.C.B. Protective effect of fluoride varnish containing trimetaphosphate against dentin erosion and erosion/abrasion: An in vitro study. *Caries Res.* 2020, 54, 292–296. [CrossRef]
- Ganss, C.; Lussi, A.; Schlueter, N. Dental erosion as oral disease. insights in etiological factors and pathomechanisms, and current strategies for prevention and therapy. *Am. J. Dent.* 2012, 25, 351–364.
- Frese, C.; Wohlrab, T.; Sheng, L.; Kieser, M.; Krisam, J.; Wolff, D. Clinical effect of stannous fluoride and amine fluoride containing oral hygiene products: A 4-year randomized controlled pilot study. *Sci. Rep.* 2019, *9*, 7681. [CrossRef] [PubMed]
- 32. De Carvalho Leal, I.; Costa, W.K.D.F.; Passos, V.F. Fluoride dentifrice containing calcium silicate and sodium phosphate salts on dental erosion: In vitro study. *Arch. Oral Biol.* 2020, *118*, 1–6. [CrossRef]
- West, N.X.; He, T.; Hellin, N.; Claydon, N.; Seong, J.; Macdonald, E.; Farrell, S.; Eusebio, R.; Wilberg, A. Randomized in situ clinical trial evaluating erosion protection efficacy of a 0.454% stannous fluoride dentifrice. *Int. J. Dent. Hyg.* 2019, 17, 261–267. [CrossRef]
- Schlueter, N.; Klimek, J.; Ganss, C. Efficacy of tin-containing solutions on erosive mineral loss in enamel and dentine in situ. *Clin.* Oral Investig. 2011, 15, 361–367. [CrossRef]
- 35. Huysmans, M.C.; Jager, D.H.; Ruben, J.L.; Unk, D.E.; Klijn, C.P.; Vieira, A.M. Reduction of erosive wear in situ by stannous fluoride-containing toothpaste. *Caries Res.* **2011**, *45*, 518–523. [CrossRef] [PubMed]
- Olivan, S.R.G.; Sfalcin, R.A.; Fernandes, K.P.S.; Ferrari, R.A.M.; Horliana, A.C.R.T.; Motta, L.J.; Ortega, S.M.; Pinto, M.M.; Deana, A.M.; Bussadori, S.K. Preventive effect of remineralizing materials on dental erosion lesions by speckle technique: An in vitro analysis. *Photodiagn. Photodyn. Ther.* 2020, 29, 1–6. [CrossRef] [PubMed]
- Arnold, W.H.; Dorow, A.; Langenhorst, S.; Gintner, Z.; Bánóczy, J.; Gaengler, P. Effect of fluoride toothpastes on enamel demineralization. BMC Oral Health 2006, 6, 1–8. [CrossRef]
- Ganss, C.; Neutard, L.; von Hinckeldey, J.; Klimek, J.; Schlueter, N. Efficacy of a Tin/Fluoride rinse: A randomized in situ trial on erosion. J. Dent. Res. 2010, 89, 1214–1218. [CrossRef]
- 39. Wegehaupt, F.J.; Sener, B.; Attin, T.; Schmidlin, P.R. Anti-erosive potential of amine fluoride, cerium chloride and laser irradiation application on dentine. *Arch. Oral Biol.* 2011, *56*, 1541–1547. [CrossRef]
- 40. Da Silva Ávila, D.M.; Zanatta, R.F.; Scaramucci, T.; Aoki, I.V.; Torres, C.R.G.; Borges, A.B. Influence of bioadhesive polymers on the protective effect of fluoride against erosion. *J. Dent.* **2016**, *56*, 45–52. [CrossRef] [PubMed]
- 41. Bellamy, P.G.; Harris, R.; Date, R.F.; Mussett, A.J.; Manly, A.; Barker, M.L.; Hellin, N.; West, N.X. In situ clinical evaluation of a stabilised, stannous fluoride dentifrice. *Int. Dent. J.* **2014**, *64*, 43–50. [CrossRef]
- West, N.X.; Hooper, S.M.; O'Sullivan, D.; Hughes, N.; North, M.; MacDonald, E.L.; Davies, M.; Claydon, N.C.A. In situ randomised trial investigating abrasive effects of two desensitising toothpastes on dentine with acidic challenge prior to brushing. *J. Dent.* 2012, 40, 77–85. [CrossRef]
- 43. West, N.X.; Hellin, N.; Eusebio, R.; He, T. The erosion protection efficacy of a stabilized stannous fluoride dentifrice: An in situ randomized clinical trial. *Am. J. Dent.* **2019**, *32*, 138–142.

- Zhao, X.; He, T.; He, Y.; Chen, H. Efficacy of a stannous-containing dentifrice for protecting against combined erosive and abrasive tooth wear in situ. Oral Health Prev. Dent. 2020, 18, 619–624. [CrossRef] [PubMed]
- Schlueter, N.; Neutard, L.; von Hinckeldey, J.; Klimek, J.; Ganss, C. Tin and fluoride as anti-erosive agents in enamel and dentine in vitro. *Acta Odontol. Scand.* 2010, 68, 180–184. [CrossRef] [PubMed]
- 46. West, N.X.; Seong, J.; Hellin, N.; Eynon, H.; Barker, M.L.; He, T. A clinical study to measure anti-erosion properties of a stabilized stannous fluoride dentifrice relative to a sodium fluoride/triclosan dentifrice. *Int. J. Dent. Hyg.* 2017, *15*, 113–119. [CrossRef]
- Machado, A.; Sakae, L.; Niemeyer, S.H.; Carvalho, T.S.; Amaechi, B.; Scaramucci, T. Anti-erosive effect of rinsing before or after toothbrushing with a fluoride/stannous ions solution: An in situ investigation: Application order of fluoride/tin products for erosive tooth wear. J. Dent. 2020, 101, 103450. [CrossRef] [PubMed]
- 48. Körner, P.; Wiedemeier, D.B.; Attin, T.; Wegehaupt, F.J. Prevention of enamel softening by rinsing with a calcium solution before dental erosion. *Caries Res.* 2020, 54, 127–133. [CrossRef]
- Lussi, A.; Carvalho, T.S. The future of fluorides and other protective agents in erosion prevention. *Caries Res.* 2015, 49, 18–29. [CrossRef]
- 50. De Souza, B.M.; Santi, L.R.P.; de Souza Silva, M.; Buzalaf, M.A.R.; Magalhães, A.C. Effect of an experimental mouth rinse containing NaF and TiF4 on tooth erosion and abrasion in situ. *J. Dent.* **2018**, *73*, 45–49. [CrossRef]
- 51. Hove, L.H.; Stenhagen, K.R.; Holme, B.; Tveit, A.B. The protective effect of SnF2 containing toothpastes and solution on enamel surfaces subjected to erosion and abrasion in situ. *Eur. Arch. Paediatr. Dent.* **2014**, *15*, 237–243. [CrossRef] [PubMed]
- 52. Barlow, A.; Sufi, F.; Mason, S. Evaluation of different fluoridated dentifrice formulations using an in situ erosion remineralization model. *J. Clin. Dent.* **2009**, *20*, 192–198. [PubMed]
- 53. Wang, X.; Lussi, A. Assessment and management of dental erosion. Dent. Clin. N. Am. 2010, 54, 565–578. [CrossRef] [PubMed]
- 54. Magalhães, A.C.; Levy, F.M.; Rizzante, F.A.; Rios, D.; Buzalaf, M.A.R. Effect of NaF and TiF4 varnish and solution on bovine dentin erosion plus abrasion in vitro. *Acta Odontol. Scand.* **2012**, *70*, 160–164. [CrossRef]
- De Souza, B.M.; Santi, L.R.P.; João-Souza, S.H.; Carvalho, T.S.; Magalhães, A.C. Effect of titanium tetrafluoride/sodium fluoride solutions containing chitosan at different viscosities on the protection of enamel erosion in vitro. *Arch. Oral Biol.* 2020, 120, 1–5. [CrossRef]
- 56. Yamashita, J.M.; Torres, N.M.; Moura-Grec, P.G.; Marsicano, J.A.; Sales-Peres, A.; Sales-Peres, S.H.C. Role of arginine and fluoride in the prevention of eroded enamel: An in vitro model. *Aust. Dent. J.* **2013**, *58*, 478–482. [CrossRef]
- 57. Mantilla, T.F.; Turssi, C.P.; Ramos-Oliveira, T.M.; Silva, C.V.D.; Suzuki, L.C.; Freitas, P.M.D. The in situ effect of titanium tetrafluoride gel on erosion/abrasion progression in human dentin. *Braz. Dent. J.* **2017**, *28*, 337–345. [CrossRef]
- Mantilla, T.F.; Silva, C.V.D.; Ramos-Oliveira, T.M.; Turssi, C.P.; Ribeiro, M.S.; Freitas, P.M.D. TiF₄ gel effects on tubular occlusion of eroded/abraded human dentin. *Microsc. Res. Tech.* 2017, *80*, 1182–1188. [CrossRef] [PubMed]
- Magalhães, A.C.; Rios, D.; Honório, H.M.; Delbem, A.C.B.; Buzalaf, M.A.R. Effect of 4% titanium tetrafluoride solution on the erosion of permanent and deciduous human enamel: An in situ/ex vivo study. J. Appl. Oral Sci. 2009, 17, 56–60. [CrossRef]
- 60. Wiegand, A.; Waldheim, E.; Sener, B.; Magalhães, A.C.; Attin, T. Comparison of the effects of TiF₄ and NaF solutions at pH 1.2 and 3.5 on enamel erosion in vitro. *Caries Res.* **2009**, *43*, 269–277. [CrossRef]
- 61. Yu, H.; Attin, T.; Wiegand, A.; Buchalla, W. Effects of various fluoride solutions on enamel erosion in vitro. *Caries Res.* 2010, 44, 390–401. [CrossRef] [PubMed]
- 62. Hove, L.H.; Holme, B.; Stenhagen, K.R.; Tveit, A.B. Protective effect of TiF₄ solutions with different concentrations and pH on development of erosion-like lesions. *Caries Res.* **2011**, *45*, 64–68. [CrossRef] [PubMed]
- 63. Wiegand, A.; Hiestand, B.; Sener, B.; Magalhães, A.C.; Roos, M.; Attin, T. Effect of TiF₄, ZrF₄, HfF₄ and AmF on erosion and erosion/abrasion of enamel and dentin in situ. *Arch. Oral Biol.* **2009**, *55*, 223–228. [CrossRef]
- 64. Hove, L.H.; Holme, B.; Young, A.; Tveit, A.B. The protective effect of TiF4, SnF2 and NaF against erosion-like lesions in situ. *Caries Res.* 2008, 42, 68–72. [CrossRef] [PubMed]
- 65. Mei, M.L.; Nudelman, F.; Marzec, B.; Walker, J.M.; Lo, E.C.M.; Walls, A.W.; Chu, C.H. Formation of fluorohydroxyapatite with silver diamine fluoride. *J. Dent. Res.* 2017, *96*, 1122–1128. [CrossRef] [PubMed]
- Zhao, I.S.; Mei, M.L.; Burrow, M.F.; Lo, E.C.-M.; Chu, C.-H. Effect of silver diamine fluoride and potassium iodide treatment on secondary caries prevention and tooth discolouration in cervical glass ionomer cement restoration. *Int. J. Mol. Sci.* 2017, *18*, 340. [CrossRef] [PubMed]
- 67. Soares-Yoshikawa, A.L.; Cury, J.A.; Tabchoury, C.P.M. Fluoride concentration in SDF commercial products and their bioavailability with demineralized dentine. *Braz. Dent. J.* 2020, *31*, 257–263. [CrossRef] [PubMed]
- Ahn, J.-H.; Kim, J.-W.; Yoon, Y.-M.; Lee, N.-Y.; Lee, S.-H.; Jih, M.-K. Time-dependent anti-demineralization effect of silver diamine fluoride. *Children* 2020, 7, 251. [CrossRef] [PubMed]
- Cardenas, A.F.M.; Siqueira, F.S.F.; Morales, L.A.R.; Araujo, L.C.R.; Campos, V.S.; Bauer, J.R.; Reis, A.; Loguercio, A.D. Influence of silver diamine fluoride on the adhesive properties of interface resin-eroded dentin. *Int. J. Adhes. Adhes.* 2021, 106, 1–9. [CrossRef]
- 70. Crystal, Y.O.; Niederman, R. Evidence-based dentistry update on silver diamine fluoride. *Dent. Clin. N. Am.* 2018, 63, 45–68. [CrossRef] [PubMed]
- Viana, Í.E.L.; Lopes, R.M.; Silva, F.R.O.; Lima, N.B.; Aranha, A.C.C.; Feitosa, S.; Scaramucci, T. Novel fluoride and stannousfunctionalized β-tricalcium phosphate nanoparticles for the management of dental erosion. J. Dent. 2020, 92, 1–6. [CrossRef]

- 72. Somani, R.; Jaidka, S.; Singh, D.J.; Arora, V. Remineralizing potential of various agents on dental erosion. *J. Oral Biol. Craniofac. Res.* **2014**, *4*, 104–108. [CrossRef]
- 73. Yu, H.; Jiang, N.-W.; Ye, X.-Y.; Zheng, H.-Y.; Attin, T.; Cheng, H. In situ effect of tooth mousse containing CPP-ACP on human enamel subjected to in vivo acid attacks. *J. Dent.* **2018**, *76*, 40–45. [CrossRef]
- Jordão, M.C.; Alencar, C.R.B.; Mesquita, I.M.; Buzalaf, M.A.R.; Magalhães, A.C.; Machado, M.A.A.M.; Honório, H.M.; Rios, D. In situ effect of chewing gum with and without CPP-ACP on enamel surface hardness subsequent to ex vivo acid challenge. *Caries Res.* 2016, 50, 325–330. [CrossRef]
- 75. Wiegand, A.; Attin, T. Randomised in situ trial on the effect of milk and CPP-ACP on dental erosion. *J. Dent.* **2014**, *42*, 1210–1215. [CrossRef]
- 76. Agrawal, N.; Shashikiran, N.D.; Singla, S.; Ravi, K.S.; Kulkarni, V.K. Effect of remineralizing agents on surface microhardness of primary and permanent teeth after erosion. *J. Dent. Child.* **2014**, *81*, 117–121.
- 77. Grewal, N.; Kudupudi, V.; Grewal, S. Surface remineralization potential of casein phosphopeptide-amorphous calcium phosphate on enamel eroded by cola-drinks: An in-situ model study. *Contemp. Clin. Dent.* **2013**, *4*, 331–337. [CrossRef]
- De Alencar, C.R.B.; Magalhães, A.C.; de Andrade Moreira Machado, M.A.; de Oliveira, T.M.; Honório, H.M.; Rios, D. In situ effect of a commercial CPP-ACP chewing gum on the human enamel initial erosion. J. Dent. 2014, 42, 1502–1507. [CrossRef]
- 79. Zawaideh, F.I.; Owais, A.I.; Mushtaha, S. Effect of CPP-ACP or a potassium nitrate sodium fluoride dentifrice on enamel erosion prevention. *J. Clin. Pediatr. Dent.* 2017, *41*, 135–140. [CrossRef]
- Fernandes, L.H.F.; Alencar, C.R.B.D.; Melo, J.B.D.C.A.D.; Rios, D.; Honório, H.M.; Cavalcanti, A.L. In situ effect of intra-oral application of pastes containing CPP-ACP or CPP-ACPF against initial enamel erosion. *Pesqui. Bras. Odontopediatria Clín. Integr.* 2019, 19, 1–9. [CrossRef]
- 81. Srinivasan, N.; Kavitha, M.; Loganathan, S.C. Comparison of the remineralization potential of CPP–ACP and CPP–ACP with 900 ppm fluoride on eroded human enamel: An in situ study. *Arch. Oral Biol.* **2010**, *55*, 541–544. [CrossRef]
- 82. Fanfoni, L.; Costantinides, F.; Berton, F.; Marchesi, G.; Polo, L.; di Lenarda, R.; Nicolin, V. From erosion to remineralization: The possible role of two topic home devices used as combined treatment. *Appl. Sci.* **2020**, *10*, 4093. [CrossRef]
- Ionta, F.Q.; dos Santos, N.M.; Mesquita, I.M.; Dionísio, E.J.; Cruvinel, T.; Honório, H.M.; Rios, D. Is the dentifrice containing calcium silicate, sodium phosphate, and fluoride able to protect enamel against chemical mechanical wear? An in situ/ex vivo study. *Clin. Oral Investig.* 2019, 23, 3713–3720. [CrossRef]
- 84. João-Souza, S.H.; Scaramucci, T.; Bühler Borges, A.; Lussi, A.; Saads Carvalho, T.; Corrêa Aranha, A.C. Influence of desensitizing and anti-erosive toothpastes on dentine permeability: An in vitro study. *J. Dent.* **2019**, *89*, 1–7. [CrossRef] [PubMed]
- Joiner, A.; Schäfer, F.; Naeeni, M.M.; Gupta, A.K.; Zero, D.T. Remineralisation effect of a dual-phase calcium silicate/phosphate gel combined with calcium silicate/phosphate toothpaste on acid-challenged enamel in situ. J. Dent. 2014, 42, 53–59. [CrossRef]
- Marianna Lopes, R.; Scaramucci, T.; Walker, C.L.; Feitosa, S.A.; Aranha, A.C.C. In situ evaluation of desensitizing toothpastes for protecting against erosive tooth wear and its characterization. *Clin. Oral Investig.* 2021, 25, 6857–6870. [CrossRef]
- 87. De Oliveira, A.F.B.; Mathews, S.M.; Ramalingam, K.; Amaechi, B. The effectiveness of an NaF rinse containing fTCP on eroded enamel remineralization. *J. Public Health* **2016**, *24*, 147–152. [CrossRef]
- Amaechi, B.T.; Karthikeyan, R.; Mensinkai, P.K.; Najibfard, K.; Mackey, A.C.; Karlinsey, R.L. Remineralization of eroded enamel by a NaF rinse containing a novel calcium phosphate agent in an in situ model: A pilot study. *Clin. Cosmet. Investig. Dent.* 2010, 2, 93–100. [CrossRef]
- 89. Mathews, M.S.; Amaechi, B.T.; Ramalingam, K.; Ccahuana-Vasquez, R.A.; Chedjieu, I.P.; Mackey, A.C.; Karlinsey, R.L. In situ remineralisation of eroded enamel lesions by NaF rinses. *Arch. Oral Biol.* **2012**, *57*, 525–530. [CrossRef] [PubMed]
- 90. Ince, S.G.; Banu Ermis, R. The in situ potential of synthetic nano-hydroxyapatite for tooth enamel repair. *Bioinspired Biomim. Nanobiomater.* **2021**, *10*, 78–86. [CrossRef]
- 91. Esteves-Oliveira, M.; Santos, N.M.; Meyer-Lueckel, H.; Wierichs, R.J.; Rodrigues, J.A. Caries-preventive effect of anti-erosive and nano-hydroxyapatite-containing toothpastes in vitro. *Clin. Oral Investig.* **2016**, *21*, 291–300. [CrossRef]
- Tschoppe, P.; Zandim, D.L.; Martus, P.; Kielbassa, A.M. Enamel and dentine remineralization by nano-hydroxyapatite toothpastes. J. Dent. 2011, 39, 430–437. [CrossRef]
- Bordea, I.R.; Candrea, S.; Alexescu, G.T.; Bran, S.; Băciuț, M.; Băciuț, G.; Lucaciu, O.; Dinu, C.M.; Todea, D.A. Nano-hydroxyapatite use in dentistry: A systematic review. *Drug Metab. Rev.* 2020, 52, 319–332. [CrossRef]
- 94. Shellis, R.P.; Featherstone, J.D.B.; Lussi, A. Understanding the chemistry of dental erosion. *Monogr. Oral Sci.* 2014, 25, 163–179. [CrossRef] [PubMed]
- Abou Neel, E.A.; Bakhsh, T.A. An eggshell-based toothpaste as a cost-effective treatment of dentin hypersensitivity. *Eur. J. Dent.* 2021, 15, 733–740. [CrossRef] [PubMed]
- Missel, E.M.C.; Cunha, R.F.; Vieira, A.E.M.; Cruz, N.V.S.; Castilho, F.C.N.; Delbem, A.C.B. Sodium trimetaphosphate enhances the effect of 250 ppm. fluoride toothpaste against enamel demineralization in vitro. *Eur. J. Oral Sci.* 2016, 124, 343–348. [CrossRef] [PubMed]
- Lussi, A. Dental erosion—Novel remineralizing agents in prevention or repair. Adv. Dent. Res. 2009, 21, 13–16. [CrossRef] [PubMed]
- 98. Pancote, L.P.; Manarelli, M.M.; Danelon, M.; Delbem, A.C.B. Effect of fluoride gels supplemented with sodium trimetaphosphate on enamel erosion and abrasion: In vitro study. *Arch. Oral Biol.* **2013**, *59*, 336–340. [CrossRef]

- Danelon, M.; Pessan, J.P.; Santos, V.R.d.; Chiba, E.K.; Garcia, L.S.G.; de Camargo, E.R.; Delbem, A.C.B. Fluoride toothpastes containing micrometric or nano-sized sodium trimetaphosphate reduce enamel erosion in vitro. *Acta Odontol. Scand.* 2018, 76, 119–124. [CrossRef] [PubMed]
- Moretto, M.J.; Delbem, A.C.B.; Manarelli, M.M.; Pessan, J.P.; Martinhon, C.C.R. Effect of fluoride varnish supplemented with sodium trimetaphosphate on enamel erosion and abrasion: An in situ/ex vivo study. J. Dent. 2013, 41, 1302–1306. [CrossRef]
- 101. Manarelli, M.M.; Vieira, A.E.M.; Matheus, A.A.; Sassaki, K.T.; Delbem, A.C.B. Effect of mouth rinses with fluoride and trimetaphosphate on enamel erosion: An in vitro study. *Caries Res.* 2011, 45, 506–509. [CrossRef]
- Scaramucci, T.; João-Souza, S.H.; Lippert, F.; Eckert, G.J.; Aoki, I.V.; Hara, A.T. Influence of toothbrushing on the antierosive effect of film-forming agents. *Caries Res.* 2016, 50, 104–110. [CrossRef] [PubMed]
- 103. Scaramucci, T.; Borges, A.B.; Lippert, F.; Zero, D.T.; Aoki, I.V.; Hara, A.T. Anti-erosive properties of solutions containing fluoride and different film-forming agents. *J. Dent.* 2015, *43*, 458–465. [CrossRef]
- 104. João-Souza, S.H.; Bezerra, S.J.C.; de Freitas, P.M.; de Lima, N.B.; Aranha, A.C.C.; Hara, A.T.; Scaramucci, T. In situ evaluation of fluoride-, stannous- and polyphosphate-containing solutions against enamel erosion. J. Dent. 2017, 63, 30–35. [CrossRef]
- 105. Creeth, J.E.; Parkinson, C.R.; Burnett, G.R.; Sanyal, S.; Lippert, F.; Zero, D.T.; Hara, A.T. Effects of a sodium fluoride- and phytate-containing dentifrice on remineralisation of enamel erosive lesions—An in situ randomised clinical study. *Clin. Oral Investig.* 2018, 22, 2543–2552. [CrossRef]
- 106. Parkinson, C.R.; Burnett, G.R.; Creeth, J.E.; Lynch, R.J.M.; Budhawant, C.; Lippert, F.; Hara, A.T.; Zero, D.T. Effect of phytate and zinc ions on fluoride toothpaste efficacy using an in situ caries model. *J. Dent.* **2018**, *73*, 24–31. [CrossRef]
- Borges, A.B.; Scaramucci, T.; Lippert, F.; Zero, D.T.; Hara, A.T. Erosion protection by calcium lactate/sodium fluoride rinses under different salivary flows in vitro. *Caries Res.* 2014, 48, 193–199. [CrossRef]
- 108. Bezerra, S.J.; João-Souza, S.H.; Aoki, I.V.; Borges, A.B.; Hara, A.T.; Scaramucci, T. Anti-erosive effect of solutions containing sodium fluoride, stannous chloride, and selected film-forming polymers. *Caries Res.* **2019**, *53*, 305–313. [CrossRef]
- Beyer, M.; Reichert, J.; Sigusch, B.W.; Watts, D.C.; Jandt, K.D. Morphology and structure of polymer layers protecting dental enamel against erosion. *Dent. Mater.* 2012, 28, 1089–1097. [CrossRef]
- Wang, W.; Xie, Q.; Xu, T.; Wang, Q.; Malmstrom, H.S.; Ren, Y.-F. Fluoride release and anti-erosive effects of dentifrices containing PVM/MA copolymers. J. Dent. 2012, 41, 148–154. [CrossRef] [PubMed]
- 111. Creeth, J.E.; Burnett, G.R.; Souverain, A.; Gomez-Pereira, P.; Zero, D.T.; Lippert, F.; Hara, A.T. In situ efficacy of an experimental toothpaste on enamel rehardening and prevention of demineralisation: A randomised, controlled trial. *BMC Oral Health* **2020**, *20*, 118–127. [CrossRef] [PubMed]
- 112. Da Silva Ávila, D.M.; Augusto, M.G.; Zanatta, R.F.; Scaramucci, T.; Aoki, I.V.; Torres, C.R.G.; Borges, A.B. Enhancing the anti-erosive properties of fluoride and stannous with the polymer carbopol. *Caries Res.* **2020**, *54*, 250–257. [CrossRef]
- 113. Sakae, L.O.; Niemeyer, S.H.; Bezerra, S.J.C.; Borges, A.B.; Turssi, C.P.; Scaramucci, T. The addition of propylene glycol alginate to a fluoride solution to control enamel wear: An in situ study. *Caries Res.* **2020**, *54*, 517–523. [CrossRef]
- Olley, R.C.; Pilecki, P.; Hughes, N.; Jeffery, P.; Austin, R.S.; Moazzez, R.; Bartlett, D. An in situ study investigating dentine tubule occlusion of dentifrices following acid challenge. J. Dent. 2012, 40, 585–593. [CrossRef]
- 115. West, N.X.; He, T.; Macdonald, E.L.; Seong, J.; Hellin, N.; Barker, M.L.; Eversole, S.L. Erosion protection benefits of stabilized SnF2 dentifrice versus an arginine–sodium monofluorophosphate dentifrice: Results from in vitro and in situ clinical studies. *Clin. Oral Investig.* 2017, 21, 533–540. [CrossRef] [PubMed]
- 116. Pinto, S.C.S.; Bandéca, M.C.; Pinheiro, M.C.; Cavassim, R.; Tonetto, M.R.; Borges, A.H.; Sampaio, J.E.C. Preventive effect of a high fluoride toothpaste and arginine-carbonate toothpaste on dentinal tubules exposure followed by acid challenge: A dentine permeability evaluation. *BMC Res. Notes* 2014, 7, 385. [CrossRef] [PubMed]
- 117. West, N.; He, T.; Hellin, N.; Claydon, N.; Seong, J.; Macdonald, E.; Eusebio, R. Erosion protection efficacy of a 0.454% stannous fluoride dentifrice versus an arginine-containing dentifrice. *Am. J. Dent.* **2018**, *31*, 63–66.
- Seong, J.; Macdonald, E.; Newcombe, R.G.; Davies, M.; Jones, S.B.; Johnson, S.; West, N.X. In situ randomised trial to investigate the occluding properties of two desensitising toothpastes on dentine after subsequent acid challenge. *Clin. Oral Investig.* 2013, 17, 195–203. [CrossRef]
- 119. Ionta, F.; Bassoto, M.A.; dos Santos, N.; di Campli, F.; Honório, H.; Cruvinel, T.; Buzalaf, M.; Rios, D. Effect of in situ aspartame mouthwash to prevent intrinsic and extrinsic erosive tooth wear. *J. Clin. Exp. Dent.* **2020**, *12*, 638–643. [CrossRef]
- 120. Rios, D.; Ionta, F.-Q.; Rebelato, R.; Jordão, M.-C.; Wang, L.; Magalhães, A.-C.; Honório, H.-M. The effect of aspartame and pH changes on the erosive potential of cola drinks in bovine enamel: An in vitro study. *J. Clin. Exp. Dent.* 2018, 10, 933–937. [CrossRef] [PubMed]
- 121. Pelá, V.T.; Lunardelli, J.G.Q.; Tokuhara, C.K.; Gironda, C.C.; Silva, N.D.G.; Carvalho, T.S.; Santiago, A.C.; Souza, B.M.; Moraes, S.M.; Henrique-Silva, F.; et al. Safety and in situ antierosive effect of CaneCPI-5 on dental enamel. *J. Dent. Res.* 2021, 100, 1344–1350. [CrossRef]
- 122. Santos, L.A.; Martini, T.; Câmara, J.V.F.; Reis, F.N.; Cassia Ortiz, A.D.; Camiloti, G.D.; Levy, F.M.; Shibao, P.Y.T.; Honorio, H.M.; Henrique-Silva, F.; et al. Solutions and gels containing a sugarcane-derived cystatin (CaneCPI-5) reduce enamel and dentin erosion in vitro. *Caries Res.* 2021, 55, 594–602. [CrossRef] [PubMed]

- 123. Santiago, A.C.; Khan, Z.N.; Miguel, M.C.; Gironda, C.C.; Soares-Costa, A.; Pelá, V.T.; Leite, A.L.; Edwardson, J.M.; Buzalaf, M.A.R.; Henrique-Silva, F. A new sugarcane cystatin strongly binds to dental enamel and reduces erosion. *J. Dent. Res.* 2017, 96, 1051–1057. [CrossRef] [PubMed]
- 124. Chawhuaveang, D.D.; Yu, O.Y.; Yin, I.X.; Lam, W.Y.-H.; Mei, M.L.; Chu, C.-H. Acquired salivary pellicle and oral diseases: A literature review. *J. Dent. Sci.* 2021, *16*, 523–529. [CrossRef] [PubMed]
- 125. Jager, D.H.J.; Vissink, A.; Timmer, C.J.; Bronkhorst, E.; Vieira, A.M.; Huysmans, M.C.D.N.J.M. Reduction of erosion by proteincontaining toothpastes. *Caries Res.* 2013, 47, 135–140. [CrossRef]
- 126. White, A.J.; Gracia, L.H.; Barbour, M.E. Inhibition of dental erosion by casein and casein-derived proteins. *Caries Res.* 2011, 45, 13–20. [CrossRef]
- 127. Beltrame, A.P.C.; Suchyta, D.; Abd Alraheam, I.; Mohammed, A.; Schoenfisch, M.; Walter, R.; Almeida, I.C.; Souza, L.C.; Miguez, P.A. Effect of phosphorylated chitosan on dentin erosion: An in vitro study. *Caries Res.* **2018**, *52*, 378–386. [CrossRef]
- 128. Schlueter, N.; Klimek, J.; Ganss, C. Randomised in situ study on the efficacy of a Tin/Chitosan toothpaste on erosive-abrasive enamel loss. *Caries Res.* **2013**, *47*, 574–581. [CrossRef]
- 129. Schlueter, N.; Klimek, J.; Ganss, C. Effect of a chitosan additive to a Sn²⁺-containing toothpaste on its anti-erosive/anti-abrasive efficacy—A controlled randomised in situ trial. *Clin. Oral Investig.* **2014**, *18*, 107–115. [CrossRef]
- 130. Souza, B.M.; Machado, P.F.; Vecchia, L.R.P.; Magalhães, A.C. Effect of chitosan solutions with or without fluoride on the protection against dentin erosion in vitro. *Eur. J. Oral Sci.* **2020**, *128*, 495–500. [CrossRef]
- 131. Aykut-Yetkiner, A.; Attin, T.; Wiegand, A. Prevention of dentine erosion by brushing with anti-erosive toothpastes. *J. Dent.* **2014**, 42, 856–861. [CrossRef] [PubMed]
- Ionta, F.Q.; Alencar, C.R.B.D.; Santos, N.M.D.; Bergantin, B.T.P.; Val, P.P.; Honório, H.M.; Oliveira, T.M.D.; Rios, D. Effect of palm oil alone or associated to stannous solution on enamel erosive-abrasive wear: A randomized in situ/ex vivo study. *Arch. Oral Biol.* 2018, 95, 68–73. [CrossRef]
- 133. Ionta, F.Q.; Alencar, C.R.B.D.; Val, P.P.; Boteon, A.P.; Jordão, M.C.; Honório, H.M.; Buzalaf, M.A.R.; Rios, D. Effect of vegetable oils applied over acquired enamel pellicle on initial erosion. J. Appl. Oral Sci. 2017, 25, 420–426. [CrossRef] [PubMed]
- 134. Wang, Y.-L.; Chang, H.-H.; Chiang, Y.-C.; Lu, Y.-C.; Lin, C.-P. Effects of fluoride and epigallocatechin gallate on soft-drink-induced dental erosion of enamel and root dentin. *J. Formos. Med. Assoc.* **2018**, *117*, 276–282. [CrossRef] [PubMed]
- 135. De Moraes, M.D.R.; Carneiro, J.R.M.; Passos, V.F.; Santiago, S.L. Effect of green tea as a protective measure against dental erosion in coronary dentine. *Braz. Oral Res.* 2016, 30, 1–6. [CrossRef]
- Kato, M.T.; Magalhães, A.C.; Rios, D.; Hannas, A.R.; Attin, T.; Buzalaf, M.A.R. Protective effect of green tea on dentin erosion and abrasion. J. Appl. Oral Sci. 2009, 17, 560–564. [CrossRef]
- De Souza-E-Silva, C.M.; da Silva Ventura, T.M.; de Paila Silva Cassiano, L.; de Lima Leite, A.; Buzalaf, M.A.R. Effect of gels containing chlorhexidine or epigallocatechin-3-gallate on the protein composition of the acquired enamel pellicle. *Arch. Oral Biol.* 2017, *82*, 92–98. [CrossRef] [PubMed]
- 138. Kato, M.T.; Hannas, A.R.; Cardoso, C.D.A.B.; Cassiano, L.D.P.S.; Naves, P.A.; Magalhães, A.C.; Tjäderhane, L.; Buzalaf, M.A.R. Dentifrices or gels containing MMP inhibitors prevent dentine loss: In situ studies. *Clin. Oral Investig.* 2021, 25, 2183–2190. [CrossRef] [PubMed]
- 139. Sales-Peres, S.H.D.C.; Xavier, C.N.H.; Mapengo, M.A.A.; Forim, M.R.; Silva, M.D.F.; Sales-Peres, A. Erosion and abrasioninhibiting in situ effect of the Euclea natalensis plant of African regions. *Braz. Oral Res.* **2016**, *30*, 1–7. [CrossRef] [PubMed]
- Sales-Peres, S.H.D.C.; Brianezzi, L.F.D.F.; Marsicano, J.A.; Forim, M.R.; da Silva, M.F.D.G.F.; Sales-Peres, A. Evaluation of an experimental gel containing Euclea natalensis: An in vitro study. *Evid. Based Complementary Altern. Med.* 2012, 2012, 1–6. [CrossRef]
- 141. Cardoso, F.; Boteon, A.P.; Silva, T.A.P.D.; Prakki, A.; Wang, L.; HonÓrio, H.M. In situ effect of a proanthocyanidin mouthrinse on dentin subjected to erosion. *J. Appl. Oral Sci.* 2020, *28*, 1–7. [CrossRef] [PubMed]
- 142. Boteon, A.P.; Kato, M.T.; Buzalaf, M.A.R.; Prakki, A.; Wang, L.; Rios, D.; Honório, H.M. Effect of proanthocyanidin-enriched extracts on the inhibition of wear and degradation of dentin demineralized organic matrix. *Arch. Oral Biol.* 2017, 84, 118–124. [CrossRef]
- 143. Fawzy, A.S.; Priyadarshini, B.M.; Selvan, S.T.; Lu, T.B.; Neo, J. Proanthocyanidins-loaded nanoparticles enhance dentin degradation resistance. J. Dent. Res. 2017, 96, 780–789. [CrossRef] [PubMed]
- 144. Jawale, K.; Kamat, S.; Patil, J.; Nanjannawar, G.; Chopade, R. Grape seed extract: An innovation in remineralization. *J. Conserv. Dent.* 2017, 20, 415–418. [CrossRef] [PubMed]
- 145. Onwubu, S.C.; Mdluli, P.S.; Singh, S.; Nyembe, S.; Thakur, R. Corrigendum to "An in situ evaluation of the protective effect of nano eggshell/titanium dioxide against erosive acids". *Int J. Dent.* 2019, 2019, 7209168. [CrossRef] [PubMed]
- Onwubu, S.C.; Mdluli, P.S.; Singh, S.; Tlapana, T. A novel application of nano eggshell/titanium dioxide composite on occluding dentine tubules: An in vitro study. *Braz. Oral Res.* 2019, 33, 1–10. [CrossRef] [PubMed]
- 147. Onwubu, S.C.; Mdluli, P.S.; Singh, S.; Madikizela, L.; Ngombane, Y. Characterization and in vitro evaluation of an acid resistant nanosized dental eggshell-titanium dioxide material. *Adv. Powder Technol.* **2019**, *30*, 766–773. [CrossRef]
- 148. Passos, V.F.; Rodrigues, L.K.A.; Santiago, S.L. The effect of magnesium hydroxide-containing dentifrice using an extrinsic and intrinsic erosion cycling model. *Arch. Oral Biol.* **2018**, *86*, 46–50. [CrossRef]

- Park, K.S.; Kim, B.J.; Lih, E.; Park, W.; Lee, S.H.; Joung, Y.K.; Han, D.K. Versatile effects of magnesium hydroxide nanoparticles in PLGA scaffold–mediated chondrogenesis. *Acta Biomater.* 2018, 73, 204–216. [CrossRef]
- 150. Passos, V.F.; Rodrigues Gerage, L.K. Magnesium hydroxide-based dentifrice as an anti-erosive agent in an in situ intrinsic erosion model. *Am. J. Dent.* **2017**, *30*, 137–141.
- Abdallah, M.-N.; Eimar, H.; Bassett, D.C.; Schnabel, M.; Ciobanu, O.; Nelea, V.; McKee, M.D.; Cerruti, M.; Tamimi, F. Diagenesisinspired reaction of magnesium ions with surface enamel mineral modifies properties of human teeth. *Acta Biomater.* 2016, 37, 174–183. [CrossRef]
- 152. Kato, M.T.; Leite, A.L.; Hannas, A.R.; Oliveira, R.C.; Pereira, J.C.; Tjäderhane, L.; Buzalaf, M.A.R. Effect of iron on matrix metalloproteinase inhibition and on the prevention of dentine erosion. *Caries Res.* **2010**, *44*, 309–316. [CrossRef]
- 153. Kato, M.T.; Leite, A.L.; Hannas, A.R.; Calabria, M.P.; Magalhães, A.C.; Pereira, J.C.; Buzalaf, M.A.R. Impact of protease inhibitors on dentin matrix degradation by collagenase. J. Dent. Res. 2012, 91, 1119–1123. [CrossRef]
- 154. Sales-Peres, S.H.C.; Pessan, J.P.; Buzalaf, M.A.R. Effect of an iron mouthrinse on enamel and dentine erosion subjected or not to abrasion: An in situ/ex vivo study. *Arch. Oral Biol.* 2006, *52*, 128–132. [CrossRef] [PubMed]
- 155. Kato, M.T.; Buzalaf, M.A.R. Iron supplementation reduces the erosive potential of a cola drink on enamel and dentin in situ. *J. Appl. Oral Sci.* **2012**, *20*, 318–322. [CrossRef]
- 156. Messias, D.C.F.; Turssi, C.P.; Hara, A.T.; Serra, M.C. Sodium bicarbonate solution as an anti-erosive agent against simulated endogenous erosion. *Eur. J. Oral Sci.* **2010**, *118*, 385–388. [CrossRef]
- 157. Messias, D.; Serra, M.C.; Turssi, C.P. Potential effect of sodium bicarbonate-containing dentifrice in controlling enamel erosion in situ. *Am. J. Dent.* **2008**, *21*, 300–302.
- 158. Messias, D.C.F.; Martins, M.E.N.; Serra, M.C.; Turssi, C.P. Feasibility of using sodium bicarbonate solution as a damage-limiting strategy for erosion lesions. *Oral Health Prev. Dent.* **2008**, *6*, 155–158.
- 159. Magalhães, A.C.; Wiegand, A.; Rios, D.; Hannas, A.; Attin, T.; Buzalaf, M.A.R. Chlorhexidine and green tea extract reduce dentin erosion and abrasion in situ. *J. Dent.* **2009**, *37*, 994–998. [CrossRef]
- Charone, S.; Cardoso, C.D.A.B.; Kato, M.T.; Ducati, P.; Fukushima, R.; Gennaro, G.; Magalhães, A.C.; Buzalaf, M.A.R. The effect of mouthwashes containing biguanides on the progression of erosion in dentin. *BMC Oral Health* 2014, 14, 131–136. [CrossRef] [PubMed]
- 161. Ozan, G.; Sar Sancakli, H.; Yucel, T. Effect of black tea and matrix metalloproteinase inhibitors on eroded dentin in situ. *Microsc. Res. Tech.* **2020**, *83*, 834–842. [CrossRef]
- 162. Yu, O.Y.; Zhao, I.S.; Mei, M.L.; Lo, E.C.-M.; Chu, C.-H. A review of the common models used in mechanistic studies on demineralization-remineralization for cariology research. *Dent. J.* 2017, *5*, 20. [CrossRef]