Quantification of Oral Candidal Carriage Rate and Prevalence of Oral Candidal Species in HIV Patients with and Without Highly Active Antiretroviral Therapy

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Abstract

Background: Most documents review decrease in the prevalence of HIV related oral lesions to the tune of 10-50% following the advent of HAART. However long term use of HAART on oral health status of HIV infected subjects is poorly documented. Also antifungal agents can effectively treat mucosal candidiasis. However their use can lead to colonization with less susceptible strains among normal susceptible strains. Aims and Objectives: To know the candidal carriage rate (i.e. presence/absence of candidal growth), candidal density(CFUs/ml) & species variations (species diversity) in HIV positive individuals with and without highly active anti-retroviral therapy (HAART), attending the regional voluntary Counselling and Confidential Testing Centre (VCCTC). Materials and Methods: The study population were categorized into 3 groups. Method followed were Germ tube test, Chlamydospore formation test, CHROM-Agar test. Results: Quantification comparison study of candidal carriage rate, density with detection of various candidal species in the oral cavity of HIV-positive individuals with and without HAART therapy treatment prooved higher candidal carriage rate and lower density than Non-HAART category.

Keywords: AIDS, candidiasis, carriage rate, HIV, Voluntary Counseling and Confidential Testing Centre

INTRODUCTION

Due to of the emergence of other and newer species of *Candida* as pathogens and a development of change in the susceptibility pattern of *Candida albicans*, it necessitates the isolation and identification of the causative species.^[1-6]

The advent of highly active antiretroviral therapy (HAART) has changed the scenario of HIV infection and has become a standard treatment for HIV infection.^[7] It induces a marked reduction in viral load and increase in CD4⁺ cell count leading to decline in the morbidity and mortality of HIV-infected patients.^[8,9] In HAART therapy, a range of different combination of drugs are used and each combination of drugs has advantages and disadvantages. They are administered simultaneously to bring about sustained block in viral replication and restore immune function as well as to minimize resistance to drugs.^[8]

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Aims and objectives

- 1. To analyze the oral candidal carriage rates in HIV-positive individuals who are undergoing antiretroviral therapy and in patients who are not on antiretroviral therapy
- 2. To quantitatively assess the candidal density in the above-mentioned groups
- 3. To assess the strain diversity in the above mentioned groups.

MATERIALS AND METHODS

The study population included 30 patients each of Group I and Group II attending the Voluntary Counseling and Confidential

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Testing Centre (VCCTC), ART center.

- 1. Group I: Included 30 patients who were people living with HIV/AIDS naive to HAART
- 2. Group II: Included 30 patients who were people living with HIV/AIDS who were registered for HAART and started on drug regimen

The patients from Group I and Group II were direct walk-in patients in the VCCTC, who were positive for HIV by three tests (COOMBS AIDS; TRI DOT; and TRI LINE), according to the guidelines of the National AIDS Control Organization.

3. Group III: Included 30 HIV seronegative healthy subjects as controls.

Inclusion criteria

- 30 HIV-positive patients who were being treated with HAART regimen, at least for a duration of 1 month and with a known CD4⁺ T-lymphocyte count
- 2. 30 HIV-positive patients who were not yet initiated with the treatment of HAART and with a known CD4⁺ T-lymphocyte count.

Exclusion criteria

- 1. Patients with a history of tuberculosis, diabetes mellitus, cardiovascular diseases, rheumatoid arthritis, and any systemic ailments were excluded. Pregnant females and denture wearers were also excluded from the study
- 2. HIV-positive patients with HAART duration less than a month were also excluded.

Methods

- 1. All the patients included in this study were asked to rinse with 10 ml of normal saline for 60 s before expectorating into the sterile container [Figure 1: Figure 2]
- 2. The oral rinse sample was immediately taken to the microbiology department for inoculation of the sample on a Sabouraud dextrose agar (SDA), specific for candidal growth
- 3. 0.1 ml of undiluted oral rinse sample was inoculated on two plates of SDA containing chloramphenicol
- 4. 0.1 ml of diluted (10^{-1}) oral rinse sample of HIV-positive



Figure 1: Armamentarium used for clinical examination of patients

patients was also inoculated on two plates of SDA plates containing chloramphenicol.

Note: 10^{-1} dilution of the oral rinse sample is prepared by mixing 0.1 ml of oral rinse sample of HIV-positive patients with 0.9 ml of sterile normal saline.

- 5. The above plates were then incubated aerobically at 37°C for 48–72 h
- 6. The growth appeared in 2–3 days as creamy white, smooth, pasty colonies. In a few, the growth was observed within 24 h, i.e., overnight incubation
- 7. The complete growth of any candidal colonies on the culture plates was recorded as a positive growth and the subject as positive candidal carrier (i.e., positive candidal carriage rate)
- 8. The number of colonies on each plate was counted manually, and an average count of both the diluted (10⁻¹) plates was taken
- 9. The number of colony-forming units (CFU's) per ml was calculated to indicate the candidal density [Figures 3-6].

The calculation was as follows:

- N no of colonies in 0.1 ml of 10⁻¹ dilution (since 0.1 ml of 10⁻¹ was spread on the agar plate)
- 10N no of colonies in1 ml of 10⁻¹ dilution of the normal saline
- 100N colonies in 1 ml of sterile saline which gives the CFU's/ml.

The representative colonies of *Candida* species on SDA plate were then purified on blood agar with a streak method. Further identification of species was done by VITEK test using the purified colonies grown on blood agar [Figures 7 and 8].

Some of the samples were tested retrospectively with the help of germ tube test, chlamydospore formation test, and with CHROM agar.

Germ tube test

The principle of this test is the ability of *C. albicans* and its variants to produce germ tubes when incubated with various



Figure 2: Sterile bottle containing10ml of sterile normal saline for patients oral rinse sample



Figure 3: Armamentarium used for inoculation of oral rinse sample



Figure 4: Working place for the aseptic inoculation of oral rinse sample



Figure 5: Positive growth of *Candida* species in undiluted and diluted (10^{-1}) plates



Figure 6: Negative growth of *Candida* species in undiluted and diluted (10^{-1}) plates



Figure 7: VITEK machine for rapid identification of different strains of *Candida* species

substances such as human or sheep serum, rabbit plasma, egg albumin, saliva, tissue culture medium, thioglycolate trypticase soya broth, and various peptone mediums. This is a rapid screening procedure for differentiating *C. albicans* from other *Candida* species.

Chlamydospore formation test

Cultivation on cornmeal agar facilitates and appreciates chlamydospore formation. This property is peculiar to



Figure 8: Yeast identification card after performing the test in VITEK machine

C. albicans and to very rare isolates of *Candida tropicalis* and *Candida stellatoidea*. The chlamydospore has been defined as a thick walled nondeciduous intercalary or asexual spore formed by rounding off of a cell or cells.

CHROMagar test

CHORMagar is a novel differential culture medium for isolation and presumptive to identification of different species

of *Candida* and has revealed mixtures of *Candida* species in many types of clinical samples more often than would have been expected. The species of *Candida* can be identified by different colored colonies. Different colored colonies produced on the CHROM agar are as follows:

 C. albicans: light green; 2. Candida glabrata: purple 3; C. tropicalis: blue with pink hallow; 4. Candida parapsilosis: cream; 5. Candida krusei: pink (rough, fuzzy spreading) l; and 6. Candida dubliniensis: dark green.^[10-89]

Statistical analysis

- 1. To test the association between the candidal carriage rate, candidal density, and species diversity among the HIV-positive individuals with and without HAART, Chi-square test was applied
- To test the association between the candidal carriage rate, candidal density, and species diversity with the CD4 count ≤200 and >200 cells/mm³, again, the Chi-square test was applied.

Results and Observations

The present study was aimed at quantification of candidal carriage rate, candidal density, and identification of different species of *Candida* in the oral cavity of HIV-positive individuals with and without HAART therapy.

The study included patients visiting the outpatient department of the institute hospital, as well as outpatients visiting private HIV clinics (direct walk-in clients of VCCTC, ART center).

To test if there was any association between normal individuals and HIV-positive individuals and the presence and absence (i.e., prevalence) of *Candida*, the contingencies are prepared as shown in the Table 1a.

To analyze if there was any association, the Chi-square test of independence of attributes was applied at 95% confidence level.

It was observed that the calculated value of Chi-square was 4.364 which was significant with P = 0.037, indicating that there was a positive relation between the two attributes (i.e., between normal and HIV-positive individuals).

Further, it was also verified from the percentage that among the HIV-positive individuals, 60% have shown the presence of *Candida*, while among the normal individuals, 36.66% have shown the presence of *Candida*.

The subjects who were chosen for the study, i.e., HIV-positive individuals were further divided into two groups, those who were on HAART treatment and those who were not on HAART treatment.

Table 1b shows the cross-contingency for this classification, i.e., individuals receiving HAART or not and whether *Candida* is present or absent.

To understand if there was any association between the treatment and prevalence of *Candida*, once again, Chi-square test was applied at 95% confidence level. It showed that

calculated value of Chi-square was 6.944 which was significant with P = 0.008.

Regarding the patients who were on HAART treatment, 76.66% showed the presence of *Candida*. Analysis of patients who were not on HAART treatment revealed that 43.33% showed the prevalence of *Candida*.

Thus, it can be concluded that the prevalence of *Candida* was more in patients receiving HAART than those who were naïve to HAART therapy.

To identify the relationship between the candidal density and the presence or absence of HIV, the cross-tabulation, as shown in Table 2a, is used. This shows *Candida* density count between 1–2000 CFU's/ml and more than 2000 CFU's/ml. The Chi-square test at 95% of confidence level applied to the data showed that the calculated value of Chi-square was 4.96 with P = 0.084 which indicated that there was no significant relationship between the two, i.e., between the normal and HIV-positive individuals.

Of the patients who showed positive candidal carriage rate, 36 were in HIV-positive patients and 11 were in normal individuals. It was found that 41.66% of HIV-positive patients had candidal density >2000 CFU's/ml compared to 27.27% of normal individuals.

Table 1a: Correlation of candidal carriage rate between normal individuals and HIV-positive patients [Graph 1a]

Inidviduals	<i>Candida</i> species (present)	<i>Candida</i> species (absent)	Total
HIV (<i>n</i> =60)	60% (36)	40% (24)	60
Normal (n=30)	36.66% (11)	63.33% (19)	30
Total (n=90)	52.22% (47)	47.77% (43)	90

Table 1b: Correlation of candidal carriage rate betweenHIV-positive patients with and without highly activeantiretroviral therapy [Graph 1b]

HIV individuals	<i>Candida</i> species (present)	<i>Candida</i> species (absent)	Total
With HAART (n=30)	76.66% (23)	23.33% (7)	30
Without HAART (n=30)	43.33% (13)	56.66% (17)	30
Total (<i>n</i> =60)	60% (36)	40% (24)	60
Total (<i>n</i> =60)	60% (36)	40% (24)	60

HAART: Highly active antiretroviral therapy

Table 2a: Correlation of candidal density (colony-forming unit's/ml) between normal individuals and HIV-positive patients [Graph 2a]

Individuals	1-2000 (CFU's/ml)	>2000 (CFU's/ml)	Total
HIV	58.33% (21)	41.66% (15)	36
Normal	72.72% (08)	27.27% (03)	11
Total	29	18	47
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CFU's: Colony-forming unit's

Further, we also tried to understand whether there was any association between the HAART treatment and the candidal density count, as seen in Table 2b, and applying Chi-square test at 95% confidence level. This showed that the Chi-square calculated value was 9.995 with P = 0.007 which was significant. Thus, it was observed that there was a relationship between the candidal density and the HAART treatment.

It was seen that 69.56% of the patients who were on HAART treatment had candidal density count between 1 and 2000 CFU's/ml, compared to 38.46% of individuals who were not on HAART.

To test if there was any association between the normal and HIV-positive individuals and the candidal diversity, the contingencies are prepared, as shown Table 3a.

To assess whether there was a significant relationship between the species diversity between the normal individuals and HIV seropositive patients, the Chi-square test at 95% of confidence level was applied to the data which showed that the calculated value of Chi-square was 6.39 with P = 0.041. This showed that the correlation of the species diversity between the normal individuals and HIV-positive patients was significant.

Table 3b shows the total *Candida* species isolated in the present study which included the HIV-positive individuals with and



Graph 1: (a) Correlation of candidal carriage rate between normal individuals and HIV-positive patients. (b) Correlation of candidal carriage rate between HIV-positive patients with and without highly active antiretroviral therapy

without HAART. The Chi-square test at 95% confidence level was applied to the data. The calculated value of Chi-square was 8.033 with P = 0.18, which indicated that there was a significant relationship between the species diversity in HAART and non-HAART HIV-seropositive individuals.

Table 2b: Correlation of candidal density (colony-forming unit's/ml) between HIV-positive patients with and without highly active antiretroviral therapy [Graph 2b]

HIV individuals	1-2000 (CFU's/ml)	>2000 (CFU's/ml)	Total
With HAART	69.56% (16)	30.43% (07)	23
Without HAART	38.46% (05)	61.53% (08)	13
Total	21	15	36

HAART: Highly active antiretroviral therapy, CFU's: Colony-forming unit's

Table 3a: Correlation of candidal species diversity between normal individuals and HIV-positive patients [Graph 3a]

Individuals	Albicans	Nonalbicans	Total
HIV	80.55% (29)	19.44% (07)	36
Normal	100% (11)	0% (0)	11
Total	40	7	47

Table 3b: Correlation of Candidal species diversity between HIV-positive patients with and without highly active antiretroviral therapy [Graph 3b]

HIV individuals	Albicans	Nonalbicans	Total
With HAART	78.26% (18)	21.73% (5)	23
Without HAART	92.30% (12)	7.69% (1)	13
Total	30	6	36

HAART: Highly active antiretroviral therapy

Table 4a: Correlation between CD4 count (\leq 200, >200 cells/mm³) in HIV-positive patients and candidal carriage rate [Graph 4a]

CD4 count (cells/mm³)	<i>Candida</i> species (present)	<i>Candida</i> species (absent)	Total
≤200	65.51% (19)	34.48% (10)	29
>200	54.83% (17)	45.16% (14)	31
Total	36	24	60

Table 4b: Correlation between CD4 count (≤ 200 , > 200 cells/mm³) in HIV-positive patients and candidal density (colony-forming unit's/ml) [Graph 4b]

CD4 count	Candidal density (CFU's/ml)		
(cells/mm³)	1-2000 (CFU's/ml)	>2000 (CFU's/ml)	
≤200	52.63% (10)	47.36% (9)	19
>200	64.70% (11)	35.29% (6)	17
Total	21	15	36
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CFU's: Colony-forming unit's

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To know the association between the prevalence of *Candida* and the CD4 count, Chi-square test was applied at 95% confidence level. The standard classification of CD4 count \leq 200 and >200 cells/mm³ was used for HIV-positive individuals under the study. The calculated value of Chi-square was 0.712 with *P* = 0.399 which showed no significant relation between the prevalence of *Candida* carriage rate and CD4 count of HIV-positive individuals.

Chi-square was applied at 95% confidence level to check whether there was any association between CD4 count and candidal density. The calculated value of Chi-square was 1.249 with P = 0.536 which was not significant. Thus, it can be stated that there was no relation between CD4 count and candidal density of the individuals.

To know the relationship between CD4 count and candidal diversity, the Chi-square test was applied at 95% of confidence

Table 4c: Correlation between CD4 count (\leq 200, >200 cells/mm³) in HIV-positive patients and candidal species diversity [Graph 4c]

CD4 count (cells/mm³)	Candidal species diversity		Total
	Albicans	Nonalbicans	
≤200	78.94% (15)	21.05% (4)	19
>200	82.35% (14)	17.64% (3)	17
Total	29	7	36



Graph 2: (a) Correlation of candidal density (CFU's/ml) between normal individuals and HIV-positice patients. (b) Correlation of candidal density (CFU's/ml) between HIV-positive patients with and without highly active antiretroviral therapy

level. The calculated value of Chi-square was 0.066 with P = 0.799. The results indicated that there was no significant relationship between CD4 count and candidal diversity [Table 4a-c and Graphs 1-4].

CONCLUSION

HIV-positive individuals undergoing HAART therapy showed higher candidal carriage rate and lower candidal density than the non-HAART group.

Candidal density is a more valuable marker in predicting the development of OC. Hence, we conclude that HAART definitely has a role in preventing HIV-infected seropositive individuals from developing overt candidiasis. Furthermore, the emergence of increased number of NCAC species in individuals undergoing HAART suggests one to explore the dynamics of HAART action on *C. albicans* species in more detail and with different parameters.

Further studies should be conducted to gain insight into the effect of HAART on the albicans and nonalbicans species and the resistance to it at the molecular level. Furthermore, no significant association was found between OPC and CD4 count in our study, and hence, HIV viral load should be taken in consideration as a parameter.

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Graph 3: (a) Correlation of candidal species diversity between normal individuals and HIV-positive patients. (b) correlation of candidal species diversity between HIV +ve patients with and without highly active antiretroviral therapy



Graph 4: (a) Correlation between CD4 count (≤ 200 , > 200 cells/mm³) in HIV-positive patients and candidal carriage rate. (b) Correlation between CD4 count (≤ 200 , > 200) in HIV-positive patients and candidal density (CFU's/ml). (c) Correlation between CD4 count (≤ 200 , > 200 cells/mm³) in HIV-positive patients and candidal species diversity

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- 1. Douek DC, Roederer M, Koup RA. Emerging concepts in the immunopathogenesis of AIDS. Annu Rev Med 2009;60:471-84.
- Cunningham AL, Donaghy H, Harman AN, Kim M, Turville SG. Manipulation of dendritic cell function by viruses. Curr Opin Microbiol 2010;13:524-9.
- Meurman JH, Siikala E, Richardson M, Rautemaa R. Non-Candida albicans Candida yeasts of the oral cavity. In: Communicating Current Research and Educational Topics and Trends in Applied Microbiology. 2nd edition, vol. 5; 2007. p. 719-30.
- Carl JF, Judith AA. Candidiasis and HIV. HIV In Site Knowledge; 2006. p. 1-11. Available from: http://hivinsite.ucsf.edu. [Last accessed on 2010 Dec 21].

- Dodd CL, Greenspan D, Katz MH, Westenhouse JL, Feigal DW, Greenspan JS. Oral candidiasis in HIV infection: Pseudomembranous and erythematous candidiasis show similar rates of progression to AIDS. AIDS 1991;5:1339-43.
- Schorling SR, Kortinga HC, Froschb M, Mühlschlegel FA. The role of *Candida dubliniensis* in oral candidiasis in human immunodeficiency virus-infected individuals. Crit Rev Microbiol 2000;26:59-68.
- Kantheti LP, Reddy B, Ravikumar S, Anuradha CH, Chandrasekhar P, Rajeswari MR. Isolation, identification, and carriage of Candidal species in PHLAs and their correlation with immunological status in cases with and without HAART. J Oral Maxillofac Pathol 2012;16:38-44.
- Nittayananta W, Talungchit S, Jaruratanasirikul S, Silpapojakul K, Chayakul P, Nilmanat A, *et al.* Effects of long-term use of HAART on oral health status of HIV-infected subjects. J Oral Pathol Med 2010;39:397-406.
- Lederman MM. Immune restoration and CD4+ T-cell function with antiretroviral therapies. AIDS 2001;15 Suppl 2:S11-5.
- Patton LL, McKaig R, Strauss R, Rogers D, Eron JJ Jr. Changing prevalence of oral manifestations of human immuno-deficiency virus in the era of protease inhibitor therapy. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2000;89:299-304.
- Tappuni AR, Fleming GJ. The effect of antiretroviral therapy on the prevalence of oral manifestations in HIV-infected patients: A UK study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2001;92:623-8.
- Centers for Disease Control (CDC). Kaposi's sarcoma and pneumocystis pneumonia among homosexual men – New York City and California. MMWR Morb Mortal Wkly Rep 1981;30:305-8.
- Gottlieb MS, Schroff R, Schanker HM, Weisman JD, Fan PT, Wolf RA, et al. Pneumocystis carinii pneumonia and mucosal candidiasis in previously healthy homosexual men: Evidence of a new acquired cellular immunodeficiency. N Engl J Med 1981;305:1425-31.
- Coffin J, Haase A, Levy JA, Montagnier L, Oroszlan S, Teich N, et al. What to call the AIDS virus? Nature 1986;321:10.
- Rainey L. In The Search for the Cause of AIDS, Two Labs and the Titanic Egos of the Men that Ran them Took Centre Stage. Dallas Voice Print Edition; 2006.
- Prabhu SR, Kolhi A, Rao B. Human immunodeficiency virus. In: Jack N, editor. HIV and AIDS in Dental Practice. 1st ed., Ch. 11. Vol.2: Thomas Press; 2007. p. 85-92.
- Hoffmann C, Rockstroh JK, Kamps BS. Pathogenesis of HIV infection. In: Andrea R, Georg B, Mario O, editors. HIV Medicine. 15th ed., Ch. 4. Paris: Flying Publisher; 2007. p. 60-1.
- Spira AI, Marx PA, Patterson BK, Mahoney J, Koup RA, Wolinsky SM, et al. Cellular targets of infection and route of viral dissemination after an intravaginal inoculation of simian immunodeficiency virus into rhesus macaques. J Exp Med 1996;183:215-25.
- Chun TW, Carruth L, Finzi D, Shen X, DiGiuseppe JA, Taylor H, *et al.* Quantification of latent tissue reservoirs and total body viral load in HIV-1 infection. Nature 1997;387:183-8.
- McMichael AJ, Rowland-Jones SL. Cellular immune response to HIV. Nature 2000;410:980-7.
- Pantaleo G, Menzo S, Vaccarezza M, Graziosi C, Cohen O, Deramrest JF, et al. Studies in subjects with long term non progressive human immunodeficiency virus infection. N Engl J Med 1995;332:209-16.
- Downs AM, Salamina G, Ancelle-Park RA. Incubation period of vertically acquired AIDS in Europe, before wide spread use of prophylactic therapies. J Acquir Immunodefic Syndr 1995;9:297-304.
- Alder MW. Natural history and management of early HIV infection. In: Mindel A, Tenant-Flowers M, editors. ABC of AIDS. 5th ed., Ch. 4. London: BMJ Publishing Group; 2001. p. 17-22.
- Mayo Clinic. Available from: http://www.mayoclinic.com/health/ HIV-aids/DS 00005/DSECTION. [Last accessed on Last accessed on 2019 Aug 02].
- 25. Sterling TR, Chaisson RE. General clinical manifestations of human immunodeficiency virus infection (including the acute retroviral syndrome and oral, cutaneous, renal, ocular, metabolic, and cardiac diseases). In: Mandell GL, Bennett JE, Dolin R, editors. Principles and Practice of Infectious Diseases. 7th ed., Ch. 121. Philadelphia, Pa: Elsevier Churchill Livingstone; 2009.
- 26. Centre for Disease Control and Prevention. Revised classification system

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for HIV infection and expanded surveillance case definition for AIDS among adolescents and adults. Morb Mortal Wkly Rep 1992;41:1-19.

- Levy JA. HIV and Pathogenesis of AIDS. Washington DC: American Society for Microbiology; 1998.
- Schupbach J. Human immunodeficiency viruses. In: Murray PR, editor. Manual of Clinical Microbiology. Washington DC: ASM Press; 2003. p. 1253-81.
- Anneroth G, Anneroth I, Lyunch DP. Acquired immune deficiency syndrome in United States: Etiology, epidemiology, clinical manifestation. J Oral Maxillofac Surg 1986;44:956-64.
- Puelacher W, Zangerle R, Kulmer S, Waldhart E, Fritsch P, Hintner H. Oral manifestations of HIV infection. Z Stomatol 1989;86:539-48.
- Arendorf T, Holmes H. Oral manifestations associated with human immunodeficiency virus (HIV) infection in developing countries – Are there differences from developed countries? Oral Dis 2000;6:133-5.
- Kwon-Chung KJ, Bennet JE. Principles of antifungal therapy. In: Lea N, Febiger M, editors. Medical Mycology. Philadelphia 1992. p. 81.
- Neville BW, Damm DD, Allen CM, Bouquot JE. Oral and Maxillofacial Pathology. 3rd ed., Ch. 6. Missouri: Elsevier; 2009. p. 213-38.
- 34. Sono E, Masuda T, Takesko K, Kato I, Uchida K, Murayama SY, et al. Comparision of secretary acid proteinases from *Candida tropicalis*, *Candida parapsilosis*, *Candida albicans*. Microbiol Immunol 1992;36:1099-104.
- 35. Fusek M, Smith EA, Monod M, Dunn BM, Foundling SI. Extracellular aspartic proteinases from *Candida albicans*, *Candida tropicalis*, and *Candida parapsilosis* yeasts differ substantially in their specificities. Biochemistry 1994;33:9791-9.
- Vitkov L, Krautgartner WD, Hannig M, Weitgasser R, Stoiber W. Candida attachment to oral epithelium. Oral Microbiol Immunol 2002;17:60-4.
- Chander J. A Text Book of Medical Mycology. 2nd ed. New Delhi: Mehta; 2002. p. 212-31.
- Esthar S, Elad D. Candidiasis. In: Morpz WG, Hay RJ, Arnold H, editors. Topley and Wilson's Microbiology and Microbial Infections. 10th ed., Ch. 30. 2005. p. 556.
- Klein RS, Harris CA, Small CB, Moll B, Lesser M, Friedland GH. Oral candidiasis in high-risk patients as the initial manifestation of the acquired immunodeficiency syndrome. N Engl J Med 1984;311:354-8.
- Cutler JE. Putative virulence factors of *Candida albicans*. Annu Rev Microbiol 1991;45:187-218.
- Samarnayake LP. Oral mycosis in HIV infection: A review. Oral Med Oral Pathol 1992;73:171-80.
- 42. McQuillen DP, Zingman BS, Meunier F, Levitz SM. Invasive infections due to *Candida krusei*: Report of ten cases of fungemia that include three cases of endophthalmitis. Clin Infect Dis 1992;14:472-8.
- Teanpaisan R, Nittayananta W. Prevalence of *Candida* species in AIDS patients and HIV-free subjects in Thailand. J Oral Pathol Med 1998;27:4-7.
- 44. Hamza OJ, Matlee MI, Masti MJ, Simon EN, Mugusi F, MiKx FH, *et al.* Species distribution and *in vitro* antifungal susceptibility of oral yeast isolates from Tanzania HIV infected patients with primary and recurrent oropharyngeal candidiasis. BMC Microbiol 2008;8:135.
- Baradkar VP, Kumar S. Species identification of Candida isolates obtained from oral lesions of HIV infected patients. Indian J Dermatol 2009;54:385-6.
- 46. Brilhante RS, Castelo-Branco DS, Soares GD, Astete-Medrano DJ, Monteiro AJ, Cordeiro RA, *et al.* Characterization of the gastrointestinal yeast microbiota of cockatiels (*Nymphicus hollandicus*): A potential hazard to human health. J Med Microbiol 2010;59:718-23.
- Sitheeque MA, Samaranayake LP. Chronic hyperplastic candidosis/ candidiasis (*Candidal leukoplakia*). Crit Rev Oral Biol Med 2003;14:253-67.
- Roberts GD, Goodman NL. Laboratory diagnosis. In: Morpz WG, Hay RJ, Arnold H, editors. Topley and Wilson's Microbiology and Microbial infections. 10th ed., Ch. 4. Vol. 1; 2005. p. 83-95.
- Merz WG, Robert GD. Detection and Recovery of Fungi from Clinical Specimen. In: Manual of Clinical Microbiology. 8th ed. Washington, DC: American Society for Microbiology; 2003. p. 668-85.
- Rippon JW. Candidiasis and the pathogenic yeasts. In: Medical Mycology. 3rd ed. Philadelphia: W.B. Saunders; 1998. p. 536-81.

- Koneman EW, Roberts GD, Wright SE. Practical Laboratory Mycology. 2nd ed. 1st edition: The Williams and Wilkins; 1997. p. 112.
- Gupta A, Mi H, Wroe C, Jaques B, Talbot D. Fatal *Candida famata* peritonitis complicating sclerosing peritonitis in a peritoneal dialysis patient. Nephrol Dial Transplant 2006;21:2036-7.
- Milne LJ. Fungi. In: Collee JG, editor. Mackie and McCartney, Practical Medical Microbiology. 14th ed., Ch. 41. Edinburgh: Churchill Livingstone; 1996. p. 695-717.
- Odds FC, Bernaerts R. CHROMagar Candida, a new differential isolation medium for presumptive identification of clinically important *Candida* species. J Clin Microbiol 1994;32:1923-9.
- 55. Pfaller MA, Houston A, Coffmann S. Application of CHROMagar Candida for rapid screening of clinical specimens for *Candida albicans*, *Candida tropicalis, Candida krusei*, and Candida (Torulopsis) glabrata. J Clin Microbiol 1996;34:58-61.
- Graf B, Adam T, Zill E, Göbel UB. Evaluation of the VITEK 2 system for rapid identification of yeasts and yeast-like organisms. J Clin Microbiol 2000;38:1782-5.
- 57. Young RC, Bennett JE, Geelhoed GW, Levine AS. Fungemia in compromized host resistance. Ann Intern Med 1974;80:605-12.
- Murray HW, Hillman JK, Rubin BY, Kelly CD, Jacobs JL, Tyler LW, et al. Patients at risk for AIDS-related opportunistic infections. Clinical manifestations and impaired gamma interferon production. N Engl J Med 1985;313:1504-10.
- Singh K, Arunalooke C, Narang A, Sarala G. Yeast colonization and fungemia in preterm neonates in a tertiary care center. Indian J Med Res 1996;110:169-73.
- Narang A, Agrawal PB, Chakrabarti A, Kumar P. Epidemiology of systemic candidiasis in a tertiary care neonatal unit. J Trop Pediatr 1998;44:104-8.
- Narain S, Sastry JS, Mathur M, Mehta PR. Neonatal systemic candidiasis in a tertiary care centre. Indian J Med Microbiol 2003;21:56-8.
- Samaranayake LP, Fidel PL, Naglik JR, Sweet SP, Teanpaisan R, Coogan MM, *et al.* Fungal infections associated with HIV infection. Oral Dis 2002;8 Suppl 2:151-60.
- Campisi G, Pizzo G, Milici ME, Mancuso S, Margiotta V. Candidal carriage in the oral cavity of human immunodeficiency virus-infected subjects. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2002;93:281-6.
- Thompson GR 3rd, Patel PK, Kirkpatrick WR, Westbrook SD, Berg D, Erlandsen J, *et al.* Oropharyngeal candidiasis in the era of antiretroviral therapy. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010;109:488-95.
- Ranganathan K, Reddy BV, Kumarasamy N, Solomon S, Viswanathan R, Johnson NW. Oral lesions and conditions associated with human immunodeficiency virus infection in 300 South Indian patients. Oral Dis 2000;6:152-7.
- Greenspan D, Canchola AJ, MacPhail LA, Cheikh B, Greenspan JS. Effect of highly active antiretroviral therapy on frequency of oral warts. Lancet 2001;357:1411-2.
- Mocroft A, Lundgren JD. Starting highly active antiretroviral therapy: Why, when and response to HAART. J Antimicrob Chemother 2004;54:10-3.
- Ceballos-Saloberna A, Gaitan-Cepeda L, Ceballos-Gracia L, Samarnayake LP. The effect of highly active antiretroviral therapy on the prevalence of HIV associated oral candidiasis in a Spanish cohort. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2004;97:345-50.
- Hung CC, Yang YL, Lauderdale TL, McDonald LC, Hsiao CF, Cheng HH. Colonization of human immunodeficiency virus-infected patients in Taiwan with *Candida* species. Am Soc Microbiol 2005;43:1600-3.
- Gaitan-Cepeda LA, Martinez-Gonazaiez M, Ceballos-Salobrena A. Oral candidiasis as a clinical marker of immune failure in patients with HIV/ AIDS on HAART. AIDS Patient Care STDS 2005;19:70-7.
- Costa CR, Cohen AJ, Fernandes OF, Miranda K, Passos S, Souza LK, et al. Asymptomatic oral carriage of *Candida* species in HIV-infected patients in the highly active antiretroviral therapy era. Rev Inst Med Trop Sao Paulo 2006;48:257-61.
- Yang YL, Lo HJ, Hung CC, Li Y. Effects of prolonged HAART on oral colonisation with Candida and candidiasis. BMC Infect Dis 2006;6:8.

- Umadevi KM, Ranganathan K, Pavithra S, Hemalatha R, Saraswathi TR, Kumarasamy N, *et al.* Oral lesions among persons with HIV disease with and without highly active antiretroviral therapy in Southern India. J Oral Pathol Med 2007;36:136-41.
- 74. Pomarico L, Cerqueira DF, de Araujo Soares RM, de Souza IP, de Araujo Castro GF, Socransky S, *et al.* Associations among the use of highly active antiretroviral therapy, oral candidiasis, oral *Candida* species and salivary immunoglobulin A in HIV-infected children. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2009;108:203-10.
- Ortega KL, Vale DA, Magalhaes MH. Imapet of PI and NNRTI HAART-based therapy on oral lesions of Brazilian HIV – Infected patients. J Oral Pathol Med 2009;38:489-94.
- Ananthalakshmi R, Murali S, Sekar B. Association of asymptomatic oral candidal carriage, oral candidiasis with CD4 lymphocyte count in HIV/AIDS patients – A comparative study. JIADS 2011;2:6-10.
- Munro CA, Hube B. Antifungal therapy at the HAART of viral therapy. Trends Microbiol 2002;10:173-7.
- Badiee P, Alborzi A, Davarpanah MA, Shakiba E. Distribution and antifungal susceptibility of *Candida* species from mucosal sites in HIV positive patients. Arch Iran Med 2010;13:282-7.
- Pignato S, Salvo S, Coniglio MA, Marranzano M, Faro G, Giammanco G. Persistent oral and urinary *Candida* spp. carriage in Italian HIV-seropositive asymptomatic subjects. J Prev Med Hyg 2009;50:232-5.
- Jain PA, Veerabhadrudu K, Kulkarni RD, Ajantha GS, Shubhada C, Amruthkishan U. Comparative study of adherence of oral *Candida albicans* isolates from HIV sero-positive individuals and HIV sero-negative individuals to human buccal epithelial cells. Indian J Pathol Microbiol 2010;53:513-7.

- Lar PM, Pam KV, Yop T, Olukose S, Yusuf A, Dashen MM, et al. Prevalence and distribution of *Candida* species in HIV infected persons on antiretroviral therapy in Jos. J Med Med Sci 2012;3:254-9.
- Martins M, Henriques M, Ribero AP. Oral Candia carriage of patients attending a dental clinic in Braga, Portugal. Rev Iberoam Micol 2010;27:119-24.
- Tsang CS, Samarnayake LP. Oral yeast and coliforms in HIV infected individuals in Hong Kong. Mycoses 2000;43:303-8.
- Dgnani MC, Solomkin J, Anaisse EJ. Candida. In: Anaisse F, Meginnis M, Pfaller T, editors. Textbook of Clinical Mycology. 1st ed., Ch. 8. Philadelphia: Churchill Livingston; 2003. p. 195-239.
- Patel M, Shackleton JT, Coogan MM. Effect of antifungal treatment on the prevalence of yeasts in HIV-infected subjects. J Med Microbiol 2006;55:1279-84.
- Cauda R, Tacconelli E, Tumbarello M, Morace G, De Bernardis F, Torosantucci A, *et al.* Role of protease inhibitors in preventing recurrent oral candidosis in patients with HIV infection: A prospective case-control study. J Acquir Immune Defic Syndr 1999;21:20-5.
- 87. Cassone A, Tacconelli E, De Bernardis F, Tumbarello M, Torosantucci A, Chiani P, et al. Antiretroviral therapy with protease inhibitors has an early, immune reconstitution-independent beneficial effect on *Candida* virulence and oral candidiasis in human immunodeficiency virus-infected subjects. J Infect Dis 2002;185:188-95.
- Cartledge JD, Midgley J, Gazzard BG. Non-albicans oral candidosis in HIV-positive patients. J Antimicrob Chemother 1999;43:419-22.
- De Melo NR, Vilela MM, Jorge JJ, Kamei K, Miyaji M, Fukushima K, et al. HIV-1 Antiretroviral drug effect on the C. albicans hyphal growth rate by a bio-cell tracer system. Brazilian J Microbiol 2006;37:225-9.