



Genome Sequence of the Early 20th-Century Extreme Halophile *Halobacterium* sp. Strain NRC-34001

Priya DasSarma,^a Brian P. Anton,^b Jessie M. Griffith,^{a*} Karina S. Kunka,^{a§} Richard J. Roberts,^b Shiladitya DasSarma^a

^aInstitute of Marine and Environmental Technology, Department of Microbiology and Immunology, University of Maryland, Baltimore, Maryland, USA

^bNew England Biolabs, Ipswich, Massachusetts, USA

ABSTRACT *Halobacterium* sp. strain NRC-34001 is a red, extremely halophilic archaeon isolated in Canada in 1934. Single-molecule real-time sequencing revealed a 2.3-Mbp genome with a 2-Mbp chromosome and two plasmids (235 kb and 43 kb). The genome encodes all conserved core haloarchaeal groups (cHOGs) and a highly acidic proteome.

The red haloarchaeon *Halobacterium* sp. strain NRC-34001 was originally isolated as “*Serratia cutirubra*” (subsequently, *Pseudomonas*/“*Halobacter*”/*Halobacterium cutirubrum*) from Canadian salted buffalo hide by A. G. Lochhead (1890 to 1980) while investigating damaging red stains on the flesh side of salted hide (“red heat”), similar to fish spoilage (1–5). It was vital to the discovery of classical haloarchaeal traits such as hypotonic lysis, spheroplast formation, high-GC content genome, compartmentalization, and endemic halophage infection, all prior to the classification of *Halobacterium* species as members of the Archaea in the third domain of life (6–10).

The strain in this study, obtained from the American Type Culture Collection (ATCC 33170), was grown in CM⁺ (complete medium plus trace metals) at 37°C with shaking at 220 rpm (New Brunswick Innova 4230 instrument; New Brunswick, NJ, USA), as previously described (11, 12). Cells were lysed by osmotic shock, and nucleic acids were isolated using standard phenol extraction-based methods (12–15).

Single-molecule real-time (SMRT) sequencing was performed using the PacBio Sequel platform (Menlo Park, CA). A SMRTbell sequencing library was prepared from 3.9 µg genomic DNA, without further shearing, using the SMRTbell Express template prep kit 2.0. The library was purified by removing SMRTbell reads of <15 kb using the BluePippin size selection system (Sage Science, Beverly, MA) and sequenced on a single SMRT cell (Sequel Binding kit 3.0, Sequel Sequencing Plate 3.0) with a 20-h collection time. Sequencing reads were filtered and assembled *de novo* using the Microbial Assembly pipeline (SMRT Link 9.0.0.92188), run with default parameters. A total of 28,995 subreads aligned to the draft assembly, with a mean sub-read length of 15,890 bp, and the polished assembly resolved automatically into three circular contigs, with 200× mean coverage. The GC-rich genome was found to consist of a large circular chromosome (2,012,898 bp; GC content, 67.9%) and 2 plasmids, pHcu_235 (235,323 bp; GC content, 59.8%) and pHcu_43 (42,817 bp, GC content, 57.8%).

Genome annotation was performed at NCBI using the Prokaryotic Genome Annotation Pipeline (PGAP) build 3190, and the taxonomy was determined by NCBI Taxonomy (16, 17). The genome includes 1 rRNA operon and 47 tRNA genes and encodes 2,341 proteins with a calculated mean isoelectric point (pI) value of 4.9, a highly acidic proteome characteristic of haloarchaea (18–20). Based on additional in-house analysis using GeneMark.hmm 2, FeatureExtract, Sequence Manipulation Suite, and EMBOSS (21–24), all 799 conserved core haloarchaeal groups (cHOGs), 6 Orc/Cdc6 proteins, 3 TATA-binding proteins, and 4 TFB proteins (25–28) were found, as well as genes coding for bacteriorhodopsin, halorhodopsin, and sensory rhodopsins 1 and 2 (29, 30). The genome also includes a complete buoyant gas vesicle

Editor Kenneth M. Stedman, Portland State University

Copyright © 2022 DasSarma et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Shiladitya DasSarma, sdassarma@som.umaryland.edu.

*Present address: Jessie M. Griffith, Washington University, St. Louis, Missouri, USA.

§Present address: Karina S. Kunka, University of Pittsburgh, School of Dentistry, Pittsburgh, Pennsylvania, USA.

The authors declare a conflict of interest.

Received 7 December 2021

Accepted 13 December 2021

Published 13 January 2022

nanoparticle (GVNP) gene cluster, although the strain has not been shown to produce GVNPs (31, 32).

Methylation patterns were determined using the Base Modification and Motif Analysis application within the same SMRT Link environment using default settings (minimum Qmod score = 100). A single methylated DNA motif was identified (^{m4}CTAG, the product of M.Hsp34001, which is commonly found in *Halobacterium* species) (33).

Data availability. The *Halobacterium* sp. NRC-34001 genome sequence has been deposited at GenBank (accession numbers CP085884.1, CP085882.1, CP085883.1). The raw data are available in the NCBI Sequence Read Archive under the accession number SRR16600243 and the BioProject accession number PRJNA412908.

ACKNOWLEDGMENTS

Work in the DasSarma laboratory is supported by NASA Exobiology grant 80NSSC17K0263 and NIH grant AI139808. We thank Joan Slonczewski, at Kenyon College, Gambier, OH, whose students (J.M.G. and K.S.K.) were supported in the DasSarma laboratory through a NASA Astrobiology Program Early Career Collaboration Award.

B.P.A. and R.J.R. work for New England Biolabs, a company that sells research reagents, including restriction enzymes and DNA methyltransferases, to the scientific community.

REFERENCES

1. DasSarma P, DasSarma S. 2008. On the origin of prokaryotic "species": the taxonomy of halophilic Archaea. *Saline Syst* 4:5. <https://doi.org/10.1186/1746-1448-4-5>.
2. Lochhead AG. 1934. Bacteriological studies on the red discolouration in salted hides. *Can J Res* 10:275–286. <https://doi.org/10.1139/cjr34-026>.
3. Lochhead AG. 1943. Notes on the taxonomic position of the red chromogenic halophilic bacteria. *J Bacteriol* 45:574–575. <https://doi.org/10.1128/jb.45.574-575.1943>.
4. Anderson H. 1954. The reddening of salted hides and fish. *Appl Microbiol* 2:64–69. <https://doi.org/10.1128/am.2.2.64-69.1954>.
5. DasSarma P, Klebahn G, Klebahn H. 2010. Translation of Henrich Klebahn's 'Damaging agents of the klipfish—a contribution to the knowledge of the salt-loving organisms.' *Saline Syst* 6:7. <https://doi.org/10.1186/1746-1448-6-7>.
6. DasSarma P, Capes MD, DasSarma S. 2019. Comparative genomics of *Halobacterium* strains from diverse locations, p 285–322. In Das S, Dash HR (ed), *Microbial diversity in the genomic era*, 1st ed. Academic Press, Cambridge, MA. <https://doi.org/10.1016/B978-0-12-814849-5.00017-4>.
7. Kushner DJ, Masson G, Gibbons NE. 1965. Simple method for killing halophilic bacteria in contaminated solar salt. *Appl Microbiol* 13:288. <https://doi.org/10.1128/am.13.2.288-288.1965>.
8. Wais AC, Kon M, MacDonald RE, Stollar BD. 1975. Salt-dependent bacteriophage infecting *Halobacterium cutirubrum* and *H. halobium*. *Nature* 256:314–315. <https://doi.org/10.1038/256314a0>.
9. Joshi JG, Guild WR, Handler P. 1963. The presence of two species of DNA in some halobacteria. *J Mol Biol* 6:34–38. [https://doi.org/10.1016/S0022-2836\(63\)80079-0](https://doi.org/10.1016/S0022-2836(63)80079-0).
10. Moore RL, McCarthy BJ. 1969. Characterization of the deoxyribonucleic acid of various strains of halophilic bacteria. *J Bacteriol* 99:248–254. <https://doi.org/10.1128/jb.99.1.248-254.1969>.
11. Berquist BR, Müller JA, DasSarma S. 2006. Chapter 27. Genetic systems for halophilic archaea, p 649–680. In Oren A, Rainey F (ed), *Methods in microbiology*, vol 35. Elsevier Academic Press, San Diego, CA.
12. Ng WL, Yang CF, Halladay JT, Arora P, DasSarma S. 1995. Protocol 25. Isolation of genomic and plasmid DNAs from *Halobacterium halobium*, p 179–184. In DasSarma S, Fleischmann EM (ed), *Archaea, a laboratory manual: halophiles*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
13. DasSarma S. 1984. The bacterio-opsin gene in *Halobacterium halobium*: high-frequency inactivation by insertion sequences and structure of the messenger RNA. PhD dissertation. Massachusetts Institute of Technology, Cambridge, MA. https://mit.primo.exlibrisgroup.com/permalink/01MIT_INST/ejdckj/alma990002112390106761.
14. Fomenkov A, DasSarma P, Kennedy SP, Roberts RJ, DasSarma S. 2021. Complete genome and methylome analysis of the box-shaped halophilic archaeon *Haloarcula sinaiensis* ATCC 33800. *Microbiol Resour Announc* 10:e00619-21. <https://doi.org/10.1128/MRA.00619-21>.
15. DasSarma P, Anton BP, Ehrenheim HAL, Martínez FL, Guzmán D, Roberts RJ, DasSarma S. 2021. Genome sequence of *Halobacterium* sp. strain BOL4-2 isolated and cultured from Salar de Uyuni. *Microbiol Resour Announc* 10:e01045-21. <https://doi.org/10.1128/MRA.01045-21>.
16. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
17. Schoch CL, Ciuffo S, Domrachev M, Hotton CL, Kannan S, Khovanskaya R, Leipe D, McVeigh R, O'Neill K, Robbertse B, Sharma S, Soussou V, Sullivan JP, Sun L, Turner S, Karsch-Mizrachi I. 2020. NCBI taxonomy: a comprehensive update on curation, resources and tools. *Database (Oxford)* 2020: baaa062. <https://doi.org/10.1093/database/baaa062>.
18. Kozlovskii LP. 2016. IPC—Isoelectric Point Calculator. *Biol Direct* 11:55. <https://doi.org/10.1186/s13062-016-0159-9>.
19. Kennedy SP, Ng WV, Salzberg SL, Hood L, DasSarma S. 2001. Understanding the adaptation of *Halobacterium* species NRC-1 to its extreme environment through computational analysis of its genome sequence. *Genome Res* 11:1641–1650. <https://doi.org/10.1101/gr.190201>.
20. DasSarma S, DasSarma P. 2015. Halophiles and their enzymes: negativity put to good use. *Curr Opin Microbiol* 25:120–126. <https://doi.org/10.1016/j.mib.2015.05.009>.
21. Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Res* 33:W451–W454. <https://doi.org/10.1093/nar/gki487>.
22. Wernersson R. 2005. FeatureExtract—extraction of sequence annotation made easy. *Nucleic Acids Res* 33:W567–W569. <https://doi.org/10.1093/nar/gki388>.
23. Stothard P. 2000. The Sequence Manipulation Suite: JavaScript programs for analyzing and formatting protein and DNA sequences. *Biotechniques* 28:1102–1104. <https://doi.org/10.2144/00286i01>.
24. Madeira F, Park YM, Lee J, Buso N, Gur T, Madhusoodanan N, Basutkar P, Tivey ARN, Potter SC, Finn RD, Lopez R. 2019. The EMBL-EBI search and sequence analysis tools APIs in 2019. *Nucleic Acids Res* 47:W636–W641. <https://doi.org/10.1093/nar/gkz268>.
25. Capes MD, DasSarma P, DasSarma S. 2012. The core and unique proteins of haloarchaea. *BMC Genomics* 13:39. <https://doi.org/10.1186/1471-2164-13-39>.
26. DasSarma S, Capes M, DasSarma P. 2008. Chapter 1: Haloarchaeal megaplasmids, p 3–30. In Schwartz E (ed), *Microbiology monographs*. Springer, Berlin, Germany.
27. Capes MD, Coker JA, Gessler R, Grinblat-Huse V, DasSarma SL, Jacob CG, Kim J-M, DasSarma P, DasSarma S. 2011. The information transfer system of halophilic archaea. *Plasmid* 65:77–101. <https://doi.org/10.1016/j.plasmid.2010.11.005>.
28. Berquist BR, Soneja J, DasSarma S. 2005. Comparative genomic survey of information transfer systems in two diverse extremely halophilic archaea, *Halobacterium* sp. strain NRC-1 and *Haloarcula marismortui*. In Gundlach Cimerman N, Oren A, Plemenitaš A (ed), *Adaptation to life at high salt concentrations in archaea, bacteria, and eukarya. Cellular origin, life in extreme environments*, p 1–16. Springer, Berlin, Germany.

- habitats and astrobiology, vol 9. Springer, Dordrecht, Netherlands. <https://doi.org/10.1007/1-4020-3633-7>.
29. DasSarma S, Kennedy SP, Berquist B, Ng W-LV, Baliga NS, Spudich JL, Krebs MP, Eisen JA, Johnson CH, Hood L. 2001. Genomic perspective on the photobiology of *Halobacterium* species NRC-1, a phototrophic, photo-tactic, and UV-tolerant haloarchaeon. *Photosynth Res* 70:3–17. <https://doi.org/10.1023/A:1013879706863>.
30. DasSarma S, DasSarma P, Laye VJ, Schwieterman EW. 2020. Extremophilic models for astrobiology: haloarchaeal survival strategies and pigments for remote sensing. *Extremophiles* 24:31–41. <https://doi.org/10.1007/s00792-019-01126-3>.
31. DasSarma S, DasSarma P. 2015. Gas vesicle nanoparticles for antigen display. *Vaccines (Basel)* 3:686–702. <https://doi.org/10.3390/vaccines3030686>.
32. DasSarma P, DasSarma S. 2021. Gas vesicle nanoparticles, p 1–17. In eLS, vol 2. John Wiley & Sons, Hoboken, NJ. <https://doi.org/10.1002/9780470015902.a0029044>.
33. Roberts RJ, Vincze T, Posfai J, Macelis D. 2015. REBASE—a database for DNA restriction and modification: enzymes, genes and genomes. *Nucleic Acids Res* 43:D298–D299. <https://doi.org/10.1093/nar/gku1046>.