



REVIEW

The physiological functions of central nervous system pericytes and a potential role in pain [version 1; referees: 2 approved]

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Abstract

Central nervous system (CNS) pericytes regulate critical functions of the neurovascular unit in health and disease. CNS pericytes are an attractive pharmacological target for their position within the neurovasculature and for their role in neuroinflammation. Whether the function of CNS pericytes also affects pain states and nociceptive mechanisms is currently not understood. Could it be that pericytes hold the key to pain associated with CNS blood vessel dysfunction? This article reviews recent findings on the important physiological functions of CNS pericytes and highlights how these neurovascular functions could be linked to pain states.

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Introduction

Other than the retina, the central nervous system (CNS) contains the highest ratio of pericytes to endothelial cells in the body¹, yet the contribution, if any, of CNS pericytes to pain states and nociception is not understood. CNS pericytes inhabit a perivascular niche within the neurovascular unit (NVU)², a unique position interfacing the circulatory and peripheral immune systems and the central nervous parenchyma. CNS pericytes regulate critical functions of the NVU: blood-brain barrier/blood-spinal cord barrier (BBB/BSCB) integrity, cerebral (and presumably spinal cord) blood flow, clearance of toxic substances, angiogenesis, mesenchymal stem cell activity, and neuroinflammation³. As such, severe neuronal defects are observed with CNS pericyte deficiency^{4,5}. CNS pericytes have attracted interest in neuropathology, particularly with respect to their involvement in neuroinflammation, yet on the basis of a scan of the published literature on CNS pericytes, it is evident that understanding of their potential influence(s) in pain states and nociceptive mechanisms is currently severely lacking. This article reviews recent literature on the physiological functions of CNS pericytes that, when awry, could contribute or lead to the development of pain.

The multipotent nature of CNS pericytes

Pericytes, first identified and labelled as Rouget cells in 1873 by the French physiologist Charles-Marie Benjamin Rouget, are a heterogeneous population of cells and, as such, have proven a challenge to characterise both functionally and biochemically. A lack of pericyte-specific markers has significantly hindered consistency within pericyte research, and many debates discussing what constitutes a pericyte have played out⁶. Pericytes are capable of self-renewal, and express markers and behave like mesenchymal stem cells. For example, throughout the body, pericytes have the ability to replace specialized tissue-specific cells such as adipocytes⁷, myocytes⁸, myofibroblasts⁹ and odontoblasts¹⁰ in repair processes. Pericytes can also facilitate repair processes indirectly through the release of factors^{11,12}, and some of these – for example, nerve growth factor (NGF), vascular endothelial growth factor-A (VEGF-A), tumour necrosis factor-alpha (TNF α), interleukin 1 beta (IL1 β), IL6, NAD(P)H oxidase-4 (NOX4) and matrix metalloproteinase 2 (MMP2)^{13–18} – are direct neuronal sensitizers or increased levels are associated with pain states. CNS pericytes have been shown to migrate into the cortex parenchyma and differentiate into a microglia-like phenotype in a model of stroke¹⁹. The authors observed pericyte migration, proliferation, a morphological change resembling reactive microglia, and expression of IBA-1 and CD11b, the latter being an integrin strongly expressed by reactive microglia and macrophages in pain models^{20–22}. However, it is not known whether such pericyte-to-microglia differentiation occurs in pain states in which microglial activation and central sensitization occur. Microglial blockage (with minocycline, for example) can exhibit anti-nociceptive actions in pain models²³. A pericytic transformation into a pro-nociceptive microglial phenotype would present a novel mechanism to target for alleviation of microglial-driven neuroinflammation and neuronal sensitization known to underpin some chronic pain states, in preclinical models and in humans^{24–27}.

To complicate matters further, CNS pericytes can differentiate into a neuronal-like phenotype with basic fibroblast growth factor (bFGF) stimulation and are also capable of self-renewal²⁸, indicating that pericytes may be a source of pluripotent progenitor cells. Forebrain pericytes are of neuroectodermal origin²⁹ and it may be this pericyte subtype that gives rise to neuronal phenotypes. The heterogeneous and pluripotent nature of pericytes appears to allow diverse differentiation responses in different situations. How CNS pericytes behave in pain states and preclinical pain models and whether they present a novel target for the alleviation of pain are not yet known.

CNS pericytes in vessel barrier integrity

The BBB and BSCB are selective barriers that limit cell and molecular access into the CNS from the blood. The barriers maintain the microenvironment within the CNS required for physiological neuronal function. The CNS microvasculature is comprised of endothelial cells, pericytes, perivascular macrophages, microglia, and astrocytic end-feet (Figure 1). Unlike in the periphery, CNS endothelial cells are not fenestrated but are connected via tight junction proteins such as occludin, junctional adhesion molecules (JAMs), vascular endothelial cadherin (VE-cadherin), and claudins, which restrict the inter-endothelial space. Pericytes are embedded in the basement membrane (perivascular niche), which surrounds the endothelial cells. Pericytes are polymorphs with an oval to elongated morphology and extend processes along capillaries, pre-capillary arterioles and post-capillary venules. In the CNS, these processes encircle the endothelium and cover endothelial tight junction regions³⁰. Astrocytic end-feet wrap around the basement membrane encircling perivascular cells and vessels and provide another barrier (glia limitans), further limiting access to the nervous parenchyma.

Pericytes are key modulators of the BBB/BSCB and participate in neuroinflammation^{3,31}. Platelet-derived growth factor receptor-beta (PDGFR β) is predominantly expressed by pericytes in the CNS³² and, via mice with genetically disrupted PDGFR β signalling, demonstrated the necessity for pericytes in BBB formation during embryogenesis³³. In addition, in both development and adulthood, barrier permeability is inversely correlated with pericyte coverage^{5,33}. There is lower pericyte capillary coverage in the spinal cord compared with the brain, which correlates with increased permeability, and lower expression of two tight junction proteins: ZO-1 and occludin³⁴.

Mice with deficient PDGFR β signalling (*pdgfr β ^{F7/F7}*) demonstrated region-dependent losses in pericytes that related to BBB breakdown³⁵. Conversely, disrupted PDGFR β signalling through a mutation in the retention motif of PDGF-B (*pdgfb^{ret/ret}*), one of two ligands for the receptor, caused homogenous pericyte loss across the brain, but the extent of pericyte loss in this experiment did not correlate with increased BBB permeability³⁶. The authors hypothesise that this may be due to the phenotypic diversity of pericytes and alternative local signalling mechanisms controlling BBB permeability. In addition, the difference in mutation strategy (receptor versus ligand) could have contributed to the contrasting results.

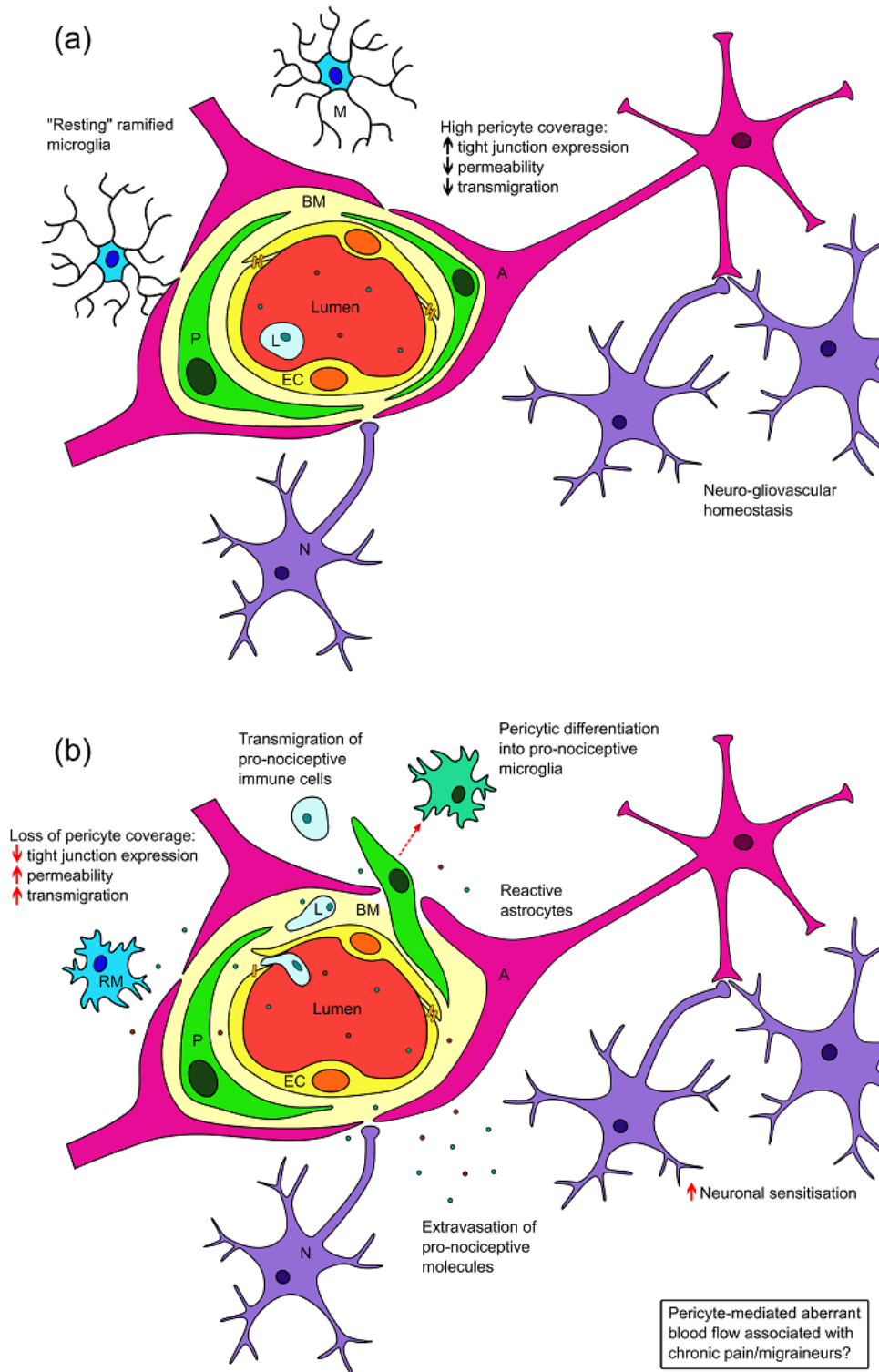


Figure 1. A diagrammatic overview of the physiological roles of central nervous system (CNS) pericytes and possible links of pericyte function to neuronal sensitization and pain. (A) Under physiological conditions, the high pericyte-vessel coverage in the CNS promotes high tight junction protein expression, consequently maintaining vessel integrity and reduced vessel permeability. Pericytes influence the low level of blood cell transmigration into the parenchyma under physiological conditions. **(B)** Reduced pericyte coverage leads to decreased tight junction protein expression, loss of vessel integrity, and increased vessel permeability. Ensuing pro-nociceptive molecule extravasation and pro-nociceptive and pro-inflammatory immune cell transmigration are likely to lead to neuronal sensitization. In addition, there is emerging evidence that multipotent CNS pericytes are able to migrate out of their peri-vascular niche and differentiate into a microglia-like phenotype in preclinical pain models, which in turn could have a neuronal sensitizing effect. A, astrocyte; BM, basement membrane; EC, endothelial cell; L, leukocyte; M, microglia; N, neuron; P, pericyte.

Foxf2, a transcription factor, is specifically expressed in cerebral pericytes derived from the neural crest (neuroectodermal cells)³⁷. Loss of Foxf2 caused cerebral haemorrhage, increased pericyte densities in embryonic cerebral capillaries, and induced BBB disruption in both development and adulthood. There was also a decrease in PDGFR β and transforming growth factor beta (TGF β) (implicated in pericyte and endothelial proliferation, migration and differentiation) signalling despite an increased number of pericytes³⁷. This suggests that the correct differentiation of pericytes is key to BBB development and maintenance, and there are cues other than PDGFR β which are involved in pericyte recruitment to the endothelium. For example, loss of glial laminin resulted in BBB breakdown, concluded to be due to the observed altered pericytic differentiation into a contractile phenotype, consequently disrupting the barrier³⁸. In addition, CD146 has been implicated in regulating PDGFR β /PDGF-B and TGF β signalling in barrier formation and maintenance. Pericyte-secreted CD146 acts as a co-receptor for PDGFR β during pericyte-vascular recruitment, and in the mature barrier, endothelial cell-secreted CD146 is downregulated by pericyte production of TGF β ³⁹.

Pericyte-endothelial cell signalling is paramount in the maintenance of the BBB/BSCB, especially through PDGFR β /PDGF-B signalling⁴⁰. However, many of the specific mechanisms of how pericyte-endothelial cell signalling affects barrier function are still largely unknown. *In vitro* culture techniques offer the ability to study pericyte function in detail. Indeed, much of the knowledge gained about pericytes has been from combined *in vivo* and *in vitro* techniques. Recently, Herland *et al.*⁴¹ developed a dynamic flow model within a microfluidic device that permits co-culturing human endothelial cells in an engineered lumen with pericytes or astrocytes embedded in the surrounding extracellular matrix. In this model, the presence of pericytes reduced the permeability of the engineered vessel and increased the production of both basal and TNF α -induced cytokines compared with endothelial cells alone. The development of sophisticated *in vitro* models of the BBB/BSCB will allow more detailed and specific research into the contribution of pericytes and other cell types to barrier permeability and function.

In many preclinical models of painful neuropathy, the BBB/BSCB is altered^{42–46}. Leakage of neurotoxic blood-derived molecules into the nervous parenchyma (for example, erythrocytic free iron, fibrinogen, plasminogen and thrombin) can lead to a detrimental neuronal response, including sensitization, and may contribute to an increased pain state in various painful diseases (Figure 1b). Gaining a better understanding of pericytic function (or indeed pericytic dysfunction) in the loss of barrier integrity in the context of pain may present an opportunity to intervene and limit the possibly painful consequences.

Pericytes in haemodynamic regulation

The precise roles of contractile pericytes, despite their isolation and identification in the 1870s, in regulating haemodynamic control of CNS blood flow are only now being probed effectively. Smooth muscle cell (SMC) contraction in pial and penetrating arterioles is, as in other tissues, the primary control

on CNS blood flow⁴⁷. Capillaries are devoid of SMC and evidence indicates that pericytes contribute to blood flow regulation in capillaries, most likely through electrical coupling with capillary endothelial cells^{48,49}. Pericytes are able to regulate bi-directional control of CNS capillary diameter independent of arterioles⁵⁰, and pericyte stimulation propagates signals that cause downstream pericytes to constrict, indicative of a pericyte-pericyte signalling network⁵¹. Furthermore, there is evidence of an electrical endothelial network: CNS capillary endothelial cells expressing the potassium channel K_v2.1 caused vasodilatation of distant upstream arterioles in the CNS microvasculature in the absence of pericytes⁵². The authors conclude that a hyperpolarising signal is transmitted through endothelial gap junctions, inhibiting calcium influx, and causes SMC relaxation and vessel dilation. Evidence points towards pericytes being electrically coupled to capillary endothelial cells and therefore possibly being able to regulate this novel electrical endothelial network^{47,48}. Further evidence of the intricate relationship between pericytes and capillaries being responsible for control of cerebral blood flow (CBF) following neuronal innervation derives from knockout animals, in which decreased pericyte numbers resulted in a reduction in capillary coverage and dysregulation of the microvasculature^{35,53,54}. Potential signalling networks between pericytes and myocytes in uterine smooth muscle also point to multi-cellular interactions in blood flow control, as pericyte constrictions persist longer following stimulation compared with myocytes⁵⁵. Exaggerated pericyte constriction, persisting longer than SMC constriction, has been linked to a loss of reperfusion in ischaemia and stroke, even when occluded arteries have been dilated^{56–60}. This supports the role of pericytes having an influence on CBF which can be detrimental.

In keeping with a pericyte contribution to the NVU⁴⁷, several neuro-glial transmitters modulate pericyte influence on microvasculature in cerebellar slices. Pericyte populations are heterogeneous depending on pericyte locus in the microcirculation^{6,40,53,61,62}. Pericyte constriction is stimulated by noradrenaline and blocked by glutamate, transmitters involved in neurovascular coupling. HETE-20 is a known CNS vasoconstrictor that is inhibited by glutamate-driven nitric oxide (NO) release. Block of synthesis of both HETE-20 and NO resulted in pericyte dilation, mediated by prostaglandin E₂, a known CNS vasodilator⁵⁰.

Although the exact contribution of pericytes in maintaining and altering CBF requires further elucidation, evidence suggests that they have a much more significant role in CBF than assumed since their initial discovery. Emerging evidence points to pericytes acting as major players in the NVU which involved a “sensory web” of microvasculature⁵². Pericytes preside over profound changes in capillary tone and may be able to initiate upstream effects on arteriolar smooth muscle, contrary to initial opinion. These findings implicate pericytes as key players in pain that arises from altered CBF, for example in migraine and chronic pain conditions associated with altered blood vessel function⁶³. Blood oxygen level-dependent technology has linked generalised cerebral hypoperfusion with severe pain in

migraineurs, which was associated with concurrent vasospasm⁶⁴. Induced hypoxia worsened pain in response to stimuli designed to invoke an episode. Such stimuli could be linked to aberrant neuronal signalling causing detrimental pericytic action⁶⁵. These intriguing studies highlight how aberrant neurovascular coupling and detrimental pericytic function could contribute to pain.

Pericytes in CNS angiogenesis

Pericytes are vital for vascular function, including the control of angiogenesis. Angiogenesis is important in the development and maintenance of CNS function and involves several cell types in the NVU. Developmental CNS angiogenesis is initially dependent on neural VEGF-A expression leading to the formation of endothelial-derived tip cells and enhanced endothelial cell survival. Pericytes form part of the NVU⁶⁶. They are recruited to sprouting vessels through endothelial secretion of PDGF- β , promote tube formation, and later secrete angiostatic substances that contribute to the termination of CNS angiogenesis and vascular stabilisation⁶⁷. The reduced permeability of the BBB compared with the systemic vasculature is not intrinsic to endothelial cells; the presence of neuronal precursors is required for BBB induction, and CNS pericytes and astrocytes are required for BBB maturation⁶⁶.

The contribution of pericytes to BSCB and angiogenesis is less well understood, but evidence suggests that it is important as activated pericytes stimulate increased vascular density (interpreted as angiogenesis) in spinal cord explant cultures⁶⁸. Altered BSCB function is evident both in people with amyotrophic lateral sclerosis (ALS) and in animal models of ALS⁶⁹. Patients with ALS have increased spinal cord ventral horn microvascular density (also interpreted as angiogenesis) with decreased spinal cord pericyte coverage; those patients requiring respiratory support displayed an increased incidence of spinal cord angiogenesis²⁷. These human observations imply that spinal cord vascular dysfunction, with increased angiogenesis and decreased pericyte function, contribute to the disease.

Do pericytes contribute to migration of immune cells into the CNS and the generation of pain?

The BBB/BSCB normally restricts leukocyte entry and as a result the CNS is considered an immune-privileged site under normal conditions. However, under many pathological conditions, leukocyte transendothelial migration into the CNS occurs and pericytes contribute to this process. First, pericyte dilatation increases blood flow to specific areas, thereby increasing leukocyte delivery to the NVUs in question. Second, pericytes are able to release factors into the circulation which promote leukocyte chemoattraction, including TNF α , interleukins (including IL-1 β , IL-6 and IL-10), interferon gamma (IFN γ), TGF β 1, and members of the CC (denoting 2 adjacent cysteines) chemokines, including monocyte-chemoattractant protein-1 (MCP-1)^{70,71}. Pro-inflammatory secreted factors are, however, species-dependent, and rodents differ significantly from human pericytes in their secretome³. Third, CNS pericyte-derived

chemokines stimulate leukocyte integrins, allowing interaction with endothelial adhesion molecules in the vascular lumen, and pericytes also express intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1) contributing to leukocyte transmigration into the perivascular space³. Lastly, once leukocytes are in the perivascular niche, without pericyte-mediated adhesion molecule guidance, leukocytes can be cleared by a perivascular clearance mechanism and not breach the astrocytic end-feet (glia limitans) and reach the nervous parenchyma².

In preclinical models of painful neuropathy such as peripheral nerve injury model, there is evidence that immune cells transmigrate into the CNS and these may contribute to the development of CNS neuronal sensitization (central sensitization)⁷²⁻⁷⁴. A recent report shows that peripheral nerve injury results in disruption of the BSCB, and loss of both tight junction proteins and spinal pericyte coverage⁷⁵. Therefore, if pericytes regulate the passage of immune cells into the nervous tissue parenchyma (Figure 1), then altering this process may be a viable intervention with the aim of lessening central sensitizing processes that lead to increased pain. Pericytes are crucial to the development of the CNS and in central neurodegenerative disorders, and these findings suggest that they also contribute to spinal processing of sensory information and pain.

Summary

This article highlights the key areas of CNS pericyte physiology that, when dysregulated in pathology, could lead to neuronal sensitization and an increased pain state (Figure 1b). Pericytes are a more attractive pharmacological target than those that are beyond the second barrier of the BBB/BSCB, the glia limitans. An agent targeting CNS perivascular cells will not need to penetrate the glia limitans thereby reducing possible off target and detrimental side effects within the CNS parenchyma. However, whether CNS pericytic actions affect pain is currently severely under-researched; more focussed research into the actions of pericytes in the context of neuronal sensitization and pain could present many potential therapeutic opportunities.

Competing interests

The authors declare that they have no competing interests.

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References



1. Sims DE: **Recent advances in pericyte biology--implications for health and disease.** *Can J Cardiol.* 1991; 7(10): 431–43.
[PubMed Abstract](#)
2. Owens T, Bechmann I, Engelhardt B: **Perivascular spaces and the two steps to neuroinflammation.** *J Neuropathol Exp Neurol.* 2008; 67(12): 1113–21.
[PubMed Abstract](#) | [Publisher Full Text](#)
3. **F** Rustenhoven J, Jansson D, Smyth LC, *et al.*: **Brain Pericytes As Mediators of Neuroinflammation.** *Trends Pharmacol Sci.* 2017; 38(3): 291–304.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
4. Winkler EA, Sagare AP, Zlokovic BV: **The pericyte: a forgotten cell type with important implications for Alzheimer's disease?** *Brain Pathol.* 2014; 24(4): 371–86.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
5. **F** Bell RD, Winkler EA, Sagare AP, *et al.*: **Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging.** *Neuron.* 2010; 68(3): 409–27.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
6. **F** Attwell D, Mishra A, Hall CN, *et al.*: **What is a pericyte?** *J Cereb Blood Flow Metab.* 2016; 36(2): 451–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
7. **F** Tang W, Zeve D, Suh JM, *et al.*: **White fat progenitor cells reside in the adipose vasculature.** *Science.* 2008; 322(5901): 583–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
8. Dellavalle A, Maroli G, Covarello D, *et al.*: **Pericytes resident in postnatal skeletal muscle differentiate into muscle fibres and generate satellite cells.** *Nat Commun.* 2011; 2: 499.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. **F** Humphreys BD, Lin SL, Kobayashi A, *et al.*: **Fate tracing reveals the pericyte and not epithelial origin of myofibroblasts in kidney fibrosis.** *Am J Pathol.* 2010; 176(1): 85–97.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
10. **F** Feng J, Mantesso A, De Bari C, *et al.*: **Dual origin of mesenchymal stem cells contributing to organ growth and repair.** *Proc Natl Acad Sci U S A.* 2011; 108(16): 6503–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
11. Chen CW, Montelatici E, Crisan M, *et al.*: **Perivascular multi-lineage progenitor cells in human organs: regenerative units, cytokine sources or both?** *Cytokine Growth Factor Rev.* 2009; 20(5–6): 429–34.
[PubMed Abstract](#) | [Publisher Full Text](#)
12. Gaceb A, Ozen I, Padel T, *et al.*: **Pericytes secrete pro-regenerative molecules in response to platelet-derived growth factor-BB.** *J Cereb Blood Flow Metab.* 2018; 38(1): 45–57.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
13. Boettger MK, Weber K, Grossmann D, *et al.*: **Spinal tumor necrosis factor alpha neutralization reduces peripheral inflammation and hyperalgesia and suppresses autonomic responses in experimental arthritis: a role for spinal tumor necrosis factor alpha during induction and maintenance of peripheral inflammation.** *Arthritis Rheum.* 2010; 62(5): 1308–18.
[PubMed Abstract](#) | [Publisher Full Text](#)
14. Geis C, Geuss E, Sommer C, *et al.*: **NOX4 is an early initiator of neuropathic pain.** *Exp Neurol.* 2017; 288: 94–103.
[PubMed Abstract](#) | [Publisher Full Text](#)
15. Hulse RP, Beazley-Long N, Hua J, *et al.*: **Regulation of alternative VEGF-A mRNA splicing is a therapeutic target for analgesia.** *Neurobiol Dis.* 2014; 71: 245–59.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Ishitsuka K, Ago T, Arimura K, *et al.*: **Neurotrophin production in brain pericytes during hypoxia: a role of pericytes for neuroprotection.** *Microvasc Res.* 2012; 83(3): 352–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
17. Ross JL, Queme LF, Lamb JE, *et al.*: **Interleukin 1 β inhibition contributes to the antinociceptive effects of voluntary exercise on ischemia/reperfusion-induced hypersensitivity.** *Pain.* 2018; 159(2): 380–392.
[PubMed Abstract](#) | [Publisher Full Text](#)
18. Tian G, Luo X, Tang C, *et al.*: **Astrocyte contributes to pain development via MMP2-JNK1/2 signaling in a mouse model of complex regional pain syndrome.** *Life Sci.* 2017; 170: 64–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Ozen I, Deierborg T, Miharada K, *et al.*: **Brain pericytes acquire a microglial phenotype after stroke.** *Acta Neuropathol.* 2014; 128(3): 381–96.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
20. Miro-Mur F, Perez-de-Puig I, Ferrer-Ferrer M, *et al.*: **Immature monocytes recruited to the ischemic mouse brain differentiate into macrophages with features of alternative activation.** *Brain Behav Immun.* 2016; 53: 18–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. Papageorgiou IE, Lewen A, Galow LV, *et al.*: **TLR4-activated microglia require IFN- γ to induce severe neuronal dysfunction and death in situ.** *Proc Natl Acad Sci U S A.* 2016; 113(1): 212–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Vacca V, Marinelli S, Pieroni L, *et al.*: **Higher pain perception and lack of recovery from neuropathic pain in females: a behavioural, immunohistochemical, and proteomic investigation on sex-related differences in mice.** *Pain.* 2014; 155(2): 388–402.
[PubMed Abstract](#) | [Publisher Full Text](#)
23. Chen HS, Wang JX, Zhang JH, *et al.*: **Contribution of the spinal microglia to bee venom-induced inflammatory pain in conscious rats.** *Neurosci Lett.* 2013; 534: 301–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. **F** Del Valle L, Schwartzman RJ, Alexander G: **Spinal cord histopathological alterations in a patient with longstanding complex regional pain syndrome.** *Brain Behav Immun.* 2009; 23(1): 85–91.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
25. Loggia ML, Chonde DB, Akeju O, *et al.*: **Evidence for brain glial activation in chronic pain patients.** *Brain.* 2015; 138(Pt 3): 604–15.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
26. Shi Y, Gelman BB, Lisinicchia JG, *et al.*: **Chronic-pain-associated astrocytic reaction in the spinal cord dorsal horn of human immunodeficiency virus-infected patients.** *J Neurosci.* 2012; 32(32): 10833–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
27. Yamadera M, Fujimura H, Inoue K, *et al.*: **Microvascular disturbance with decreased pericyte coverage is prominent in the ventral horn of patients with amyotrophic lateral sclerosis.** *Amyotroph Lateral Scler Frontotemporal Degener.* 2015; 16(5–6): 393–401.
[PubMed Abstract](#) | [Publisher Full Text](#)
28. Dore-Duffy P, Katyshev A, Wang X, *et al.*: **CNS microvascular pericytes exhibit multipotential stem cell activity.** *J Cereb Blood Flow Metab.* 2006; 26(5): 613–24.
[PubMed Abstract](#) | [Publisher Full Text](#)
29. Winkler EA, Bell RD, Zlokovic BV: **Central nervous system pericytes in health and disease.** *Nat Neurosci.* 2011; 14(11): 1398–405.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. Abbott NJ: **Inflammatory mediators and modulation of blood-brain barrier permeability.** *Cell Mol Neurobiol.* 2000; 20(2): 131–47.
[PubMed Abstract](#) | [Publisher Full Text](#)
31. Armulik A, Genove G, Mae M, *et al.*: **Pericytes regulate the blood-brain barrier.** *Nature.* 2010; 468(7323): 557–61.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Guillemin GJ, Brew BJ: **Microglia, macrophages, perivascular macrophages, and pericytes: a review of function and identification.** *J Leukoc Biol.* 2004; 75(3): 388–97.
[PubMed Abstract](#) | [Publisher Full Text](#)
33. **F** Daneman R, Zhou L, Kebede AA, *et al.*: **Pericytes are required for blood-brain barrier integrity during embryogenesis.** *Nature.* 2010; 468(7323): 562–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
34. Winkler EA, Sengillo JD, Bell RD, *et al.*: **Blood-spinal cord barrier pericyte reductions contribute to increased capillary permeability.** *J Cereb Blood Flow Metab.* 2012; 32(10): 1841–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
35. Nikolakopoulou AM, Zhao Z, Montagne A, *et al.*: **Regional early and progressive loss of brain pericytes but not vascular smooth muscle cells in adult mice with disrupted platelet-derived growth factor receptor- β signaling.** *PLoS One.* 2017; 12(4): e0176225.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
36. Villaseñor R, Kuennecke B, Ozmen L, *et al.*: **Region-specific permeability of the blood-brain barrier upon pericyte loss.** *J Cereb Blood Flow Metab.* 2017; 37(12): 3683–94.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
37. Reyahi A, Nik AM, Ghiami M, *et al.*: **Foxf2 Is Required for Brain Pericyte Differentiation and Development and Maintenance of the Blood-Brain Barrier.** *Dev Cell.* 2015; 34(1): 19–32.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Yao Y, Chen ZL, Norris EH, *et al.*: **Astrocytic laminin regulates pericyte differentiation and maintains blood brain barrier integrity.** *Nat Commun.* 2014; 5: 3413.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. **F** Chen J, Luo Y, Hui H, *et al.*: **CD146 coordinates brain endothelial cell-pericyte communication for blood-brain barrier development.** *Proc Natl Acad Sci U S A.* 2017; 114(36): E7622–E31.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
40. **F** Sweeney MD, Ayyadurai S, Zlokovic BV: **Pericytes of the neurovascular unit: key functions and signaling pathways.** *Nat Neurosci.* 2016; 19(6): 771–83.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
41. **F** Herland A, van der Meer AD, FitzGerald EA, *et al.*: **Distinct Contributions of Astrocytes and Pericytes to Neuroinflammation Identified in a 3D Human**

- Blood-Brain Barrier on a Chip.** *PLoS One.* 2016; 11(3): e0150360.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
42. Lochhead JJ, Ronaldson PT, Davis TP: **Hypoxic Stress and Inflammatory Pain Disrupt Blood-Brain Barrier Tight Junctions: Implications for Drug Delivery to the Central Nervous System.** *AAPS J.* 2017; 19(4): 910–20.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
43. **F**eggs S, Liu XJ, Kwan C, *et al.*: **Peripheral nerve injury and TRPV1-expressing primary afferent C-fibers cause opening of the blood-brain barrier.** *Mol Pain.* 2010; 6: 74.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
44. Souza PS, Goncalves ED, Pedrosa GS, *et al.*: **Physical Exercise Attenuates Experimental Autoimmune Encephalomyelitis by Inhibiting Peripheral Immune Response and Blood-Brain Barrier Disruption.** *Mol Neurobiol.* 2017; 54(6): 4723–37.
[PubMed Abstract](#) | [Publisher Full Text](#)
45. Reinhold AK, Ritter HL: **Barrier function in the peripheral and central nervous system—a review.** *Pflügers Arch.* 2017; 469(1): 123–34.
[PubMed Abstract](#) | [Publisher Full Text](#)
46. Hulse RP, Beazley-Long N, Ved N, *et al.*: **Vascular endothelial growth factor-165 prevents diabetic neuropathic pain and sensory neuronal degeneration.** *Clin Sci (Lond).* 2015; 129(8): 741–56.
[PubMed Abstract](#) | [Publisher Full Text](#)
47. **F**lodecola C: **The Neurovascular Unit Coming of Age: A Journey through Neurovascular Coupling in Health and Disease.** *Neuron.* 2017; 96(1): 17–42.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
48. Wu DM, Minami M, Kawamura H, *et al.*: **Electrotonic transmission within pericyte-containing retinal microvessels.** *Microcirculation.* 2006; 13(5): 353–63.
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Matsushita K, Puro DG: **Topographical heterogeneity of K_m currents in pericyte-containing microvessels of the rat retina: effect of diabetes.** *J Physiol.* 2006; 573(Pt 2): 483–95.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
50. **F** Hall CN, Reynell C, Gesslein B, *et al.*: **Capillary pericytes regulate cerebral blood flow in health and disease.** *Nature.* 2014; 508(7494): 55–60.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
51. **F** Peppiatt CM, Howarth C, Mobbs P, *et al.*: **Bidirectional control of CNS capillary diameter by pericytes.** *Nature.* 2006; 443(7112): 700–4.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
52. **F** Longden TA, Dabertrand F, Koide M, *et al.*: **Capillary K^+ -sensing initiates retrograde hyperpolarization to increase local cerebral blood flow.** *Nat Neurosci.* 2017; 20(5): 717–26.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
53. Kisler K, Nelson AR, Montagne A, *et al.*: **Cerebral blood flow regulation and neurovascular dysfunction in Alzheimer disease.** *Nat Rev Neurosci.* 2017; 18(7): 419–34.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
54. **F** Li Y, Lucas-Osma AM, Black S, *et al.*: **Pericytes impair capillary blood flow and motor function after chronic spinal cord injury.** *Nat Med.* 2017; 23(6): 733–41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
55. Borysova L, Burdyga T: **Evidence that NO/cGMP/PKG signalling cascade mediates endothelium dependent inhibition of IP_3R mediated Ca^{2+} oscillations in myocytes and pericytes of ureteric microvascular network *in situ*.** *Cell Calcium.* 2015; 58(6): 535–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
56. Abba M, Patil N, Allgayer H: **MicroRNAs in the Regulation of MMPs and Metastasis.** *Cancers (Basel).* 2014; 6(2): 625–45.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
57. Traissac L, Baudet J: **[Reconstitution of the digestive cervical axis, so-called circular reconstruction, in ORL oncology. Apropos of 26 cases].** *Rev Laryngol Otol Rhinol (Bord).* 1986; 107(2): 113–5.
[PubMed Abstract](#)
58. Zehendner CM, Wedler HE, Luhmann HJ: **A novel *in vitro* model to study pericytes in the neurovascular unit of the developing cortex.** *PLoS One.* 2013; 8(11): e81637.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
59. **F** Yemisci M, Gursoy-Ozdemir Y, Vural A, *et al.*: **Pericyte contraction induced by oxidative-nitrative stress impairs capillary reflow despite successful opening of an occluded cerebral artery.** *Nat Med.* 2009; 15(9): 1031–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
60. O'Farrell FM, Mastitskaya S, Hammond-Haley M, *et al.*: **Capillary pericytes mediate coronary no-reflow after myocardial ischaemia.** *eLife.* 2017; 6: pii: e29280.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
61. Hartmann DA, Underly RG, Grant RI, *et al.*: **Pericyte structure and distribution in the cerebral cortex revealed by high-resolution imaging of transgenic mice.** *Neurophotonics.* 2015; 2(4): 041402.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
62. Jansson D, Rustenhoven J, Feng S, *et al.*: **A role for human brain pericytes in neuroinflammation.** *J Neuroinflammation.* 2014; 11: 104.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
63. Honda T, Maruta T, Takahashi K: **Brain perfusion abnormality in patients with chronic pain.** *Keio J Med.* 2007; 56(2): 48–52.
[PubMed Abstract](#) | [Publisher Full Text](#)
64. Cadiot D, Longuet R, Bruneau B, *et al.*: **Magnetic resonance imaging in children presenting migraine with aura: Association of hypoperfusion detected by arterial spin labelling and vasospasm on MR angiography findings.** *Cephalalgia.* 2017; 33(102417723570).
[PubMed Abstract](#) | [Publisher Full Text](#)
65. Arngrim N, Hougaard A, Schytz HW, *et al.*: **Effect of hypoxia on BOLD fMRI response and total cerebral blood flow in migraine with aura patients.** *J Cereb Blood Flow Metab.* 2017; 271678X17719430.
[PubMed Abstract](#)
66. Sá-Pereira I, Brites D, Brito MA: **Neurovascular unit: a focus on pericytes.** *Mol Neurobiol.* 2012; 45(2): 327–47.
[PubMed Abstract](#) | [Publisher Full Text](#)
67. van Hinsbergh VW, Koolwijk P: **Endothelial sprouting and angiogenesis: matrix metalloproteinases in the lead.** *Cardiovasc Res.* 2008; 78(2): 203–12.
[PubMed Abstract](#) | [Publisher Full Text](#)
68. Mayo JN, Bearden SE: **Driving the Hypoxia-Inducible Pathway in Human Pericytes Promotes Vascular Density in an Exosome-Dependent Manner.** *Microcirculation.* 2015; 22(8): 711–23.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
69. Rosenberg GA: **Neurological diseases in relation to the blood-brain barrier.** *J Cereb Blood Flow Metab.* 2012; 32(7): 1139–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
70. Guijarro-Muñoz I, Compte M, Álvarez-Cienfuegos A, *et al.*: **Lipopolysaccharide activates Toll-like receptor 4 (TLR4)-mediated NF- κ B signaling pathway and proinflammatory response in human pericytes.** *J Biol Chem.* 2014; 289(4): 2457–68.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
71. Pieper C, Marek JJ, Unterberg M, *et al.*: **Brain capillary pericytes contribute to the immune defense in response to cytokines or LPS *in vitro*.** *Brain Res.* 2014; 1550: 1–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
72. Zhang J, Shi XQ, Echeverry S, *et al.*: **Expression of CCR2 in both resident and bone marrow-derived microglia plays a critical role in neuropathic pain.** *J Neurosci.* 2007; 27(45): 12396–406.
[PubMed Abstract](#) | [Publisher Full Text](#)
73. **F** Newton VL, Guck JD, Cotter MA, *et al.*: **Neutrophils Infiltrate the Spinal Cord Parenchyma of Rats with Experimental Diabetic Neuropathy.** *J Diabetes Res.* 2017; 2017: 4729284.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
74. Huber JD, Campos CR, Mark KS, *et al.*: **Alterations in blood-brain barrier ICAM-1 expression and brain microglial activation after lambda-carrageenan-induced inflammatory pain.** *Am J Physiol Heart Circ Physiol.* 2006; 290(2): H732–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
75. **F** Sauer RS, Kirchner J, Yang S, *et al.*: **Blood-spinal cord barrier breakdown and pericyte deficiency in peripheral neuropathy.** *Ann N Y Acad Sci.* 2017; 1405(1): 71–88.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)

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- 1 **Thomas P Davis** Department of Pharmacology, University of Arizona, Arizona, USA
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