



Physical Activity, Immune System, and the Microbiome in Cardiovascular Disease

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Cardiovascular health is a primary research focus, as it is a leading contributor to mortality and morbidity worldwide, and is prohibitively costly for healthcare. Atherosclerosis, the main driver of cardiovascular disease, is now recognized as an inflammatory disorder. Physical activity (PA) may have a more important role in cardiovascular health than previously expected. This review overviews the contribution of PA to cardiovascular health, the inflammatory role of atherosclerosis, and the emerging evidence of the microbiome as a regulator of inflammation.

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AN OVERVIEW OF ATHEROSCLEROTIC CARDIOVASCULAR DISEASE

Cardiovascular disease (CVD) is the worldwide leading cause of death and is a global economic burden (Cook et al., 2014). Most of the disease is driven by a process known as atherosclerosis—the buildup of plaque which occludes arterial vessels (Rajamani and Fisher, 2017). Throughout disease progression, the atherosclerotic plaque loses stability and becomes prone to rupture, a sudden event that can lead to arterial thrombosis and cause deleterious acute ischemic events like myocardial infarction (MI) and stroke (Gisterå and Hansson, 2017).

Atherosclerosis is influenced by a number of risk factors including, lifestyle choices (i.e., diet, physical activity, and cigarette smoking), advancing age, and associated disorders like hypertension, diabetes, obesity, and dyslipidemia (Tegos et al., 2001). However, traditional risk factors alone are inadequate at predicting atherosclerotic CVD. According to the Participants of Early Subclinical Atherosclerosis (PESA) study, subclinical atherosclerosis was detected in almost 50% of participants that were free of the conventional cardiovascular risk factors (Fernández-Friera et al., 2017). Statins, the gold standard treatment for lowering lipids (Grundy, 2016), have proven effective at reducing cardiovascular events, yet their contribution to reducing mortality remains questionable (Cholesterol Treatment Trialists' [CTT] Collaborators, 2012; DuBroff and de Lorgeril, 2015). The recently developed proprotein convertase subtilisin-kexin type 9 (PCSK9) inhibitors effectively lowered cholesterol levels and reduced cardiovascular events when taken in conjunction with statins (Waters and Hsue, 2017). However, some of these studies failed to meet expectations based on the linear relationship between LDL reduction and percentage reduction of cardiovascular events based on data from 14 clinical trials using statins (Waters and Hsue, 2017). A possible explanation is that PCSK9 inhibitors failed to reduce C-reactive protein (CRP) levels, a clinical biomarker of inflammation and cardiovascular risk (Waters and Hsue, 2017).

Atherosclerosis is recognized as a chronic systemic inflammatory disease with focal manifestations at the vascular site (Ross, 1999; Conti and Shaik-Dasthagirisaeab, 2015). CVD is often pronounced in other immune disorders such as rheumatoid arthritis (RA) or systemic lupus erythematosus (SLE) (Gabriel, 2008; Abu-Shakra and Novack, 2012; Skeoch and Bruce, 2015; Henrot et al., 2018). Recent studies propose the gut microbiome as an emerging regulator of inflammatory conditions including atherosclerosis (Ridaura et al., 2013; Romano et al., 2015; Halfvarson et al., 2017; Kreznar et al., 2017; Slingerland et al., 2017).

The World Health Organization, American Heart Association, and European Society of Cardiology recommend several significant behavioral changes for the preventative care of CVD including smoking cessation, dietary changes, weight control, alcohol intake, and physical activity (PA) (Goff et al., 2014; Piepoli et al., 2016; Whelton et al., 2018). Of particular interest is the contribution of PA to CVD (Lin et al., 2015; Murtagh et al., 2015). Utilizing PA as therapy is particularly appealing because its implementation is cost-effective for the patient and the benefits can ease the global economic burden by reducing the cost of care (Valero-Elizondo et al., 2016). Recently, PA has been the focus of intense investigation for its ability to regulate the underlying immune system. This review will focus on the impact of PA as it relates to the regulation of systemic inflammation and the contribution of the microbiome.

PHYSICAL ACTIVITY AND CARDIOVASCULAR HEALTH

Regular PA is associated with many health benefits, including improving cholesterol levels, reducing body weight and blood pressure, increasing insulin sensitivity, and neuroprotective effects (Myers, 2003; Chieffi et al., 2017). Indeed, some of these benefits have been attributed to in part by the small neuropeptides called Orexins (Chieffi et al., 2017). Orexin A is released into plasma upon exercise, and contributes to regulating energy balance (Messina et al., 2016, 2017).

Numerous studies have established that PA is beneficial for reducing the risk and effect of CVD (Diaz et al., 2016; Vella et al., 2017; Florido et al., 2018). Physically fit individuals have a reduced risk of developing CVD (Kokkinos, 2012). A 10 year follow up study that surveyed senior participants found that leisure time PA reduced the risk of CVD incidents and mortality in a dose-dependent manner (Barengo et al., 2017). The Multi-Ethnic Study of Atherosclerosis (MESA) suggests that moderate to vigorous PA accompanies a more favorable inflammatory marker profile (Vella et al., 2017). The Atherosclerotic Risk in Communities (ARIC) Study evaluated participants with no history of CVD, and found that maintenance and engagement of PA were effective in decreasing heart failure risk (Florido et al., 2018). Additionally, the HUNT Study (Nord-Trondelag Health Study) showed that sustained PA, and not weight loss, substantially improved survival at 30 years follow up in individuals with coronary artery disease (Moholdt et al., 2018).

Physical activity has profound effects on vascular function and lumen dimension, and structural cardiac modifications (Green and Smith, 2017; Francavilla et al., 2018). For example PA regulates heart rate variability (HRV), a predictive factor for sudden cardiac death and MI (Sessa et al., 2018). Elderly athletes display less Carotid Intima Thickening and a more favorable HRV compared to their sedentary counterparts (Galletta et al., 2013; Soares-Miranda et al., 2016). Animal models have also demonstrated that old rats subjected to exercise training had reversed age-related microvascular dysfunction (Hotta et al., 2017).

Sedentary behavior (SB) is described as the lowest energy expenditure for waking activities (e.g., sitting or lying down), and is measured by metabolic equivalents (METs). Although uncoupling SB from associated illness and other risk factors like obesity and RA is difficult, SB promotes a pro-inflammatory status (Fenton et al., 2017). Biomarker analysis from a cross-sectional study of senior men found that higher levels of SB correlated with higher levels of pro-inflammatory markers IL-6, CRP, and tPA (Parsons et al., 2017). There is evidence that reallocation of SB with moderate to vigorous PA promotes a better inflammatory profile, with increased adiponectin levels and lower IL-6, C3, leptin, and leukocyte concentrations (Phillips et al., 2017). Moreover, patients with an inflammatory disease like RA, that have extended sedentary bouts have an increased risk of developing cardiovascular events and could benefit by interrupting sedentary time with leisurely PA (Fenton et al., 2017). Long-term studies, like the 15-years long Tanushimaru Study, confirm that decreased sitting time reduces the risk of mortality (Sakaue et al., 2017).

INFLAMMATION AND ATHEROSCLEROSIS

The first critical step of atherosclerosis development is endothelial dysfunction and increased endothelial permeability that facilitates the build-up and deposition of low-density lipoproteins (LDLs) into the intima layer of the arterial wall where they become oxidized (oxLDL) (Lusis, 2000). Monocytes from the circulation infiltrate the arterial wall and differentiate into macrophages that engulf oxLDL becoming foam cells. Foam cells trapped in the intima layer become apoptotic and necrotic, which form the basis of a necrotic core (Ross, 1999; Gisterå and Hansson, 2017). Immune cells express cytokines and chemokines that are critical modulators of inflammatory signaling during atherogenesis (Turner et al., 2014; Ramji and Davies, 2015). Cytokines are also highly regulated during and as a consequence of exercise (Pedersen, 2017). Although there are many cytokines implicated in atherosclerosis, we will address the top key players, which are summarized in **Table 1**.

The initial injury to endothelial cells (ECs) by LDL typically occurs at the arterial branching points where laminar flow becomes disturbed, which results in morphological and functional changes that promote the permeability of the EC layer and allow retention of LDL (Moore et al., 2013). Activated ECs recruit immune cells by enhancing expression of adhesion

TABLE 1 | Regulation of key chemokines and cytokines in atherosclerosis and physical activity.

Cytokine	CVD effect	Regulation in atherosclerosis	Reference	PA effect	Regulation in physical activity	Reference
CCL2/MCP-1	↑	Upregulated in atherosclerosis	Lin et al., 2014	↓	Low intensity training for 8 weeks decreased mRNA levels of MCP-1 in leukocytes	Yakeu et al., 2010
CCL5	↑	Blocking receptor binding reduces Atherosclerotic Plaque Formation	Veillard et al., 2004	↓	Circulating CCL5 was decreased in obese patients subjected to 3 months of physical activity	Baturcam et al., 2014
CX3CL1	↑	Upregulated on Monocytes from Coronary Artery Disease patients	Apostolakis et al., 2007	↑	Increased after a single bout of exercise	Strömberg et al., 2016
IFN- γ	↑	Induces macrophage gene expression. Mice lacking IFN-gamma receptor have reduced atherosclerotic plaque	Gupta et al., 1997	↑	Moderate exercise increased levels on mononuclear cells	Zamani et al., 2017
IL-10	↓	Overexpression inhibits plaque progression in mice and decreases cholesterol levels	Eefting et al., 2007	↑	Increased by 940% on mononuclear cells in high-risk CVD patients subjected to long term exercise	Smith et al., 1999
IL-1b	↑	Inhibition decreases severity of atherosclerosis in mice and in humans	Kiri et al., 2003; Taylor et al., 2011; Shah et al., 2018	↑	Plasma concentrations increase immediately following exercise and remain elevated for 24 h	Moldoveanu et al., 2000
IL-2	↑	Blocking antibodies reduce atherosclerosis	Upadhyaya et al., 2004	↓	Levels decrease following strenuous exercise	Shephard and Shek, 1994
IL-4	↑	Conflicting Reports. IL-4 deficiency in mice reduces atherosclerosis, but exogenous delivery showed no involvement in the disease	Davenport and Tipping, 2003; King et al., 2007	↑	Increased by 94% on mononuclear cells in high-risk CVD patients subjected to long term exercise	Smith et al., 1999
IL-5	↓	Macrophage expression of IL-5 in mice reduced lesion size by 43%	Zhao et al., 2015	↑	Higher expression in the plasma profile of exercise trained individuals	Schild et al., 2016
IL-6	↑	Upregulated in cardiovascular disease. Exogenous expression of IL-6 increases plaque size	Huber et al., 1999	↑	100-fold increase after acute exercise	Pedersen et al., 2001
TGF-b	↓	Reduces atherosclerosis by weakening T cell activation	Robertson et al., 2003	↑	Increased by 43% on mononuclear cells in high-risk CVD patients subjected to long term exercise	Smith et al., 1999
TNF- α	↑	Inhibition reduces atherosclerosis in ApoE $^{-/-}$ mice	Branen et al., 2004	↓	Reduces circulating levels in patients with metabolic syndrome	Palomo et al., 2010

molecules (e.g., VCAM and ICAM) and chemokines (Gimbrone and García-Cardena, 2016). Pro-inflammatory chemokines are crucial for atherogenesis, for example, inhibition of a three chemokine axis (CCL2, CCL5, and CX3CL1) in mouse models leads to almost a complete attenuation of atherosclerosis (Ramji and Davies, 2015). Although chemokine pathways have been exploited as therapeutic targets (Sheikine and Hansson, 2006), few drugs have been FDA approved, but none for the treatment of atherosclerosis. For instance, the CCR5 antagonist Maraviroc

was developed for the treatment of HIV and the low molecular weight CXCR4 antagonist Plerixafor (AMD3100) for stem cell mobilization (Koenen and Weber, 2011). In fact, modulating these pathways in complex disorders like CVD may be difficult due to the high risk of side effects like increased infection rate (Koenen and Weber, 2010).

Interferon- γ (IFN- γ) is a pro-inflammatory cytokine that functions as the primary activator of macrophages and has been shown to influence many stages of atherogenesis

(Gupta et al., 1997; Voloshyna et al., 2014). IFN- γ enhances Monocyte Chemoattractant Protein-1 (MCP-1) levels, a chemokine that recruits monocytes (Struyf et al., 1998) and the expression of Interferon-induced protein 35 (IFI35), a protein that contributes to EC proliferation and migration (Jian et al., 2018). Moreover, IFN- γ acts synergistically with other pro-inflammatory cytokines, like TNF- α to enhance chemokine production in monocytes and T-cells (Mehta et al., 2017).

Interleukin-1 β (IL-1 β) is a pro-inflammatory cytokine and a prime mediator of inflammation in CVD (Moss and Ramji, 2016). Therapeutic targeting of IL-1 β in the Canakinumab Anti-inflammatory Thrombosis Outcomes Study (CANTOS) trial has provided the first proof-of-concept that inflammation is a key driver of cardiovascular events in high risk populations (Ridker et al., 2017). Inhibition of IL-1 β reduced the risk of non-fatal CV events by 17% in patients with recent MI and elevated high sensitivity CRP (hs-CRP) (Ridker et al., 2017). Remarkably, a subgroup analysis of the CANTOS revealed that patients who achieved hs-CRP levels less than 2 mg/dl had a 25% reduction in major adverse cardiovascular events (Ridker et al., 2018), suggesting that the residual CV risk in this subpopulation was due to inflammation.

Interleukin-6 is a pleiotropic cytokine that is induced by IL-1 β (Cahill and Rogers, 2008) and modulated by Canakinumab (Ridker et al., 2012). IL-6 can act as both a pro- or anti-inflammatory molecule, and thus its function is context dependent (Reiss et al., 2017). It participates in two distinct signaling mechanisms, binding to either the membrane-tethered IL-6 receptor (classical pathway) or a soluble form of the IL-6 receptor (trans-pathway). Trans-signaling is regarded as pro-inflammatory, while classical signaling is as anti-inflammatory (Scheller et al., 2011). Coronary artery disease patients have high levels of IL-6 which are also a predictive biomarker of the disease (Wainstein et al., 2017). However, mouse studies show that IL-6 depletion in ApoE $^{-/-}$ mice promotes atherosclerosis (Schieffer et al., 2004). Physical activity regulates IL-6 levels and acute exercise induces the release of IL-6 from the skeletal muscles into the circulatory system (Pedersen, 2017). Of note, IL-6 induced during exercise stimulates the release of anti-inflammatory cytokines IL-10 and IL-1ra (Pedersen, 2006, 2017). IL-6 is also a mediator of lipid metabolism that stimulates lipolysis and fat oxidation (van Hall et al., 2003) and has been shown to increase after a meal (Payette et al., 2009).

IMMUNE CELLS AND PHYSICAL ACTIVITY

Exercise has a profound effect on both the adaptive and innate immune system by regulating immune cell populations.

On the innate arm, natural killer (NK) cells, neutrophils, and monocyte are all regulated by exercise (Koelwyn et al., 2015). NK cells are modulated during exercise by increasing in number and cytotoxic activity (Pedersen and Ullum, 1994; Na et al., 2000) and increased NK cell infiltration of tumors was observed upon exercise (Pedersen et al., 2016). Neutrophils, a hallmark of acute inflammation, in aging become dysfunctional

(Jackaman et al., 2017). One study showed that PA promotes the migratory function of neutrophils in older adults (Bartlett et al., 2016). PA can also influence monocyte polarization with a decrease of classical monocytes, and an increase of intermediate and non-classical monocytes (Slusher et al., 2017). Another study found that strenuous, anaerobic exercise leads to the acute mobilization of intermediate monocytes by a catecholamine-dependent mechanism (Steppich et al., 2000). Particularly, acute exercise was associated with a reduction of monocyte-platelets aggregates, which are associated with increased risk of cardiovascular clinical events (Michelson et al., 2001).

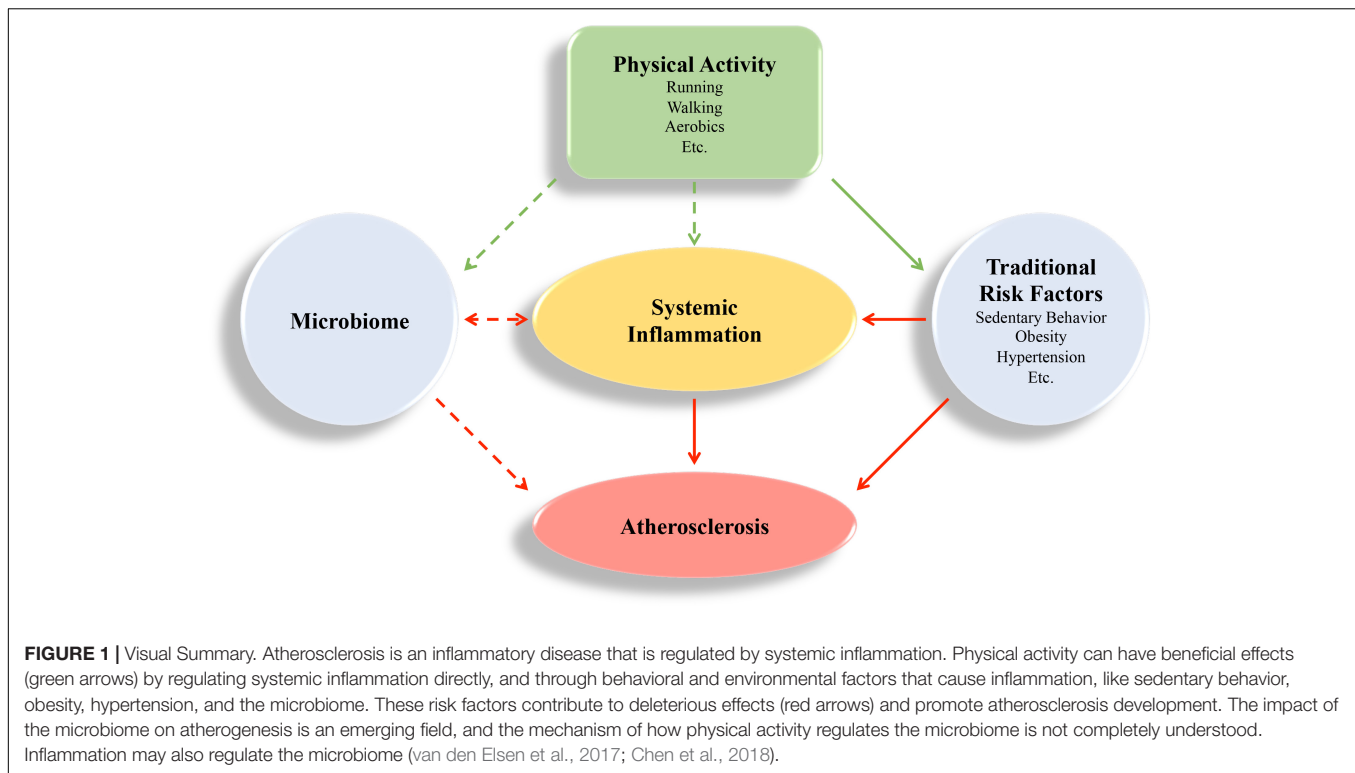
Physical activity also regulates the expression of inflammatory markers and this effect is dependent on the exercise intensity. For example, studies in obese individuals subjected to either medium or high intensity exercise regimes demonstrated that the training intensity can modulate differently the expression of chemokine receptors. Moderate training promoted expression of CCR2 and CXCR2 on monocytes, while higher intensity training promoted CCR5 expression on monocytes, neutrophils, and T-cells (Barry et al., 2017).

On the adaptive side, either prolonged or strenuous exercise affects the balance of types of T-cells. Exercise promotes the predominance of Th2 cells versus Th1 cells. Th1 cells produce IL-2 and IFN- γ , while Th2 cells produce IL-4, IL-5, IL-6, and IL-10 (Sharif et al., 2018). Regulatory T-cells (Treg) belong to a subset of T-helper cells, are anti-inflammatory, atheroprotective (Foks et al., 2015), and are also affected by exercise. A study of elite Olympic athletes from various disciplines, found that athletes displayed higher frequencies of Tregs compared to age and sex-matched controls and this effect was associated with PA intensity (Weinhold et al., 2016). Tregs release immunosuppressive cytokines like IL-10, IL-35, and TGF- β (Olson et al., 2013), and decrease the production of pro-inflammatory cytokines like IFN- γ (Mallat et al., 2003; Ou et al., 2018).

THE MICROBIOME AND CVD: THE ROLE OF PHYSICAL ACTIVITY AND LINKS TO INFLAMMATION

There is growing evidence indicating that the gut microbiome is a critical player in modulating host physiology (Clemente et al., 2012). Deviations from microbial homeostasis have been associated with various diseases, such as inflammatory bowel disease (Berg et al., 2015), arthritis (Clemente et al., 2018), or food allergies (Bunyavanich et al., 2016). Importantly, the microbiome has also been shown to play a critical role in obesity (Karlsson et al., 2012), atherosclerosis (Karlsson et al., 2012), and in the pathogenesis and progression of CVD (Kelly et al., 2016; Jie et al., 2017).

Karlsson et al. (2012) demonstrated that the genus *Collinsella* was enriched in patients with symptomatic atherosclerosis, while *Roseburia* and *Eubacterium* were enriched in healthy controls. These changes in bacterial composition were paralleled by enrichment in patients of bacterial genes encoding peptidoglycan synthesis (which might contribute to atherosclerosis by enhancing neutrophil function) and depletion of phytoene



dehydrogenase and serum levels of b-carotene (hypothesized to have beneficial health effects) (Karlsson et al., 2012). A more recent study examined the association between gut microbiota and lifetime CVD risk in 112 participants in the Bogalusa Heart Study (Kelly et al., 2016). High risk participants had lower microbial diversity, as well as an increase in the abundance of *Prevotella* and *Tyzzarella*, and a decrease in *Alloprevotella* and *Catenibacterium*. A study in a cohort of 405 Chinese subjects found an enrichment in *Enterobacteriaceae* and *Streptococcus* spp. in atherosclerosis patients, suggesting that at least some of the associations between the gut microbiome and CVD might be population-specific (Jie et al., 2017).

The mechanisms by which the microbiome plays a role in CVD are however still poorly understood. One possible mechanism is the production of bacterial metabolites that induce the differentiation of pro- and anti-inflammatory cytokines, which has been demonstrated to be of critical importance in mouse models of colitis (Atarashi et al., 2013). Alternatively, the gut microbiota has also been shown to contribute to atherosclerosis through the conversion of choline or L-carnitine into TMAO (trimethylamine-*N*-oxide) (Koeth et al., 2013). Plasma TMAO is a biomarker of CVD risk associated with increased atherosclerotic stenosis, risk of major cardiovascular events, and mortality (Senthong et al., 2016). The deleterious effects of TMAO are hypothesized to be due to its promotion of platelet aggregation (Ridaura et al., 2013; Romano et al., 2015; Kreznar et al., 2017; Chen et al., 2018).

Given the reported associations between microbiome and CVD, several groups have investigated whether the beneficial

effects of PA in CVD risk could be partially mediated by the changes induced in microbial composition. A study of professional rugby athletes and matched controls found that athletes had lower inflammatory status, and enrichment in bacterial diversity (Clarke et al., 2014). Differences were also observed in the abundance of 48 bacterial taxa, with notable enrichment in *Ruminococcaceae*, *Succinivibrio*, and *Akkermansia* in athletes compared to controls. However, diet was significantly distinct between the groups, and so the differences in microbial diversity and composition might be attributable both to PA and nutritional intake.

A more recent study aimed at disentangling the correlation between exercise, diet, and obesity status in shaping the gut microbiome. Eighteen lean and 14 obese subjects underwent 6 weeks of supervised endurance-based exercise training, followed by a washout period of 6 weeks in which they returned to a sedentary lifestyle (Allen et al., 2018b). Interestingly, changes in gut microbiome were dependent on obesity status, and short-chain fatty acids concentrations increased in lean but not in obese participants. Additionally, gut microbiome alterations disappeared once exercise ceased, suggesting that sustainment of PA is required for these changes to persist.

The causality of these associations is difficult to assess in human studies. Germ-free (GF) animals provide a suitable model to determine whether exercise-mediated changes in gut microbiome can induce specific phenotypes. Allen et al. (2018a) showed that transplanting the gut microbiome from either exercised or not exercised mice to GF mice induced specific distinct changes in the microbiome, metabolome, colonic inflammation, and body

mass of the recipient mice (Allen et al., 2018a). Furthermore, colonization from exercised mice resulted in an attenuated response to DSS-induced colitis. These results demonstrate that exercise can alter gut microbiome and that those changes can result in beneficial health outcomes for the host.

Overall, current evidence points to an association between CVD and gut microbiome through the production of bacterial metabolites that induce potent host pro- or anti-inflammatory responses. Strategies to alter bacterial content are therefore of high interest for their potential therapeutic value. Together with diet (Llewellyn et al., 2017) and fecal transplants (Paramsothy et al., 2017), PA represents an additional approach through which the beneficial effects of gut microbiome modulation can be achieved.

CONCLUSION

Ongoing research focusing on the immune system and CVD continues to probe the molecular intricacies that contribute to atherosclerosis, including the individual contribution of immune cells, cytokines, and the microbiome (summarized in **Figure 1**). While an association between systemic inflammation, gut microbiome, and CVD is emerging, there is a great need to further our understanding of what constitutes normal biological variation versus pathological changes.

Immune modulatory therapeutics like Canakinumab show great promise to aid in the treatment of atherosclerosis,

yet, the development of molecularly targeted pharmacological intervention is a challenging process due to the multifactorial nature of atherosclerosis. Additionally, the use of drugs to control atherosclerosis can be very costly posing a growing economic burden on the society. PA has proven to be a general modulator of systemic inflammation with some emerging effects on the gut microbiome that may be beneficial to overall cardiovascular health. While more research is needed to understand the implications of these changes, improving healthy behaviors by incorporating PA as part of a healthy lifestyle is a promising way to combat CVD.

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REFERENCES

- Abu-Shakra, M., and Novack, V. (2012). Mortality and multiple causes of death in systemic lupus erythematosus – role of the death certificate. *J. Rheumatol.* 39, 458–460. doi: 10.3899/jrheum.111556
- Allen, J. M., Mailing, L. J., Cohrs, J., Salmonson, C., Fryer, J. D., Nehra, V., et al. (2018a). Exercise training-induced modification of the gut microbiota persists after microbiota colonization and attenuates the response to chemically-induced colitis in gnotobiotic mice. *Gut Microbes* 9, 115–130. doi: 10.1080/19490976.2017.1372077
- Allen, J. M., Mailing, L. J., Niemi, G. M., Moore, R., Cook, M. D., White, B. A., et al. (2018b). Exercise alters gut microbiota composition and function in lean and obese humans. *Med. Sci. Sports Exerc.* 50, 747–757. doi: 10.1249/MSS.0000000000001495
- Apostolakis, S., Krambovitis, E., Vlata, Z., Kochiadakis, G. E., Baritaki, S., and Spandidos, D. A. (2007). CX3CR1 receptor is up-regulated in monocytes of coronary artery diseased patients: impact of pre-inflammatory stimuli and renin-angiotensin system modulators. *Thromb. Res.* 121, 387–395. doi: 10.1016/j.thromres.2007.04.005
- Atarashi, K., Tanoue, T., Oshima, K., Suda, W., Nagano, Y., Nishikawa, H., et al. (2013). Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. *Nature* 500, 232–236. doi: 10.1038/nature12331
- Barengo, N. C., Antikainen, R., Borodulin, K., Harald, K., and Jousilahti, P. (2017). Leisure-time physical activity reduces total and cardiovascular mortality and cardiovascular disease incidence in older adults. *J. Am. Geriatr. Soc.* 65, 504–510. doi: 10.1111/jgs.14694
- Barry, J. C., Simtchouk, S., Durrer, C., Jung, M. E., and Little, J. P. (2017). Short-term exercise training alters leukocyte chemokine receptors in obese adults. *Med. Sci. Sport Exerc.* 49, 1631–1640. doi: 10.1249/MSS.0000000000001261
- Bartlett, D. B., Fox, O., McNulty, C. L., Greenwood, H. L., Murphy, L., Sapey, E., et al. (2016). Habitual physical activity is associated with the maintenance of neutrophil migratory dynamics in healthy older adults. *Brain Behav. Immun.* 56, 12–20. doi: 10.1016/j.bbi.2016.02.024
- Baturcam, E., Abubaker, J., Tiss, A., Abu-Farha, M., Khadir, A., Al-Ghimlas, F., et al. (2014). Physical exercise reduces the expression of RANTES and its CCR5 receptor in the adipose tissue of obese humans. *Mediators Inflamm.* 2014:627150. doi: 10.1155/2014/627150
- Berg, D., Clemente, J. C., and Colombel, J.-F. (2015). Can inflammatory bowel disease be permanently treated with short-term interventions on the microbiome? *Expert Rev. Gastroenterol. Hepatol.* 9, 781–795. doi: 10.1586/17474124.2015.1013031
- Branen, L., Hovgaard, L., Nitulescu, M., Bengtsson, E., Nilsson, J., and Jovinge, S. (2004). Inhibition of tumor necrosis factor- reduces atherosclerosis in apolipoprotein e knockout mice. *Arterioscler. Thromb. Vasc. Biol.* 24, 2137–2142. doi: 10.1161/01.ATV.0000143933.20616.1b
- Bunyavanich, S., Shen, N., Grishin, A., Wood, R., Burks, W., Dawson, P., et al. (2016). Early-life gut microbiome composition and milk allergy resolution. *J. Allergy Clin. Immunol.* 138, 1122–1130. doi: 10.1016/j.jaci.2016.03.041
- Cahill, C. M., and Rogers, J. T. (2008). Interleukin (IL) 1 β induction of IL-6 is mediated by a novel phosphatidylinositol 3-kinase-dependent AKT/I κ B kinase α pathway targeting activator protein-1. *J. Biol. Chem.* 283, 25900–25912. doi: 10.1074/jbc.M707692200
- Chen, J., Guo, Y., Gui, Y., and Xu, D. (2018). Physical exercise, gut, gut microbiota, and atherosclerotic cardiovascular diseases. *Lipids Health Dis.* 17:17. doi: 10.1186/s12944-017-0653-9
- Chieffo, S., Carotenuto, M., Monda, V., Valenzano, A., Villano, I., Precenzano, F., et al. (2017). Orexin system: the key for a healthy life. *Front. Physiol.* 8:357. doi: 10.3389/fphys.2017.00357
- Cholesterol Treatment Trialists' [CTT] Collaborators (2012). The effects of lowering LDL cholesterol with statin therapy in people at low risk of vascular disease: meta-analysis of individual data from 27 randomised trials. *Lancet* 380, 581–590. doi: 10.1016/S0140-6736(12)60367-5
- Clarke, S. F., Murphy, E. F., O'Sullivan, O., Lucey, A. J., Humphreys, M., Hogan, A., et al. (2014). Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 63, 1913–1920. doi: 10.1136/gutjnl-2013-306541

- Clemente, J. C., Manasson, J., and Scher, J. U. (2018). The role of the gut microbiome in systemic inflammatory disease. *BMJ* 360:j5145. doi: 10.1136/BMJ.j5145
- Clemente, J. C., Ursell, L. K., Parfrey, L. W., and Knight, R. (2012). The impact of the gut microbiota on human health: an integrative view. *Cell* 148, 1258–1270. doi: 10.1016/j.cell.2012.01.035
- Conti, P., and Shaik-Dasthagirisae, Y. (2015). Atherosclerosis: a chronic inflammatory disease mediated by mast cells. *Cent. J. Immunol.* 40, 380–386. doi: 10.5114/ceji.2015.54603
- Cook, C., Cole, G., Asaria, P., Jabbour, R., and Francis, D. P. (2014). The annual global economic burden of heart failure. *Int. J. Cardiol.* 171, 368–376. doi: 10.1016/j.ijcard.2013.12.028
- Davenport, P., and Tipping, P. G. (2003). The role of interleukin-4 and interleukin-12 in the progression of atherosclerosis in apolipoprotein E-deficient mice. *Am. J. Pathol.* 163, 1117–1125. doi: 10.1016/S0002-9440(10)63471-2
- Diaz, K. M., Booth, J. N., Seals, S. R., Hooker, S. P., Sims, M., Dubbert, P. M., et al. (2016). Sedentary behavior and subclinical atherosclerosis in African Americans: cross-sectional analysis of the Jackson heart study. *Int. J. Behav. Nutr. Phys. Act.* 13:31. doi: 10.1186/s12966-016-0349-y
- DuBroff, R., and de Lorgeril, M. (2015). Cholesterol confusion and statin controversy. *World J. Cardiol.* 7, 404–409. doi: 10.4330/wjc.v7.i7.404
- Eefting, D., Schepers, A., De Vries, M. R., Pires, N. M. M., Grimbergen, J. M., Lagerweij, T., et al. (2007). The effect of interleukin-10 knock-out and overexpression on neointima formation in hypercholesterolemic APOE*3-Leiden mice. *Atherosclerosis* 193, 335–342. doi: 10.1016/j.atherosclerosis.2006.09.032
- Fenton, S. A. M., Veldhuijzen van Zanten, J. J. C. S., Kitas, G. D., Duda, J. L., Rouse, P. C., Yu, C.-A., et al. (2017). Sedentary behaviour is associated with increased long-term cardiovascular risk in patients with rheumatoid arthritis independently of moderate-to-vigorous physical activity. *BMC Musculoskelet. Disord.* 18:131. doi: 10.1186/s12891-017-1473-9
- Fernández-Friera, L., Fuster, V., López-Melgar, B., Oliva, B., García-Ruiz, J. M., Mendiguren, J., et al. (2017). Normal LDL-cholesterol levels are associated with subclinical atherosclerosis in the absence of risk factors. *J. Am. Coll. Cardiol.* 70, 2979–2991. doi: 10.1016/j.jacc.2017.10.024
- Florido, R., Kwak, L., Lazo, M., Nambi, V., Ahmed, H. M., Hegde, S. M., et al. (2018). Six-year changes in physical activity and the risk of incident heart failure: the atherosclerosis risk in communities (ARIC) study. *Circulation* 137, 2142–2151. doi: 10.1161/CIRCULATIONAHA.117.030226
- Foks, A. C., Lichtman, A. H., and Kuiper, J. (2015). Treating atherosclerosis with regulatory T cells. *Arterioscler. Thromb. Vasc. Biol.* 35, 280–287. doi: 10.1161/ATVBAHA.114.303568
- Françavilla, C. V., Sessa, F., Salerno, M., Albano, G. D., Villano, I., Messina, G., et al. (2018). Influence of football on physiological cardiac indexes in professional and young athletes. *Front. Physiol.* 9:153. doi: 10.3389/fphys.2018.00153
- Gabriel, S. E. (2008). Cardiovascular morbidity and mortality in rheumatoid arthritis. *Am. J. Med.* 121, S9–S14. doi: 10.1016/j.amjmed.2008.06.011
- Galetta, F., Franzoni, F., Tocchini, L., Camici, M., Milanese, D., Belatti, F., et al. (2013). Effect of physical activity on heart rate variability and carotid intima-media thickness in older people. *Intern. Emerg. Med.* 8, 27–29. doi: 10.1007/s11739-013-0919-9
- Gimbrone, M. A., and García-Cardeña, G. (2016). Endothelial cell dysfunction and the pathobiology of atherosclerosis. *Circ. Res.* 118, 620–636. doi: 10.1161/CIRCRESAHA.115.306301
- Gisterà, A., and Hansson, G. K. (2017). The immunology of atherosclerosis. *Nat. Rev. Nephrol.* 13, 368–380. doi: 10.1038/nrneph.2017.51
- Goff, D. C., Lloyd-Jones, D. M., Bennett, G., Coady, S., D'Agostino, R. B., Gibbons, R., et al. (2014). 2013 ACC/AHA guideline on the assessment of cardiovascular risk: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. *Circulation* 129, S49–S73. doi: 10.1161/01.cir.0000437741.48606.98
- Green, D. J., and Smith, K. J. (2017). Effects of exercise on vascular function, structure, and health in humans. *Cold Spring Harb. Perspect. Med.* 8:a029819. doi: 10.1101/cshperspect.a029819
- Grundey, S. M. (2016). Primary prevention of cardiovascular disease with statins: assessing the evidence base behind clinical guidance. *Clin. Pharm.* 8. doi: 10.1211/CP.2016.20200568
- Gupta, S., Pablo, A. M., Jiang, X. C., Wang, N., Tall, A. R., and Schindler, C. (1997). IFN-gamma potentiates atherosclerosis in ApoE knock-out mice. *J. Clin. Invest.* 99, 2752–2761. doi: 10.1172/JCI119465
- Halfvarson, J., Brislaw, C. J., Lamendella, R., Vázquez-Baeza, Y., Walters, W. A., Bramer, L. M., et al. (2017). Dynamics of the human gut microbiome in inflammatory bowel disease. *Nat. Microbiol.* 2:17004. doi: 10.1038/nmicrobiol.2017.4
- Henrot, P., Foret, J., Barnetche, T., Lazaro, E., Duffau, P., Seneschal, J., et al. (2018). Assessment of subclinical atherosclerosis in systemic lupus erythematosus: a systematic review and meta-analysis. *Joint Bone Spine* 85, 155–163. doi: 10.1016/j.jbspin.2017.12.009
- Hotta, K., Chen, B., Behnke, B. J., Ghosh, P., Stabley, J. N., Bramy, J. A., et al. (2017). Exercise training reverses age-induced diastolic dysfunction and restores coronary microvascular function. *J. Physiol.* 595, 3703–3719. doi: 10.1113/JP274172
- Huber, S. A., Sakkinen, P., Conze, D., Hardin, N., and Tracy, R. (1999). Interleukin-6 exacerbates early atherosclerosis in mice. *Arterioscler. Thromb. Vasc. Biol.* 19, 2364–2367. doi: 10.1161/01.ATV.19.10.2364
- Jackaman, C., Tomay, F., Duong, L., Abdol Razak, N. B., Pixley, F. J., Metharom, P., et al. (2017). Aging and cancer: the role of macrophages and neutrophils. *Ageing Res. Rev.* 36, 105–116. doi: 10.1016/j.arr.2017.03.008
- Jian, D., Wang, W., Zhou, X., Jia, Z., Wang, J., Yang, M., et al. (2018). Interferon-induced protein 35 inhibits endothelial cell proliferation, migration and re-endothelialization of injured arteries by inhibiting the nuclear factor-kappa B pathway. *Acta Physiol.* doi: 10.1111/apha.13037 [Epub ahead of print].
- Jie, Z., Xia, H., Zhong, S.-L., Feng, Q., Li, S., Liang, S., et al. (2017). The gut microbiome in atherosclerotic cardiovascular disease. *Nat. Commun.* 8:845. doi: 10.1038/s41467-017-00900-1
- Karlsson, F. H., Fåk, F., Nookaew, I., Tremaroli, V., Fagerberg, B., Petranovic, D., et al. (2012). Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat. Commun.* 3:1245. doi: 10.1038/ncomms2266
- Kelly, T. N., Bazzano, L. A., Ajami, N. J., He, H., Zhao, J., Petrosino, J. F., et al. (2016). Gut microbiome associates with lifetime cardiovascular disease risk profile among Bogalusa heart study participants. *Circ. Res.* 119, 956–964. doi: 10.1161/CIRCRESAHA.116.309219
- King, V. L., Cassis, L. A., and Daugherty, A. (2007). Interleukin-4 does not influence development of hypercholesterolemia or angiotensin II-induced atherosclerotic lesions in mice. *Am. J. Pathol.* 171, 2040–2047. doi: 10.2353/ajpath.2007.060857
- Kirii, H., Niwa, T., Yamada, Y., Wada, H., Saito, K., Iwakura, Y., et al. (2003). Lack of Interleukin-1beta decreases the severity of atherosclerosis in ApoE-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* 23, 656–660. doi: 10.1161/01.ATV.0000064374.15232.C3
- Koelwyn, G. J., Wennerberg, E., Demaria, S., and Jones, L. W. (2015). Exercise in regulation of inflammation-immune axis function in cancer initiation and progression. *Oncology* 29, 908–920. doi: 10.1016/j.semcancer.2015.04.010. Targeting
- Koehn, R. R., and Weber, C. (2010). Therapeutic targeting of chemokine interactions in atherosclerosis. *Nat. Rev. Drug Discov.* 9, 141–153. doi: 10.1038/nrd3048
- Koehn, R. R., and Weber, C. (2011). Chemokines: established and novel targets in atherosclerosis. *EMBO Mol. Med.* 3, 713–725. doi: 10.1002/emmm.201100183
- Koeth, R. A., Wang, Z., Levison, B. S., Buffa, J. A., Org, E., Sheehy, B. T., et al. (2013). Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat. Med.* 19, 576–585. doi: 10.1038/nm.3145
- Kokkinos, P. (2012). Physical activity, health benefits, and mortality risk. *ISRN Cardiol.* 2012:718789. doi: 10.5402/2012/718789
- Kreznar, J. H., Keller, M. P., Traeger, L. L., Rabaglia, M. E., Schueler, K. L., Stapleton, D. S., et al. (2017). Host genotype and gut microbiome modulate insulin secretion and diet-induced metabolic phenotypes. *Cell Rep.* 18, 1739–1750. doi: 10.1016/j.celrep.2017.01.062
- Lin, J., Kakkur, V., and Lu, X. (2014). Impact of MCP-1 in atherosclerosis. *Curr. Pharm. Des.* 20, 4580–4588. doi: 10.2174/1381612820666140522115801
- Lin, X., Zhang, X., Guo, J., Roberts, C. K., McKenzie, S., Wu, W.-C., et al. (2015). Effects of exercise training on cardiorespiratory fitness and biomarkers of cardiometabolic health: a systematic review and meta-analysis of randomized controlled trials. *J. Am. Heart Assoc.* 4, 1–28. doi: 10.1161/JAHA.115.002014

- Llewellyn, S. R., Britton, G. J., Contijoch, E. J., Vennaro, O. H., Mortha, A., Colombel, J.-F., et al. (2017). Interactions between diet and the intestinal microbiota alter intestinal permeability and colitis severity in mice. *Gastroenterology* 154, 1037.e2–1046.e2. doi: 10.1053/j.gastro.2017.11.030
- Lusis, A. J. (2000). Atherosclerosis. *Nature* 407, 233–241. doi: 10.1038/35025203
- Mallat, Z., Gojova, A., Brun, V., Esposito, B., Fournier, N., Cottrez, F., et al. (2003). Induction of a regulatory T cell type 1 response reduces the development of atherosclerosis in apolipoprotein E-knockout mice. *Circulation* 108, 1232–1237. doi: 10.1161/01.CIR.0000089083.61317.A1
- Mehta, N. N., Teague, H. L., Swindell, W. R., Baumer, Y., Ward, N. L., Xing, X., et al. (2017). IFN- γ and TNF- α synergism may provide a link between psoriasis and inflammatory atherogenesis. *Sci. Rep.* 7:13831. doi: 10.1038/s41598-017-14365-1
- Messina, G., Di Bernardo, G., Viggiano, A., De Luca, V., Monda, V., Messina, A., et al. (2016). Exercise increases the level of plasma orexin A in humans. *J. Basic Clin. Physiol. Pharmacol.* 27, 611–616. doi: 10.1515/jbcp-2015-0133
- Messina, G., Valenzano, A., Moscatelli, F., Salerno, M., Lonigro, A., Esposito, T., et al. (2017). Role of autonomic nervous system and orexinergic system on adipose tissue. *Front. Physiol.* 8:137. doi: 10.3389/fphys.2017.00137
- Michelson, A. D., Barnard, M. R., Krueger, L. A., Valeri, C. R., and Furman, M. I. (2001). Circulating monocyte-platelet aggregates are a more sensitive marker of in vivo platelet activation than platelet surface P-selectin: studies in baboons, human coronary intervention, and human acute myocardial infarction. *Circulation* 104, 1533–1537. doi: 10.1161/hc3801.095588
- Moholdt, T., Lavie, C. J., and Nauman, J. (2018). Sustained physical activity, not weight loss, associated with improved survival in coronary heart disease. *J. Am. Coll. Cardiol.* 71, 1094–1101. doi: 10.1016/j.jacc.2018.01.011
- Moldoveanu, A. I., Shephard, R. J., and Shek, P. N. (2000). Exercise elevates plasma levels but not gene expression of IL-1 β , IL-6, and TNF- α in blood mononuclear cells. *J. Appl. Physiol.* 89, 1499–1504. doi: 10.1152/jappl.2000.89.4.1499
- Moore, K. J., Sheedy, F. J., and Fisher, E. A. (2013). Macrophages in atherosclerosis: a dynamic balance. *Nat. Rev. Immunol.* 13, 709–721. doi: 10.1038/nri3520
- Moss, J. W., and Ramji, D. P. (2016). Cytokines: roles in atherosclerosis disease progression and potential therapeutic targets. *Future Med. Chem.* 8, 1317–1330. doi: 10.4155/fmc-2016-0072
- Murtagh, E. M., Nichols, L., Mohammed, M. A., Holder, R., Nevill, A. M., and Murphy, M. H. (2015). The effect of walking on risk factors for cardiovascular disease: an updated systematic review and meta-analysis of randomised control trials. *Prev. Med.* 72, 34–43. doi: 10.1016/j.jpmed.2014.12.041
- Myers, J. (2003). Cardiology patient pages. Exercise and cardiovascular health. *Circulation* 107, e2–e5. doi: 10.1161/01.CIR.0000048890.59383.8D
- Na, Y. M., Kim, M. Y., Kim, Y. K., Ha, Y. R., and Yoon, D. S. (2000). Exercise therapy effect on natural killer cell cytotoxic activity in stomach cancer patients after curative surgery. *Arch. Phys. Med. Rehabil.* 81, 777–779. doi: 10.1053/apmr.2000.3871
- Olson, B. M., Sullivan, J. A., and Burlingham, W. J. (2013). Interleukin 35: a key mediator of suppression and the propagation of infectious tolerance. *Front. Immunol.* 4:315. doi: 10.3389/fimmu.2013.00315
- Ou, H.-X., Guo, B.-B., Liu, Q., Li, Y.-K., Yang, Z., Feng, W.-J., et al. (2018). Regulatory T cells as a new therapeutic target for atherosclerosis. *Acta Pharmacol. Sin.* doi: 10.1038/aps.2017.140 [Epub ahead of print].
- Palomo, I., Herrera, J., Cancino, V., Alarcón, M., Mujica, V., Leiva, E., et al. (2010). Physical activity reduces circulating TNF-alpha but not pro-thrombotic factors levels in patients with metabolic syndrome. *Diabetes Metab. Syndr. Clin. Res. Rev.* 4, 234–238. doi: 10.1016/J.DSX.2010.05.004
- Paramsothy, S., Kamm, M. A., Kaakoush, N. O., Walsh, A. J., van den Bogaerde, J., Samuel, D., et al. (2017). Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet* 389, 1218–1228. doi: 10.1016/S0140-6736(17)30182-4
- Parsons, T. J., Sartini, C., Welsh, P., Sattar, N., Ash, S., Lennon, L. T., et al. (2017). Physical activity, sedentary behavior, and inflammatory and hemostatic markers in men. *Med. Sci. Sports Exerc.* 49, 459–465. doi: 10.1249/MSS.0000000000001113
- Payette, C., Blackburn, P., Lamarche, B., Tremblay, A., Bergeron, J., Lemieux, I., et al. (2009). Sex differences in postprandial plasma tumor necrosis factor- α , interleukin-6, and C-reactive protein concentrations. *Metabolism* 58, 1593–1601. doi: 10.1016/j.metabol.2009.05.011
- Pedersen, B. K. (2006). The anti-inflammatory effect of exercise: its role in diabetes and cardiovascular disease control. *Essays Biochem.* 42, 105–117. doi: 10.1042/bse0420105
- Pedersen, B. K. (2017). Anti-inflammatory effects of exercise: role in diabetes and cardiovascular disease. *Eur. J. Clin. Invest.* 47, 600–611. doi: 10.1111/eci.12781
- Pedersen, B. K., Steensberg, A., and Schjerling, P. (2001). Muscle-derived interleukin-6: possible biological effects. *J. Physiol.* 536, 329–337. doi: 10.1111/J.1469-7793.2001.0329C.XD
- Pedersen, B. K., and Ullum, H. (1994). NK cell response to physical activity: possible mechanisms of action. *Med. Sci. Sports Exerc.* 26, 140–146. doi: 10.1249/00005768-199402000-00003
- Pedersen, L., Idorn, M., Olofsson, G. H., Lauenborg, B., Nookaew, I., Hansen, R. H., et al. (2016). Voluntary running suppresses tumor growth through epinephrine- and IL-6-dependent NK cell mobilization and redistribution. *Cell Metab.* 23, 554–562. doi: 10.1016/j.cmet.2016.01.011
- Phillips, C. M., Dillon, C. B., and Perry, I. J. (2017). Does replacing sedentary behaviour with light or moderate to vigorous physical activity modulate inflammatory status in adults? *Int. J. Behav. Nutr. Phys. Act.* 14:138. doi: 10.1186/s12966-017-0594-8
- Piepoli, M. F., Hoes, A. W., Agewall, S., Albus, C., Brotons, C., Catapano, A. L., et al. (2016). 2016 European Guidelines on cardiovascular disease prevention in clinical practice. *Eur. Heart J.* 37, 2315–2381. doi: 10.1093/eurheartj/ehw106
- Rajamani, K., and Fisher, M. (2017). “An overview of atherosclerosis,” in *Primer on Cerebrovascular Diseases*, 2nd Edn, eds L. R. Caplan, J. Biller, M. Leary, E. Lo, A. Thomas, M. Yenari, et al. (Cambridge, MA: Academic Press), 105–108. doi: 10.1016/B978-0-12-803058-5.00020-5
- Ramji, D. P., and Davies, T. S. (2015). Cytokines in atherosclerosis: key players in all stages of disease and promising therapeutic targets. *Cytokine Growth Factor Rev.* 26, 673–685. doi: 10.1016/j.cytogfr.2015.04.003
- Reiss, A. B., Siegart, N. M., and De Leon, J. (2017). Interleukin-6 in atherosclerosis: atherogenic or atheroprotective? *Clin. Lipidol.* 12, 14–23. doi: 10.1080/17584299.2017.1319787
- Ridaura, V. K., Faith, J. J., Rey, F. E., Cheng, J., Duncan, A. E., Kau, A. L., et al. (2013). Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science* 341:1241214. doi: 10.1126/science.1241214
- Ridker, P. M., Everett, B. M., Thuren, T., MacFadyen, J. G., Chang, W. H., Ballantyne, C., et al. (2017). Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N. Engl. J. Med.* 377, 1119–1131. doi: 10.1056/NEJMoa1707914
- Ridker, P. M., Howard, C. P., Walter, V., Everett, B., Libby, P., Hensen, J., et al. (2012). Effects of interleukin-1 β inhibition with canakinumab on hemoglobin A1c, lipids, C-reactive protein, interleukin-6, and fibrinogen: a phase IIb randomized, placebo-controlled trial. *Circulation* 126, 2739–2748. doi: 10.1161/CIRCULATIONAHA.112.122556
- Ridker, P. M., MacFadyen, J. G., Everett, B. M., Libby, P., Thuren, T., Glynn, R. J., et al. (2018). Relationship of C-reactive protein reduction to cardiovascular event reduction following treatment with canakinumab: a secondary analysis from the CANTOS randomised controlled trial. *Lancet* 391, 319–328. doi: 10.1016/S0140-6736(17)32814-3
- Robertson, A.-K. L., Rudling, M., Zhou, X., Gorelik, L., Flavell, R. A., and Hansson, G. K. (2003). Disruption of TGF-beta signaling in T cells accelerates atherosclerosis. *J. Clin. Invest.* 112, 1342–1350. doi: 10.1172/JCI18607
- Romano, K. A., Vivas, E. I., Amador-Noguez, D., and Rey, F. E. (2015). Intestinal microbiota composition modulates choline bioavailability from diet and accumulation of the proatherogenic metabolite trimethylamine-N-oxide. *MBio* 6:e02481. doi: 10.1128/mBio.02481-14
- Ross, R. (1999). Atherosclerosis — An inflammatory disease. *N. Engl. J. Med.* 340, 115–126. doi: 10.1056/NEJM199901143400207
- Sakaue, A., Adachi, H., Enomoto, M., Fukami, A., Kumagai, E., Nakamura, S., et al. (2017). Association between physical activity, sitting time and mortality in a general population: the 15-year prospective survey in the Tanushimaru study. *Atherosclerosis* 263, e186–e187. doi: 10.1016/J.ATHEROSCLEROSIS.2017.06.598
- Scheller, J., Chalaris, A., Schmidt-Arras, D., and Rose-John, S. (2011). The pro- and anti-inflammatory properties of the cytokine interleukin-6. *Biochim. Biophys. Acta* 1813, 878–888. doi: 10.1016/J.BBAMCR.2011.01.034
- Schieffer, B., Selle, T., Hilfiker, A., Hilfiker-Kleiner, D., Grote, K., Tietge, U. J. F., et al. (2004). Impact of interleukin-6 on plaque development and morphology

- in experimental atherosclerosis. *Circulation* 110, 3493–3500. doi: 10.1161/01.CIR.0000148135.08582.97
- Schild, M., Eichner, G., Beiter, T., Zügel, M., Krumholz-Wagner, I., Hudemann, J., et al. (2016). Effects of acute endurance exercise on plasma protein profiles of endurance-trained and untrained individuals over time. *Mediators Inflamm.* 2016:4851935. doi: 10.1155/2016/4851935
- Senthong, V., Wang, Z., Fan, Y., Wu, Y., Hazen, S. L., and Tang, W. H. W. (2016). Trimethylamine N-oxide and mortality risk in patients with peripheral artery disease. *J. Am. Heart Assoc.* 5:e004237. doi: 10.1161/JAHA.116.004237
- Sessa, F., Anna, V., Messina, G., Cibelli, G., Monda, V., Marsala, G., et al. (2018). Heart rate variability as predictive factor for sudden cardiac death. *Aging* 10, 166–177. doi: 10.18632/aging.101386
- Shah, S. R., Abbasi, Z., Fatima, M., Ochani, R. K., Shah Nawaz, W., Asim Khan, M., et al. (2018). Canakinumab and cardiovascular outcomes: results of the CANTOS trial. *J. Community Hosp. Intern. Med. Perspect.* 8, 21–22. doi: 10.1080/20009666.2018.1428023
- Sharif, K., Watad, A., Bragazzi, N. L., Lichtbroun, M., Amital, H., and Shoenfeld, Y. (2018). Physical activity and autoimmune diseases: get moving and manage the disease. *Autoimmun. Rev.* 17, 53–72. doi: 10.1016/j.autrev.2017.11.010
- Sheikine, Y. A., and Hansson, G. K. (2006). Chemokines as potential therapeutic targets in atherosclerosis. *Curr. Drug Targets* 7, 13–27. doi: 10.2174/138945006775270240
- Shephard, R. J., and Shek, P. N. (1994). Potential impact of physical activity and sport on the immune system—a brief review. *Br. J. Sports Med.* 28, 247–255. doi: 10.1136/BJSM.28.4.247
- Skeoch, S., and Bruce, I. N. (2015). Atherosclerosis in rheumatoid arthritis: is it all about inflammation? *Nat. Rev. Rheumatol.* 11, 390–400. doi: 10.1038/nrrheum.2015.40
- Slingerland, A. E., Schwabkey, Z., Wiesnoski, D. H., and Jenq, R. R. (2017). Clinical evidence for the microbiome in inflammatory diseases. *Front. Immunol.* 8:400. doi: 10.3389/fimmu.2017.00400
- Slusher, A. L., Zúñiga, T. M., and Acevedo, E. O. (2017). Maximal exercise alters the inflammatory phenotype and response of mononuclear cells. *Med. Sci. Sport Exerc.* 1, doi: 10.1249/MSS.0000000000001480
- Smith, J. K., Dykes, R., Douglas, J. E., Krishnaswamy, G., and Berk, S. (1999). Long-term exercise and atherogenic activity of blood mononuclear cells in persons at risk of developing ischemic heart disease. *JAMA* 281, 1722–1727. doi: 10.1001/jama.281.18.1722
- Soares-Miranda, L., Siscovick, D. S., Psaty, B. M., Longstreth, W. T., Mozaffarian, D., and Mozaffarian, D. (2016). Physical activity and risk of coronary heart disease and stroke in older adults: the cardiovascular health study. *Circulation* 133, 147–155. doi: 10.1161/CIRCULATIONAHA.115.018323
- Steppich, B., Dayyani, F., Gruber, R., Lorenz, R., Mack, M., and Ziegler-Heitbrock, H. W. L. (2000). Selective mobilization of CD14(+)CD16(+) monocytes by exercise. *Am. J. Physiol. Physiol.* 279, C578–C586. doi: 10.1152/ajpcell.2000.279.3.C578
- Strömberg, A., Olsson, K., Dijksterhuis, J. P., Rullman, E., Schulte, G., and Gustafsson, T. (2016). CX3CL1—a macrophage chemoattractant induced by a single bout of exercise in human skeletal muscle. *Am. J. Physiol. Integr. Comp. Physiol.* 310, R297–R304. doi: 10.1152/ajpregu.00236.2015
- Struyf, S., van Coillie, E., Paemen, L., Put, W., Lenaerts, J.-P., Proost, P., et al. (1998). Synergistic induction of MCP-1 and -2 by IL-1 β and interferons in fibroblasts and epithelial cells. *J. Leukoc. Biol.* 63, 364–372. doi: 10.1002/jlb.63.3.364
- Taylor, F., Ward, K., Moore, T. H., Burke, M., Davey Smith, G., Casas, J.-P., et al. (2011). Statins for the primary prevention of cardiovascular disease. *Cochrane Database Syst. Rev.* CD004816. doi: 10.1002/14651858.CD004816.pub4
- Tegos, T. J., Kalodiki, E., Sabetai, M. M., and Nicolaides, A. N. (2001). The genesis of atherosclerosis and risk factors: a review. *Angiology* 52, 89–98.
- Turner, M. D., Nedjai, B., Hurst, T., and Pennington, D. J. (2014). Cytokines and chemokines: at the crossroads of cell signalling and inflammatory disease. *Biochim. Biophys. Acta* 1843, 2563–2582. doi: 10.1016/J.BBAMCR.2014.05.014
- Upadhyaya, S., Mooteri, S., Peckham, N., and Pai, R. G. (2004). Atherogenic effect of interleukin-2 and antiatherogenic effect of interleukin-2 antibody in apo-E-deficient mice. *Angiology* 55, 289–294. doi: 10.1177/000331970405500308
- Valero-Elizondo, J., Salami, J. A., Osondu, C. U., Ogunmoroti, O., Arrieta, A., Spatz, E. S., et al. (2016). Economic impact of moderate-vigorous physical activity among those with and without established cardiovascular disease: 2012 medical expenditure panel survey. *J. Am. Heart Assoc.* 5:e003614. doi: 10.1161/JAHA.116.003614
- van den Elsen, L. W., Poyntz, H. C., Weyrich, L. S., Young, W., and Forbes-Blom, E. E. (2017). Embracing the gut microbiota: the new frontier for inflammatory and infectious diseases. *Clin. Transl. Immunol.* 6:e125. doi: 10.1038/cti.2016.91
- van Hall, G., Steensberg, A., Sacchetti, M., Fischer, C., Keller, C., Schjerling, P., et al. (2003). Interleukin-6 stimulates lipolysis and fat oxidation in humans. *J. Clin. Endocrinol. Metab.* 88, 3005–3010. doi: 10.1210/jc.2002-021687
- Veillard, N. R., Kwak, B., Pelli, G., Mulhaupt, F., James, R. W., Proudfoot, A. E. I., et al. (2004). Antagonism of RANTES receptors reduces atherosclerotic plaque formation in mice. *Circ. Res.* 94, 253–261. doi: 10.1161/01.RES.0000109793.175914.E
- Vella, C. A., Allison, M. A., Cushman, M., Jenny, N. S., Miles, M. P., Larsen, B., et al. (2017). Physical activity and adiposity-related inflammation: the MESA. *Med. Sci. Sports Exerc.* 49, 915–921. doi: 10.1249/MSS.0000000000001179
- Voloshyna, I., Littlefield, M. J., and Reiss, A. B. (2014). Atherosclerosis and interferon- γ : new insights and therapeutic targets. *Trends Cardiovasc. Med.* 24, 45–51. doi: 10.1016/j.tcm.2013.06.003
- Wainstein, M. V., Mossman, M., Araujo, G. N., Gonçalves, S. C., Gravina, G. L., Sangalli, M., et al. (2017). Elevated serum interleukin-6 is predictive of coronary artery disease in intermediate risk overweight patients referred for coronary angiography. *Diabetol. Metab. Syndr.* 9:67. doi: 10.1186/s13098-017-0266-5
- Waters, D. D., and Hsue, P. Y. (2017). PCSK9 inhibition to reduce cardiovascular risk: tempering expectations. *Circ. Res.* 120, 1537–1539. doi: 10.1161/CIRCRESAHA.117.311015
- Weinhold, M., Shimabukuro-Vornhagen, A., Franke, A., Theurich, S., Wahl, P., Hallek, M., et al. (2016). Physical exercise modulates the homeostasis of human regulatory T cells. *J. Allergy Clin. Immunol.* 137, 1607.e8–1610.e8. doi: 10.1016/j.jaci.2015.10.035
- Whelton, P. K., Carey, R. M., Aronow, W. S., Casey, D. E., Collins, K. J., Dennison Himmelfarb, C., et al. (2018). 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APHA/ASH/ASPC/NMA/PCNA guideline for the prevention, detection, evaluation, and management of high blood pressure in adults: executive summary: a report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. 71, 1269–1324. doi: 10.1016/j.jacc.2017.11.006
- Yakeu, G., Butcher, L., Isa, S., Webb, R., Roberts, A. W., Thomas, A. W., et al. (2010). Low-intensity exercise enhances expression of markers of alternative activation in circulating leukocytes: roles of PPAR γ and Th2 cytokines. *Atherosclerosis* 212, 668–673. doi: 10.1016/j.atherosclerosis.2010.07.002
- Zamani, A., Salehi, I., and Alahgholi-Hajibehzad, M. (2017). Moderate exercise enhances the production of interferon- γ and interleukin-12 in peripheral blood mononuclear cells. *Immune Netw.* 17, 186–191. doi: 10.4110/in.2017.17.3.186
- Zhao, W., Lei, T., Li, H., Sun, D., Mo, X., Wang, Z., et al. (2015). Macrophage-specific overexpression of interleukin-5 attenuates atherosclerosis in LDL receptor-deficient mice. *Gene Ther.* 22, 645–652. doi: 10.1038/gt.2015.33

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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