

Review Article



Unusual or Uncommon Histology of Gastric Cancer

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Conflict of Interest

No potential conflict of interest relevant to this article was reported.

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ABSTRACT

This review comprehensively examines the diverse spectrum of gastric cancers, focusing on unusual or uncommon histology that presents significant diagnostic and therapeutic challenges. While the predominant form, tubular adenocarcinoma, is well-characterized, this review focuses on lesser-known variants, including papillary adenocarcinoma, micropapillary carcinoma, adenosquamous carcinoma, squamous cell carcinoma (SCC), hepatoid adenocarcinoma, gastric choriocarcinoma, gastric carcinoma with lymphoid stroma, carcinosarcoma, gastroblastoma, parietal cell carcinoma, oncocytic adenocarcinoma, Paneth cell carcinoma, gastric adenocarcinoma of the fundic gland type, undifferentiated carcinoma, and extremely well-differentiated adenocarcinoma. Although these diseases have different nomenclatures characterized by distinct histopathological features, these phenotypes often overlap, making it difficult to draw clear boundaries. Furthermore, the number of cases was limited, and the unique histopathological nature and potential pathogenic mechanisms were not well defined. This review highlights the importance of understanding these rare variants for accurate diagnosis, effective treatment planning, and improving patient outcomes. This review emphasizes the need for ongoing research and case studies to enhance our knowledge of these uncommon forms of gastric cancer, which will ultimately contribute to more effective treatments and better prognostic assessments. This review aimed to broaden the pathological narrative by acknowledging and addressing the intricacies of all cancer types, regardless of their rarity, to advance patient care and improve prognosis.

Keywords: Pathology; Classification; Diagnosis

INTRODUCTION

Gastric cancer remains a significant clinical and diagnostic challenge and is characterized by various histological types and the occasional occurrence of metastases from other malignancies. The most prevalent form of stomach cancer is tubular adenocarcinoma, which accounts for over half of all cases and has been reported to be as high as 72% in South Korea [1,2]. This histological type is well characterized in terms of epidemiology, pathogenesis, and clinical management. However, within the spectrum of gastric malignancies, there is a subset of unusual gastric cancers that exhibit unique pathological features that defy the typical presentation patterns. These rare entities often present diagnostic conundrums and therapeutic challenges due to their atypical presentation and sometimes aggressive nature.

However, the unusual forms of gastric cancers are poorly understood. This type of cancer includes papillary adenocarcinoma, micropapillary carcinoma, adenosquamous carcinoma, SCC, hepatoid adenocarcinoma, gastric choriocarcinoma, gastric carcinoma with lymphoid stroma (GCLS), carcinosarcoma, gastroblastoma, parietal cell carcinoma, oncocytic adenocarcinoma, Paneth cell carcinoma, gastric adenocarcinoma of the fundic gland, undifferentiated carcinoma, and extremely well-differentiated adenocarcinoma. Each unusual form of stomach cancer has distinct histopathological characteristics, potentially unique pathogenic mechanisms, and molecular profiles.

This review aimed to synthesize the current knowledge and case reports regarding unusual and uncommon gastric cancers, providing insight into their histology and diagnostic challenges.

By shedding light on these lesser-known facets of gastric cancer, this review contributes to a broader oncological narrative that acknowledges and addresses the intricacies of all cancer types, regardless of their rarity, to advance patient care and improve prognosis.

PAPILLARY ADENOCARCINOMA

Papillary adenocarcinoma of the stomach is a distinct and relatively rare subtype, accounting for 2.7%–9.9% of cases [3]. Grossly, these tumors appear as exophytic polypoid masses. Tumor cells form well-differentiated, elongated, finger-like, or papillary processes lined with columnar or cuboidal epithelial cells, maintaining their polarity (**Fig. 1**). These cells are supported by fibrovascular connective tissue cores that sometimes contain tubular structures in a tubulopapillary configuration.

Moreover, papillary adenocarcinomas are associated with a higher risk of adverse outcomes [4,5]. This is highlighted by their tendency to metastasize to the liver and the resulting poor survival rates linked to this subtype.

Clear cell variants have been reported in papillary adenocarcinoma, which share basic histologic features with papillary carcinoma, including a glycogen-rich clear cytoplasm and frequent tubulopapillary patterns [6,7]. However, this subtype continues to be reported as the alpha-fetoprotein (AFP)-producing tumor and designated as gastric adenocarcinoma with enteroblastic differentiation (GAED) [8] (discussed in “Hepatoid adenocarcinoma”).

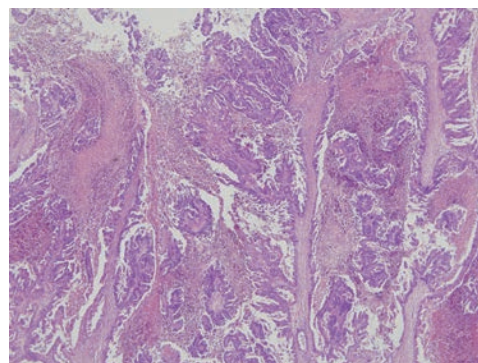


Fig. 1. Papillary adenocarcinoma. The tumor cells form well-differentiated, elongated, finger-like, or papillary processes that are lined by columnar or cuboidal epithelial cells maintaining their polarity (hematoxylin and eosin, original magnification $\times 40$). Image courtesy of Dr. Jihun Kim.

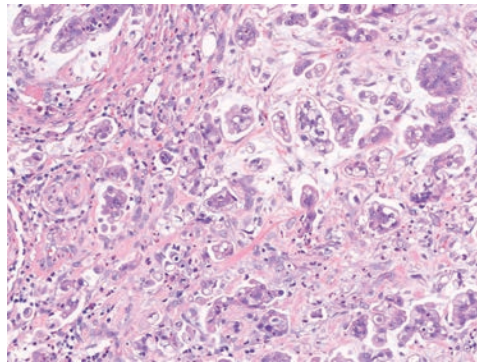


Fig. 2. Micropapillary carcinoma. Small nests of tumor cells show the absence of fibrovascular cores within transparent lacunar spaces that mimic the architecture of lymphatic or vascular channels (hematoxylin and eosin, original magnification $\times 100$). Image courtesy of Dr. Soomin Ahn.

MICROPAPILLARY CARCINOMA

Micropapillary adenocarcinoma is rare and has a worse prognosis due to higher lymphovascular invasion and lymph node metastasis rates [9,10], and it is characterized by the presence of small tumor cell nests without fibrovascular cores within transparent lacunar spaces that mimic the architecture of lymphatic or vascular channels (**Fig. 2**), micropapillary adenocarcinoma arises in conventional tumors, including tubular or papillary adenocarcinoma, and its proportion varies from 5% to 90% [9,11].

ADENOSQUAMOUS CARCINOMA

Adenosquamous carcinoma of the stomach forms 0.25%–2% of all cases [12,13]. It is a primary carcinoma that exhibits both glandular and squamous cell elements, with the latter accounting for approximately 25% of the tumor volume [14]. Clinically, these tumors often present as large masses and are most frequently located in the lower third of the stomach, although they can occur throughout the organ [15].

Histopathologically, adenosquamous carcinoma demonstrates a distinct squamous component characterized by keratin pearl formation and intercellular bridges along with the glandular element of adenocarcinoma (**Fig. 3A**) [16]. The transition between these 2

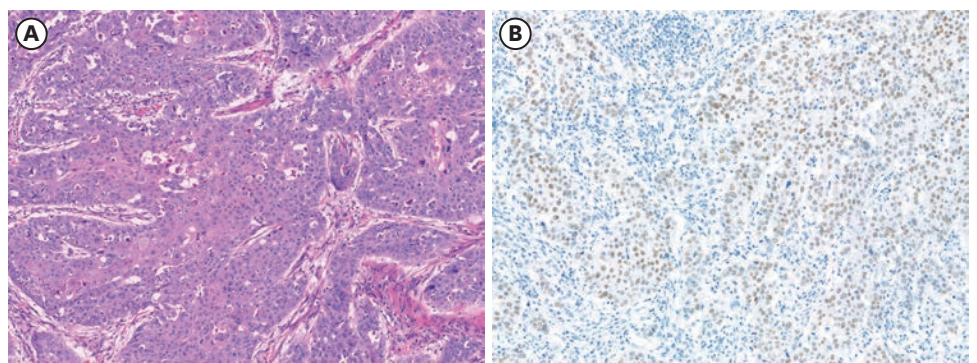


Fig. 3. (A) Adenosquamous carcinoma shows a distinct squamous component alongside the glandular element of adenocarcinoma (hematoxylin and eosin, original magnification, $\times 40$). (B) Squamous components of the tumor are positive in p40 immunohistochemistry (p40, original magnification, $\times 40$). Case courtesy of Dr. Su-Jin Shin.

components can be abrupt or intermingled, suggesting a potential origin for multipotentially common stem cells [17]. The squamous cell component exhibited features similar to those observed in esophageal SCC, and p63/p40 immunohistochemistry confirmed squamous differentiation (**Fig. 3B**). These tumors have a poor prognosis owing to their deep penetration and high propensity for lymphovascular invasion, which often results in an advanced stage at diagnosis and a higher likelihood of metastasis [18,19].

SCC

Gastric SCC is even rarer than adenosquamous carcinoma, accounting for 0.04%–0.07% of gastric cancers, with under 100 cases reported in the literature [20–25]. It is characterized by the presence of keratinocyte-type cells with features such as intercellular bridges and keratinization and lacks the glandular components observed in adenosquamous carcinomas (**Fig. 4**).

The origin of gastric SCC is not definitively known; however, it has been hypothesized to arise from squamous metaplasia within adenocarcinomas, ectopic squamous nests, or multipotent stem cells in the gastric mucosa capable of bidirectional differentiation [24,26,27]. Gastric SCC is typically diagnosed at an advanced stage, and its prognosis is generally poor, although certain case reports have demonstrated better responses to chemotherapy than their adenocarcinoma counterparts [28,29].

HEPATOID ADENOCARCINOMA

Hepatoid adenocarcinoma is characterized by the presence of tumor cells that exhibit features reminiscent of hepatocellular carcinoma (HCC), notable for eosinophilic polygonal tumor cells (**Fig. 5A**), and AFP production, a biomarker typically associated with HCC and fetal hepatic differentiation, detectable both serologically and immunohistochemically (**Fig. 5B**) in tumor tissues [30,31].

Hepatoid adenocarcinoma typically affects patients aged >50 years and is predominantly located in the stomach [32]. Morphologically, hepatoid adenocarcinoma exhibits large polygonal cells with eosinophilic cytoplasm, often arranged in trabecular or acinar

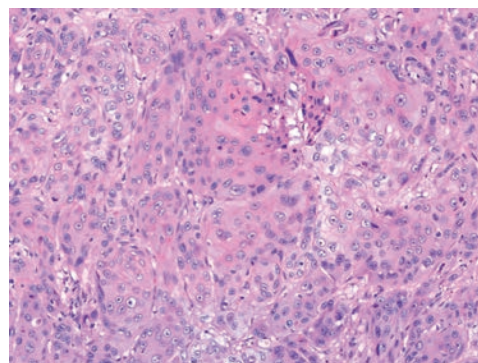


Fig. 4. Squamous cell carcinoma. This cancer is characterized by the presence of keratinocyte-type cells, with features such as intercellular bridges and keratinization, and lacks glandular components (hematoxylin and eosin, original magnification, $\times 100$). Image courtesy of Dr. Soomin Ahn.

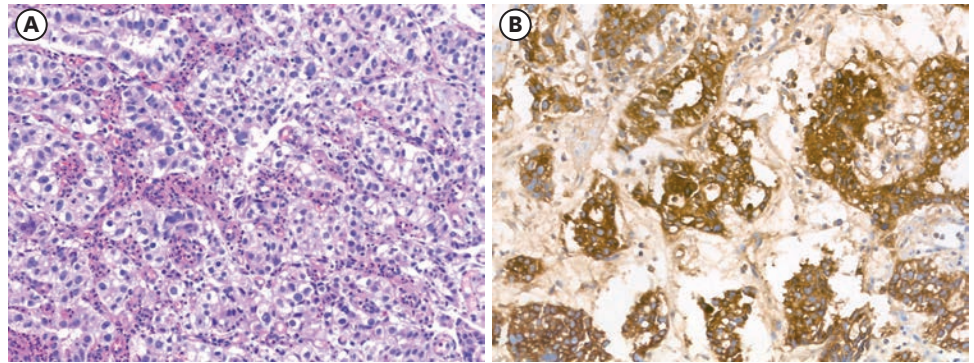


Fig. 5. (A) Hepatoid adenocarcinoma is characterized by the presence of tumor cells that exhibit features reminiscent of hepatocellular carcinoma, notable for eosinophilic polygonal tumor cells (hematoxylin and eosin, original magnification, $\times 100$). (B) Tumor cells are positive for alpha-fetoprotein immunohistochemistry (alpha-fetoprotein, original magnification, $\times 200$). Images courtesy of Dr. Soomin Ahn.

structures, similar to those seen in HCC [31,32]. Hepatoid adenocarcinoma is known for its Periodic acid-Schiff-diastase-resistant intracytoplasmic hyaline globules and may exhibit less differentiated regions with bizarre giant and spindle cells [32,33]. These tumors frequently exhibit mixed histological features of hepatoid areas with traditional tubulopapillary adenocarcinomatous patterns.

The disease is aggressive, often infiltrating the venous and lymphatic vessels, leading to a high incidence of liver and lymph node metastases and culminating in a poorer prognosis, regardless of AFP production [32,34–36]. Diagnosing hepatoid adenocarcinoma with multiple liver metastases can be challenging. Cases with multiple hepatic mass lesions with high serum AFP levels and the absence of background chronic liver disease should raise the possibility of hepatoid adenocarcinoma [37].

Immunohistochemically, hepatoid adenocarcinoma can be differentiated from HCC by the expression of markers such as Hep-Par-1, which typically shows extensive staining in HCC, but only focal staining in hepatoid adenocarcinoma [38]. In addition, significant presence of cytokeratin (CK) 19 and CK20 were detected in hepatoid carcinomas (HACs) at rates of 94% and 47%, respectively. These rates are considerably higher than those of HCCs, which suggests that CK19 is present in 0%–50% and CK20 in 0%–42% [39–41].

AFP-expressing carcinomas appear in 2.6%–5.4% of gastric cancers [42,43]. They include GAED, well-differentiated papillary/tubular adenocarcinoma with a clear cytoplasm, and yolk sac tumor-like carcinoma [44–46]. GAED is characterized by a tubular papillary structure and columnar cancer cells with clear cytoplasm. This differentiation is reminiscent of the early developmental stages of fetal gastrointestinal epithelium in fetuses [8,47]. These morphological features lead to the expression of several fetal gut markers, including SALL4, claudin-6, and glypican-3, which are useful for differentiating them from HCC [47,48]. GAED has a relatively favorable prognosis compared to HAC [45,49].

CHORIOCARCINOMA

Gastric choriocarcinoma exhibits adenocarcinomatous variable degrees of differentiation and germ cell elements with syncytiotrophoblasts and cytotrophoblasts (**Fig. 6A**) [50,51].

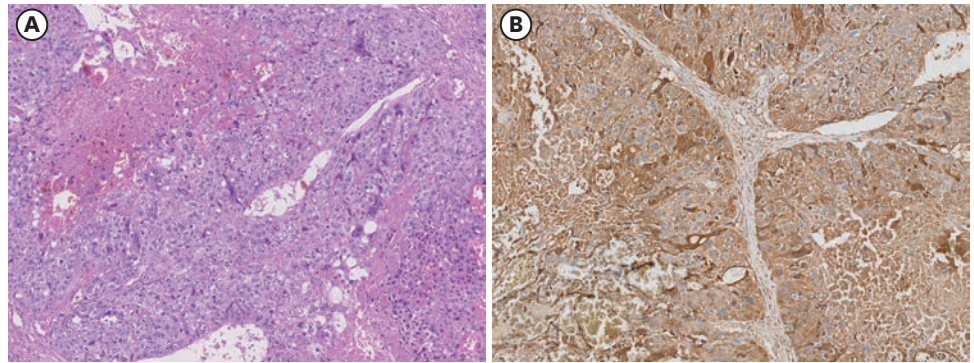


Fig. 6. (A) Gastric choriocarcinomas exhibit both adenocarcinomatous of variable degrees of differentiation and germ cell elements with syncytiotrophoblasts and cytotrophoblasts (hematoxylin and eosin, original magnification, $\times 100$). (B) Tumor cells are positive for beta-hCG. (beta-hCG, original magnification, $\times 100$). Case courtesy of Dr. Jihun Kim.
hCG = human chorionic gonadotropin.

The pathogenesis of gastric choriocarcinoma is thought to involve the choriocarcinomatous transformation of conventional adenocarcinoma [51]. Macroscopically, the lesion typically presents as an exophytic mass with significant necrosis and hemorrhage [52,53]. Immunohistochemistry often revealed human chorionic gonadotropin (hCG) expression within the trophoblastic components of the tumor (**Fig. 6B**), and patients typically exhibited markedly elevated circulating hCG levels. Serum hCG serves as both a prognostic indicator and a postoperative marker of potential tumor recurrence [52,54-56].

The management of gastric choriocarcinoma is challenging owing to its rarity, aggressive behavior, and complex histological presentation. With a high metastatic disease rate at the time of diagnosis, therapeutic approaches are often limited, and overall treatment outcomes remain poor. Their propensity to metastasize hematogenously often leads to hepatic failure due to extensive liver metastasis. Metastatic spread is rapid and widespread, significantly affecting prognosis, with most patients surviving for only a few months post-diagnosis [51,52,57].

Other germ cell tumor elements may present alongside embryonal carcinoma, yolk sac tumors, and hepatoid adenocarcinoma [58,59]. Only a few cases involving pure gastric yolk sac tumors have been documented [60-63].

GCLS

GCLS, also known as lymphoepithelioma-like carcinoma or medullary carcinoma, is an uncommon subtype, accounting for approximately 1%–8% of gastric cancers [64,65]. Typically, GCLS are histologically diverse, with patterns ranging from well-defined cellular trabeculae to less-organized syncytial structures (**Fig. 7A**). They are marked by dense lymphoid cell infiltration within the tumor architecture, mainly comprising cytotoxic CD8-positive T cells, although other immune cells contribute to dense infiltration [66,67].

Epstein-Barr virus (EBV) infection causes an interplay between carcinoma cells and immune elements within the gastric tissue. EBV positivity rates vary from 22.5% to 100% depending on the report, and it is worth noting that studies reporting rates as high as 80% or more used

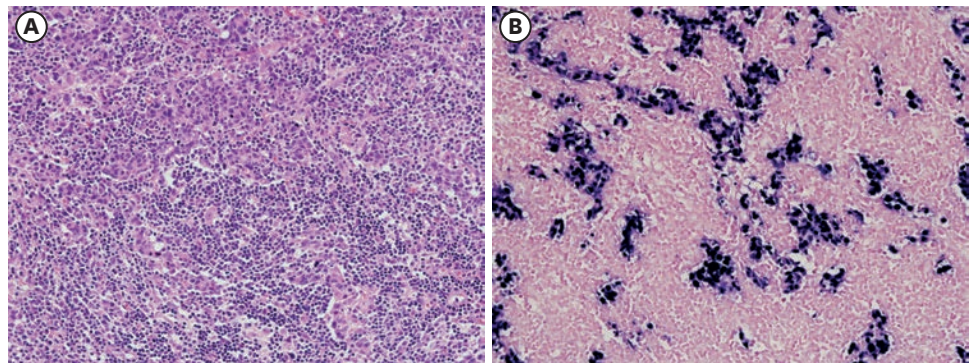


Fig. 7. (A) Gastric carcinoma with lymphoid stroma. Typically, these cases are histologically diverse, with a range of patterns from well-defined cellular trabeculae to less organized syncytial structures. They are marked by dense infiltration of lymphoid cells within the tumor architecture (hematoxylin and eosin, original magnification, $\times 200$). (B) The tumor cells, not the tumor-infiltrating lymphocytes, are positive in EBER *in situ* hybridization (EBER *in situ*, original magnification, $\times 200$).
EBER = Epstein-Barr virus-encoded small RNA.

EBV-encoded small RNA (EBER) (**Fig. 7B**) [64,68-72]. The role of EBV in GCLS carcinogenesis is multifaceted, with evidence suggesting that the virus contributes to carcinogenesis through a cascade of molecular events [73,74]. These include the induction of CpG island methylation and alterations in signaling pathways potentially mediated by viral gene products [75,76].

A subset of EBV-negative GCLS cases has been identified as microsatellite instability-high (MSI-H) [77]. MSI-H tumors are characterized by a significant number of alterations in DNA microsatellite sequences. This instability can arise because of defects in the DNA mismatch repair (MMR) system, which is responsible for correcting errors in DNA replication. This type of cancer can be detected by immunolabeling with MMR protein heterodimers (loss of MLH1/PMS2 vs. MSH2/MSH6 expression).

Predominantly occurring in the proximal stomach, GCLS demonstrates a male predominance and is often diagnosed at an earlier stage than other gastric cancers [75,78]. Early detection and the associated immune response may contribute to a comparatively improved prognosis in patients with GCLS [70,79].

The management of GCLS may benefit from the targeted application of checkpoint inhibitors due to the frequent overexpression of immunomodulatory molecules such as programmed cell death ligand 1/2 in these tumors [80-82], suggesting that GCLC is a possible immune checkpoint inhibitor and opens up a new treatment option.

CARCINOSARCOMA

Carcinosarcoma of the stomach is a biphasic malignancy that represents a distinct histological subtype characterized by the coexistence of adenocarcinoma and sarcoma components within the same tumor. These tumors exhibit striking heterogeneity in their histological composition, often incorporating sarcomatous elements. The sarcomatous component commonly displays spindle-cell morphology and frequently exhibits features of various types of sarcomas, including osteosarcoma, chondrosarcoma, rhabdomyosarcoma, and leiomyosarcoma [83-86]. Reported cases have demonstrated neuroendocrine or adenosquamous differentiation, underscoring the remarkable heterogeneity within these tumors [87-90].

Most gastric carcinosarcomas typically manifest as large polypoid or fungating masses and are associated with poor prognosis, partly due to their aggressive behavior and advanced stage at diagnosis in many cases [91].

It is crucial to distinguish true gastric carcinosarcomas from benign ossification or osteoid production, which occasionally occurs in gastric carcinomas [92]. The presence of mature bone or osteoids in the tumor stroma must be carefully evaluated to rule out true carcinosarcoma.

GASTROBLASTOMA

Gastroblastoma is a biphasic tumor that primarily affects children and young adults, with a male predominance. This unique neoplasm is composed of mixed spindle and epithelial cellular elements, both of which lack sufficient atypia to be diagnosed as carcinosarcoma [93-97].

Most commonly, these tumors are located in the antrum of the stomach and present as submucosal tumors centered in the proper muscle layer [96,97]. Clinical symptoms often include abdominal pain, epigastric discomfort, fatigue, or gastrointestinal bleeding [93-97]. Physical examination revealed a mass in the epigastric area.

A gastroblastoma hallmark is its biphasic histology, which is characterized by the presence of 2 distinct cellular components: spindle cells and nests of epithelial cells. Epithelial cells in gastroblastomas exhibit specific features, including scant pale cytoplasm, round nuclei, and inconspicuous nucleoli. The spindle cell component was monotonous, with long, slender cells often observed in the myxoid background (**Fig. 8**). Importantly, mitotic activity in these tumors is generally low; however, a case demonstrating up to 30 mitoses per 50 high-power fields has been reported [93].

A defining molecular feature is the presence of the MALAT1-GLI1 fusion gene [97,98]. This fusion gene drives GLI1 oncogenic properties and activates the Hh signaling pathway. Immunohistochemical staining revealed that the mesenchymal component of gastroblastoma was positive for CD10, CD56, and vimentin, whereas the epithelial component expressed CK markers (AE1/AE3 and CAM 5.2) and may show focal positivity for

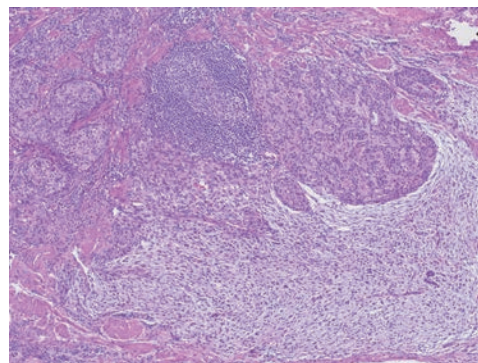


Fig. 8. The hallmark of gastroblastoma is its biphasic histology, characterized by the presence of two distinct cellular components - spindle cells and nests of epithelial cells. The epithelial cells in gastroblastomas exhibit specific features, including scant pale cytoplasm, round nuclei, and inconspicuous nucleoli. The spindle cell component is monotonous, with long and slender cells often observed in a myxoid background (hematoxylin and eosin, original magnification, $\times 10$). Case courtesy of Dr. Jihun Kim.

CD56 and CD10 [97,99]. Importantly, both components are strongly and diffusely positive for GLI1 immunolabeling [97].

Gastroblastomas are associated with a generally favorable prognosis following surgical resection, which is typically performed as a partial or subtotal gastrectomy. Although regional and distant lymph node metastases and local recurrences are rare, liver and lymph node metastases have been reported [96,97,99].

PARIETAL CELL CARCINOMA AND ONCOCYTIC ADENOCARCINOMA

Parietal cell carcinoma resembles acid-secreting parietal cells, which are normally observed in the gastric lining, with an eosinophilic granular cytoplasm. Histologically, parietal cell carcinomas exhibit an expanding growth pattern and are composed of solids and sheets of cells, a few of which may form small gland-like clefts [100,101]. The characteristic eosinophilic granular cytoplasm can be highlighted by special stains such as phosphotungstic acid-hematoxylin and Luxol fast blue. Furthermore, parietal cell carcinoma cells are positive for specific antibodies associated with parietal cells, including H⁺/K⁺ ATPase and human milk fat globule-2. The ultrastructural evaluation of these cells reveals the presence of numerous mitochondria and intracellular canaliculi [100,102].

Although exceedingly rare cases have been reported, it is assumed that parietal cell carcinoma has a more favorable prognosis than typical gastric carcinomas [101,103]. However, because available data are limited, further research is needed to establish a clear prognosis and potential therapeutic strategies.

Besides parietal cell carcinomas, oncocytic adenocarcinomas have been reported. These tumors share features with parietal cell carcinoma, in which both tumors have abundant eosinophilic and granular cytoplasm, numerous mitochondria in their cytoplasm, and occasional intracytoplasmic lumina with associated long microvilli [104]. However, these tumors do not express parietal cell-specific anti-H⁺/K⁺-ATPase antibodies, and the authors suggested the term oncocytic adenocarcinoma [104]. Caruso et al. [105] analyzed 9 gastric adenocarcinoma cases and demonstrated an abundance of eosinophilic cytoplasm. They proposed the term mitochondrion-rich carcinoma. They assumed that the tumors were similar to those of thyroid follicular carcinomas. The characteristics of these cases included small tumor size, infrequent lymph node metastasis, early-stage diagnosis, and favorable prognosis. The classification of parietal cell carcinomas, oncocytic adenocarcinomas, and mitochondria-rich carcinomas remains unclear [106]. More cases are required to clarify the terminology and classifications.

Notably, adenocarcinomas of the fundic-gland type, in which parietal cells predominate, are distinct from both parietal cell carcinomas and oncocytic carcinomas. These adenocarcinomas are typically small, well differentiated, and lack the solid sheet-like growth pattern observed in parietal cell carcinoma.

PANETH CELL CARCINOMA

Paneth cell carcinoma is characterized by the predominance of Paneth cells, demonstrating eosinophilic cytoplasmic granules that are positive for specific markers such as lysozyme and defensin-5 on immunohistochemistry [107,108].

Paneth cells, primarily located at the base crypt of the small intestine and proximal colon, secrete antimicrobial peptides, such as alpha defensin, lysozyme, and phospholipase A2, and play a significant role in defending against intestinal microbes and regulating host immunity [109]. Since Paneth cells may be present in areas of gastric intestinal metaplasia (IM), it can be assumed that Paneth cell carcinoma develops into a preneoplastic lesion with IM [110]. Paneth cell carcinomas are extremely rare, making our understanding of their clinical presentation limited. Clinical data on the effect of Paneth cell carcinomas on patient survival are scarce.

In a few cases, Paneth cells were dispersed in typical gastric adenocarcinomas [111]. This highlights the diversity of cell types encountered within gastric neoplasms and underscores the need for careful histological examination to identify and characterize these rare subtypes.

GASTRIC ADENOCARCINOMA OF FUNDIC-GLAND TYPE (GA-FG)

GA-FG is a well-differentiated neoplasm that develops from an oxyntic gland adenoma [112]. According to a single-institute surveillance study, GA-FG accounts for a small percentage (1%) of early gastric adenocarcinomas treated with endoscopic submucosal dissection [113]. Oxyntic gland differentiation, a tumor hallmark, can be further classified into 3 subcategories based on the tumor tissue composition: chief cell-predominant, parietal cell-predominant, and mixed chief and parietal cell types [112]. The chief cell-predominant type constitutes the majority of the reported cases, accounting for approximately 99% of them. The parietal cell-predominant type comprises parietal cells and another type of oxyntic gland cell. A mixed phenotype, containing a combination of chief and parietal cells, was observed within the neoplastic tissues (**Fig. 9**). In cases of the parietal cell-predominant type, differential diagnosis from parietal cell carcinoma is important. GA-FG can be distinguished by its differentiation, whereas parietal cell carcinoma is generally solid with polygonal cell sheets.

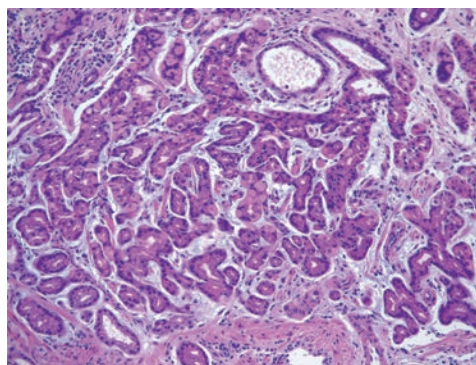


Fig. 9. Gastric adenocarcinoma of fundic-gland type. The tumor cells are composed of well-differentiated columnar cells with pale basophilic cytoplasm and mild nuclear atypia (hematoxylin and eosin, original magnification, $\times 200$).

Submucosal invasion was observed in a significant proportion (60%) of cases, emphasizing the need for careful histological evaluation. Nevertheless, most lesions appear in the early stages, and lymph node metastasis is rare [112].

These tumors are positive for specific markers including pepsinogen I and MUC6 [114]. Tumor cells exhibiting parietal cell differentiation are positive for H+/K+ ATPase immunolabeling [115].

UNDIFFERENTIATED CARCINOMA (INCLUDES SWI/SNF CHROMATIN-REMODELING COMPLEX MUTATION TYPE)

Undifferentiated carcinomas were characterized by the presence of anaplastic cells lacking specific cytological or architectural differentiation (**Fig. 10A**). These tumors are known for their heterogeneity and can resemble various other malignancies such as lymphomas, metastatic melanoma, germ cell neoplasms, and sarcomas. Undifferentiated carcinoma can manifest as several distinct histological variants, including rhabdoid carcinoma, sarcomatoid carcinoma, carcinoma with osteoclast-like giant cells, and pleomorphic giant cell carcinoma [116-122].

Immunohistochemical analyses often play a critical role in confirming epithelial phenotypes. Positive CK immunolabeling was observed at various intensities and proportions, whereas additional markers such as epithelial membrane antigen immunostaining can be useful, especially in cases with low CK expression. Vimentin, a mesenchymal marker, is consistently expressed in these tumors and often displays a perinuclear dot pattern [123,124].

A few undifferentiated carcinomas may contain areas with glandular components, suggesting that they may have originated through dedifferentiation from other gastric cancer types. The underlying genetic alterations in these tumors are unclear and may involve SWI/SNF chromatin-remodeling complex components. Loss of SMARCB1 (INI1), SMARCA4 (BRG1), and ARID1A, which are associated with chromatin regulation, has been reported in a few cases [123,124].

SMARCB1-deficient undifferentiated carcinoma, previously referred to as malignant rhabdoid tumor, is a recently studied entity in which loss of INI1 expression can be easily detected by

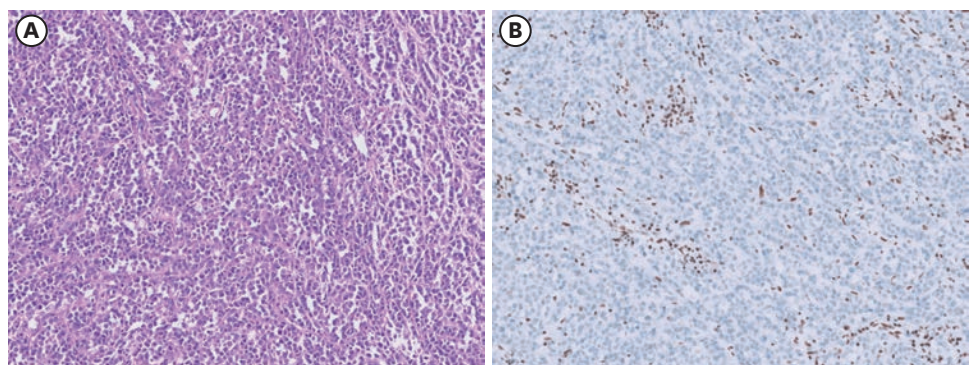


Fig. 10. (A) Undifferentiated carcinoma. The tumor lacks specific cytological or architectural differentiation (hematoxylin and eosin, original magnification, $\times 100$). (B) SMARCA4 mutant carcinoma. BRG1 immunohistochemistry is negative in the tumor cells (BRG1, original magnification, $\times 100$).

immunohistochemical staining [123]. This group of tumors has a rhabdoid phenotype and an extremely poor prognosis. SMARCA4-deficient neoplasia may present with rhabdoid features in minor cases, and the loss of BRG1 expression can be detected by immunohistochemical staining (**Fig. 10B**) [125].

Given the rarity of undifferentiated carcinoma and its highly aggressive nature, its specific prognostic features remain elusive. The disease stage at the time of diagnosis is believed to be prognostically relevant, with most patients presenting with advanced disease. Undifferentiated carcinomas are known for their rapid progression and many patients develop extensive regional metastases shortly after diagnosis and surgery. The prognosis for patients with undifferentiated stomach carcinoma is generally poor, with a high mortality rate within a year of diagnosis.

EXTREMELY WELL-DIFFERENTIATED ADENOCARCINOMA (EWDA)

EWDA is characterized by nonaggressive nuclear atypia and subtle architectural abnormalities, including tortuous, branching, anastomosing, distended glands, and lateral spreading characteristics of the neoplasm [126-129]. EWDA can be categorized into 2 types: intestinal and gastric. The intestinal type is composed of glands resembling intestinal metaplasia, containing goblet and Paneth cells [126,129]. In contrast, the gastric type features mucin-rich cells with basally located nuclei that appear benign, resembling either hyperplastic foveolar epithelium or expanded pyloric glands [130].

EWDA's innocuous appearance of EWDA often leads to its misinterpretation as regenerating atypia or inflammatory changes, particularly in endoscopic biopsy samples. Different studies have emphasized that the hallmark of EWDA is structural abnormalities rather than cytological atypia. Anastomosing glands, spiky glands, distended glands, discohesive cells, and disproportionately larger glands than the surrounding non-neoplastic glands [129,131].

Endoscopically, they often appear as ill-defined, slightly depressed, or flat lesions without significant mucosal color changes. While generally considered to have a favorable prognosis, there have been reports of certain cases evolving into a more aggressive diffuse type [128].

CONCLUSION

Gastric cancer presents a complex landscape with various histological subtypes, including papillary, squamous, AFP-producing, eosinophilic, and rhabdoid. This review highlights the distinct characteristics, clinical presentations, and challenges associated with the diagnosis and management of various unusual and uncommon gastric cancer forms. These include papillary adenocarcinoma, micropapillary carcinoma, adenosquamous carcinoma, SCC, hepatoid adenocarcinoma, gastric choriocarcinoma, GCLS, carcinosarcoma, gastroblastoma, parietal cell carcinoma, oncocytic adenocarcinoma, Paneth cell carcinoma, GA-FG, undifferentiated carcinoma, and EWDA.

Each subtype presents unique diagnostic challenges and pathological features requiring careful evaluation. Understanding these rare variants is crucial for an accurate diagnosis,

establishing treatment strategies, and improving patient outcomes. Future research and case studies will play an important role in deepening our understanding of these rare cancers, leading to more effective treatments and improved prognostic assessments. This review underscores the importance of recognizing the diversity of the histological characteristics of gastric cancer, emphasizing that these types of cancers may present unique features that impact their care and prognosis.

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REFERENCES

1. Dawson H, Novotny A, Becker K, Reim D, Langer R, Gullo I, et al. Macroscopy predicts tumor progression in gastric cancer: a retrospective patho-historical analysis based on Napoleon Bonaparte's autopsy report. *Dig Liver Dis* 2016;48:1378-1385.
[PUBMED](#) | [CROSSREF](#)
2. Park SH, Kang MJ, Yun EH, Jung KW. Epidemiology of gastric cancer in Korea: trends in incidence and survival based on Korea Central Cancer Registry Data (1999-2019). *J Gastric Cancer* 2022;22:160-168.
[PUBMED](#) | [CROSSREF](#)
3. Adachi Y, Yasuda K, Inomata M, Sato K, Shiraishi N, Kitano S. Pathology and prognosis of gastric carcinoma: well versus poorly differentiated type. *Cancer* 2000;89:1418-1424.
[PUBMED](#) | [CROSSREF](#)
4. Yu H, Fang C, Chen L, Shi J, Fan X, Zou X, et al. Worse prognosis in papillary, compared to tubular, early gastric carcinoma. *J Cancer* 2017;8:117-123.
[PUBMED](#) | [CROSSREF](#)
5. Yasuda K, Adachi Y, Shiraishi N, Maeo S, Kitano S. Papillary adenocarcinoma of the stomach. *Gastric Cancer* 2000;3:33-38.
[PUBMED](#) | [CROSSREF](#)
6. Ghotli ZA, Serra S, Chetty R. Clear cell (glycogen rich) gastric adenocarcinoma: a distinct tubulo-papillary variant with a predilection for the cardia/gastro-oesophageal region. *Pathology* 2007;39:466-469.
[PUBMED](#) | [CROSSREF](#)
7. Carr N. Tubulopapillary clear cell carcinoma of the stomach may be a type of pylorocardiac carcinoma. *Pathology* 2008;40:333.
[PUBMED](#) | [CROSSREF](#)
8. Matsunou H, Konishi F, Jalal RE, Yamamichi N, Mukawa A. Alpha-fetoprotein-producing gastric carcinoma with enteroblastic differentiation. *Cancer* 1994;73:534-540.
[PUBMED](#) | [CROSSREF](#)
9. Roh JH, Srivastava A, Lauwers GY, An J, Jang KT, Park CK, et al. Micropapillary carcinoma of stomach: a clinicopathologic and immunohistochemical study of 11 cases. *Am J Surg Pathol* 2010;34:1139-1146.
[PUBMED](#) | [CROSSREF](#)
10. Eom DW, Kang GH, Han SH, Cheon GJ, Han KH, Oh HS, et al. Gastric micropapillary carcinoma: a distinct subtype with a significantly worse prognosis in TNM stages I and II. *Am J Surg Pathol* 2011;35:84-91.
[PUBMED](#) | [CROSSREF](#)
11. Hattori T, Sentani K, Hattori Y, Oue N, Yasui W. Pure invasive micropapillary carcinoma of the esophagogastric junction with lymph nodes and liver metastasis. *Pathol Int* 2016;66:583-586.
[PUBMED](#) | [CROSSREF](#)
12. Akce M, Jiang R, Alese OB, Shaib WL, Wu C, Behera M, et al. Gastric squamous cell carcinoma and gastric adenosquamous carcinoma, clinical features and outcomes of rare clinical entities: a National Cancer Database (NCDB) analysis. *J Gastrointest Oncol* 2019;10:85-94.
[PUBMED](#) | [CROSSREF](#)

13. Chen YY, Li AF, Huang KH, Lan YT, Chen MH, Chao Y, et al. Adenosquamous carcinoma of the stomach and review of the literature. *Pathol Oncol Res* 2015;21:547-551.
[PUBMED](#) | [CROSSREF](#)
14. Saito S, Hosoya Y, Morishima K, Ui T, Haruta H, Kurashina K, et al. A clinicopathological and immunohistochemical study of gastric cancer with squamous cell carcinoma components: a clinically aggressive tumor. *J Dig Dis* 2012;13:407-413.
[PUBMED](#) | [CROSSREF](#)
15. Feng F, Zheng G, Qi J, Xu G, Wang F, Wang Q, et al. Clinicopathological features and prognosis of gastric adenosquamous carcinoma. *Sci Rep* 2017;7:4597.
[PUBMED](#) | [CROSSREF](#)
16. Mori M, Iwashita A, Enjoji M. Adenosquamous carcinoma of the stomach. A clinicopathologic analysis of 28 cases. *Cancer* 1986;57:333-339.
[PUBMED](#) | [CROSSREF](#)
17. Mori M, Fukuda T, Enjoji M. Adenosquamous carcinoma of the stomach. Histogenetic and ultrastructural studies. *Gastroenterology* 1987;92:1078-1082.
[PUBMED](#) | [CROSSREF](#)
18. Chen H, Shen C, Yin R, Yin Y, Chen J, Han L, et al. Clinicopathological characteristics, diagnosis, treatment, and outcomes of primary gastric adenosquamous carcinoma. *World J Surg Oncol* 2015;13:136.
[PUBMED](#) | [CROSSREF](#)
19. Aoki Y, Tabuse K, Wada M, Katsumi M, Uda H. Primary adenosquamous carcinoma of the stomach: experience of 11 cases and its clinical analysis. *Gastroenterol Jpn* 1978;13:140-145.
[PUBMED](#) | [CROSSREF](#)
20. Gao S, Chen D, Huang L, Dai R, Shan Y. Primary squamous cell carcinoma of the stomach: a case report and literature review. *Int J Clin Exp Pathol* 2015;8:9667-9671.
[PUBMED](#)
21. Boswell JT, Helwig EB. Squamous cell carcinoma and adenoacanthoma of the stomach. A clinicopathologic study. *Cancer* 1965;18:181-192.
[PUBMED](#) | [CROSSREF](#)
22. Bonnheim DC, Sarac OK, Fett W. Primary squamous cell carcinoma of the stomach. *Am J Gastroenterol* 1985;80:91-94.
[PUBMED](#)
23. Muto M, Hasebe T, Muro K, Boku N, Ohtsu A, Fujii T, et al. Primary squamous cell carcinoma of the stomach: a case report with a review of Japanese and Western literature. *Hepatogastroenterology* 1999;46:3015-3018.
[PUBMED](#)
24. Wakabayashi H, Matsutani T, Fujita I, Kanazawa Y, Nomura T, Hagiwara N, et al. A rare case of primary squamous cell carcinoma of the stomach and a review of the 56 cases reported in Japan. *J Gastric Cancer* 2014;14:58-62.
[PUBMED](#) | [CROSSREF](#)
25. González-Sánchez JA, Vitón R, Collantes E, Rodríguez-Montes JA. Primary squamous cell carcinoma of the stomach. *Clin Med Insights Oncol* 2017;11:1179554916686076.
[PUBMED](#) | [CROSSREF](#)
26. Oono Y, Fu K, Nagahisa E, Kuwata T, Ikematsu H, Yano T, et al. Primary gastric squamous cell carcinoma in situ originating from gastric squamous metaplasia. *Endoscopy* 2010;42 Suppl 2:E290-E291.
[PUBMED](#) | [CROSSREF](#)
27. Ahn S, Bae GE, Kim KM. Exuberant squamous metaplasia of the gastric mucosa in a patient with gastric adenocarcinoma. *Diagn Pathol* 2015;10:46.
[PUBMED](#) | [CROSSREF](#)
28. Meng Y, Zhang J, Wang H, Zhang Y, Sun R, Zhang Z, et al. Poorer prognosis in patients with advanced gastric squamous cell carcinoma compared with adenocarcinoma of the stomach: case report. *Medicine (Baltimore)* 2017;96:e9224.
[PUBMED](#) | [CROSSREF](#)
29. Marubashi S, Yano H, Monden T, Tateishi H, Kanoh T, Iwazawa T, et al. Primary squamous cell carcinoma of the stomach. *Gastric Cancer* 1999;2:136-141.
[PUBMED](#) | [CROSSREF](#)
30. Inagawa S, Shimazaki J, Hori M, Yoshimi F, Adachi S, Kawamoto T, et al. Hepatoid adenocarcinoma of the stomach. *Gastric Cancer* 2001;4:43-52.
[PUBMED](#) | [CROSSREF](#)

31. Ishikura H, Kirimoto K, Shamoto M, Miyamoto Y, Yamagiwa H, Itoh T, et al. Hepatoid adenocarcinomas of the stomach. An analysis of seven cases. *Cancer* 1986;58:119-126.
[PUBMED](#) | [CROSSREF](#)
32. Nagai E, Ueyama T, Yao T, Tsuneyoshi M. Hepatoid adenocarcinoma of the stomach. A clinicopathologic and immunohistochemical analysis. *Cancer* 1993;72:1827-1835.
[PUBMED](#) | [CROSSREF](#)
33. Petrella T, Montagnon J, Roignot P, Van Nieuvanhuysse A, Matagrín C, Michiels-Marzais D, et al. Alphafetoprotein-producing gastric adenocarcinoma. *Histopathology* 1995;26:171-175.
[PUBMED](#) | [CROSSREF](#)
34. Baek SK, Han SW, Oh DY, Im SA, Kim TY, Bang YJ. Clinicopathologic characteristics and treatment outcomes of hepatoid adenocarcinoma of the stomach, a rare but unique subtype of gastric cancer. *BMC Gastroenterol* 2011;11:56.
[PUBMED](#) | [CROSSREF](#)
35. Xiao C, Wu F, Jiang H, Teng L, Song F, Wang Q, et al. Hepatoid adenocarcinoma of the stomach: nine case reports and treatment outcomes. *Oncol Lett* 2015;10:1605-1609.
[PUBMED](#) | [CROSSREF](#)
36. Chang YC, Nagasue N, Kohno H, Taniura H, Uchida M, Yamanoi A, et al. Clinicopathologic features and long-term results of alpha-fetoprotein-producing gastric cancer. *Am J Gastroenterol* 1990;85:1480-1485.
[PUBMED](#)
37. Fakhruddin N, Bahmad HF, Aridi T, Yammine Y, Mahfouz R, Boulos F, et al. Hepatoid adenocarcinoma of the stomach: a challenging diagnostic and therapeutic disease through a case report and review of the literature. *Front Med (Lausanne)* 2017;4:164.
[PUBMED](#) | [CROSSREF](#)
38. Terracciano LM, Glatz K, Mhawech P, Vasei M, Lehmann FS, Vecchione R, et al. Hepatoid adenocarcinoma with liver metastasis mimicking hepatocellular carcinoma: an immunohistochemical and molecular study of eight cases. *Am J Surg Pathol* 2003;27:1302-1312.
[PUBMED](#) | [CROSSREF](#)
39. Van Eyken P, Sciort R, Paterson A, Callea F, Kew MC, Desmet VJ. Cytokeratin expression in hepatocellular carcinoma: an immunohistochemical study. *Hum Pathol* 1988;19:562-568.
[PUBMED](#) | [CROSSREF](#)
40. Maeda T, Kajiyama K, Adachi E, Takenaka K, Sugimachi K, Tsuneyoshi M. The expression of cytokeratins 7, 19, and 20 in primary and metastatic carcinomas of the liver. *Mod Pathol* 1996;9:901-909.
[PUBMED](#)
41. Tickoo SK, Zee SY, Obiekwe S, Xiao H, Koea J, Robiou C, et al. Combined hepatocellular-cholangiocarcinoma: a histopathologic, immunohistochemical, and in situ hybridization study. *Am J Surg Pathol* 2002;26:989-997.
[PUBMED](#) | [CROSSREF](#)
42. Chang YC, Nagasue N, Abe S, Taniura H, Kumar DD, Nakamura T. Comparison between the clinicopathologic features of AFP-positive and AFP-negative gastric cancers. *Am J Gastroenterol* 1992;87:321-325.
[PUBMED](#)
43. Yamazawa S, Ushiku T, Shinozaki-Ushiku A, Hayashi A, Iwasaki A, Abe H, et al. Gastric cancer with primitive enterocyte phenotype: an aggressive subgroup of intestinal-type adenocarcinoma. *Am J Surg Pathol* 2017;41:989-997.
[PUBMED](#) | [CROSSREF](#)
44. Motoyama T, Aizawa K, Watanabe H, Fukase M, Saito K. alpha-Fetoprotein producing gastric carcinomas: a comparative study of three different subtypes. *Acta Pathol Jpn* 1993;43:654-661.
[PUBMED](#)
45. Kinjo T, Taniguchi H, Kushima R, Sekine S, Oda I, Saka M, et al. Histologic and immunohistochemical analyses of α -fetoprotein--producing cancer of the stomach. *Am J Surg Pathol* 2012;36:56-65.
[PUBMED](#) | [CROSSREF](#)
46. Akazawa Y, Saito T, Hayashi T, Yanai Y, Tsuyama S, Akaike K, et al. Next-generation sequencing analysis for gastric adenocarcinoma with enteroblastic differentiation: emphasis on the relationship with hepatoid adenocarcinoma. *Hum Pathol* 2018;78:79-88.
[PUBMED](#) | [CROSSREF](#)
47. Murakami T, Yao T, Mitomi H, Morimoto T, Ueyama H, Matsumoto K, et al. Clinicopathologic and immunohistochemical characteristics of gastric adenocarcinoma with enteroblastic differentiation: a study of 29 cases. *Gastric Cancer* 2016;19:498-507.
[PUBMED](#) | [CROSSREF](#)

48. Ushiku T, Shinozaki A, Shibahara J, Iwasaki Y, Tateishi Y, Funata N, et al. SALL4 represents fetal gut differentiation of gastric cancer, and is diagnostically useful in distinguishing hepatoid gastric carcinoma from hepatocellular carcinoma. *Am J Surg Pathol* 2010;34:533-540.
[PUBMED](#) | [CROSSREF](#)
49. Kwon MJ, Byeon S, Kang SY, Kim KM. Gastric adenocarcinoma with enteroblastic differentiation should be differentiated from hepatoid adenocarcinoma: a study with emphasis on clear cells and clinicopathologic spectrum. *Pathol Res Pract* 2019;215:152525.
[PUBMED](#) | [CROSSREF](#)
50. Wurzel J, Brooks JJ. Primary gastric choriocarcinoma: immunohistochemistry, postmortem documentation, and hormonal effects in a postmenopausal female. *Cancer* 1981;48:2756-2761.
[PUBMED](#) | [CROSSREF](#)
51. Kobayashi A, Hasebe T, Endo Y, Sasaki S, Konishi M, Sugito M, et al. Primary gastric choriocarcinoma: two case reports and a pooled analysis of 53 cases. *Gastric Cancer* 2005;8:178-185.
[PUBMED](#) | [CROSSREF](#)
52. Saigo PE, Brigati DJ, Sternberg SS, Rosen PP, Turnbull AD. Primary gastric choriocarcinoma. An immunohistological study. *Am J Surg Pathol* 1981;5:333-342.
[PUBMED](#) | [CROSSREF](#)
53. Imai Y, Kawabe T, Takahashi M, Matsumura M, Komatsu Y, Hamada E, et al. A case of primary gastric choriocarcinoma and a review of the Japanese literature. *J Gastroenterol* 1994;29:642-646.
[PUBMED](#) | [CROSSREF](#)
54. Liu AY, Chan WY, Ng EK, Zhang X, Li BC, Chow JH, et al. Gastric choriocarcinoma shows characteristics of adenocarcinoma and gestational choriocarcinoma: a comparative genomic hybridization and fluorescence in situ hybridization study. *Diagn Mol Pathol* 2001;10:161-165.
[PUBMED](#) | [CROSSREF](#)
55. Smith FR, Barkin JS, Hensley G. Choriocarcinoma of the stomach. *Am J Gastroenterol* 1980;73:45-48.
[PUBMED](#)
56. Noguchi T, Takeno S, Sato T, Takahashi Y, Uchida Y, Yokoyama S. A patient with primary gastric choriocarcinoma who received a correct preoperative diagnosis and achieved prolonged survival. *Gastric Cancer* 2002;5:112-117.
[PUBMED](#) | [CROSSREF](#)
57. Jindrak K, Bochetto JF, Alpert LI. Primary gastric choriocarcinoma: case report with review of world literature. *Hum Pathol* 1976;7:595-604.
[PUBMED](#) | [CROSSREF](#)
58. Satake N, Chikakiyo M, Yagi T, Suzuki Y, Hirose T. Gastric cancer with choriocarcinoma and yolk sac tumor components: case report. *Pathol Int* 2011;61:156-160.
[PUBMED](#) | [CROSSREF](#)
59. Wook Hwang S, Jae Lee S, Eun Park P, Seon Kim M, Bae HI. Gastric adenocarcinoma with choriocarcinomatous and hepatoid differentiation. *Basic Appl Pathol* 2012;5:22-25.
[CROSSREF](#)
60. Kim YS, Kim SH, Seong JK, Lee BS, Jeong HY, Song KS. Gastric yolk sac tumor: a case report and review of the literature. *Korean J Intern Med* 2009;24:143-146.
[PUBMED](#) | [CROSSREF](#)
61. Kanai M, Torii A, Hamada A, Endo Y, Takeda Y, Yamakawa M, et al. Pure gastric yolk sac tumor that was diagnosed after curative resection: case report and review of literature. *Int J Gastrointest Cancer* 2005;35:77-81.
[PUBMED](#) | [CROSSREF](#)
62. Ibrahim A, MacDermid E, Nguyen HP, Ashrafy AH. Massive pure gastric yolk sac tumour: a unique presentation of a rare pathology. *ANZ J Surg* 2019;89:E417-E419.
[PUBMED](#) | [CROSSREF](#)
63. Magni E, Sonzogni A, Zampino MG. Primary pure gastric yolk sac tumor. *Rare Tumors* 2010;2:e10.
[PUBMED](#) | [CROSSREF](#)
64. Hissong E, Ramrattan G, Zhang P, Zhou XK, Young G, Klimstra DS, et al. Gastric carcinomas with lymphoid stroma: an evaluation of the histopathologic and molecular features. *Am J Surg Pathol* 2018;42:453-462.
[PUBMED](#) | [CROSSREF](#)
65. Watanabe H, Enjoji M, Imai T. Gastric carcinoma with lymphoid stroma. Its morphologic characteristics and prognostic correlations. *Cancer* 1976;38:232-243.
[PUBMED](#) | [CROSSREF](#)
66. Camargo MC, Murphy G, Koriyama C, Pfeiffer RM, Kim WH, Herrera-Goepfert R, et al. Determinants of Epstein-Barr virus-positive gastric cancer: an international pooled analysis. *Br J Cancer* 2011;105:38-43.
[PUBMED](#) | [CROSSREF](#)

67. Gullo I, Oliveira P, Athelougou M, Gonçalves G, Pinto ML, Carvalho J, et al. New insights into the inflamed tumor immune microenvironment of gastric cancer with lymphoid stroma: from morphology and digital analysis to gene expression. *Gastric Cancer* 2019;22:77-90.
[PUBMED](#) | [CROSSREF](#)
68. Murphy G, Pfeiffer R, Camargo MC, Rabkin CS. Meta-analysis shows that prevalence of Epstein-Barr virus-positive gastric cancer differs based on sex and anatomic location. *Gastroenterology* 2009;137:824-833.
[PUBMED](#) | [CROSSREF](#)
69. Chang MS, Lee HS, Kim CW, Kim YI, Kim WH. Clinicopathologic characteristics of Epstein-Barr virus-incorporated gastric cancers in Korea. *Pathol Res Pract* 2001;197:395-400.
[PUBMED](#) | [CROSSREF](#)
70. Wang HH, Wu MS, Shun CT, Wang HP, Lin CC, Lin JT. Lymphoepithelioma-like carcinoma of the stomach: a subset of gastric carcinoma with distinct clinicopathological features and high prevalence of Epstein-Barr virus infection. *Hepatogastroenterology* 1999;46:1214-1219.
[PUBMED](#)
71. Abe H, Saito R, Ichimura T, Iwasaki A, Yamazawa S, Shinozaki-Ushiku A, et al. CD47 expression in Epstein-Barr virus-associated gastric carcinoma: coexistence with tumor immunity lowering the ratio of CD8⁺/Foxp3⁺ T cells. *Virchows Arch* 2018;472:643-651.
[PUBMED](#) | [CROSSREF](#)
72. Ichimura T, Abe H, Morikawa T, Yamashita H, Ishikawa S, Ushiku T, et al. Low density of CD204-positive M2-type tumor-associated macrophages in Epstein-Barr virus-associated gastric cancer: a clinicopathologic study with digital image analysis. *Hum Pathol* 2016;56:74-80.
[PUBMED](#) | [CROSSREF](#)
73. Fukayama M, Chong JM, Kaizaki Y. Epstein-Barr virus and gastric carcinoma. *Gastric Cancer* 1998;1:104-114.
[PUBMED](#) | [CROSSREF](#)
74. Shibata D, Weiss LM. Epstein-Barr virus-associated gastric adenocarcinoma. *Am J Pathol* 1992;140:769-774.
[PUBMED](#)
75. Lee JH, Kim SH, Han SH, An JS, Lee ES, Kim YS. Clinicopathological and molecular characteristics of Epstein-Barr virus-associated gastric carcinoma: a meta-analysis. *J Gastroenterol Hepatol* 2009;24:354-365.
[PUBMED](#) | [CROSSREF](#)
76. Matsunou H, Konishi F, Hori H, Ikeda T, Sasaki K, Hirose Y, et al. Characteristics of Epstein-Barr virus-associated gastric carcinoma with lymphoid stroma in Japan. *Cancer* 1996;77:1998-2004.
[PUBMED](#) | [CROSSREF](#)
77. Gullo I, Carvalho J, Martins D, Lemos D, Monteiro AR, Ferreira M, et al. The transcriptomic landscape of gastric cancer: insights into Epstein-Barr virus infected and microsatellite unstable tumors. *Int J Mol Sci* 2018;19:2079.
[PUBMED](#) | [CROSSREF](#)
78. Huang SC, Ng KF, Chen KH, Hsu JT, Liu KH, Yeh TS, et al. Prognostic factors in Epstein-Barr virus-associated stage I-III gastric carcinoma: implications for a unique type of carcinogenesis. *Oncol Rep* 2014;32:530-538.
[PUBMED](#) | [CROSSREF](#)
79. Minamoto T, Mai M, Watanabe K, Ooi A, Kitamura T, Takahashi Y, et al. Medullary carcinoma with lymphocytic infiltration of the stomach. Clinicopathologic study of 27 cases and immunohistochemical analysis of the subpopulations of infiltrating lymphocytes in the tumor. *Cancer* 1990;66:945-952.
[PUBMED](#) | [CROSSREF](#)
80. Derks S, Liao X, Chiaravalli AM, Xu X, Camargo MC, Solcia E, et al. Abundant PD-L1 expression in Epstein-Barr Virus-infected gastric cancers. *Oncotarget* 2016;7:32925-32932.
[PUBMED](#) | [CROSSREF](#)
81. Ma C, Patel K, Singhi AD, Ren B, Zhu B, Shaikh F, et al. Programmed death-ligand 1 expression is common in gastric cancer associated with Epstein-Barr virus or microsatellite instability. *Am J Surg Pathol* 2016;40:1496-1506.
[PUBMED](#) | [CROSSREF](#)
82. Saito R, Abe H, Kunita A, Yamashita H, Seto Y, Fukayama M. Overexpression and gene amplification of PD-L1 in cancer cells and PD-L1⁺ immune cells in Epstein-Barr virus-associated gastric cancer: the prognostic implications. *Mod Pathol* 2017;30:427-439.
[PUBMED](#) | [CROSSREF](#)
83. Cho KJ, Myong NH, Choi DW, Jang JJ. Carcinosarcoma of the stomach. A case report with light microscopic, immunohistochemical, and electron microscopic study. *Acta Pathol Microbiol Scand Suppl* 1990;98:991-995.
[PUBMED](#) | [CROSSREF](#)

84. Nakayama Y, Murayama H, Iwasaki H, Iwanaga S, Kikuchi M, Ikeda S, et al. Gastric carcinosarcoma (sarcomatoid carcinoma) with rhabdomyoblastic and osteoblastic differentiation. *Pathol Int* 1997;47:557-563.
[PUBMED](#) | [CROSSREF](#)
85. Sato Y, Shimozono T, Kawano S, Toyoda K, Onoe K, Asada Y, et al. Gastric carcinosarcoma, coexistence of adenosquamous carcinoma and rhabdomyosarcoma: a case report. *Histopathology* 2001;39:543-544.
[PUBMED](#) | [CROSSREF](#)
86. Randjelovic T, Filipovic B, Babic D, Cemerikic V, Filipovic B. Carcinosarcoma of the stomach: a case report and review of the literature. *World J Gastroenterol* 2007;13:5533-5536.
[PUBMED](#) | [CROSSREF](#)
87. Tsuneyama K, Sasaki M, Sabit A, Yokoi K, Arano Y, Imai T, et al. A case report of gastric carcinosarcoma with rhabdomyosarcomatous and neuroendocrinal differentiation. *Pathol Res Pract* 1999;195:93-97.
[PUBMED](#) | [CROSSREF](#)
88. Yamazaki K. A gastric carcinosarcoma with neuroendocrine cell differentiation and undifferentiated spindle-shaped sarcoma component possibly progressing from the conventional tubular adenocarcinoma; an immunohistochemical and ultrastructural study. *Virchows Arch* 2003;442:77-81.
[PUBMED](#) | [CROSSREF](#)
89. Teramachi K, Kanomata N, Hasebe T, Ishii G, Sugito M, Ochiai A. Carcinosarcoma (pure endocrine cell carcinoma with sarcoma components) of the stomach. *Pathol Int* 2003;53:552-556.
[PUBMED](#) | [CROSSREF](#)
90. Kuroda N, Oonishi K, Iwamura S, Ohara M, Hirouchi T, Mizumo K, et al. Gastric carcinosarcoma with neuroendocrine differentiation as the carcinoma component and leiomyosarcomatous and myofibroblastic differentiation as the sarcomatous component. *Acta Pathol Microbiol Scand Suppl* 2006;114:234-238.
[PUBMED](#) | [CROSSREF](#)
91. Ikeda Y, Kosugi S, Nishikura K, Ohashi M, Kanda T, Kobayashi T, et al. Gastric carcinosarcoma presenting as a huge epigastric mass. *Gastric Cancer* 2007;10:63-68.
[PUBMED](#) | [CROSSREF](#)
92. Kypson AP, Morpew E, Jones R, Gottfried MR, Seigler HF. Heterotopic ossification in rectal cancer: rare finding with a novel proposed mechanism. *J Surg Oncol* 2003;82:132-136.
[PUBMED](#) | [CROSSREF](#)
93. Miettinen M, Dow N, Lasota J, Sobin LH. A distinctive novel epitheliomesenchymal biphasic tumor of the stomach in young adults ("gastroblastoma"): a series of 3 cases. *Am J Surg Pathol* 2009;33:1370-1377.
[PUBMED](#) | [CROSSREF](#)
94. Shin DH, Lee JH, Kang HJ, Choi KU, Kim JY, Park DY, et al. Novel epitheliomesenchymal biphasic stomach tumour (gastroblastoma) in a 9-year-old: morphological, ultrastructural and immunohistochemical findings. *J Clin Pathol* 2010;63:270-274.
[PUBMED](#) | [CROSSREF](#)
95. Centonze G, Mangogna A, Salviato T, Belmonte B, Cattaneo L, Monica MA, et al. Gastroblastoma in adulthood-a rarity among rare cancers-a case report and review of the literature. *Case Rep Pathol* 2019;2019:4084196.
[PUBMED](#) | [CROSSREF](#)
96. Toumi O, Ammar H, Korbi I, Ayed M, Gupta R, Nasr M, et al. Gastroblastoma, a biphasic neoplasm of stomach: a case report. *Int J Surg Case Rep* 2017;39:72-76.
[PUBMED](#) | [CROSSREF](#)
97. Graham RP, Nair AA, Davila JI, Jin L, Jen J, Sukov WR, et al. Gastroblastoma harbors a recurrent somatic MALAT1-GLI1 fusion gene. *Mod Pathol* 2017;30:1443-1452.
[PUBMED](#) | [CROSSREF](#)
98. Chetty R. Gene of the month: GLI-1. *J Clin Pathol* 2020;73:228-230.
[PUBMED](#) | [CROSSREF](#)
99. Wey EA, Britton AJ, Sferra JJ, Kasunic T, Pepe LR, Appelman HD. Gastroblastoma in a 28-year-old man with nodal metastasis: proof of the malignant potential. *Arch Pathol Lab Med* 2012;136:961-964.
[PUBMED](#) | [CROSSREF](#)
100. Capella C, Frigerio B, Cornaggia M, Solcia E, Pinzon-Trujillo Y, Chejfec G. Gastric parietal cell carcinoma-a newly recognized entity: light microscopic and ultrastructural features. *Histopathology* 1984;8:813-824.
[PUBMED](#) | [CROSSREF](#)
101. Mavroeidis VK, Gkegkes ID, Saffioti F, Kandilaris K, Alexiou K, Horti M, et al. Parietal cell/oncocytic gastric carcinoma: systematic review and first-time assessment of HER2 status in two new cases. *Ann R Coll Surg Engl* 2020;102:300-307.
[PUBMED](#) | [CROSSREF](#)

102. Yang GY, Liao J, Cassai ND, Smolka AJ, Sidhu GS. Parietal cell carcinoma of gastric cardia: immunophenotype and ultrastructure. *Ultrastruct Pathol* 2003;27:87-94.
[PUBMED](#) | [CROSSREF](#)
103. Byrne D, Holley MP, Cuschieri A. Parietal cell carcinoma of the stomach: association with long-term survival after curative resection. *Br J Cancer* 1988;58:85-87.
[PUBMED](#) | [CROSSREF](#)
104. Takubo K, Honma N, Sawabe M, Arai T, Izumiyama-Shimomura N, Kammori M, et al. Oncocytic adenocarcinoma of the stomach: parietal cell carcinoma. *Am J Surg Pathol* 2002;26:458-465.
[PUBMED](#) | [CROSSREF](#)
105. Caruso RA, Napoli P, Nania A, Parisi A, Fedele F, Zuccalà V. Mitochondrion-rich differentiated adenocarcinomas of the stomach: clinicopathological, immunohistochemical and electron microscopy study of nine cases. *Virchows Arch* 2010;456:499-505.
[PUBMED](#) | [CROSSREF](#)
106. Mavroeidis VK, Saffioti F. Parietal cell/oncocytic/mitochondrion-rich gastric carcinoma: a need for clarity in terminology. *Pathology* 2021;53:278-279.
[PUBMED](#) | [CROSSREF](#)
107. Kazzaz BA, Eulderink F. Paneth cell-rich carcinoma of the stomach. *Histopathology* 1989;15:303-305.
[PUBMED](#) | [CROSSREF](#)
108. Inada KI, Mizoshita T, Tsukamoto T, Porter EM, Tatematsu M. Paneth type gastric cancer cells exhibit expression of human defensin-5. *Histopathology* 2005;47:330-331.
[PUBMED](#) | [CROSSREF](#)
109. Nakamura K, Sakuragi N, Takakuwa A, Ayabe T. Paneth cell α -defensins and enteric microbiota in health and disease. *Biosci Microbiota Food Health* 2016;35:57-67.
[PUBMED](#) | [CROSSREF](#)
110. Luo W, Hofstetter WL, Tan D. Gastroesophageal junction Paneth cell carcinoma with extensive cystic and secretory features - case report and literature review. *Diagn Pathol* 2019;14:1.
[PUBMED](#) | [CROSSREF](#)
111. Ooi A, Nakanishi I, Itoh T, Ueda H, Mai M. Predominant Paneth cell differentiation in an intestinal type gastric cancer. *Pathol Res Pract* 1991;187:220-225.
[PUBMED](#) | [CROSSREF](#)
112. Benedict MA, Lauwers GY, Jain D. Gastric adenocarcinoma of the fundic gland type: update and literature review. *Am J Clin Pathol* 2018;149:461-473.
[PUBMED](#) | [CROSSREF](#)
113. Miyazawa M, Matsuda M, Yano M, Hara Y, Arihara F, Horita Y, et al. Gastric adenocarcinoma of fundic gland type: five cases treated with endoscopic resection. *World J Gastroenterol* 2015;21:8208-8214.
[PUBMED](#) | [CROSSREF](#)
114. Ota H, Yamaguchi D, Iwaya M, Kobayashi M, Tateiwa N, Iwaya Y, et al. Principal cells in gastric neoplasia of fundic gland (chief cell predominant) type show characteristics of immature chief cells. *Pathol Int* 2015;65:202-204.
[PUBMED](#) | [CROSSREF](#)
115. Ueyama H, Yao T, Nakashima Y, Hirakawa K, Oshiro Y, Hirahashi M, et al. Gastric adenocarcinoma of fundic gland type (chief cell predominant type): proposal for a new entity of gastric adenocarcinoma. *Am J Surg Pathol* 2010;34:609-619.
[PUBMED](#) | [CROSSREF](#)
116. Shomori K, Sugamura K, Adachi K, Shiomi T, Nanba E, Ito H. Gastric adenocarcinoma with rhabdoid morphology. *Gastric Cancer* 2011;14:290-294.
[PUBMED](#) | [CROSSREF](#)
117. Ueyama T, Nagai E, Yao T, Tsuneyoshi M. Vimentin-positive gastric carcinomas with rhabdoid features. A clinicopathologic and immunohistochemical study. *Am J Surg Pathol* 1993;17:813-819.
[PUBMED](#) | [CROSSREF](#)
118. Rivera-Hueto F, Ríos-Martín JJ, Domínguez-Triano R, Herreras-Gutierrez JM. Early gastric stump carcinoma with rhabdoid features. Case report. *Pathol Res Pract* 1999;195:841-846.
[PUBMED](#) | [CROSSREF](#)
119. Zheng LD, Yang XP, Pan HX, Nie X, He J, Lv Q, et al. Gastric carcinoma with osteoclast-like giant cells: a case report and review of the literature. *J Zhejiang Univ Sci B* 2009;10:237-241.
[PUBMED](#) | [CROSSREF](#)
120. Jing H, Geng M, Meng Q, Tai Y. Sarcomatoid carcinoma of the stomach with osteoclast-like giant cells. *Tumori* 2012;98:82e-85e.
[PUBMED](#) | [CROSSREF](#)

121. Agaimy A. Pleomorphic (giant cell) carcinoma revisited: a historical perspective and conceptual reappraisal. *Semin Diagn Pathol* 2021;38:187-192.
[PUBMED](#) | [CROSSREF](#)
122. Wang YK, Ma L, Wang ZQ, Wang Y, Li P, Jiang B, et al. Clinicopathological features and differential diagnosis of gastric pleomorphic giant cell carcinoma. *Open Life Sci* 2023;18:20220683.
[PUBMED](#) | [CROSSREF](#)
123. Agaimy A, Rau TT, Hartmann A, Stoeck R. SMARCB1 (INI1)-negative rhabdoid carcinomas of the gastrointestinal tract: clinicopathologic and molecular study of a highly aggressive variant with literature review. *Am J Surg Pathol* 2014;38:910-920.
[PUBMED](#) | [CROSSREF](#)
124. Agaimy A, Daum O, Märkl B, Lichtmanegger I, Michal M, Hartmann A. SWI/SNF complex-deficient undifferentiated/rhabdoid carcinomas of the gastrointestinal tract: a series of 13 cases highlighting mutually exclusive loss of SMARCA4 and SMARCA2 and frequent co-inactivation of SMARCB1 and SMARCA2. *Am J Surg Pathol* 2016;40:544-553.
[PUBMED](#) | [CROSSREF](#)
125. Huang SC, Ng KF, Yeh TS, Cheng CT, Chen MC, Chao YC, et al. The clinicopathological and molecular analysis of gastric cancer with altered SMARCA4 expression. *Histopathology* 2020;77:250-261.
[PUBMED](#) | [CROSSREF](#)
126. Endoh Y, Tamura G, Motoyama T, Ajioka Y, Watanabe H. Well-differentiated adenocarcinoma mimicking complete-type intestinal metaplasia in the stomach. *Hum Pathol* 1999;30:826-832.
[PUBMED](#) | [CROSSREF](#)
127. Okamoto N, Kawachi H, Yoshida T, Kitagaki K, Sekine M, Kojima K, et al. "Crawling-type" adenocarcinoma of the stomach: a distinct entity preceding poorly differentiated adenocarcinoma. *Gastric Cancer* 2013;16:220-232.
[PUBMED](#) | [CROSSREF](#)
128. Yao T, Utsunomiya T, Oya M, Nishiyama K, Tsuneyoshi M. Extremely well-differentiated adenocarcinoma of the stomach: clinicopathological and immunohistochemical features. *World J Gastroenterol* 2006;12:2510-2516.
[PUBMED](#) | [CROSSREF](#)
129. Ushiku T, Arnason T, Ban S, Hishima T, Shimizu M, Fukayama M, et al. Very well-differentiated gastric carcinoma of intestinal type: analysis of diagnostic criteria. *Mod Pathol* 2013;26:1620-1631.
[PUBMED](#) | [CROSSREF](#)
130. Joo M, Han SH. Gastric-type extremely well-differentiated adenocarcinoma of the stomach: a challenge for preoperative diagnosis. *J Pathol Transl Med* 2016;50:71-74.
[PUBMED](#) | [CROSSREF](#)
131. Lee J, Lee IS, Ahn JY, Park YS, Kim J. Extremely well-differentiated adenocarcinoma of the stomach: diagnostic pitfalls in endoscopic biopsy. *J Pathol Transl Med* 2022;56:63-72.
[PUBMED](#) | [CROSSREF](#)