

ATRT-16. MODELLING ATRT THROUGH SWI/SNF COMPLEX DEFICIENCY IN GENETICALLY-ENGINEERED MOUSE MODELS

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Atypical Teratoid/Rhabdoid Tumours (ATRT) are highly malignant neoplasms arising primarily in the CNS of children. They are defined by loss of function mutations in *smarcb1*, a gene serving a vital role in neurogenesis and differentiation. In order to recapitulate ATRT in the mouse, we used a Cre-Lox recombination system to conditionally knockout *smarcb1* in specific cell compartments. Loss of *smarcb1* in BLBP-expressing cells of the developing brain led to severe neurologic defects. Mice exhibited seizures, ataxia, and median 12-day survival. Histological analysis revealed severe thinning of the cerebral cortex and cerebellum. Temporally-targeted *smarcb1* loss in BLBP/Nestin-expressing embryonic compartments did not result in tumour formation. Similarly, BLBP-expressing, *smarcb1*-deficient neural stem/progenitor cells (NSC/NPCs) were isolated and allografted but did not form tumours. These cells demonstrated decreased proliferation, higher apoptosis, and upregulation of p53, CDKN1A, and CDKN2A. In contrast, ubiquitous *smarcb1* loss at the earlier embryonic day 6.5 produced widespread tumorigenicity in the forebrain, hindbrain, skullbase, and spine; each with unique phenotypes, survival, and morphology. We employed a clinically-relevant Nanostring gene-panel screen to stratify tumours into genetically distinct subgroups. Our findings indicate that *smarcb1* plays an important role in CNS development. Loss of *smarcb1* in NSC/NPCs is lethal, and its developmental context influences cell fate. Targeted *smarcb1* loss likely plays a tumorigenic role at an earlier developmental stage than previously determined, in a diverse array of primitive stem cells. These data support the generation of a murine ATRT model capable of producing distinct tumour entities that recapitulate the human disease.

ATRT-17. TARGETING GLUTAMINE METABOLISM LOWERS METHYLATION POTENTIALS IN AT/RT AND SYNERGIZE WITH TAZEMETOSTAT

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Atypical teratoid/rhabdoid tumors (AT/RT) have a single recurring genetic mutation in *SMARCB1*. This deletion leads to an abnormal SWI/SNF chromatin remodeling complex and the constitutive activation of *EZH2*. S-adenosyl-L-methionine (SAM) donates a methyl group to *EZH2* which then methylates DNA and histones leading to the abnormal gene expression responsible for AT/RT's aggressive phenotype. We have previously shown that glutamine metabolic inhibition with 6-diazo-5-oxo-L-norleucine (DON) confers a survival advantage in AT/RT. In this study, we identified with ultra-high performance liquid chromatography mass spectrometry that DON treatment lowered the methylation potential in AT/RT (Decreased SAM:SAH ratio, t-test in 5 AT/RT human-derived cell models comparing DON treatment to DMSO control, $p < 0.05$). AT/RT cell lines grown in glutamine deplete media compared to normal growth conditions also had a reduced methylation potential (decreased SAM:SAH, t-test, $p < 0.05$). DON treatment over 5 days decreased histone methylation (as determined by western blot for H3K27me3). Tazemetostat is a small molecule inhibitor that blocks the SAM methyl donor site on *EZH2*. We find that DON combines synergistically with Tazemetostat to slow AT/RT cell growth (MTS assay, $p < 0.01$ t-test; MUSE viability assay, $p < 0.01$ ANOVA) and enhances cytotoxicity (MUSE Annexin-V, $p < 0.01$ by ANOVA). Synergies were especially pronounced at low concentrations of Tazemetostat which is significant given that Tazemetostat's efficacy in AT/RT has been limited by poor CNS penetration. These studies identify a novel treatment strategy that has potential to improve survival in AT/RT.

ATRT-18. SHH-SUBTYPE ATYPICAL TERATOID/RHABDOID TUMORS ARE SELECTIVELY SENSITIVE TO GEMCITABINE TREATMENT

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Atypical Teratoid Rhabdoid Tumors (ATRT) are highly malignant embryonal tumors of the central nervous system with a dismal prognosis. ATRT

can be divided into three molecular subgroups of which the Sonic Hedgehog (SHH) subgroup is most prevalent. In this study, we developed and validated a novel patient-derived ATRT model, which we used along a panel of other primary ATRT models for large scale drug discovery assays. We found that ATRTs are selectively sensitive to the nucleoside analogue gemcitabine, with SHH-subtype ATRTs being the most sensitive subgroup. Gene expression profiles and protein analysis indicated that gemcitabine treatment causes degradation of Sirtuin 1 (SIRT1), which causes ATRT specific cell-death through NF- κ B and p53 activation. Furthermore, we found that this gemcitabine induced loss of SIRT1 results in a nucleus-to-cytoplasm shift of the SHH signaling activator Gli, explaining the additional gemcitabine sensitivity in SHH-subtype ATRT. Treatment of SHH-subgroup ATRT xenograft-bearing mice resulted in a >40% increase in median survival ($p < 0.01$, log-rank test) and long-term survivors in two independent models. To prepare translation of our findings to the clinic, we investigated potential gemcitabine induced resistance mechanisms by conducting kinome-wide CRISPR/Cas9 knockout screens in primary ATRT cells. Through these experiments we found that low-dose gemcitabine treatment combined with inhibition of protein kinase C zeta (PKC ζ) prevents regrowth of resistant ATRT subclones. Together, these findings show that ATRT are highly sensitive to gemcitabine treatment; and as such we suggest that gemcitabine may be rapidly incorporated into future treatment regimens for SHH-ATRT.

ATRT-19. EPIGENETIC REPROGRAMMING LEADS TO INNATE IMMUNE PATHWAY ACTIVATION IN AT/RT

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BACKGROUND: Atypical teratoid/rhabdoid tumors (AT/RT) are highly aggressive brain tumors affecting early childhood and are characterized by bi-allelic inactivation of the *SMARCB1* gene. Though patients benefit from multimodal therapy, there is no improvement in overall survival necessitating exploration of alternative approaches including innate-based immune therapy and epigenetic therapy, which have shown promise in treating adult brain tumors and other cancers. Though reconstitution of *SMARCB1* in *SMARCB1*-deficient cells leads to activation of interferon-stimulated genes, the role of innate immune signaling has not been investigated in AT/RTs. METHODS: Our data from a panel of AT/RT cell lines indicates loss of expression of key innate signaling components, like RIG-I, MDA-5, cGAS and STING that are required for sensing extracellular dsRNA and dsDNA. These cell lines also do not respond to dsDNA-based or dsRNA-based innate agonists. However, co-treatment of the BT-16 cell line with two epigenetic drugs, panobinostat and 5-azacytidine leads to re-expression of STING and RIG-I. Panobinostat/5-azacytidine co-treatment followed by either genomic DNA (dsDNA agonist) or poly(I:C) (dsRNA agonist) treatment results in induction of innate responses, measured by STAT1 phosphorylation and production of ISG-15 and IPIT-1. CONCLUSION: Our data suggests that AT/RT cell lines are unresponsive to innate agonists possibly due to the loss of expression of key innate immune components. However, these pathways can be reactivated by epigenetic drugs and further potentiated by dsDNA/dsRNA-based innate agonists. Combined epigenetic reprogramming and innate pathway stimulation may serve as a potential therapy option for treating AT/RT.

ATRT-20. CDK7 INHIBITION IN AT/RT

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Atypical teratoid/rhabdoid tumors (AT/RT) are characterized by loss-of-function mutations in the *SMARCB1* component (and less commonly *SMARCA4*) of the SWI/SNF chromatin-remodeling complex. AT/RT demonstrate an overall silent genomic landscape with epigenetic dysregulation of the genome. CDK7 is a key transcriptional regulator that preferentially phosphorylates the Ser5 and Ser7 positions on RNA Polymerase C terminal domain and is involved early in transcription. In tumor cells, CDK7 is enriched at super enhancers which preferentially regulate genes involved in cell transformation, and expressed at significantly higher levels in transformed tissues than the surrounding normal brain. Our preliminary data shows that CDK7 is expressed in a number of AT/RT tumor cell lines and patient-derived tumor cultures, and that loss of CDK7 function through exposure to the novel CDK7 inhibitor THZ2 results in lack of proliferation at lower doses, and caspase-mediated apoptosis at higher concentrations. shRNA-based inhibition confirms that this effect is due specifically to loss of CDK7. RNA sequencing of cells treated with lower doses of THZ2 show significant alterations in transcript expression consistent with altered balance between antagonistic SWI/SNF and PRC2 chromatin-modeling complex activities, as