

## Research Article

# Bioinformatics Analysis: The Regulatory Network of hsa\_circ\_0007843 and hsa\_circ\_0007331 in Colon Cancer

Zeping Han <sup>1</sup>, Huafang Chen <sup>2</sup>, Zhonghui Guo,<sup>1</sup> Jianxia Zhu,<sup>1</sup> Xingyi Xie,<sup>1</sup> Yuguang Li <sup>1</sup> and Jinhua He <sup>1</sup>

<sup>1</sup>Department of Laboratory Medicine, Central Hospital of Panyu District, Guangzhou, Guangdong 511400, China

<sup>2</sup>Leizhou Center for Disease Control and Prevention, Leizhou, Guangdong 524200, China

Correspondence should be addressed to Yuguang Li; [lyg\\_py@126.com](mailto:lyg_py@126.com) and Jinhua He; [332518579@qq.com](mailto:332518579@qq.com)

Received 5 October 2020; Revised 8 June 2021; Accepted 5 July 2021; Published 23 July 2021

Academic Editor: Federico Zambelli

Copyright © 2021 Zeping Han et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Objective.** To analyze the molecular regulation network of circular RNA (circRNA) in colon cancer (CC) by bioinformatics method. **Methods.** hsa\_circ\_0007843 and hsa\_circ\_0007331 proved to be associated with CC in previous studies were chosen as the research object. ConSite database was used to predict the transcription factors associated with circRNA, and the CC-associated transcription factors were screened out after intersection. The CircInteractome database was used to predict the RNA-binding proteins (RBPs) interacting with circRNAs and screen out the CC-associated RBPs after an intersection. Furthermore, the CircInteractome database was used to predict the miRNAs interrelated with circRNAs, and the HMDD v3.2 database was used to search for miRNAs associated with CC. The target mRNAs of miRNA were predicted by the miRWalk v3.0 database. CC-associated target genes were screened out from the GeneCards database, and the upregulated genes were enriched and analyzed by the FunRich 3.1.3 software. Finally, the molecular regulatory network diagram of circRNA in CC was plotted. **Results.** The ConSite database predicted a total of 14 common transcription factors of hsa\_circ\_0007843 and hsa\_circ\_0007331, among which Snail, SOX17, HNF3, C-FOS, and ROR $\alpha$ -1 were related to CC. The CircInteractome database predicted that the RBPs interacting with these two circRNAs were AGO2 and EIF4A3, and both of them were related to CC. A total of 17 miRNAs interacting with hsa\_circ\_0007843 and hsa\_circ\_0007331 were predicted by CircInteractome database. miR-145-5p, miR-21, miR-330-5p, miR-326, and miR-766 were associated with CC according to the HMDDv3.2 database. miR-145-5p and miR-330-5p, lowly expressed in CC, were analyzed in the follow-up study. A total of 676 common target genes of these two miRNAs were predicted by the miRWalk3.0 database. And 57 target genes were involved in the occurrence and development of CC from the GeneCards database, with 23 genes downregulated and 34 genes upregulated. Additionally, GO analysis showed that the 34 upregulated genes were mainly enriched in biological processes such as signal transduction and cell communication. KEGG pathway analysis showed that the upregulated genes were closely related to integrin, ErbB receptor, and ALK1 signal pathways. Finally, a complete regulatory network of hsa\_circ\_0007843 and hsa\_circ\_0007331 in CC was proposed, whereby each one of the participants was either directly or indirectly associated and whose deregulation may result in CC progression. **Conclusion.** Predicting the molecular regulatory network of circRNAs by bioinformatics provides a new theoretical basis for further occurrence and development pathogenesis of CC and good guidance for future experimental research.

## 1. Background

Colon cancer (CC) is the most common lethal tumor of the digestive tract, which seriously threatens human life and health [1]. Its incidence rate is the third highest in females and the second highest in males. The clinical symptom of the patient with early CC is more hidden, and most patients

are not discovered until the middle and late stages. Nowadays, surgery is still the first choice for the treatment of CC, supplemented by radiotherapy and chemotherapy, targeted therapy, and immunotherapy [2]. However, the problems of high recurrence rate and high metastasis rate after operation have not been effectively solved. The prognosis of patients with CC is still not very ideal, so it is necessary to

find new treatments as soon as possible [3]. In recent years, a large number of studies have found that circular RNA (circRNA) plays an important role in the occurrence, development, metastasis, and invasion of CC [4–10] and can also be used as biomolecule markers for the diagnosis and prognosis [11]. circRNA is a new type of endogenous noncoding RNA, without a 5'-end cap and 3'-end poly (A) tail structure, showing a covalently closed loop [12]. Currently, it has been proven to act as sponges of microRNA (miRNA) or proteins, regulating the expression and alternative splicing of host genes, translating peptides, and participating in physiological and pathological processes of the body through these functions [13].

Our previous study found that circRNA was closely related to CC. *hsa\_circ\_0007843* and *hsa\_circ\_0007331* upregulated in CC were widely involved in the occurrence and development of CC, but the specific mechanism needed to be further elucidated [14, 15]. *hsa\_circ\_0007843* is encoded by the ARHGAP32 gene (Rho GTPase-activating protein 32arhgap32), which locates at chr11:128993340-129034322 (<http://www.circbase.org/>). ARHGAP32 encodes a neuron-associated GTPase-activating protein that regulates dendritic spine morphology and strength by modulating Rho GTPase [16]. According to the literature, ARHGAP32 is widely involved in the occurrence and development of gastric cancer and liver cancer [17, 18], but its relationship with colon cancer has not been reported. *hsa\_circ\_0007331* located at chr3:195101737-195112876 is encoded by the ACAP2 gene, a homolog of *Caenorhabditis elegans* CNT-1, which has a proapoptotic function and an identical phosphoinositide-binding pattern to that of CNT-1. It was reported that knockdown of ACAP2 blocks apoptosis in cancer cells in response to the chemotherapeutic antimetabolite 5-fluorouracil and that ACAP2 expression is downregulated in some esophageal cancers, leukemia, and lymphomas suggesting that ACAP2 inactivation or downregulation in human cells may contribute to cancer development [19]. However, the association of ACAP2 in colon cancer has not been reported.

Bioinformatics plays an important role in the field of life science. It mainly includes the generation, management, and analysis of multiple sets of high-throughput data in the field of biology and then comprehensively uses the theories and tools of mathematics, computer science, and life science to clarify the biological significance of these data [20]. Moreover, bioinformatics can be used to predict the regulatory network of circRNA in the occurrence and development of CC, which can better study the pathogenesis of CC and make the next research more targeted and clearer, so bioinformatics has become an indispensable research means in genome research. In this study, using bioinformatics methods, with *hsa\_circ\_0007843* and *hsa\_circ\_0007331* as the research object, we explore the regulatory relationship between it and upstream transcription factors, downstream RNA-binding proteins (RBPs), target miRNA, and miRNA target genes (Figure 1), to better understand the mechanism of circRNA in the occurrence, development, invasion, and metastasis of CC and provide new clues for the next experiment to verify its molecular regulatory network mechanism.

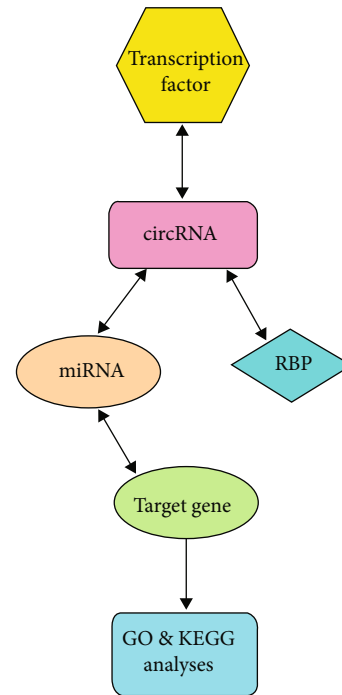


FIGURE 1: The schematic diagram of the research route.

## 2. Materials and Methods

**2.1. Identification of Transcription Factors Associated with circRNAs.** ConSite database (<http://consite.genereg.net/>) is a web-based tool for predicting transcription factors by gene-binding sites, which can be used to find cisregulatory elements in gene sequences [21]. The transcription factors of *hsa\_circ\_0007843* and *hsa\_circ\_0007331* were predicted and analyzed by the ConSite database. The common transcription factors of the abovementioned circRNA were screened. The relationship between transcription factors and CC was queried by the PubMed database (<http://www.ncbi.nlm.nih.gov/pubmed>).

**2.2. Prediction of RBPs Interacting with circRNAs.** CircInteractome database (<https://circinteractome.irp.nia.nih.gov/>), for mapping RBP- and miRNA-binding sites on human circRNAs, searches public circRNA, miRNA, and RBP databases to provide bioinformatics analyses of binding sites on circRNAs and additionally analyzes miRNA and RBP sites on junction and junction-flanking sequences. Furthermore, it can also identify potential circRNAs that can act as RBP sponges, and design primers and siRNAs for circRNA [22]. The target circRNAs were submitted in the “circular RNA” of the CircInteractome database to obtain the RBPs that interact with circRNAs. Then, the common RBP of these two circular RNAs associated with CC was obtained through the PubMed database.

**2.3. Prediction of miRNA Interacting with *hsa\_circ\_0007843* and *hsa\_circ\_0007331*.** Enter the prediction interface through “miRNA target sites,” and then, submit the target circRNAs to get the predicted miRNAs in the CircInteractome database.

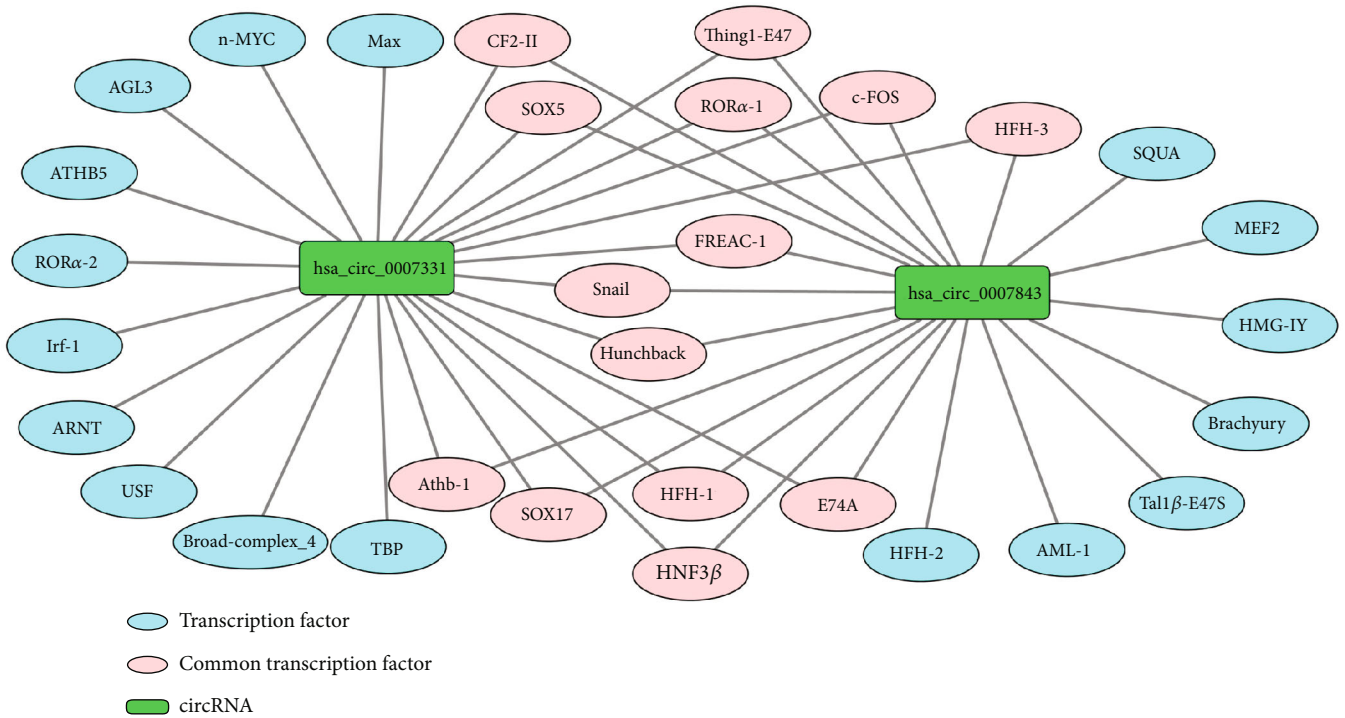


FIGURE 2: Transcription factors associated with circRNAs.

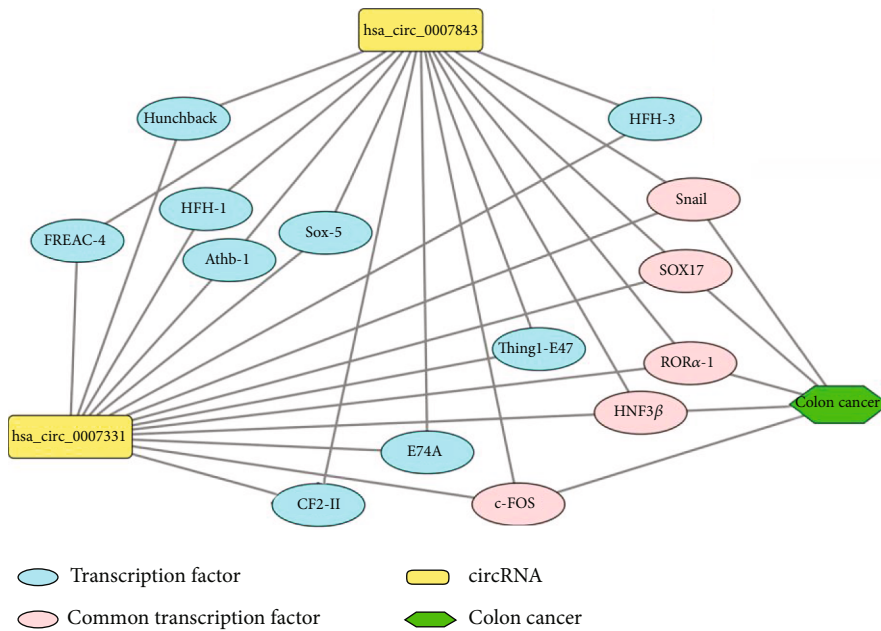


FIGURE 3: Transcription factors associated with colon cancer.

2.4. *Identification of CC-Associated miRNAs.* The HMDD database is an experimentally confirmed database related to human miRNA diseases [23]. We searched for miRNAs that had been confirmed to be related to CC using HMDD v3.2 (<http://www.cuilab.cn/hmdd/>).

2.5. *Target mRNA Prediction.* miRWalk is an online software that covers the latest information about the interaction between mRNAs and miRNAs in humans, mice, dogs, and cattle. It can easily identify important miRNA targets to better understand the role of multiple miRNAs and optimize

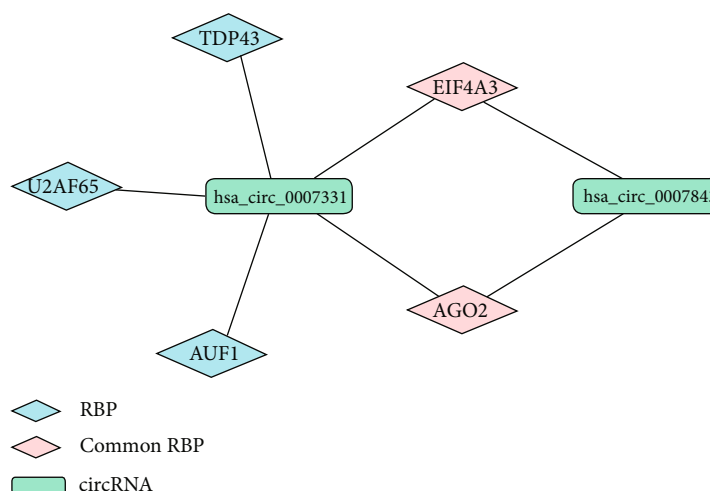


FIGURE 4: RBPs interacting with circRNAs.

TABLE 1: Colon cancer-associated miRNAs.

miRNA name	PMID	Description	Expression
hsa-miR-145	22766504	miR-145 regulates PAK4 via the MAPK pathway and exhibits an antitumor effect in human colon cells.	Downregulation
hsa-miR-330	22132977	Expression of miR-330 in various colon cancer cell lines varied fivefold between samples and correlated with in vitro gemcitabine resistance.	Downregulation
hsa-miR-326	22043014	The development of colonic inflammation in IL-10(-/-) mice was accompanied by upregulation in the expression of 10 miRNAs (miR-19a, miR-21, miR-31, etc.).	Upregulation
hsa-miR-766	30127618	Verified by qRT-PCR, the expression of hsa-miR-766 was upregulated in colon cancer cells.	Upregulation
hsa-miR-21	21279518	PDCD4 nuclear loss inversely correlates with miR-21 levels in colon carcinogenesis.	Upregulation

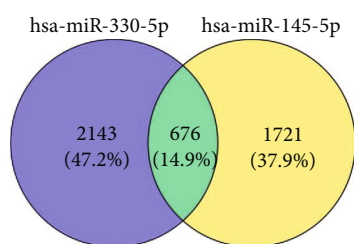


FIGURE 5: The common target mRNAs of hsa-miR-145-5p and hsa-miR-330-5p.

their genetic targets [24]. The target mRNAs of the above miRNAs are predicted by miRwalk v3.0 (<http://mirwalk.umm.uni-heidelberg.de/>), and the common target genes were screened after intersection.

**2.6. Identification of CC-Related Target Genes.** GeneCards database (<https://www.genecards.org/>) provides concise genome, proteome, transcriptome, disease, and functional data of all known and predicted human genes. It is a comprehensive database of human genes, which can query not only specific gene details but also all genes related to one disease [25]. All genes associated with CC can be obtained by simply typing “colon cancer” above “keywords.”

**2.7. Functional Enrichment Analysis.** FunRich is an open bioinformatics analysis system capable of functional enrichment and network analysis of genes and proteins [26]. GO and KEGG enrichment analysis of target genes can be performed by FunRich version 3.1.3 software (<http://www.funrich.org/download>).

**2.8. Construction of the Regulatory Network of circRNAs in CC.** Based on the genomic information above, the regulatory network diagram of circRNA in CC was constructed by Cytoscape software version 3.7.2 (<http://www.cytoscape.org/index.html>).

### 3. Results

**3.1. Transcription Factors Associated with circRNAs.** As transcription factors can act as upstream regulatory molecules of circRNA and affect its expression, we used the ConSite database to predict the upstream transcription factors of each circRNA. Under the condition of setting “cut-off” to 95%, 21 transcription factors of has\_circ\_0007843 and 25 transcription factors of has\_circ\_0007331 were predicted by the ConSite database. After intersection, there were 14 common transcription factors of the two circRNAs (Figure 2).

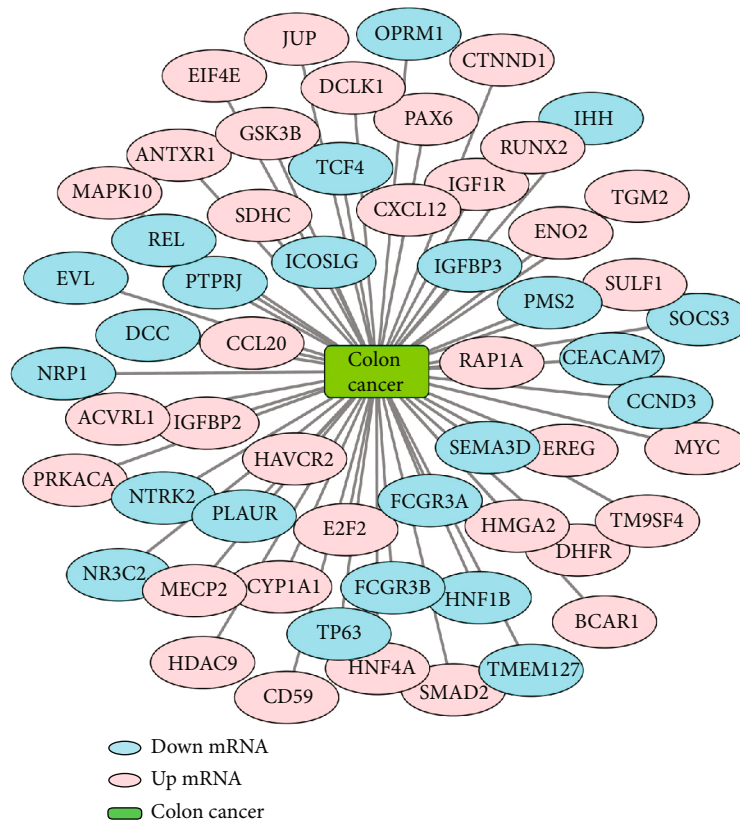


FIGURE 6: Intersection target genes associated with colon cancer.

3.2. *Transcription Factors Associated with CC.* As shown in Figure 3, only 5 of the 14 transcription factors (Snail, SOX17, HNF3  $\beta$ , c-FOS, and ROR $\alpha$ -1) are involved in the occurrence and development of colon cancer, consulting the literature through the PubMed database.

3.3. *RBPs Interacting with circRNAs.* After identifying the upstream regulatory transcription factors of circRNA, we continued to predict the downstream regulatory pathways of circRNA. The interaction with RBPs is also considered an important factor for investigating the function of circRNAs. The RBPs of has\_circ\_0007843 were EIF4A3 and AGO2, while those of has\_circ\_0007331 were AGO2, EIF4A3, AUF1, TDP43, and U2AF65 predicted by the CircInteractome database. Figure 4 showed that EIF4A3 and AGO2 were the common RBPs of the two circRNAs, and both of them were associated with colon cancer [27, 28].

3.4. *circRNA-Associated miRNAs.* The CircInteractome database predicted that the miRNAs interacting with has\_circ\_0007843 were hsa-miR-1299, hsa-miR-326, hsa-miR-330-5p, hsa-miR-485-3p, hsa-miR-516a-5p, hsa-miR-543, hsa-miR-665, hsa-miR-766, hsa-miR-892b, and hsa-miR-921. Additionally, there were 7 miRNAs interacting with has\_circ\_0007331, such as hsa-miR-145-5p, hsa-miR-21, hsa-miR-590-5p, hsa-miR-433, hsa-miR-526b, hsa-miR-561, and hsa-miR-578.

3.5. *CC-Associated miRNAs.* A total of 139 miRNAs were confirmed to be associated with CC according to HMDD v3.2, which were intersected with the above 17 miRNAs to filter out five common miRNAs, including hsa-miR-326, hsa-miR-766, hsa-miR-330-5p, hsa-miR-145-5p, and hsa-miR-21. As known to all, acting as miRNA sponges, circRNAs can bind miRNAs through a miRNA response element (MRE) and negatively regulate their activity [29]. Since has\_circ\_0007843 and has\_circ\_0007331 were upregulated in CC, hsa-miR-145-5p and hsa-miR-330-5p, downregulated in CC, were selected as the objects of further research (Table 1).

3.6. *Target mRNA Binding to miRNA.* Click “miRNAs” from the main page of miRwalk v3.0, then enter hsa-miR-145-5p and hsa-miR-330-5p and predict the target mRNAs, respectively. Under the condition of setting the score to 1, a total of 2397 and 2819 target mRNAs were identified for hsa-miR-145-5p and hsa-miR-330-5p, respectively. As shown in Figure 5, a total of 676 overlaps were identified for these two miRNAs.

3.7. *The CC-Associated mRNAs.* 1762 mRNAs associated with CC were obtained under the condition of “relevance” not less than 10 in the GeneCards database. A total of 57 mRNAs related to CC, with 34 mRNAs upregulated and 23 mRNAs downregulated, were obtained after taking an intersection with the 676 mRNAs above. As miRNAs are single-

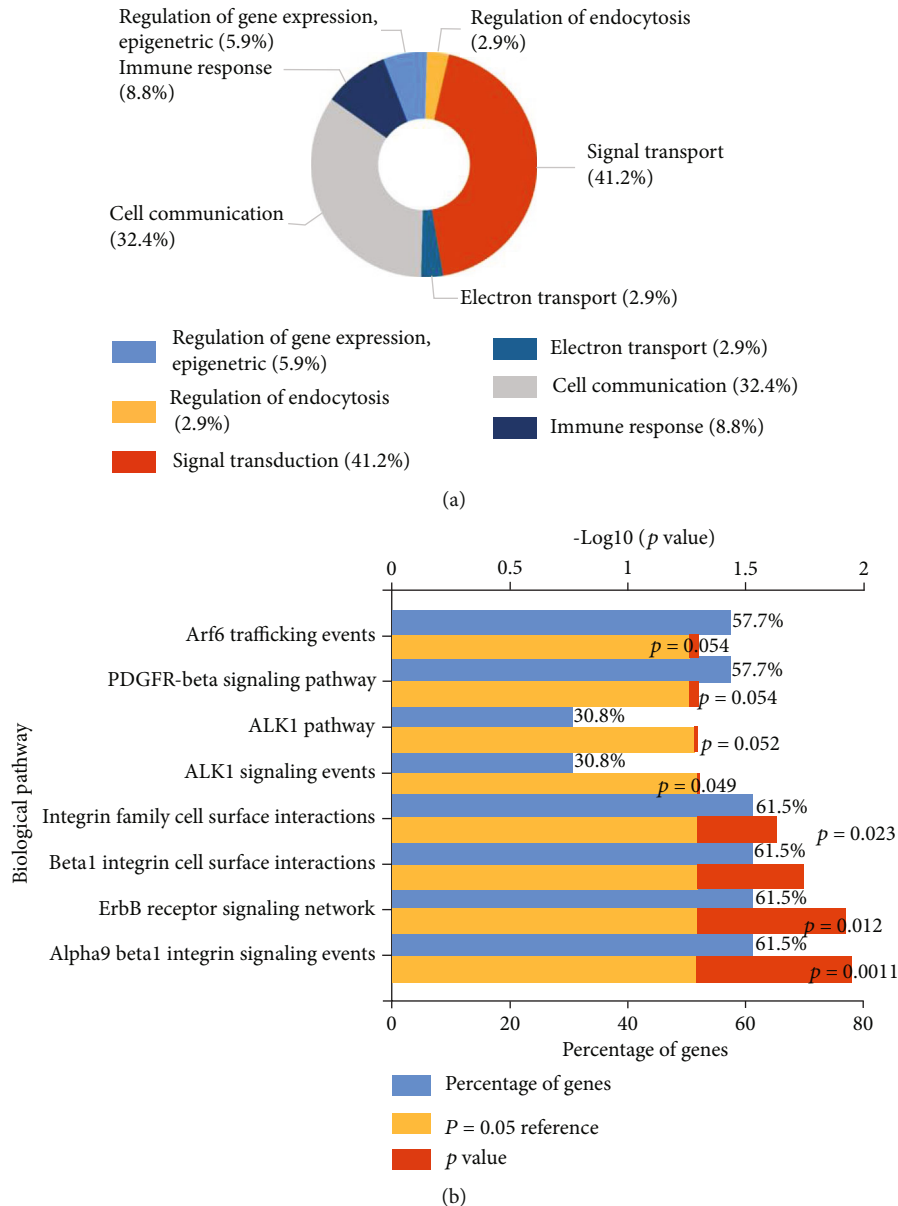


FIGURE 7: Enrichment analysis of target genes: (a) GO function enrichment; (b) KEGG pathway enrichment.

stranded RNA molecules that bind to targets in a base pair-mediated manner, resulting in the degradation or inhibition of the expression and function of target mRNAs, the expression of the target mRNAs was inverse to that of the miRNAs [30]. Then, 34 upregulated target mRNAs were included in the follow-up study (Figure 6).

**3.8. GO and KEGG Enrichment Analysis.** Enrichment analysis of the 34 upregulated target mRNAs was performed by FunRich 3.1.3. GO enrichment includes cellular components (CC), molecular functions (MF), and biological processes (BP). In biological processes, target genes were mainly concentrated in signal transduction and cell communication (Figure 7(a)). In KEGG pathway analysis, the most abundant pathways were  $\alpha 9\beta 1$  integrin signaling pathway, ErbB receptor signaling network,  $\beta 1$  integrin cell surface interaction,

integrin family cell surface interaction, ALK1 signaling pathway, etc. (Figure 7(b)).

**3.9. Functional Regulatory Network of circRNA in CC.** Figure 8 presents the proposed regulatory network of circRNAs in CC.

## 4. Discussion

Bioinformatics is an interdisciplinary field of science that combines molecular biology, genetics, computer science, information engineering, mathematics, and statistics to solve data-intensive, large-scale biological problems from a computational perspective. It mainly focuses on modelling biological processes at the molecular level and making inferences from the data collected [31]. Moreover, bioinformatics can

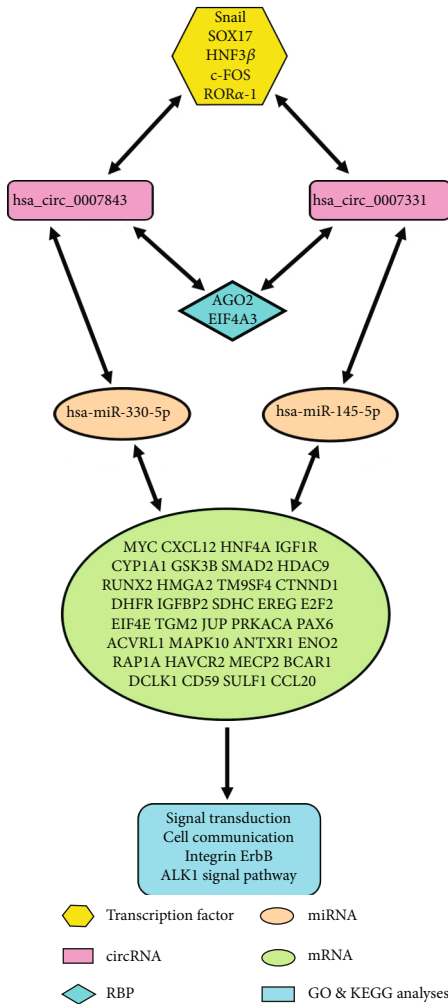


FIGURE 8: The regulatory network of circRNA in colon cancer.

seamlessly store, mine, retrieve, and analyze data from genomics and proteomics using existing and emerging computing technologies and then construct computational models that speed up research and shorten the time of scientific research. The conclusions obtained from the analysis of the experimental data can be used to design the next stage of the experiment, and the computer can also be used to manage the experimental data and predict the structure and function of new genes and then to decode the complex biological information step by step, to provide a more clear direction for the diagnosis and treatment of various diseases.

In this study, we analyzed the molecular regulatory network of circRNA in the occurrence and development of CC by bioinformatics. *hsa\_circ\_0007843* and *hsa\_circ\_0007331*, which were associated with CC in the previous study, were selected as the subjects of research. The expression of *hsa\_circ\_0007843* and *hsa\_circ\_0007331* in CC tissue was significantly higher than that in normal colon tissue. *hsa\_circ\_0007843* acted as a miRNA sponge to regulate the expression of matrix metalloproteinase 2 (MMP2) by eliminating the inhibitory effect of miR-518c-5p on the translation of MMP2 gene, thus promoting the migration and invasion ability of SW480 cell [15]. And circRNA-ACAP2 (*hsa\_circ\_*

0007331) could also affect the proliferation, migration, and invasion of CC SW480 cells by removing the inhibitory effect of miR-21-5p on Tiam1 expression [14].

As a competing endogenous RNA (ceRNA), circRNA can competitively bind to miRNA, thereby regulating the expression of its downstream target genes [32]. In this study, a total of 17 miRNAs interacting with *hsa\_circ\_0007843* and *hsa\_circ\_0007331* were predicted by the CircInteractome database. A total of 139 miRNAs associated with CC were found in HMDD v3.2, and the common miRNAs screened after taking the intersection with the above 17 miRNAs were *hsa-miR-326*, *hsa-miR-766*, *hsa-miR-330-5p*, *hsa-miR-145-5p*, and *hsa-miR-21*. Researches confirm that *hsa-miR-326*, *hsa-miR-766*, and *hsa-miR-21* are upregulated in CC [33–36]. The expression of *hsa-miR-330-5p* in colon cancer tissue is significantly lower than that in adjacent tissues and negatively regulates the expression of integrin  $\alpha 5$  (ITGA5), which affects the development of CC [37]. *hsa-miR-145-5p*, low-expressed in CC, targets CDCA3 to play a tumor suppressor role, acting as a biomarker for diagnosis and treatment [38]. It can also regulate RHBDD1 through EGFR-related signaling pathways to inhibit cell growth, invasion, migration, and tumorigenesis in colon cancer [39].

The miRwalk v3.0 online software predicted that 57 common target mRNAs of the two miRNAs (*hsa-miR-330-5p* and *hsa-miR-145-5p*) were closely associated with CC, with 23 downregulated and 34 upregulated according to literatures from the PubMed database. Enrichment analysis results showed that upregulated genes were mainly involved in related biological processes such as signal transduction and cell communication, while the KEGG pathway was mainly closely related to integrin, ErbB receptor, ALK1, and other signaling pathways.

It is well known that transcription factors are indispensable regulatory factors in gene expression. It was predicted by ConSite that there were 13 common transcription factors in *hsa\_circ\_0007843* and *hsa\_circ\_0007331*. According to PubMed literature review, Snail, SOX17, HNF3 $\beta$ , c-FOS, and ROR $\alpha$ -1 have been confirmed to be associated with colon cancer. As a key transcription factor of epithelial-mesenchymal transformation (MET) in CC, Snail is upregulated in HT29 CC cells and regulates the expression of miRNA during EMT. Besides, studies have confirmed that the increased expression of Snail is significantly related to tumor size, lymph node metastasis, distant metastasis, clinical stage, and poor prognosis in CC patients [40, 41]. The SOX transcription factor family can regulate the classic Wnt signaling pathway, and when SOX17 is overexpressed in SW480 cells, it inhibits the activity of  $\beta$ -catenin/TCF in a dose-dependent manner, while also inhibiting cell proliferation [42]. HNF3 $\beta$ , an important transcription factor in gastrointestinal development, expressed in colorectal cancer, is lower than that in normal tissues. Its overexpression can inhibit the progression of CC through JAK-STAT3 signal transduction and serve as a potential target for antitumor therapy of CC [43]. The expression of c-FOS is associated with the metastasis and TNM stage of colon cancer. In the inflammatory microenvironment, GDF15 induces the invasion and metastasis of CC by activating c-FOS to regulate

EMT gene [44]. Immunoassay revealed the expression of ROR $\alpha$ -1 protein in colon adenocarcinoma cells [45], but data on the regulatory relationship between ROR $\alpha$ -1 and CC have not been found.

circRNAs can combine with RBPs playing a crucial role in a variety of biological functions. The CircInteractome database predicted that the common RBPs of hsa\_circ\_0007843 and hsa\_circ\_0007331 were EIF4A3 and AGO2. In CC, EIF4A3 can combine with the long noncoding RNA H19, blocking the recruitment of EIF4A3 to cell cycle gene mRNA [21]. Studies by Huang et al. [23] have shown that AGO2 can be expressed in CC and may be related to the pathobiology of CC.

To synthesize the above results, the circRNA molecular regulatory network related to CC was constructed by bioinformatics analysis technology. Through this relatively systematic network, we found that the expression of hsa\_circ\_0007843, hsa\_circ\_0007331, and Snail in CC was upregulated, which promoted the growth and proliferation of CC cells; while the expression of hsa-miR-145-5p, hsa-miR-330-5p, and c-FOS was downregulated and played an inhibitory role. Based on this regulatory network, further research could provide more targeted treatment methods to reduce the expression of hsa\_circ\_0007843, hsa\_circ\_0007331, and Snail or increase the expression of hsa-miR-145-5p, hsa-miR-330-5p, and c-FOS to achieve the purpose of effective treatment of CC. The results of this study provided a valuable reference for elucidating the molecular mechanism of circRNA in the occurrence and development of CC and provided a theoretical basis for the screening of CC diagnostic markers and the selection of precise targets for drug treatment.

## Data Availability

The data that support results of the present study are available from CircInteractome database (<https://circinteractome.irp.nia.nih.gov/>), HMDD v3.2 (<http://www.cuilab.cn/hmdd/>), miRwalk v3.0 (<http://mirwalk.umm.uni-heidelberg.de/>), GeneCards database (<https://www.genecards.org/>), FunRich version 3.1.3 software (<http://www.funrich.org/download>), ConSite database (<http://consite.genereg.net/>), and PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>).

## Conflicts of Interest

The authors declare that they have no competing interests.

## Authors' Contributions

Zeping Han and Huafang Chen contributed equally to this work.

## Acknowledgments

This work was supported by grants from the Medical and Health Science and Technology Project of Panyu District, Guangzhou (No. 2018-Z04-59; No. 2019-Z04-02; and No. 2020-Z04-026); the Guangzhou Municipal Health and Family Planning Commission Program (No. 20192A011027; No.

20191A011119; No. 20201A010085; and No. 20212A010025); Natural Science Foundation of Guangdong Province (No. 2018A0303130191); Guangzhou Science and Technology Plan Project (No. 201904010044; No. 202002030032); and Medical Science and Technology Research Foundation of Guangdong Province (No. A2020304).

## References

- [1] R. L. Siegel, K. D. Miller, and A. Jemal, "Cancer statistics, 2019," *CA: a Cancer Journal for Clinicians*, vol. 69, no. 1, pp. 7–34, 2019.
- [2] K. D. Miller, L. Nogueira, A. B. Mariotto et al., "Cancer treatment and survivorship statistics, 2019," *CA: a Cancer Journal for Clinicians*, vol. 69, no. 5, pp. 363–385, 2019.
- [3] Z. Jahanafrooz, J. Mosafer, M. Akbari, M. Hashemzadei, A. Mokhtarzadeh, and B. Baradaran, "Colon cancer therapy by focusing on colon cancer stem cells and their tumor micro-environment," *Journal of Cellular Physiology*, vol. 235, no. 5, pp. 4153–4166, 2020.
- [4] F. L. Tu, X. Q. Guo, H. X. Wu et al., "Circ-0001313/miRNA-510-5p/AKT2 axis promotes the development and progression of colon cancer," *American Journal of Translational Research*, vol. 12, no. 1, pp. 281–291, 2020.
- [5] Y. Sun, Z. Cao, J. Shan et al., "Hsa\_circ\_0020095 Promotes Oncogenesis and Cisplatin Resistance in Colon Cancer by Sponging miR-487a-3p and Modulating SOX9," *Frontiers in Cell and Developmental Biology*, vol. 8, article 604869, 2021.
- [6] H. Q. Ju, Q. Zhao, F. Wang et al., "A circRNA signature predicts postoperative recurrence in stage II/III colon cancer," *EMBO molecular medicine*, vol. 11, no. 10, article e10168, 2019.
- [7] W. Cui, J. Dai, J. Ma, and H. Gu, "circCDYL/microRNA-150-5p participates in modulating growth and migration of colon cancer cells," *General Physiology and Biophysics*, vol. 38, no. 6, pp. 485–495, 2019.
- [8] M. Lai, G. Liu, R. Li et al., "Hsa\_circ\_0079662 induces the resistance mechanism of the chemotherapy drug oxaliplatin through the TNF- $\alpha$  pathway in human colon cancer," *Journal of Cellular and Molecular Medicine*, vol. 24, no. 9, pp. 5021–5027, 2020.
- [9] Z. Pan, J. Cai, J. Lin et al., "A novel protein encoded by circFNDC3B inhibits tumor progression and EMT through regulating Snail in colon cancer," *Molecular Cancer*, vol. 19, no. 1, p. 71, 2020.
- [10] L. Wang, X. Peng, X. Lu, Q. Wei, M. Chen, and L. Liu, "Inhibition of hsa\_circ\_0001313 (circCCDC66) induction enhances the radio-sensitivity of colon cancer cells via tumor suppressor miR-338-3p: effects of circ\_0001313 on colon cancer radio-sensitivity," *Pathology, Research and Practice*, vol. 215, no. 4, pp. 689–696, 2019.
- [11] X. Zhao, Y. Cai, and J. Xu, "Circular RNAs: biogenesis, mechanism, and function in human cancers," *International Journal of Molecular Sciences*, vol. 20, no. 16, p. 3926, 2019.
- [12] M. I. Taborda, S. Ramírez, and G. Bernal, "Circular RNAs in colorectal cancer: possible roles in regulation of cancer cells," *World journal of gastrointestinal oncology*, vol. 9, no. 2, pp. 62–69, 2017.
- [13] D. Li, Y. Yang, Z. Q. Li, L. C. Li, and X. H. Zhu, "Circular RNAs: from biogenesis and function to diseases," *Chinese Medical Journal*, vol. 132, no. 20, pp. 2457–2464, 2019.
- [14] J. H. He, Y. G. Li, Z. P. Han et al., "The circRNA-ACAP2/Hsa-miR-21-5p/ Tiam1 regulatory feedback circuit affects the



- proliferation, migration, and invasion of colon cancer SW480 cells," *Cellular Physiology and Biochemistry*, vol. 49, no. 4, pp. 1539–1550, 2018.
- [15] J. H. He, Z. P. Han, J. G. Luo et al., "Hsa\_circ\_0007843 acts as a miR-518c-5p sponge to regulate the migration and invasion of colon cancer SW480 cells," *Frontiers in Genetics*, vol. 11, p. 9, 2020.
- [16] F. Zeidán-Chuliá, J. L. Rybarczyk-Filho, A. B. Salmina, B. H. de Oliveira, M. Noda, and J. C. Moreira, "Exploring the multifactorial nature of autism through computational systems biology: calcium and the Rho GTPase RAC1 under the spotlight," *Neuromolecular Medicine*, vol. 15, no. 2, article 8224, pp. 364–383, 2013.
- [17] I. Schulte, E. M. Batty, J. C. Pole et al., "Structural analysis of the genome of breast cancer cell line ZR-75-30 identifies twelve expressed fusion genes," *BMC Genomics*, vol. 13, no. 1, p. 719, 2012.
- [18] Q. Chen, Y. Tan, C. Zhang et al., "A weighted gene co-expression network analysis-derived prognostic model for predicting prognosis and immune infiltration in gastric cancer," *Frontiers in Oncology*, vol. 11, article 554779, 2021.
- [19] K. D. Sullivan, A. Nakagawa, D. Xue, and J. M. Espinosa, "Human ACAP2 is a homolog of *C. elegans* CNT-1 that promotes apoptosis in cancer cells," *Cell Cycle*, vol. 14, no. 12, pp. 1771–1778, 2015.
- [20] J. Gauthier, A. T. Vincent, S. J. Charette, and N. Derome, "A brief history of bioinformatics," *Briefings in Bioinformatics*, vol. 20, no. 6, pp. 1981–1996, 2019.
- [21] A. Sandelin, W. W. Wasserman, and B. Lenhard, "ConSite: web-based prediction of regulatory elements using cross-species comparison," *Nucleic acids research*, vol. 32, no. Web Server, pp. W249–W252, 2004.
- [22] D. B. Dudekula, A. C. Panda, I. Grammatikakis, S. De, K. Abdelmohsen, and M. Gorospe, "CircInteractome: a web tool for exploring circular RNAs and their interacting proteins and microRNAs," *RNA Biology*, vol. 13, no. 1, pp. 34–42, 2016.
- [23] Z. Huang, J. Shi, Y. Gao et al., "HMDD v3.0: a database for experimentally supported human microRNA-disease associations," *Nucleic Acids Research*, vol. 47, no. D1, pp. D1013–D1017, 2019.
- [24] C. Sticht, C. De La Torre, A. Parveen, and N. Gretz, "miRWalk: an online resource for prediction of microRNA binding sites," *PLoS One*, vol. 13, no. 10, article e0206239, 2018.
- [25] M. Safran, I. Dalah, J. Alexander et al., "GeneCards Version 3: the human gene integrator," *Database (Oxford)*, vol. 2010, article baq020, 2010.
- [26] A. Benito-Martin and H. Peinado, "FunRich proteomics software analysis, let the fun begin!," *Proteomics*, vol. 15, no. 15, pp. 2555–2556, 2015.
- [27] D. Han, X. Gao, M. Wang et al., "Long noncoding RNA H19 indicates a poor prognosis of colorectal cancer and promotes tumor growth by recruiting and binding to eIF4A3," *Oncotarget*, vol. 7, no. 16, article 8063, pp. 22159–22173, 2016.
- [28] D. J. Papachristou, A. Korpentinou, E. Giannopoulou et al., "Expression of the ribonucleases Droscha, Dicer, and Ago2 in colorectal carcinomas," *Virchows Archiv*, vol. 459, no. 4, pp. 431–440, 2011.
- [29] S. Memczak, M. Jens, A. Elefsinioti et al., "Circular RNAs are a large class of animal RNAs with regulatory potency," *Nature*, vol. 495, no. 7441, pp. 333–338, 2013.
- [30] T. Du and P. D. Zamore, "Beginning to understand microRNA function," *Cell Research*, vol. 17, no. 8, pp. 661–663, 2007.
- [31] C. Tolga, "Introduction to bioinformatics," *Methods in Molecular Biology*, vol. 1107, pp. 51–71, 2014.
- [32] T. B. Hansen, T. I. Jensen, B. H. Clausen et al., "Natural RNA circles function as efficient microRNA sponges," *Nature*, vol. 495, no. 7441, pp. 384–388, 2013.
- [33] B. Jia, L. Xia, and F. Cao, "The role of miR-766-5p in cell migration and invasion in colorectal cancer," *Experimental and Therapeutic Medicine*, vol. 15, no. 3, pp. 2569–2574, 2018.
- [34] J. S. Schaefer, D. Montufar-Solis, N. Vigneswaran, and J. R. Klein, "Selective upregulation of microRNA expression in peripheral blood leukocytes in IL-10<sup>-/-</sup> mice precedes expression in the colon," *Journal of Immunology*, vol. 187, no. 11, pp. 5834–5841, 2011.
- [35] F. E. Ahmed, N. C. Ahmed, M. M. Gouda, P. W. Vos, and C. Bonnerup, "RT-qPCR for fecal mature microRNA quantification and validation," *Methods in Molecular Biology*, vol. 1765, pp. 203–215, 2018.
- [36] W. Yan, W. Yang, Z. Liu, and G. Wu, "Characterization of microRNA expression in primary human colon adenocarcinoma cells (SW480) and their lymph node metastatic derivatives (SW620)," *Oncotargets and Therapy*, vol. 11, pp. 4701–4709, 2018.
- [37] H. I. Yoo, B. K. Kim, and S. K. Yoon, "MicroRNA-330-5p negatively regulates ITGA5 expression in human colorectal cancer," *Oncology Reports*, vol. 36, no. 5, pp. 3023–3029, 2016.
- [38] Q. Chen, L. Zhou, X. Ye, M. Tao, and J. Wu, "miR-145-5p suppresses proliferation, metastasis and EMT of colorectal cancer by targeting CDCA3," *Pathology-Research and Practice*, vol. 216, no. 4, article 152872, 2020.
- [39] Y. Niu, J. Zhang, Y. Tong, J. Li, and B. Liu, "miR-145-5p restrained cell growth, invasion, migration and tumorigenesis via modulating RHBDD1 in colorectal cancer via the EGFR-associated signaling pathway," *The international journal of biochemistry & cell biology*, vol. 117, article 105641, 2019.
- [40] H. Wang, J. M. Li, W. Wei et al., "Regulation of ATP-binding cassette subfamily B member 1 by Snail contributes to chemoresistance in colorectal cancer," *Cancer Science*, vol. 111, no. 1, pp. 84–97, 2020.
- [41] P. Przygodzka, I. Papiewska-Pająk, H. Bogusz-Koziarska, E. Sochacka, J. Boncela, and M. A. Kowalska, "Regulation of miRNAs by Snail during epithelial-to-mesenchymal transition in HT29 colon cancer cells," *Scientific Reports*, vol. 9, no. 1, p. 2165, 2019.
- [42] D. Sinner, J. J. Kordich, J. R. Spence et al., "Sox17 and Sox4 differentially regulate beta-catenin/T-cell factor activity and proliferation of colon carcinoma cells," *Molecular and Cellular Biology*, vol. 27, no. 22, pp. 7802–7815, 2007.
- [43] J. Wang, H. Lu, W. Wang et al., "Hepatocyte nuclear factor 3β plays a suppressive role in colorectal cancer progression," *Frontiers in Oncology*, vol. 9, p. 1096, 2019.
- [44] Y. Ding, K. Hao, Z. Li et al., "c-Fos separation from Lamin A/C by GDF15 promotes colon cancer invasion and metastasis in inflammatory microenvironment," *Journal of Cellular Physiology*, vol. 235, no. 5, pp. 4407–4421, 2020.
- [45] M. Karasek, A. Carrillo-Vico, J. M. Guerrero, K. Winczyk, and M. Pawlikowski, "Expression of melatonin MT(1) and MT(2) receptors, and ROR alpha(1) receptor in transplantable murine colon 38 cancer," *Neuro Endocrinology Letters*, vol. 23, Supplement 1, pp. 55–60, 2002.