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RESEARCH ARTICLE

Dengue Virus Hijacks a Noncanonical Oxidoreductase Function of a Cellular Oligosaccharyltransferase Complex

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ABSTRACT Dengue virus (DENV) is the most common arboviral infection globally, infecting an estimated 390 million people each year. We employed a genome-wide clustered regularly interspaced short palindromic repeat (CRISPR) screen to identify host dependency factors required for DENV propagation and identified the oligosaccharyltransferase (OST) complex as an essential host factor for DENV infection. Mammalian cells express two OSTs containing either STT3A or STT3B. We found that the canonical catalytic function of the OSTs as oligosaccharyltransferases is not necessary for DENV infection, as cells expressing catalytically inactive STT3A or STT3B are able to support DENV propagation. However, the OST subunit MAGT1, which associates with STT3B, is also required for DENV propagation. MAGT1 expression requires STT3B, and a catalytically inactive STT3B also rescues MAGT1 expression, supporting the hypothesis that STT3B serves to stabilize MAGT1 in the context of DENV infection. We found that the oxidoreductase CXXC active site motif of MAGT1 was necessary for DENV propagation, as cells expressing an AXXA MAGT1 mutant were unable to support DENV infection. Interestingly, cells expressing single-cysteine CXXA or AXXC mutants of MAGT1 were able to support DENV propagation. Utilizing the engineered peroxidase APEX2, we demonstrate the close proximity between MAGT1 and NS1 or NS4B during DENV infection. These results reveal that the oxidoreductase activity of the STT3B-containing OST is necessary for DENV infection, which may guide the development of antiviral agents targeting DENV.

IMPORTANCE The host oligosaccharyltransferase (OST) complexes have been identified as essential host factors for dengue virus (DENV) replication; however, their functions during DENV infection are unclear. A previous study showed that the canonical OST activity was dispensable for DENV replication, suggesting that the OST complexes serve as scaffolds for DENV replication. However, our work demonstrates that one function of the OST complex during DENV infection is to provide oxidoreductase activity via the OST subunit MAGT1. We also show that MAGT1 associates with DENV NS1 and NS4B during viral infection, suggesting that these nonstructural proteins may be targets of MAGT1 oxidoreductase activity. These results provide insight into the cell biology of DENV infection, which may guide the development of antivirals against DENV.

KEYWORDS dengue fever, endoplasmic reticulum, glycosylation, host-pathogen interactions, oxidoreductases

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Dengue virus (DENV) is an enveloped positive-sense single-stranded RNA flavivirus that infects an estimated 390 million people each year, making it the most commonly acquired arbovirus infection (1). An effective vaccine that protects against all four DENV serotypes remains elusive, and to date, there are no antiviral agents approved for the treatment of DENV infection (2).

DENV encodes a single open reading frame that is translated into a polyprotein, which is processed by both cellular and viral proteases at the endoplasmic reticulum (ER) (3). DENV, as an obligate intracellular parasite, is dependent on host-cell proteins to replicate its genome and produce progeny virus. The cellular factors involved in DENV replication have not been well described; however, two genome-wide screens to identify such DENV host dependency factors have recently been published, identifying the oligosaccharyltransferase (OST) complexes as essential to DENV replication (4, 5).

STT3A and STT3B, the catalytic components of the OST complexes, were identified as essential host factors for DENV replication in these screens. There are two distinct OST complexes in mammalian cells that incorporate either STT3A or STT3B as their catalytic subunit (6). Both STT3A and STT3B are oligosaccharyltransferases that catalyze the transfer of high-mannose oligosaccharides onto target proteins in the lumen of the ER (6). STT3A and STT3B can glycosylate specific asparagine residues within the Asn-X-Ser/Thr sequon on target proteins. While STT3A is likely responsible for cotranslational glycosylation of nascent proteins entering the ER lumen, STT3B is responsible for posttranslational glycosylation of proteins with sequons skipped by STT3A (7).

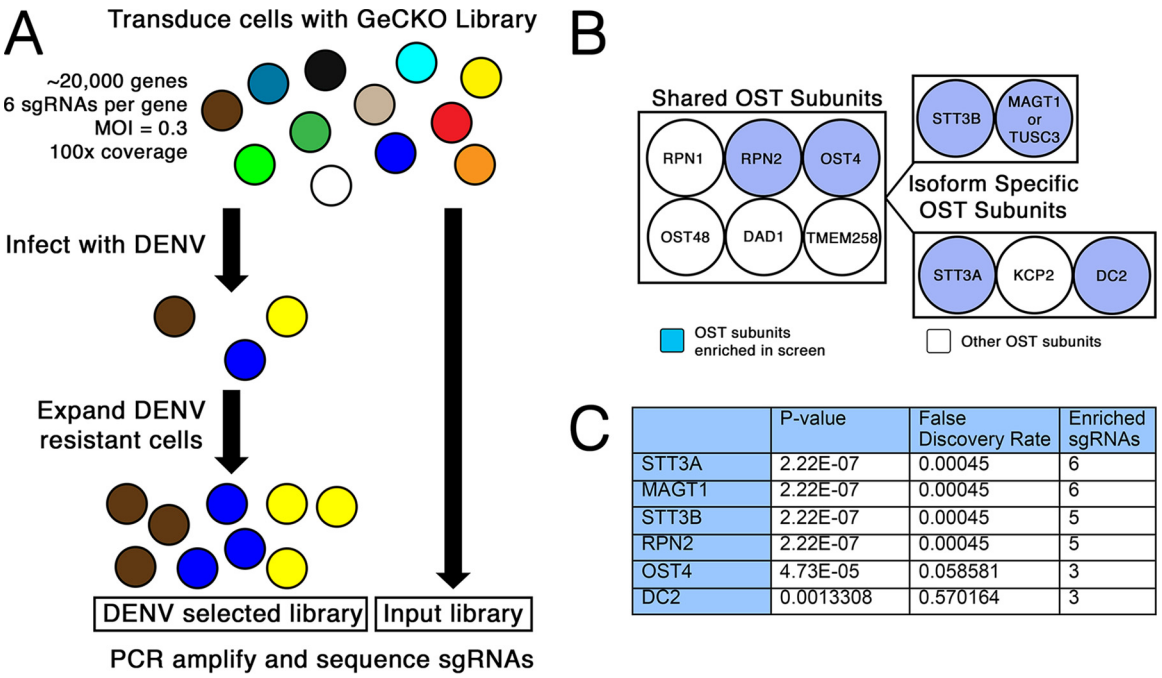
Both of the OST complexes share several noncatalytic subunits–RPN1, RPN2, OST4, OST48, DAD1, and TMEM258 (6). MAGT1 and its paralog TUSC3 are specific subunits of the STT3B complex that are important for complete glycosylation of STT3B target proteins. MAGT1 and TUSC3 are thioredoxin homologs that harbor a conserved CXXC motif important for oxidoreductase activity (8, 9). MAGT1 and TUSC3, through their CXXC active sites, may form mixed disulfides with cysteines of a target protein, delaying native disulfide bond formation, and granting STT3B access to target sequons poorly accessible to STT3A. Loss of MAGT1 and TUSC3, or abrogation of the CXXC motif, leads to hypoglycosylation of STT3B-specific substrates (8).

Marceau et al. demonstrated that DENV is dependent on both STT3A and STT3B and that the oligosaccharyltransferase activity of each of these subunits is not necessary for DENV replication (4). The authors proposed that the OST complexes act as scaffolds for DENV replication complexes to form. However, our data provide evidence that the STT3B-containing OST, through the MAGT1 subunit, provides a required catalytic activity for efficient DENV replication and not simply a scaffolding function.

In this work, a whole-genome clustered regularly interspaced short palindromic repeat (CRISPR) knockout screen was used to identify cellular factors required for DENV propagation. We confirm that both of the OST complexes are required for DENV replication and that the oligosaccharyltransferase activity of either the STT3A or STT3B OST is dispensable for DENV propagation. Importantly, however, knockout of *STT3B* leads to loss of MAGT1 expression, and *MAGT1* knockout is sufficient to block DENV propagation. Additionally, the CXXC catalytic oxidoreductase active site of MAGT1 or TUSC3 is required for DENV propagation. We provide evidence that DENV NS4B interacts with STT3B/MAGT1 OST complexes based on NS4B glycosylation and proximity labeling experiments and that NS4B synthesis is reduced in *STT3B* knockout cells. Together, these data suggest that STT3B serves to stabilize MAGT1 in the context of DENV infection and that the catalytic oxidoreductase activity of MAGT1 is required for DENV propagation, possibly through an effect on viral protein synthesis and/or folding.

RESULTS

A whole-genome CRISPR screen reveals that DENV is dependent on the oligosaccharyltransferase complexes. We employed a CRISPR-Cas9 pooled screening approach to identify host proteins necessary for DENV-mediated cell death in Huh7.5.1 human hepatoma cells (Fig. 1A). Among the top-ranked hits were multiple subunits of the host oligosaccharyltransferase (OST) complexes, including the two catalytic sub-



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	P-value	False Discovery Rate	Enriched sgRNAs
STT3A	2.22E-07	0.00045	6
MAGT1	2.22E-07	0.00045	6
STT3B	2.22E-07	0.00045	5
RPN2	2.22E-07	0.00045	5
OST4	4.73E-05	0.058581	3
DC2	0.0013308	0.570164	3

FIG 1 The CRISPR-Cas9 screen reveals the oligosaccharyltransferase complex as essential to DENV propagation. (A) Depiction of the whole-genome CRISPR screen selecting for knockout cells resistant to dengue virus infection. (B) Diagram of the two OST complexes that contain either STT3A or STT3B. Subunits of the OST complex shown in blue were significantly enriched in the screen. (C) Table of the hits from the screen showing the gene identifier (ID), *P* value, and false-discovery rate as calculated by MAGeCK analysis.

units STT3A and STT3B. Single-guide RNAs (sgRNAs) targeting genes encoding the two unique subunits of the STT3A and STT3B complexes, *DC2* and *MAGT1*, respectively, were also significantly enriched in the screen. In addition, two shared OST subunits, RPN2 and OST4, were significant hits. To validate three of the hits from the screen, we generated *STT3A*, *STT3B*, and *MAGT1* knockout Huh7.5.1 cells through stable transduction of the pLENTICRISPRv2 construct encoding both the *Streptococcus pyogenes* Cas9 protein and an sgRNA. The knockouts were confirmed by Western blotting. Importantly, in *STT3B* knockout cells, MAGT1 protein was also depleted (Fig. 2A), as its stability requires interaction with STT3B (8). We then infected these cells with a luciferase reporter dengue virus (luc-DENV), where luciferase activity directly correlates with viral propagation. Three days postinfection, we assessed luciferase activity and saw a significant and marked decrease in luciferase activity in *STT3A*, *STT3B*, and *MAGT1* knockout cells compared to control cells transduced with a control green fluorescent protein (GFP)-targeting sgRNA, demonstrating that protection from DENV-mediated cell death in these knockout cells is mediated by inhibition of DENV infection rather than by a block of cell death signaling pathways (Fig. 2A). The viability and growth of *STT3A*, *STT3B*, or *MAGT1* knockout cells were similar to those of control cells (Fig. 2E). These data validate the results from the screen and show that *STT3A*, *STT3B*, and *MAGT1* are all required for efficient DENV propagation.

Comparison of this screen's hits to two previously published whole-genome DENV screens, one using small interfering RNA (siRNA) knockdown and another using a pooled CRISPR knockout format (4, 5) (see Table S1 in the supplemental material), revealed high overlap with the CRISPR screen by Marceau et al. (4). Fifteen of the top 25 hits in our screen were identified in their screen (Table S1). The remarkable similarity between the two CRISPR screens demonstrates the technical reproducibility of pooled CRISPR screens for the identification of host dependency factors.

The OSTs are required to support efficient infection by Zika virus but not other flaviviruses. We next asked whether other arboviruses also depend on the same OST complexes and used flow cytometry to determine whether infection was impaired in STT3A or STT3B knockout cells. We infected cells with DENV serotype 2 (DENV-2), Zika

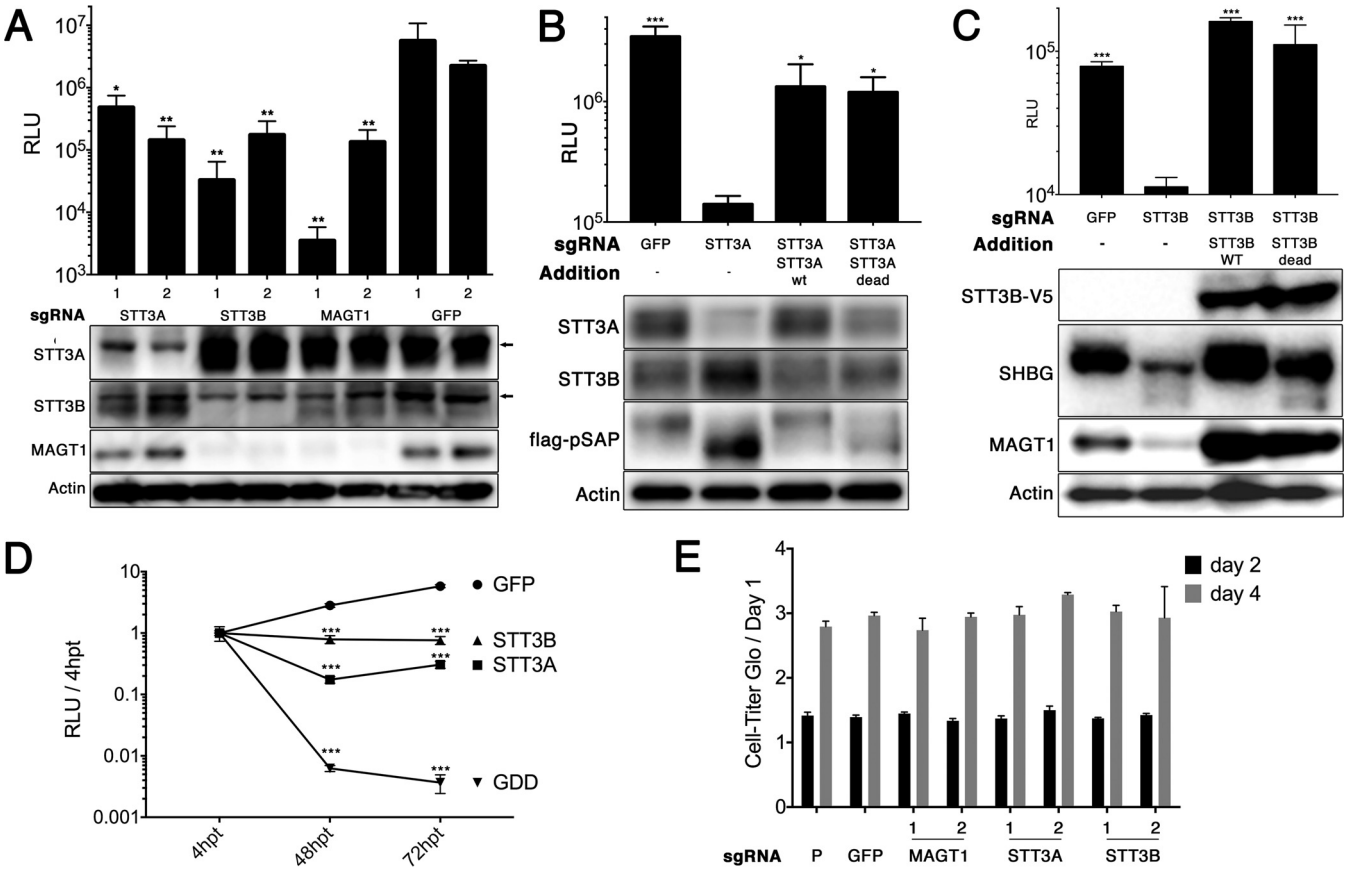

FIG 2 The catalytic activities of STT3A or STT3B are not required for DENV replication. (A) Huh7.5.1 cells were transduced with a lentivirus encoding a puromycin resistance marker, Cas9, and a single guide RNA (sgRNA) targeting the indicated genes. The sgRNAs targeting GFP act as a negative control. Two independent sgRNAs (indicated by 1 or 2 below the bars in the bar graph) were used per target gene to generate knockout cell pools. Cells were infected with luc-DENV for 3 days, and luciferase activity was measured in relative light units (RLU). Western blots for the indicated proteins are shown below the bar graph. Arrows indicate nonspecific bands. (B and C) Knockout cell pools were transduced with lentiviral vectors to rescue expression of the indicated protein. "STT3A dead" is a catalytically inactive W526A D527A double mutant. "STT3B dead" is a catalytically inactive W605A D606A double mutant with a C-terminal V5 tag. Transient transfection of constructs to express pSAP and SHBG were used to assess STT3A and STT3B catalytic activity. Western blots below the bar graph show the expression of the indicated proteins. wt and WT, wild type. (D) Knockout cell pools were transiently transfected with a luc-DENV replicon, and luciferase activity was measured at 4 h, 48 h, and 72 h posttransfection (hpt). Data are plotted as a ratio of RLU to the RLU at 4 hpt to control for transfection efficiency. GDD indicates a polymerase dead replicon with replacement of the GNN active site sequence in the NS5 RNA polymerase by GDD. (E) Cell viability for knockout cell pools was measured by Cell-Titer Glo over the course of 4 days for three independent wells. Data are normalized to day 1 values. The values for the parental Huh7.5.1 cells (P) are shown. In panels A to E, data are expressed as means plus standard deviations (SD) (error bars) for three independent biological replicates. In panels A to D, statistical significance was determined by Student's *t* test and indicated by asterisks as follows: *, $P < 0.05$; **, $P < 0.005$; ***, $P < 0.0005$. In panel A, the means were compared to the mean for GFP sgRNA 2. In panels B and C, the means were compared to the means of their respective knockouts. In panel D, the means were compared to the mean for the GFP control.

virus (ZIKV), West Nile virus (WNV), yellow fever virus (YFV), Sindbis virus (SINV), Venezuelan equine encephalitis virus (VEEV), or Chikungunya virus (CHIKV) and compared the percentage of infected cells in OST knockout cells compared to that in the control cells. Consistent with our other results, we found that DENV-2 infection was dramatically reduced in OST knockout cells compared to control cells (Fig. S1). We also found that ZIKV infection was moderately, but significantly, reduced in *STT3A* and *STT3B* knockout cells, indicating that the OSTs may also play a role during ZIKV infection (Fig. S1). However, we did not find a significant decrease in the propagation of the other viruses tested.

The catalytic oligosaccharyltransferase activity of the OST complexes is not required for DENV replication. In order to confirm the specificity of our knockout cell lines, sgRNA-resistant *STT3A* and *STT3B* were then introduced by lentiviral transduction into the corresponding knockout cell lines to rescue STT3A (Fig. 2B) or STT3B (Fig. 2C) expression. Importantly, we found that exogenous expression of STT3B in *STT3B* knockout cells led to restoration of endogenous MAGT1 expression (Fig. 2C). Exoge-

nous expression of STT3A or STT3B rescued luc-DENV infection in *STT3A* knockout or *STT3B* knockout cells, respectively (Fig. 2B and C). These data confirm that STT3A and STT3B are specifically required for efficient DENV propagation.

Both STT3A and STT3B are oligosaccharyltransferases necessary for the transfer of glycans to asparagines on target substrates (7). Mutation of a conserved WWDYG motif to WAAYG inactivates oligosaccharyltransferase activity (10). *STT3A* or *STT3B* knockout cells expressing catalytically dead STT3A (STT3A-dead) or STT3B (STT3B-dead) were able to support similar levels of DENV infection as their wild-type counterparts, demonstrating that the catalytic activity of STT3A or STT3B is not required for DENV propagation (Fig. 2B and C).

We also confirmed that these constructs were indeed catalytically inactive by transfecting the rescued cells with plasmids to express either prosaposin (pSAP) or sex hormone-binding globulin (SHBG), specific N-glycosylation substrates of STT3A and STT3B, respectively. By Western blotting, a majority of pSAP remained hypoglycosylated, as evidenced by a more rapidly migrating band on SDS-polyacrylamide gels from both STT3A knockout cells and STT3A-dead-expressing cells (Fig. 2B). Similarly, a fraction of SHBG appeared as a more rapidly migrating hypoglycosylated protein in both *STT3B* knockout cells and STT3B-dead-expressing cells (Fig. 2C). These results demonstrate that glycosylation of STT3A- and STT3B-specific substrates remains impaired in STT3A-dead and STT3B-dead-expressing cells.

We also transfected a luciferase-DENV replicon into knockout cells to assess whether the specific step of viral replication requires STT3A or STT3B. We found significant decreases in luciferase activity in *STT3A* and *STT3B* knockout cells compared to wild-type control at both 48 and 72 h posttransfection (Fig. 2D). These data confirm that both of the OST complexes are required for DENV replication, consistent with previous data (4). The viability of OST knockout cells was unchanged compared to controls (Fig. 2E).

Together, these data suggest that STT3B stabilizes MAGT1 and that the canonical oligosaccharyltransferase activity of STT3B-containing OST complexes is not required for DENV propagation. These results support a model where STT3B is required to stabilize MAGT1, which in turn has its own specific function to support DENV replication. Therefore, we explored whether the catalytic activity of MAGT1 is required for DENV replication.

The catalytic activities of TUSC3 or MAGT1 are necessary to support DENV propagation. Both MAGT1 and its paralog TUSC3 are thioredoxin homologs that harbor oxidoreductase activity through a conserved catalytic CXXC motif and interact with STT3B but not STT3A (8). TUSC3 contains a peptide-binding pocket that may interact with proteins in a sequence-specific manner (9). Cells lacking both MAGT1 and TUSC3 are unable to fully glycosylate STT3B-specific substrates (8). These data support a model where MAGT1 or TUSC3 serve to form a mixed disulfide with a protein substrate, granting STT3B access to glycosylation sites that may otherwise be inaccessible.

In some cell types, TUSC3 expression is upregulated in the absence of MAGT1 (8). Importantly, although TUSC3 has been shown to be expressed in HEK293 cells (8), we found that TUSC3 is not expressed in wild-type or *MAGT1* knockout Huh7.5.1 cells by immunoblotting (Fig. 3A) or by reverse transcription-PCR (RT-PCR) (not shown). Therefore, we asked whether TUSC3 is capable of functionally replacing MAGT1 in the context of DENV infection. We found that *MAGT1* knockout Huh7.5.1 cells transduced with a TUSC3-encoding lentiviral vector were able to support significantly increased DENV propagation compared to cells that did not express either MAGT1 or TUSC3 (Fig. 3A). These data demonstrate that the expression of either MAGT1 or TUSC3 is required for DENV propagation and that these two proteins are functionally redundant in the context of DENV propagation.

We next asked whether the enzymatic activity of MAGT1 or TUSC3 is required for DENV propagation. Expression of catalytically inactive AXXA mutants of MAGT1 or TUSC3 failed to rescue DENV propagation in *MAGT1* knockout Huh7.5.1 cells (Fig. 3A

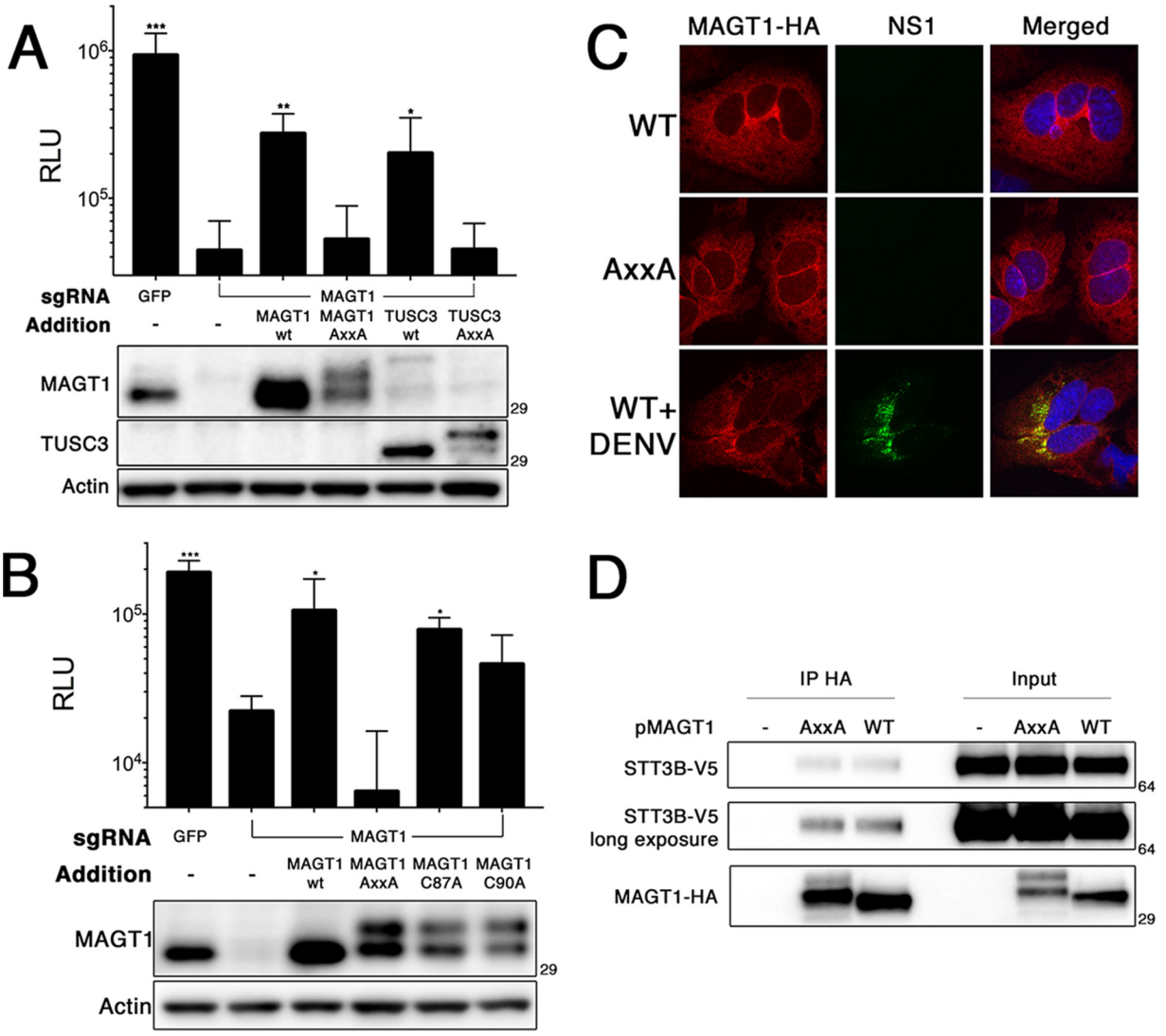

FIG 3 The oxidoreductase activity of MAGT1 is required for DENV replication. (A) CRISPR-modified Huh7.5.1 *MAGT1* knockout cell pools were transduced to express MAGT1 or TUSC3-HA. The AXXA MAGT1 and TUSC3-HA mutants were generated by mutating CXXC active sites to AXXA. (Bottom) Western blots for the indicated protein. Numbers to the right of the immunoblots in panels A, B, and D indicate molecular mass in kDa. The MAGT1 antibody cross-reacts weakly with TUSC3. (B) *MAGT1* knockout cells were transduced to express MAGT1 mutants with the indicated mutations. (Bottom) Western blots for the indicated proteins. For panels A and B, cells were infected with luc-DENV, and luciferase activity was measured at 3 days postinfection. (C) Immunofluorescence localization of MAGT1 in uninfected cells or 2 days postinfection with DENV. *MAGT1* knockout cells were stably transduced to express HA-tagged MAGT1 (wild type [WT] or mutant [AXXA]) as indicated to the left. Cells were fixed, permeabilized, and stained with antibodies to the indicated proteins. (D) 293T cells expressing the indicated MAGT1-HA mutants were transfected to express STT3B-V5. Cells were lysed, and coimmunoprecipitation was carried out using an anti-HA antibody (IP HA). Western blots detecting the specified proteins are shown. Mutants of MAGT1 migrate as two distinct bands. Data in panels A and B are expressed as means plus SD for three independent infections. Values that were statistically significant different from the values for *MAGT1* knockout cells without any additions (second bar from the left) were determined by Student's *t* test and indicated by asterisks as follows: *, $P < 0.05$; **, $P < 0.005$; ***, $P < 0.0005$.

and B), indicating that the oxidoreductase activity of MAGT1 or TUSC3 is required for DENV replication.

We also investigated whether MAGT1 knockout cells expressing single-cysteine active site MAGT1 mutants (CXXA and AXXC) are able to support DENV replication. Some single-cysteine mutants of ER resident oxidoreductases, such as protein disulfide isomerase (PDI), have been shown to retain oxidoreductase activity (11, 12), and single-cysteine MAGT1 mutants can form mixed disulfides with target proteins, demonstrating that they are reactive in cells (8). Interestingly, *MAGT1* knockout cells expressing a single-cysteine MAGT1 mutant were able to support increased levels of DENV replication compared to knockout cells, albeit at levels lower than with wild-type

MAGT1 rescue (Fig. 3B). This indicates that single-cysteine MAGT1 mutants retain significant catalytic activity to support DENV infection. Together, these data demonstrate that the oxidoreductase activity of MAGT1 is required for DENV propagation and that cells expressing single-cysteine MAGT1 are still able to support DENV replication.

MAGT1 partially localizes to DENV replication compartments. We next performed immunofluorescence microscopy on cells expressing a hemagglutinin (HA)-tagged MAGT1 construct (MAGT1-HA) to visualize the localization of MAGT1 during DENV infection. In uninfected Huh-7 cells, both wild-type MAGT1 and MAGT1 mutants were distributed in an intracellular reticular pattern consistent with ER localization (Fig. 3C). This supports the hypothesis that MAGT1 is an ER-localized component of the OST complex, rather than the proposal that MAGT1 contributes to magnesium import at the plasma membrane (8, 13). We also found that the distribution of the MAGT1-AxxA mutant was similar to that of wild-type MAGT1, demonstrating that the inability of cells expressing MAGT1-AxxA to support DENV replication is not due to defects in subcellular localization. DENV infection was associated with partial colocalization of MAGT1 with viral NS1 protein (Fig. 3C). Additionally, coimmunoprecipitation assays demonstrated MAGT1-AxxA interaction with STT3B, indicating that MAGT1-AxxA's inability to support DENV replication is not due to a defect in association with the OST complex (Fig. 3D). These data are consistent with previous data showing that OST localized to sites of DENV-induced vesicle packets and replication compartments (4).

NS1 is not detectably different in STT3A, STT3B, and MAGT1 knockout cells. We hypothesize that MAGT1 functions as an oxidoreductase, potentially affecting the folding of a DENV nonstructural protein. We asked which DENV proteins could be candidate substrates for MAGT1 oxidoreductase activity. All four DENV serotypes are impaired in their ability to propagate in *STT3B* knockout cells (4), which based on our data must also lack MAGT1 expression. Therefore, we hypothesized that a viral target of MAGT1 oxidoreductase activity should have cysteines exposed to the ER lumen that are conserved across all four serotypes.

Four of the seven nonstructural proteins have cysteine residues predicted to be exposed to the ER lumen: NS1, NS2A, NS4A, and NS4B. Of these proteins, only NS1 and NS4B have cysteines that are conserved across all four DENV serotypes. The DENV protein NS1 forms a dimer in the ER, is secreted as a soluble hexamer, and contains twelve cysteines, six disulfide bonds, and two N-glycosylation sites (14). The DENV protein NS4B contains three cysteines and two N-glycosylation sites (15).

We first investigated whether we could detect any changes in NS1 properties by immunoblotting of NS1 expressed in knockout and wild-type cells. We were unable to detect any changes in NS1 dimerization or glycosylation in *STT3A* or *STT3B* knockout cells (Fig. S2A). We also found that the pattern of NS1 glycosylation was unchanged after performing pulse-chase experiments in *STT3A*, *STT3B*, or *MAGT1* and *TUSC3* knockout cells (Fig. S2B). We were also unable to detect any interaction between NS1 and STT3B or MAGT1 by coimmunoprecipitation (not shown). In summary, these data suggest that glycosylation of NS1 is unaffected by the loss of STT3A, STT3B, or MAGT1 and that NS1 is not a target of MAGT1 oxidoreductase activity.

NS4B synthesis is altered in STT3B knockout cells. We next sought to find differences in NS4B exogenously expressed in OST subunit knockout cells by generating tagged constructs expressing C-terminally tagged DENV 2k-NS4B-HA (pNS4B-HA) or 2k-NS4B-V5 (pNS4B-V5). NS4B has been proposed to be glycosylated at two residues, N58 and N62 (15). We found that in 293T cells a fraction of transiently transfected NS4B-HA appeared as a series of more slowly migrating bands, suggestive of possible glycosylation (Fig. 4A). Notably, the pattern of higher-molecular-weight NS4B-HA species was altered in *STT3B* knockout cells, with decreased levels of higher-molecular-weight HA-reactive bands. Additionally, following transient expression of NS4B in Huh7.5.1 cells, we found three specific peptide-*N*-glycosidase F (PNGase F)-sensitive bands that migrated at a higher molecular weight than unglycosylated NS4B, confirming that NS4B can be partially glycosylated under our experimental conditions (Fig. 4B).

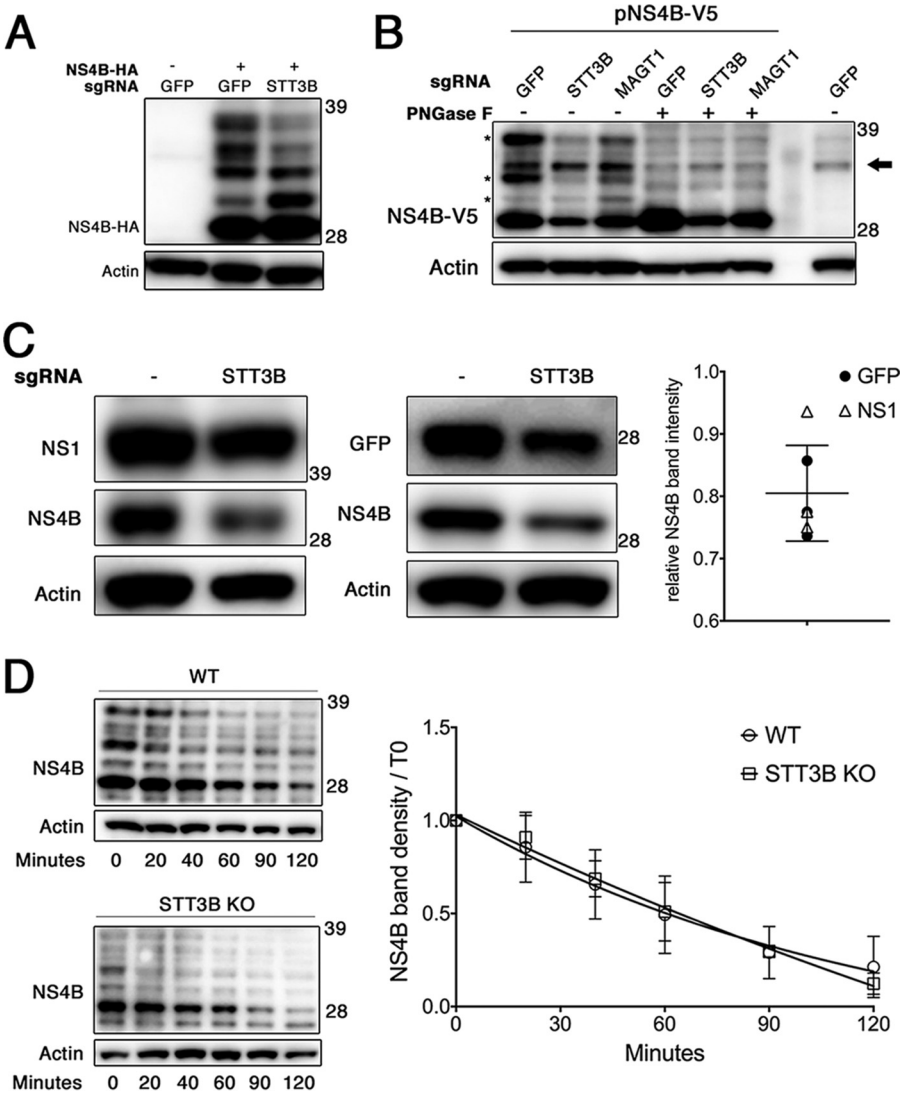

FIG 4 Effect of STT3B on NS4B glycosylation and synthesis. (A) 293T cell pools expressing Cas9 and sgRNAs targeting the indicated genes were transiently transfected to express NS4B-HA. Lysates were resolved by SDS-PAGE and visualized by Western blotting for the indicated proteins. Numbers to the right of the immunoblots in all panels indicate molecular mass in kDa. (B) Huh7.5.1 cell pools expressing Cas9 and sgRNAs targeting the indicated genes were transiently transfected to express NS4B-V5 (six leftmost lanes) or were mock transfected (rightmost lane). The lysates were subjected to PNGase F treatment (+) in the middle lanes to remove N-linked glycans from proteins. Proteins were resolved by SDS-PAGE and visualized by Western blotting for the indicated proteins. The black arrow to the right of the gel indicates a nonspecific background band, and asterisks to the left of the gel mark NS4B-specific, PNGase F-sensitive bands. (C) The indicated 293T cells were cotransfected to express NS4B-HA and either NS1 or GFP (as transfection controls). Cell lysates were treated with PNGase F to facilitate quantitation of NS4B protein by removing N-glycans, and Western blotting was performed for the indicated proteins. Chemiluminescent band intensities were quantified as NS4B-HA in *STT3B* knockout cells relative to NS4B-HA in WT cells compared to the same ratio of GFP or NS1 to control for transfection efficiency. Each point plotted is the quantification of an independent transfection, where the open triangles compare NS4B levels with NS1 and the closed circles compare NS4B levels with GFP. The mean and SD plotted are for all points. (D) The indicated 293T cells were transiently transfected to express NS4B-HA and treated with cycloheximide at 24 h posttransfection to block further translation. Cells were lysed at the indicated times after cycloheximide treatment, and proteins were resolved by Western blotting. (Left) Representative blots. (Right) Decay curves plotted with data from three independent experiments. Values are means ± SD. KO, knockout.

Together these data show that NS4B can be glycosylated and that this is mediated at least in part by STT3B. Although glycosylation of NS4B is not necessary for viral replication, as catalytically inactive STT3B rescues DENV replication in *STT3B* knockout cells, this finding suggests that NS4B physically interacts with the STT3B OST complex in the ER lumen.

We noticed a consistent reduction in NS4B protein levels in *STT3B* and *MAGT1* knockout cells and hypothesized that NS4B protein folding may be affected by the loss of STT3B or MAGT1. To measure the steady-state levels of NS4B in *STT3B* knockout cells, we cotransfected a plasmid expressing either NS1 or GFP to control for transfection efficiency and quantitated chemiluminescent band intensities to measure the relative levels of NS4B synthesis in *STT3B* knockout cells. We found a consistent and statistically significant ($P < 0.0001$) ~20% decrease in NS4B band intensity in *STT3B* knockout cells compared to wild-type controls relative to the transfection control (Fig. 4C). Despite this difference in NS4B steady-state levels, we did not find a difference in the half-life of NS4B in *STT3B* knockout cells compared to the wild type (Fig. 4D), suggesting that the rate of synthesis, and not degradation, of NS4B is altered in *STT3B* knockout cells.

Together, these data show that NS4B is an STT3B substrate that requires MAGT1 to be efficiently glycosylated. Furthermore, NS4B synthesis appears to be reduced by the loss of STT3B/MAGT1. Taken together, although NS4B glycosylation is not necessary for DENV infection, these findings suggest that NS4B physically interacts with STT3B/MAGT1 OST complexes. Furthermore, they suggest that one mechanism by which MAGT1 facilitates DENV propagation may be through promoting efficient synthesis of NS4B.

Close proximity of MAGT1 to NS4B and NS1 during DENV infection. Although our data so far suggested a possible interaction between STT3B/MAGT1 and NS4B, using conventional immunoprecipitation methods, we were unable to demonstrate a stable interaction between MAGT1 and NS4B (not shown). Therefore, we used the engineered peroxidase APEX2 to label proteins in close proximity to MAGT1 during DENV infection. APEX2 catalyzes the biotinylation of proteins within a 5- to 10-nm radius in the presence of biotin-phenol and hydrogen peroxide (16). We generated a construct in which APEX2 was inserted between the signal sequence and MAGT1, placing APEX2 in the ER lumen near the MAGT1 active site. We stably transduced *MAGT1* knockout cells to express the APEX2-MAGT1 fusion protein and induced APEX2-mediated biotinylation in DENV-infected cells. After lysis of the cells, we performed affinity purification of biotinylated proteins using streptavidin beads followed by SDS-PAGE and immunoblotting.

We found that both NS1 and NS4B were specifically biotinylated by APEX2-MAGT1 in DENV-infected cells (Fig. 5). To assess the specificity of APEX2 labeling, we probed for biotinylation of STT3A, which also associates with sites of DENV replication but does not interact with MAGT1 (4, 8), and found that STT3A is not biotinylated by APEX2-MAGT1 under these conditions (Fig. 5). Similarly, the integral ER membrane protein VAPA (vesicle-associated membrane protein-associated protein A) is not biotinylated by APEX2-MAGT1. On the other hand, EMC3, another ER protein necessary for flavivirus propagation that was a hit in our screen and interacts with the OST complex (5), was biotinylated. These data are consistent with the model that MAGT1 likely resides at sites of DENV replication and either associates with or is in close physical proximity to DENV nonstructural proteins.

The redox status of NS4B is unchanged in the absence of MAGT1. We hypothesized that the NS4B cysteine redox state is modulated by MAGT1. To test this, we used Western blots to examine changes in NS4B migration on SDS-polyacrylamide gels after treating cell lysates with methoxypolyethylene glycol maleimide 5000 (mPEG), which covalently bonds to free reduced cysteine residues. The mPEG reagent has an average molecular mass of 5 kDa, resulting in a shift in the apparent molecular mass of modified proteins visualized by Western blotting. DENV NS4B has three conserved cysteines that may participate in disulfide bonds. For a negative control, we lysed cells in the presence of *N*-ethylmaleimide (NEM), which covalently blocks all reduced cysteines, rendering them mPEG nonreactive (Fig. S3, lanes 1 and 2). For a positive control, we treated lysates with Tris(2-carboxyethyl)phosphine hydrochloride (TCEP), which reduces all cysteines, allowing subsequent mPEG modification of all cysteines (Fig. S3, lane 5).

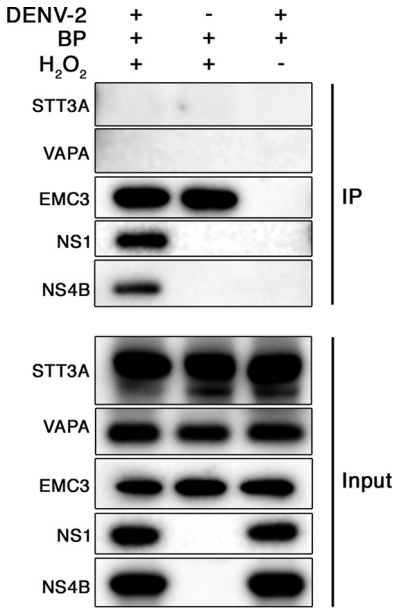

FIG 5 MAGT1 is in close proximity to NS4B and NS1 during DENV infection. Huh7.5.1 cells stably transduced to express APEX2-MAGT1 were infected with DENV-2 or mock infected for 2 days. Under all conditions, biotin-phenol (BP) was added to the cell media for 30 min. Where indicated, APEX2 was activated with hydrogen peroxide for 1 min to biotinylate APEX2-MAGT1-proximal proteins prior to quenching and lysis. The rightmost lane is a negative control where APEX2 was not activated. Biotinylated proteins were immunoprecipitated with streptavidin beads, and lysates were resolved by SDS-PAGE. Western blotting was performed for the indicated proteins for both the immunoprecipitated (IP) samples and the input.

mPEG-modified NS4B, when expressed in isolation, migrates the same in both wild-type and *STT3B* knockout cells, indicating that MAGT1 does not affect the cysteine redox state of NS4B (Fig. S3, lanes 3 and 4). Additionally, a fraction of mPEG-modified NS4B (lanes 3 and 4) runs at the same apparent molecular weight as fully reduced TCEP-treated NS4B treated with mPEG (lane 5), indicating that at steady state, a fraction of NS4B molecules contains fully reduced cysteines. However, another fraction of NS4B is modified by fewer mPEG molecules (lanes 3 and 4), demonstrating that some NS4B molecules may contain oxidized cysteines in the form of intra- or intermolecular disulfide bonds.

While our results indicate that MAGT1 does not modulate NS4B cysteine redox state when expressed in isolation, they do not rule out the possibility that MAGT1 modulates NS4B cysteine redox status transiently and/or in the context of authentic viral infection. Further experiments will have to be carried out to determine whether NS4B disulfide bonds exist during infection.

DISCUSSION

Our results demonstrate the high reproducibility of whole-genome CRISPR screens compared to the low overlap among hits from siRNA-based screens for host cofactors of viral infection. We compared the top hits from our CRISPR screen to those recently published and found that 15 of the top 25 hits from our screen were in the top 1% of the others (4). In contrast, three independent siRNA screens for host dependency factors of DENV infection performed by the same group identified more than 150 high-confidence hits; however, only 14% of hits from one independent siRNA library overlapped with hits from another siRNA library, even when performed by the same investigators using the same infection conditions (5). This is consistent with meta-analyses demonstrating the low reproducibility of siRNA screens for viral host factors (17).

While there is a high degree of overlap between CRISPR screens for DENV, the

number of significant hits is relatively low compared to those obtained from RNA interference (RNAi) screens. For example, the pooled results of the three siRNA screens for DENV yielded hundreds of hits, whereas our CRISPR screen yielded fewer than 50 significant hits. This may be due in part to some DENV host factors also being necessary for cellular survival or growth. Additionally, using cell survival from lethal viral challenge as a readout is highly stringent and will increase the false-negative rate, as only partial suppression of DENV infection may not be sufficient to protect cells from death. Thus, experimental readouts that can capture intermediate phenotypes are likely to reveal additional host dependency factors for DENV infection.

The events at the endoplasmic reticulum that direct the formation of a functional dengue virus replication organelle remain incompletely understood. In this study, a whole-genome CRISPR screen reveals a noncanonical function of the OST complex that serves to support DENV infection. STT3B, the subunit of the OST complex that provides oligosaccharyltransferase activity, is required to stabilize MAGT1 to support DENV replication. We show that MAGT1, or its homolog TUSC3, provides a catalytic oxidoreductase activity necessary for DENV to replicate.

On the other hand, we find that the N-glycosylation activity of STT3A or STT3B is dispensable for DENV propagation, in agreement with previous findings by Marceau et al. (4). On the basis of this observation, these authors suggested that the OST complexes serve as structural scaffolds for DENV to replicate. However, it is unclear why both STT3A- and STT3B-containing OST complexes would be required to serve a scaffolding function for DENV replication. Our data show that STT3B-containing complexes serve more than a structural scaffolding function to support DENV replication, and in fact contribute a direct catalytic activity for DENV replication, namely, the oxidoreductase activity of MAGT1. We propose that the dependency of DENV replication on STT3B without requiring its oligosaccharyltransferase activity is explained by the loss of MAGT1 expression in the absence of STT3B.

Analogous to the dependence of MAGT1 on STT3B for stable expression, some subunits of the STT3A complex require STT3A for stable expression (18). We propose that there may be a specific function granted by the STT3A complex subunits DC2/OSTC or KCP2 that is necessary for DENV replication. In support of this hypothesis, DC2 was a hit in both CRISPR screens (4). However, the precise cellular function of DC2 is unknown.

We have shown that the CXXC catalytic motif of MAGT1 or TUSC3 is required for DENV propagation and that TUSC3 can functionally replace MAGT1 in the context of DENV infection. Some cell types, including HEK293 cells, express both TUSC3 and MAGT1, while others, including hepatocytes and Huh7.5.1 hepatoma cells, express only MAGT1 (13). Importantly, TUSC3 expression is upregulated in HEK293 cells when *MAGT1* is knocked out (19). Thus, essential but functionally redundant functions of MAGT1 and TUSC3 in DENV replication will not be revealed in cell types capable of expressing both proteins.

We carried out several experiments directed at the hypothesis that viral NS1 and/or NS4B is a substrate of MAGT1, as these viral proteins harbor multiple cysteines conserved across all DENV serotypes that are also predicted to be accessible to MAGT1. We were unable to find any differences in NS1 dimerization or glycosylation in MAGT1 knockout cells compared to wild-type cells, suggesting that NS1 is correctly folded and processed in the absence of MAGT1. In addition, we probed the disulfide status of NS4B expressed in isolation and found no apparent differences in NS4B cysteine accessibility to mPEG modification in the absence or presence of MAGT1 activity. Typically, both cysteines in the CXXC active site motif of an oxidoreductase are involved in catalyzing disulfide bond formation in a target protein. Surprisingly, *MAGT1* knockout cells expressing CXXA or AXXC active site mutants of MAGT1 can support DENV propagation. Single-cysteine active site mutants of protein disulfide isomerase (PDI) have been shown to retain partial reductase activity *in vitro* and are still able to shuffle disulfide bonds and mediate native protein folding (11, 12). Previous studies have shown that a single-cysteine MAGT1 mutant exhibits partial catalytic activity when assessing glyco-

sylation of an STT3B substrate (8). As depicted in Fig. S4 in the supplemental material, a single-cysteine reductase can act as a disulfide isomerase by attacking a nonnative disulfide bond on a target protein, generating a mixed disulfide between the reductase (here MAGT1) and its target. An alternate cysteine from the target protein then resolves the mixed disulfide, forming the native disulfide bond and releasing the target protein from MAGT1. Single-cysteine MAGT1 could therefore serve as a disulfide isomerase in the context of DENV infection. Alternatively, single-cysteine MAGT1 could function in tandem with another ER-resident oxidoreductase to resolve the mixed disulfide and generate a correctly folded target substrate. However, this model is not easily reconciled with the observation that either single-cysteine or wild-type MAGT1 is capable of supporting DENV replication, because the majority of wild-type MAGT1 appears to be oxidized in cells, without reduced cysteines to attack disulfide bonds (8). One possibility is that the minor fraction of reduced MAGT1 or TUSC3 is sufficient to support DENV replication.

If MAGT1 does not directly catalyze disulfide bond rearrangement in a DENV protein such as NS4B, another possibility is that MAGT1, through its oxidoreductase activity, recruits a DENV protein to another cellular protein or complex. For example, the OST complex has recently been identified to associate with proteins in the ER membrane complex (EMC), which has been reported to act as a chaperone for multipass transmembrane proteins, of which NS4B is an example (20, 21). In this model, MAGT1, through its oxidoreductase activity, might transiently interact with NS4B, recruiting it to the EMC as an NS4B chaperone. Although we were unable to demonstrate a stable interaction between MAGT1 and DENV nonstructural proteins by coimmunoprecipitation, likely due to the transient association between oxidoreductases and their target substrates (22), APEX2-MAGT1 induced biotinylation of both NS4B and NS1 in DENV-infected cells, indicating that MAGT1 interacts with, or is at least in close proximity to, components of the DENV replication organelle in the ER.

Consistent with the possibility that MAGT1 might help to recruit dengue virus nonstructural proteins to the EMC complex, the rate of NS4B synthesis appears to be reduced in cells lacking MAGT1 and STT3B. While we see only relatively modest reductions in NS4B expression in cells lacking STT3B and MAGT1, one caveat is that NS4B is expressed in isolation in our system. It is possible that loss of MAGT1 may have more detrimental effects on NS4B expression (or the expression of other DENV proteins) in the context of authentic DENV infection. However, the strong inhibition of DENV infection in OST knockout cells prevents accurate assessment of protein expression.

We also found that ZIKV infection is significantly reduced in *STT3A* or *STT3B* knockout cells, suggesting that ZIKV may also require the OST complex for efficient infection or replication. However, the other flaviviruses tested did not appear to be dependent on OST complex function, despite the common dependence of most flaviviruses on the host cell ER for replication organelle formation. Interestingly, the NS4B cysteines are not conserved among flaviviruses despite being conserved across all DENV serotypes.

In conclusion, our data demonstrate that the STT3B-containing OST complex serves a catalytic and noncanonical function during DENV replication, and not merely a scaffolding function. We hypothesize that this noncanonical oxidoreductase activity of MAGT1 acts on a DENV nonstructural protein, such as NS4B, to mediate efficient synthesis, folding, and/or recruitment of nonstructural proteins to specific sites in the ER. Our results improve our understanding of the cell biology of DENV infection, and also may potentially guide the development of DENV antivirals that target MAGT1.

MATERIALS AND METHODS

Pooled CRISPR screen. The human GeCKOv2 plasmid library was a gift from Feng Zhang, acquired from Addgene. Vesicular stomatitis virus glycoprotein G (VSV-G)-pseudotyped GeCKOv2 lentiviruses were generated at the University of Michigan vector core. In brief, 16 million Huh-7 cells were transduced with the GeCKOv2 A or B lentiviral half-library at a multiplicity of infection (MOI) of 0.3 in 10-cm dishes. The cells were selected for 6 days posttransduction with 2 μg/ml puromycin, then infected with DENV-2 16681 at an MOI of 0.1 for 2 weeks. Genomic DNA from surviving cells was harvested using a Quick-gDNA

Midi kit (Zymo Research, Irvine, CA). The integrated sgRNAs were amplified using PCR, and Illumina adapters and barcodes were subsequently added by PCR as previously described (23). Sequencing was performed at the University of Michigan sequencing core on a MiSeq instrument (Illumina, San Diego, CA), and data were analyzed using MAGeCK (24).

Plasmids and lentiviral transduction. Individual sgRNAs targeting *STT3A*, *STT3B*, and *MAGT1* were generated in pLENTICRISPRv2 as previously described using the CRISPR RNA (crRNA) sequences listed in Table S2 in the supplemental material. The lentiCRISPR-EGFP (lenti stands for lentivirus, and EGFP stands for enhanced green fluorescent protein) sgRNA 1 was a gift from Feng Zhang (AddGene plasmid 51760).

The cDNA clone for *STT3A* was purchased from OriGene (RC201991; Rockville, MD). The cDNA clones for *STT3B* and *MAGT1* were described previously (19). The *TUSC3* gene was cloned by PCR from 293T cDNA. The sgRNA-resistant *STT3A*, *STT3B*, and *MAGT1* constructs were made using overlap extension PCR to introduce silent mutations either modifying the CRISPR protospacer adjacent motif or sgRNA base pair complementarity. Constructs encoding catalytically inactive *STT3B*, *STT3B*, *MAGT1*, and *TUSC3* were generated using overlap extension PCR. These constructs were cloned into the lentiviral expression vector pSMPUW (Cell Biolabs, San Diego, CA). The constructs for expression of pSAP and SHBG have been previously described (25, 26). The pNS1-flag, pNS1-NS2A-V5, and pNS4B-HA constructs were generated by PCR using pD2/IC-30P-NBX as the template (27). Detailed descriptions of construction of these plasmids are available upon request.

Lentiviral expression constructs were used to generate VSV-G-pseudotyped lentiviruses and for stable transduction of target cells as previously described (28). Knockouts and expression were confirmed by Western blotting.

Viruses. The infectious cDNA clone pD2/IC-30P-NBX encoding dengue virus serotype 2 strain 16681 was used to generate full-length viral RNA and for construction of a reporter virus and replicon (27). In brief, the luciferase reporter virus luc-DENV was generated by overlap extension PCR, by fusion of a *Renilla* luciferase (Rluc) with C-terminal self-cleaving 2A peptide to the DENV capsid in a pD2/IC-30P-NBX background. The luciferase reporter replicon was generated as previously described (29). To generate wild-type DENV-2 virus, the pD2/IC-30P-NBX plasmid was linearized with XbaI (New England Biolabs, Ipswich, MA), *in vitro* transcribed, and capped with the $m^7G(5')ppp(5')A$ cap analog (New England Biolabs) using T7 Megascript (Thermo Fisher Scientific, Waltham, MA). This RNA was transfected into Vero cells using TransIT mRNA reagent (Mirus Bio, Madison, WI). One week posttransfection, supernatant from transfected cells was 0.45 μm filtered and buffered with 20 mM HEPES. Cells were infected by replacing media on the cells with viral supernatant for 4 h at 37C and 5% CO_2. Afterward, unbound virus was aspirated and replaced with fresh media. For luciferase reporter assays, cells were infected with the luciferase reporter virus luc-DENV at an MOI of 0.1 for 2 or 3 days, and luciferase activity was measured with the *Renilla* luciferase assay system (Promega, Madison, WI) and a Synergy 2 plate reader (BioTek, Winooski, VT).

The recombinant infectious viruses used were as follows: SINV-GFP, generated from pTE/5′2J/GFP (30), YFV-Venus, generated from pYF17D-5′C25Venus2AUbi (31), WNV-GFP, generated from pBELO-WNV-GFP-RZ ic (32), CHIKV-GFP generated from pCHIKV-LR 5′GFP (33), VEEV-GFP, a TC83 vaccine strain derivative generated from pVEEV/GFP (34), and DENV2-GFP, a 16681 strain derivative generated from pDENV2-ICP30P-A-EGFP-P2AUb (35). Viral stocks were generated by electroporation of *in vitro*-transcribed RNA into WHO Vero cells (for DENV2-GFP) and BHK-21 cells (for SINV, YFV, WNV, CHIKV, and VEEV). Zika virus (ZIKV), 2015 Puerto Rican PRVABC59 strain (36), was obtained from the CDC and passaged twice in Huh-7.5 cells.

The multiplicity of infection was based on titers obtained on BHK-J cells for SINV, WNV, YFV, and VEEV infections and on Huh-7.5 cells for ZIKV. Titers were not available for YFV, CHIKV, and DENV2; therefore, different dilutions of viral stocks were tested.

Cells were seeded in a 24-well plate at 5×10^4 cells/well for ZIKV or at 1×10^5 cells/well for other viruses. The next day, cells were infected for 90 min at 37°C in phosphate-buffered saline containing 2% fetal bovine serum (2% FBS–PBS) using an MOI of 1 for SINV, an MOI of 0.01 and 0.001 for WNV, and an MOI of 0.8 for VEEV. We used a 1:4 dilution for YFV, a 1:10,000 dilution for CHIKV and a 1:5 dilution for DENV-2. Virus inoculum was removed, fresh complete medium was added to the cells, and infections were allowed to proceed for 10 h for SINV and VEEV, 24 h for CHIKV, 33 h for YFV, 48 h and 72 h for WNV, 58 h for DENV-2, and 48 h and 96 h for ZIKV. Experiments with WNV and CHIKV were carried out under biosafety level 3 containment. Infected cells were detached using Accumax cell aggregate dissociation medium (eBioscience). The cells were pelleted, fixed in 2% paraformaldehyde, and permeabilized using Cytofix/Cytoperm (BD Biosciences). For ZIKV-infected cells, E-protein expression was detected with the 4G2 monoclonal antibody (1:500 dilution), followed by incubation with Alexa Fluor 488-conjugated anti-mouse IgG antibody (Invitrogen) at 1:1,000 dilution. All samples were resuspended in 2% FBS–PBS. Fluorescence was monitored by fluorescence-activated cell sorting (FACS) using an LSRII flow cytometer (BD Biosciences). Data were analyzed with FlowJo software.

Antibodies. Western blotting was performed using antibodies against STT3A (12034-1-AP), STT3B (15323-1-AP), MAGT1 (17430-1-AP), and prosaposin (10801-1-AP) from the Proteintech Group (Rosemont, IL). Antibodies against TUSC3 (SAB4503183) and actin (A5316) were purchased from Sigma-Aldrich (St. Louis, MO). The antibody against EMC3 (sc-365903) was purchased from Santa Cruz Biotechnology (Dallas, TX). The antibody against SHBG (MAB2656) was purchased from R&D Systems (Minneapolis, MN). The antibody against HA (C29F4) was acquired from Cell Signaling Technology (Danvers, MA). The anti-V5 antibody (R960-25) was purchased from Thermo Fisher Scientific. The anti-NS1 monoclonal antibody 1F11 was a gift from P. Malasit at the National Center for Genetic Engineering and Biotechnology in Thailand.

Western blotting. Cells were lysed in a buffer containing 20 mM Tris (pH 7.5), 100 mM NaCl, 1% NP-40, 10% glycerol, and 1 mM EDTA with the addition of Halt protease inhibitor (Thermo Fisher). Lysates were cleared by centrifugation at 10,000 rpm for 10 min at 4°C. Lithium dodecyl sulfate (LDS) sample buffer was added, and lysates were resolved by SDS-PAGE on 4 to 12% bis-Tris NuPAGE Novex gels. The proteins were transferred to polyvinylidene difluoride (PVDF) membranes, which were then blocked for 30 min in 5% nonfat dry milk in Tris-buffered saline with Tween 20 (TBST). The following antibodies were added at the indicated dilutions (dilutions in parentheses): STT3A (1:1,000), STT3B (1:500), MAGT1 (1:1,000), TUSC3 (1:1,000), β-actin (1:20,000), HA (1:1,000), V5 (1:5,000), and NS1 (1:100). The membranes were incubated with primary antibody overnight at 4°C and then subsequently washed three times with TBST for 15 min each time. The membranes were then incubated in blocking buffer with 1:500 of secondary horseradish peroxidase (HRP)-conjugated antibodies (Thermo Fisher) for 1 h at room temperature. The blots were then washed, and proteins were detected by the addition of SuperSignal West Femto substrate and immediate visualization on a LI-COR (Lincoln, NE) imager or by X-ray film.

APEX labeling and immunoprecipitation. The APEX2-MAGT1 fusion construct was generated by overlap extension PCR. APEX2-mediated biotinylation and enrichment of biotinylated proteins were performed as previously described (37). A modification to the protocol was the addition of 20 mM *N*-ethylmaleimide (NEM) (Sigma-Aldrich) to the lysis buffer to preserve native disulfide bonds. Portions (10%) of the lysates were reserved to be run as input controls. One microgram of either V5 or HA antibody was added to immunoprecipitate tagged proteins. Mixtures were rotated for 2 h at 4°C. Five microliters of Dynabeads protein G (Thermo Fisher) was added to each tube and rotated for 1 h at 4°C. Complexes were washed three times in 1× PBS with 0.1% Triton X-100. Proteins were eluted in 1× LDS sample buffer with 50 mM Tris(2-carboxyethyl)phosphine hydrochloride (TCEP) (Sigma-Aldrich). Lysates were then subjected to SDS-PAGE and Western blotting as described above.

Pulse-chase analysis of NS1 glycosylation. HEK293 cells were grown to 60% confluence in 60-mm dishes and transfected with 6 μg of NS1-FLAG expression plasmids using Lipofectamine 2000 (Thermo Fisher) in Opti-MEM (Thermo Fisher) according to the manufacturer's instruction. After 24 h, NS1 substrates were labeled with Tran^{35}S label (PerkinElmer, Waltham, MA) by incubation in methionine- and cysteine-free medium containing 10% dialyzed FBS for 20 min before the addition of 200 μCi/ml of Tran^{35}S label. The cells were labeled for 5 min, then 3.7 mM unlabeled methionine and 0.75 mM unlabeled cysteine were added, and the cells were incubated for an additional 20 min. The cells were lysed at 4°C by a 30-min incubation with 1 ml of radioimmunoprecipitation assay (RIPA) lysis buffer. Lysates were clarified by centrifugation (2 min at 13,000 rpm) and precleared by incubation for 2 h with rabbit IgG and a mixture of protein A/G Sepharose beads (Santa Cruz Biotechnology) before an overnight incubation with anti-FLAG antibody (Sigma-Aldrich). Immunoprecipitates were collected with protein A/G–Sepharose beads and then washed five times with RIPA lysis buffer and twice with 10 mM Tris-HCl (pH 7.5) before eluting proteins with gel loading buffer. Where indicated, immunoprecipitated proteins were digested with endoglycosidase H (New England Biolabs). Dry gels were exposed to a phosphor screen (Fujifilm, Valhalla, NY), and scanned with a Typhoon FLA9000 laser scanner (GE Healthcare, Chicago, IL).

PNGase F digestion and maleimide-PEG assays. Lysates were subjected to PNGase F (New England Biolabs) digestion as suggested by the manufacturer's guidelines. Methoxypolyethylene glycol maleimide 5000 (mPEG; Sigma-Aldrich) was dissolved in water to a 100 mM stock immediately prior to use. Cells were lysed in lysis buffer with the addition of either 20 mM NEM, 5 mM mPEG, or 0.5 mM TCEP. Lysates were incubated at room temperature for 30 min prior to clearing by centrifugation at 10,000 rpm for 10 min. Supernatants with TCEP added were then supplemented with 5 mM mPEG to modify the now reduced cysteines. Lysates were resolved by SDS-PAGE and visualized by Western blotting as described above.

Half-life quantification and band densitometry. Cells were transfected in 6-well plates; 4 h later, the cells were trypsinized and plated in 6 wells of a 12-well plate. Twenty-four hours posttransfection, the medium was replaced with fresh medium containing 40 μg/ml cycloheximide (Cell Signaling Technology). The cells were lysed at various time points after cycloheximide treatment in RIPA buffer, and lysates were subjected to Western blot analysis as described above. Band densities were quantified using LI-COR Image Studio (LI-COR, Lincoln, NE).

Immunofluorescence. Huh7.5.1 cells were plated on poly-D-lysine-coated coverslips and infected with DENV-2 at an MOI of 0.1 or mock infected. Two days postinfection, cells were fixed in ice-cold methanol for 1 h at −20°C. Immunostaining with anti-HA (1:200) and anti-NS1 (1:50) antibodies was performed as described previously (38).

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at https://doi.org/10.1128/mBio.00939-17.

FIG S1, TIF file, 0.3 MB.

FIG S2, TIF file, 0.6 MB.

FIG S3, TIF file, 0.2 MB.

FIG S4, TIF file, 0.7 MB.

TABLE S1, DOCX file, 0.1 MB.

TABLE S2, DOCX file, 0.05 MB.

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