



Two Complementarity Immunotherapeutics in Non-Small-Cell Lung Cancer Patients—Mechanism of Action and Future Concepts

Kamila Wojas-Krawczyk ¹, Paweł Krawczyk ¹, Michał Gil ^{2,*} and Maciej Strzemski ³

- ¹ Pneumonology, Oncology and Allergology Department, Medical University of Lublin, 20-954 Lublin, Poland; kamilawojas@wp.pl (K.W.-K.); krapa@poczta.onet.pl (P.K.)
- ² Genetics and Immunology Institute, GENIM Ltd., 20-609 Lublin, Poland
- ³ Analytical Chemistry Department, Medical University of Lublin, 20-093 Lublin, Poland; maciej.strzemski@poczta.onet.pl
- Correspondence: gilu.michal@gmail.com or genim@tlen.pl

Simple Summary: Here, we focused on the most important mechanisms of action of combined immunotherapy with modern anticancer approaches in patients with non-small-cell lung cancer. This knowledge is extremely important for lung cancer clinicians. First, it facilitates proper involvement of the patient in the treatment and monitoring its effectiveness. More importantly, the knowledge of the immunotherapy mechanisms will certainly allow quick recognition of the side effects of such a therapy, which are totally different of those observed after chemotherapy. Side effects of combination therapies can occur at any stage of treatment, and even after completion thereof. This review article could particularly explain the mechanism of action of combined immunotherapy, which have different targets in patients.

Abstract: Due to the limited effectiveness of immunotherapy used as first-line monotherapy in patients with non-small-cell lung cancer (NSCLC), the concepts of combining classical immunotherapy based on immune checkpoint antibodies with other treatment methods have been developed. Pembrolizumab and atezolizumab were registered in combination with chemotherapy for the treatment of metastatic NSCLC, while durvalumab found its application in consolidation therapy after successful chemoradiotherapy in patients with locally advanced NSCLC. Exceptionally attractive, due to their relatively low toxicity and high effectiveness, are treatment approaches in which a combination of two different immunotherapy methods is applied. This method is based on observations from clinical trials in which nivolumab and ipilimumab were used as first-line therapy for advanced NSCLC. It turned out that the dual blockade of immune checkpoints activated T lymphocytes in different compartments of the immune response, at the same time affecting the downregulation of immune suppressor cells (regulatory T cells). These experiments not only resulted in the registration of combination therapy with nivolumab and ipilimumab, but also initiated other clinical trials using immune checkpoint inhibitors (ICIs) in combination with other ICIs or activators of costimulatory molecules found on immune cells. There are also studies in which ICIs are associated with molecules that modify the tumour environment. This paper describes the mechanism of the synergistic effect of a combination of different immunotherapy methods in NSCLC patients.

Keywords: immunotherapy; non-small-cell lung cancer; immune checkpoints; tumour microenvironment

1. Introduction

Checkpoint inhibitors such as anti-PD-1 (programmed death 1), anti-PD-L1 (programmed death ligand 1), or anti-CTLA-4 (cytotoxic T lymphocyte antigen 4) antibodies are widely used in cancer immunotherapy [1–4]. The effectiveness of immunotherapy used in monotherapy, compared to chemotherapy, has been proven in first- and/or second-line



Citation: Wojas-Krawczyk, K.; Krawczyk, P.; Gil, M.; Strzemski, M. Two Complementarity Immunotherapeutics in Non-Small-Cell Lung Cancer Patients—Mechanism of Action and Future Concepts. *Cancers* **2021**, *13*, 2836. https://doi.org/10.3390/ cancers13112836

Academic Editors: Roberta Alfieri and Myung-Ju Ahn

Received: 27 February 2021 Accepted: 31 May 2021 Published: 7 June 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). treatment in patients with various types of cancer (melanoma, non-small-cell lung cancer, renal cell carcinoma, head and neck region cancer, urothelial carcinomas, colorectal cancer, esophageal cancer, and lymphoma) [1–4]. This situation occurs especially in clinically selected NSCLC patients without actionable driver mutations (EGFR, ALK, ROS1, BRAF, etc.) detected in tumour cells. In patients with non-small-cell lung cancer, pembrolizumab (anti-PD-1 antibody) and atezolizumab (anti-PD-L1 antibody) used as first-line therapy may only be appropriate for patients with PD-L1 expression on \geq 50% of tumour cells (in the US, pembrolizumab can also be used in patients with a high tumour mutational burden and PD-L1 expression on $\geq 1\%$ of tumour cells) [2,5–7]. Many clinical trials have shown the effectiveness of anti-PD-1/or anti-PD-L1 immunotherapy, compared to docetaxel, in second-line treatment of NSCLC patients regardless of the PD-L1 expression. In clinical practice, atezolizumab, nivolumab, and pembrolizumab are used in this indication [1,2]. Unfortunately, the benefits of treatment with pembrolizumab or atezolizumab in monotherapy do not accrue to all PD-L1-positive patients. Indeed, the PD-L1 expression on cancer cells is the only biomarker validated in prospective immunotherapy-based clinical trials; however, it is not an ideal one [8–10]. Preclinical experiments have found synergistic effects of various treatment strategies that, when used in combination with immunotherapy, can enhance its effectiveness. The aim of combination therapy is to create a favourable environment within the cancerous tumour and maximize the potential of the immune system to eliminate cancer cells [11]. Figure 1 shows that many anticancer therapies are currently a combination of two methods of treatment employed to maximize the effectiveness of such a therapeutic approach.

Chemotherapy+ immunotherapy	Chemotherapy+ radiotherapy+ immunotherapy	Chemotherapy+ 2 methods of immunotherapy	Chemotherapy+ Radiotherapy+ 2 methods of immunotherapy	Radiotherapy+ 2 methods of immunotherapy	2 methods of immunotherapy
Pembrolizumab + platinum and pemetrexed, followed by pembrolizumab and pemetrexed in non-SCC patients (KEYNOTE-189, NCT02578680) Pembrolizumab + carboplatin and paclitaxel, followed by pembrolizumab in SCC patients (KEYNOTE-407, NCT02775435) Atezolizumab + carboplatin and nab- paclitaxel, followed by atezolizumab (IMpower131, NCT02367794) Atezolizumab + bevacizumab, carboplatin and paclitaxel, followed by atezolizumab and bevacizumab and bevacizumab and bevacizumab and bevacizumab and bevacizumab and bevacizumab in non-SCC patients (IMpower150, NCT02366143)	Durvalumab in patients with unresectable stage III NSCLC whose disease has not progressed following concurrent platinum-based chemotherapy and radiation therapy (PACIFIC, NCT02125461) ** Durvalumab or atezolizumab in patients with unresectable stage III NSCLC underwent different combination of platinum-based chemotherapy and radiation therapy	Nivolumab + ipilimumab and 2 cycles of platinum- doublet chemotherapy as 1. line treatment in patients with metastatic or recurrent NSCLC (CheckMate 9LA, NCT03215706)	* Atezolizumab + tiragolumab (anti-TIGIT) in patients with unresectable stage III NSCLC whose disease has not progressed following platinum- based chemotherapy and radiation therapy (SKYSCRAPER-03, NCT04513925)	** Atezolizumab + tiragolumab (anti-TIGIT) in patients with unresectable NSCLC underwent stereotactic body radiation therapy (SBRT)	Nivolumab + ipilimumab as 1. line treatment in patients with metastatic NSCLC with PD-L1 expression on 21% of tumor cells (CHECKMATE-227 (NCT02477826) * Atezolizumab + tiragolumab (anti-TIGIT) (SKYSCRAPER and CITYSCAPE studies) * Nivolumab + varlilumab (anti-CD27) (NCT02335918) * Bempegaldesleukin (pegylated IL-2) or ALT- 803 (IL-15 superagonist) plus nivolumab or atezolizumab * Epacadostat (IDO inhibitor) + pembrolizumab (ECH0 202, NCT02178722)
All therapies as 1. line of treatment. All patients in advanced stage of NSCLC	** planed in clinical trials		* only in clinical trials	** planed in clinical trials	** planed in clinical trials All patients in locally advanced or advanced stage of NSCLC

Figure 1. Many new options for combining cancer therapies are already available in NSCLC clinic, others as part of ongoing clinical trials.

The idea of using two different immunotherapies in cancer patients is based on the attempt to stimulate or inhibit different immune cells at different levels of their activity (e.g., in the lymph node and in the tumour) [11–15]. The most commonly used combination immunotherapy involves antibodies that target molecules capable of stimulation of the activity of lymphocytes and other immune cells and molecules that are able to inhibit this activity. Another combination immunotherapy method is the use of immune checkpoint inhibitors in combination with agents that modify the tumour microenvironment in a non-specific manner (e.g., pro-inflammatory cytokines, immunosuppressive cytokine inhibitors, and indoleamine 2,3-dioxygenase and adenosine inhibitors) [11–15].

2. Possibilities of Combining Different Immune Checkpoint Molecules

2.1. Strategies to Combine Two Different Antagonistic Antibodies against Inhibitory Immune Checkpoints

The use of various ICIs has found the widest application in clinical practice in cancer patients without the presence of actionable mutations and based on tumour histology as well as specific clinical characteristic of patients. A summary of the most important clinical trial results from phase 2/3 using combination immunotherapies and their clinical efficacy is presented in Table 1 [16–18].

Dual blockade of PD-1 and CTLA-4 with nivolumab and ipilimumab has been used to treat melanoma, renal cell carcinoma, and non-small-cell lung cancer in clinical trials [19–21]. In the CheckMate 227 study, patients with advanced NSCLC were treated with a combination of nivolumab and chemotherapy or nivolumab and ipilimumab or with chemotherapy [22,23]. Two predictive markers for immunotherapy were used: PD-L1 expression on tumour cells and the number of somatic mutations in tumour cells (tumour mutation burden, TMB) [22,23]. It was found that, in patients with high TMB (more than 10 mutations per million base pairs) even with no PD-L1 expression on tumour cells, the use of the nivolumab and ipilimumab combination prolonged progression-free survival, compared to other treatments [22,23]. During further follow-up, prolongation of patient survival was observed in patients with PD-L1 expression on $\geq 1\%$ of tumour cells using the combination of these two immunotherapies. In view of these results, the combination of nivolumab and ipilimumab for first-line therapy in NSCLC patients with high TMB was not registered and replaced by the registration of the combination of these two drugs in NSCLC patients with any PD-L1 expression on tumour cells [22,23]. In addition, the first-line combination therapy involving ipilimumab, nivolumab, and two lines of chemotherapy was registered for patients with advanced NSCLC based on the results of the CheckMate 9LA study [22–24]. However, it should be also mentioned about the results of ipilimumab with pembrolizumab combination based on Keynote-598 study. This combination does not improve clinical efficacy in metastatic NSCLC patients with PD-L1 TPS \geq 50% and no targetable EGFR or ALK aberrations. Moreover, this therapy was associated with greater toxicity than pembrolizumab monotherapy. However, in patients with high PD-L1 expression on tumor cells, immunotherapy alone appears to be a better therapeutic option [25].

Clinical Trial Identifier	Phase	Predictive Factor	Stage of NSCLC	Drugs	Number of Patients	ORR (%)	Median PFS (months)	PFS (HR, 95% CI)	Median OS	OS (HR, 95% CI)
227 3.		≥1% of PD- L1-positive TC (Part 1a)	IV	Nivolumab	396	27.5	4.2	0.82, 0.69–0.97 (nivolumab + ipilimumab vs. chemotherapy)	15.7	0.79, 0.65–0.96 (nivolumab + ipilimumab vs. chemotherapy)
	3.			Nivolumab + ipilimumab	396	35.9	5.1		17.1	
	1 C (1 alt 1a)		Chemotherapy	397	30	5.6	- 0.83, 0.71–0.97 (nivolumab + - ipilimumab vs. nivolumab)	14.9	0.90, 0.76–1.07 (nivolumab + ipilimumab vs. nivolumab)	
CheckMate 227 3. NCT02477826		<1% of PD- L1-positive TC (Part 1b)	IV	Nivolumab + chemotherapy	177	37.9	5.6	0.75, 0.59–0.96 (nivolumab + ipilimumab vs chemotherapy) 0.98, 0.77–1.24 (nivolumab + ipilimumab vs. nivolumab + chemotherapy) 0.73, 0.56–0.95 (nivolumab + chemotherapy vs. chemotherapy)	15.2	0.62, 0.48–0.78 (nivolumab + ipilimumab vs. chemotherapy) 0.77, 0.60–0.98 (nivolumab + ipilimumab vs. nivolumab + chemotherapy) 0.78, 0.60–1.02 (nivolumab + chemotherapy vs. chemotherapy)
	3			Nivolumab + ipilimumab	187	27.2	5.1		17.2	
	0.			Chemotherapy	186	33.1	4.7		12.2	
CheckMate 227 3. NCT02477826	3.	All patients	IV	Nivolumab + ipilimumab	583	33.1	5.1	0.79, 0.69–0.91	17.1	0.73, 0.64–0.84
			Chemotherapy	583	27.7	5.5		13.9		
CheckMate 9LA 3. NCT03215706	3.	All patients	IV	Nivolumab + ipilimumab + 2 cycles of chemotherapy	361	38.2	6.8	0.70, 0.57–0.86	15.6	0.66, 0.55–0.80
				Chemotherapy	358	24.9	5.0		10.9	
CITYSCAPER 2 (NCT03563716)	2.	≥1% of PD- L1-positive TC	IIIB or IV	Chemotherapy	68	21% * 23% **	3.88 * 4.11 **	0.58, 0.39–0.88 * 0.56, 0.34–0.92 **	ND	ND
				Atezolizumab + tiragolumab	67	37% * 42% **	5.55 * 10.18 **		ND	

Table 1. The Summary of the most important clinical trial results using combination immunotherapies. Abbreviations: ORR—overall response rate, PFS—progression free survival, HR—hazard ratio, CI—confidential interval, OS- overall survival, PD-L1—programmed death ligand 1, TC—tumor cells, ND—no data. (* PD-L1 expression examined by 22C3 monoclonal antibody; ** PD-L1 expression examined by SP263 monoclonal antibody) [16–18].

There are ongoing clinical trials investigating the possibility of administration of durvalumab and tremelimumab (anti-CTLA-4 antibody) combinations in NSCLC patients [26]. It should also be mentioned at this point that the PD-L1 expression on cancer cells is the only predictive factor validated in prospective clinical trials for immunotherapy in advanced NSCLC patients [2,27]. However, based on the clinical trial results, it is also known as not an ideal predictive marker. Not all patients with high PD-L1 expression can benefit from immunotherapy, but a clinical response may also be observed in patients without PD-L1 expression [2,27,28]. The anti-tumour immune response is an extremely complex multi-stage process depending on many factors. Moreover, it has been indicated that tumours have three immunoprofiles based on the activation of the immune system: (1) "hot" tumours, which are strongly infiltrated by T lymphocytes and with many inflammatory signals; (2) "cold" tumours, which are scanted of any immune cells infiltration nor inflammatory signs; (3) tumours with immune exclusion, where immune cells are at the periphery or within the stromal tissue [29,30]. The "hot" tumours are associated with denser PD-1-positive T lymphocyte infiltration, with pre-existing primed immune response, and are more likely to respond to the anti-PD-1 or anti-PD-L1 blockade used as monotherapy [29,30]. Other factors such as diet, body mass index, microbiome, lipid metabolism, and leptin activity have been shown to exert an influence on immunotherapy effectiveness [27]. What about combination therapy? In the IMpower 150 study, a significantly longer median progression-free survival was observed upon administration of combination therapy (atezolizumab, bevacizumab, chemotherapy) in patients with no PD-L1 expression but with low expression of T-effector activation genes than in patients receiving only bevacizumab with platinum doublets [31]. It may be speculated that the combination therapy triggered the release of tumour antigens, which contributed to the activation of the immune system. In addition, the PD-L1 molecule blockade may have inhibited the impact of the tumour on the immune system, stimulating it to fight effectively [29,30]. Therefore, the intensity of lymphocyte infiltration of tumour tissue, immunological analysis, or estimation of the gene expression profile in cancer tissue could be considered as a reliable biomarker in the prospective qualification for immunotherapy in different strategies.

2.2. Side Effects of Therapy Based on Combining Two Different Antagonistic Antibodies against Inhibitory Immune Checkpoints

Combination therapy with two different immunotherapy modalities is usually fairly well tolerated. Clinical trials did not identify a significant increase in the incidence of adverse events (AEs) in groups of patients treated with combination immunotherapy compared to monotherapy [16–18]. On the other hand, combination therapy with two ICIs causes a different type of side effects compared to chemotherapy. Patients receiving immunotherapy most often experience side effects related to hyperactivity of the immune system (endocrinopathies, pneumonitis, hepatotoxicity, skin reaction, and others), while patients receiving chemotherapy develop bone marrow suppression (anaemia, infections, thrombocytopenia, and febrile neutropenia) [16–18].

In the CheckMate 227 clinical trial, the frequency of grade 3 or 4 AEs was similar in the group that received nivolumab plus ipilimumab and in the chemotherapy group (32.8% vs. 36.0%) [17]. Serious treatment-related adverse events and AEs leading to discontinuation were more common in patients treated with nivolumab plus ipilimumab than with chemotherapy (24.5% vs. 13.9% and 18.1% vs. 9.1%). The most common treatment-related adverse events (TRAEs) of any grade related to the immune system in the group that received nivolumab plus ipilimumab were skin reactions (34.0% of the patients) and endocrinopathies (23.8%). Treatment-related deaths occurred in eight patients who received nivolumab plus ipilimumab and in six patients who received chemotherapy [17]. In patients with PD-L1 expression on $\geq 1\%$ of tumour cells treated with nivolumab monotherapy, grade 3 or 4 TRAEs occurred in 19.4% of the patients, and TRAEs resulted in discontinuation of the therapy in 12.3% of the patients. Two treatment-related deaths occurred in the nivolumab monotherapy group. In patients without expression of PD-L1 treated with

nivolumab plus chemotherapy, serious TRAEs occurred with a frequency of 19.2%. Four deaths were reported in this group [17].

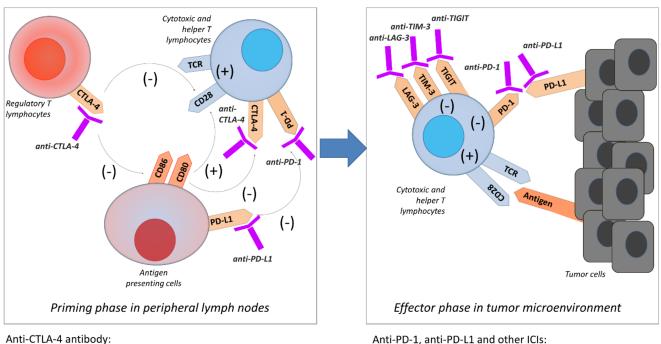
In the CheckMate 9LA clinical trial, serious TRAEs were reported in 30% of patients receiving combination therapy and in 18% of patients treated with chemotherapy [16]. Seven (2%) treatment-related deaths were observed in the former group. The following causes of death were found: acute kidney failure, colitis with diarrhoea, hepatotoxicity, hepatitis, pneumonitis, sepsis with acute renal insufficiency, and thrombocytopenia. Six (2%) deaths due to anaemia, febrile neutropenia, pancytopenia, pulmonary sepsis, respiratory failure, and sepsis occurred in the control group [16]. The most common grade 3–4 TRAEs were neutropenia (7% of patients treated with combined therapy vs. 9% of patients receiving chemotherapy), anaemia (6% vs. 14%), diarrhoea (4% vs. 1%), and febrile neutropenia (4% vs. 3%). These TRAEs were associated with the use of chemotherapy rather than immunotherapy [16].

In the CITYSCAPE clinical trial, grade \geq 3 TRAEs occurred in 19.1% of patients treated with atezolizumab monotherapy and in 14.9% of patients receiving atezolizumab in combination with tiragolumab [18]. AEs leading to treatment withdrawal occurred in 10.3% of patients from the former group and 7.5% of patients from the latter group [18].

In conclusion, the development of certain equilibrium between the effectiveness of combination therapy and its side effects should be considered. In most cases, when the side effects of combined therapy are detected at an early stage and are not very severe, it is possible to protect the patient properly against their consequences. It can be speculated that this should bring clinicians closer to the use of combination therapy in the clinic.

2.3. Molecular and Immunological Synergy of Antagonistic Antibodies against Different Inhibitory Immune Checkpoints

The effectiveness of combination therapy with nivolumab and ipilimumab is explained by the presence of interactions of these antibodies on different immunological checkpoint molecules [13,32–35]. Nivolumab inhibiting the PD-1 receptor causes activation of T lymphocytes in the tumour, lymph nodes, and peripheral tissues. This is related to the fact that the PD-L1 molecule is present on tumour cells (in primary tumours and metastases), on antigen-presenting cells infiltrating the tumour and occurring in lymph nodes (also normal, which limits the development of uncontrolled inflammatory reaction), and on most normal cells (limitation of autoimmune reaction) [13,15,32,34]. The function of the CTLA-4 molecule found on the surface of T lymphocytes is quite different [21,36]. Its stimulation plays a role during the induction of the immune response at the stage of antigen presentation. The CTLA-4 instead of CD28 molecule (the main costimulatory molecule) binds with CD80 and CD86 molecules on APC, which inhibits proliferation and activation of T helper and cytotoxic lymphocytes [15,21,36,37]. Furthermore, this interaction leads to the exfoliation of CD80 and CD86 molecules from the surface of antigen-presenting cells, causing their non-functionality. High expression of CTLA-4 on T lymphocytes also induces the intracellular FoxP3 (forkhead box P3) protein, resulting in the transformation of these cells into T regulatory lymphocytes. These reactions occur to the greatest extent in lymph nodes [15,21,36,37]. According to these considerations, the synergistic effect of nivolumab and ipilimumab consists of enhancement of the activation of T helper and cytotoxic lymphocytes by blocking one of the most potent signals inhibiting these cells (PD-1 and PD-L1 interaction) and restoring the most important, besides antigen presentation, costimulatory signal (CD28-CD80 and CD86 connections) [15,21,36,37]. Moreover, the use of ipilimumab further reduces the immunosuppressive effect of other cells of the immune system [15,19,21]. The schematic mechanism of the activity of the most important immunological checkpoints is illustrated in Figure 2.

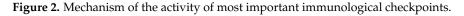


· inhibits the immunosupressive function of CTLA-4-positive regulatory T lymphocytes, mainly in peripheral lymphoid organs

 do not allow functional extinction of antigen presenting cells (DCs, macrophages) and/or CTLs and Th cells

Anti-PD-1, anti-PD-L1 and other ICIs:

restore the function of exhausted cytotoxic T lymphocytes in the tumor microenvironment, but also in peripheral tissues



In laboratory studies, this interaction between these two ICIs is strongly expressed. In the peripheral blood of patients treated with the combination therapy, compared to nivolumab or ipilimumab monotherapy, the percentage of T cytotoxic lymphocytes is significantly increased [38–40]. High levels of the proinflammatory cytokines sIL-2R α , IL-1 α , and chemokines (e.g., CXCL10) are noted in the plasma of patients undergoing combined immunotherapy, which cannot be achieved with nivolumab or ipilimumab alone. Patients with a response to combination therapy show an increase, relative to the level before the therapy, in the percentage of memory T cytotoxic lymphocytes with an EOMES+ (eomesodermin), CD69+, CD45RO+ phenotype. In addition, low expression of other negative immune checkpoints, most notably TIGIT and lymphocyte-activation gene 3 (LAG3), is observed on lymphocytes in patients responding to such treatment [38–41]. This phenomenon is not observed in patients responding to nivolumab monotherapy. The analysis of the expression of genes responsible for the immune response profile in peripheral blood leukocytes was carried out as well. Patients undergoing combination therapy express genes for granzymes A/B, Ki-67, IL-8, and HLA-DR (Human Leukocyte Antigen-DR isotype), which indicates cytolytic and proliferative activity of T cytotoxic lymphocytes and their ability to infiltrate tumour tissue. Patients receiving anti-PD-1 antibodies have increased expression of genes determining the cytolytic activity of lymphocytes (genes for granzymes A/B, KLRF1, FCRL3) [37–40]. In turn, increased expression of genes related to the capability of T lymphocytes of proliferation and production of specific cytokines (genes for Ki-67 and ICOS) is detected in patients receiving ipilimumab [38–41].

In a mouse model, tumour-infiltrating T cytotoxic lymphocytes have been divided according to their immunophenotype into 4 groups: (1) T lymphocytes with a functionally depleted cell phenotype (PD-1^{high}, LAG3⁺⁺, TIM3⁺⁺), (2) terminally differentiated T lymphocytes with an activated phenotype (PD-1+, LAG3^{int}, TIM3^{int}), (3) T lymphocytes at an early stage of differentiation (Tbet^{int}, CD86⁺, PD-1^{+/-}, Bcl2⁺), and (4) apoptosis-resistant migratory T lymphocytes (PD-1⁻, CD62L⁺, Bcl2⁺⁺) [13,38]. The use of combination immunotherapy, compared to nivolumab or ipilimumab monotherapy, significantly increases

the percentage of differentiated and activated lymphocytes and significantly decreases the percentage of functionally depleted lymphocytes [13,38]. However, the type of therapy has no effect on the percentages of other T cytotoxic lymphocyte subpopulations in the peripheral blood. Among the T helper lymphocytes, subpopulations differing in the immunophenotype have also been distinguished: Th1 lymphocytes with an effector phenotype (PD-1⁺, GATA3⁺, CD44⁺, CXCR3⁺⁺), T lymphocytes with a helper phenotype without chemokine receptors (CD44⁺, GATA3⁺, CD44⁺, CXCR3⁻), and actively migrating T lymphocytes that resist apoptosis (PD-1⁻, CD62L⁺, Bcl2⁺⁺) [13,38–42]. Combination therapy, compared to nivolumab or ipilimumab monotherapy, results in significantly increased infiltration of Th1 effector lymphocytes. T regulatory lymphocytes can be divided into three groups according to their immunophenotype: (1) Treg lymphocytes with a pro-tumour phenotype (CTLA-4⁺⁺, FoxP3⁺, CD25⁺), (2) Treg lymphocytes with an incomplete differentiation phenotype (CTLA-4⁺, FoxP3⁺⁺, CD25⁺⁺), and (3) undifferentiated and depleted Treg lymphocytes (CTLA-4⁻, FoxP3^{+/-}, CD25⁺⁺). A lower degree of infiltration of Treg lymphocytes with a pro-tumour immunophenotype was detected in mice treated with ipilimumab or combination therapy compared to nivolumab-treated or untreated mice. At the same time, it was shown that the percentage of Th1 effector lymphocytes correlated negatively and the percentage of pro-tumour Treg lymphocytes correlated positively with tumour size [41].

Based on these theoretical considerations and laboratory study results, quite new concepts of clinical trials combining antibodies that interact with different immune checkpoints have been developed. There are ongoing clinical trials in patients with advanced NSCLC, in which classical anti-PD-1 or anti-PD-L1 antibodies are attempted to be combined with antibodies against ICOS (inducible T-cell costimulator), LAG-3, TIM-3 (T-cell immunoglobulin domain and mucin domain 3), or TIGIT [43–47]. Research on new anti-LAG3 and anti-TIGIT antibodies is of particular importance. As noted above, patients without response to nivolumab and ipilimumab combination therapy had a significantly higher percentage of T lymphocytes with expression of these molecules. This suggests that their presence may have a leading role in inhibiting T lymphocyte activation and in inducing resistance to existing immunotherapies [43–45,48]. Therefore, there are indications for replacement of the anti-CTLA-4 therapy in combination therapy using anti-PD-1 or anti-PD-L1 antibodies with anti-LAG3 or anti-TIGIT antibodies [49]. A phase I trial in which tiragolumab (anti-TIGIT antibody) was used along with atezolizumab in patients with advanced NSCLC provided particularly interesting results [50–52]. Response to this type of therapy was achieved in 46% of patients, and disease stabilization occurred in 85% of patients. These encouraging results contributed to the initiation of phase II trial-CITYSCAPE and phase III trial—SKYSCRAPER-01, which used combination therapy with atezolizumab and tiragolumab compared to therapy with atezolizumab alone in advanced NSCLC patients with PD-L1 expression on tumour cells [18,53]. The CITYSCAPE trial demonstrated response in 31.3% of patients treated with the combination therapy and in 16.2% of patients receiving atezolizumab alone. The median progression-free time in these two patient groups was 5.4 months and 3.6 months, respectively [18,53].

2.4. Strategies to Combine Different Antagonistic and Agonistic Antibodies against Immune Checkpoints

On the other hand, there are ongoing early clinical trials in which agonistic antibodies that bind to costimulatory molecules on lymphocytes have been combined with antagonistic antibodies directed against negative checkpoints (usually anti-PD-1, anti-PD-L1, or anti-CTLA-4) [45,54]. Activation of CD28, CD27, OX40, CD137 (4-1BB), or GITR (glucocorticoid-induced TNFR-related) molecules increases lymphocyte proliferation and positively stimulates the development of immune response [55–58]. However, the use of agonist antibodies that bind to these molecules often causes serious side effects. Nevertheless, promising results have been obtained in cancer patients using a combination of classical ICIs with antibodies stimulating CD27 and CD137 activity [59]. The CD27 activation is a potent costimulatory factor in the first stages of immune response when it promotes T cell survival and memory T cell formation [60–62]. The only ligand for CD27 is the CD70 molecule found on APCs and on activated T lymphocytes. However, the interaction between CD27 and CD70 changes over the course of immune responses [63,64]. Chronic stimulation of CD27 by CD70 in chronic inflammation suppresses the immune response and, in the case of tumour cells expressing CD70, leads to differentiation of T lymphocytes into Treg cells [65,66]. A phase I/II clinical trial consisted in the use of variliumab, i.e., an agonistic antibody that binds to CD27, in combination with nivolumab in patients with solid tumours [67,68]. Response to the treatment was achieved in 49% of patients, although most of them did not have PD-L1 expression on tumour cells. It turned out that, after 4–6 weeks of therapy, 76% of patients acquired PD-L1 expression on antigen-presenting cells. On the one hand, T lymphocytes were stimulated by activation of the CD27 molecule and, on the other hand, a purpose for nivolumab therapy (PD-L1 expression) emerged [67,68].

3. Use of Non-Specific Immune System Stimulation and Tumour Microenvironment Modification in Immune Combination Therapies

Non-specific immunotherapy can also be associated with immune checkpoint inhibitors. Non-specific stimulation of the cytotoxic response against tumour cells can be achieved by administration of proinflammatory cytokines or by inhibition of the immunosuppressive cytokine function [69,70]. In the first case, clinical trials have been undertaken to assess combination therapy of cancer patients with anti-PD-1 and anti-PD-L1 antibodies in combination with modified cytokines IL-2 and IL-15 [69]. Pegylated IL-2 with attached polyethylene glycol chains (bempegaldesleukin) has a longer half-life in the body than recombinant IL-2 (aldesleukin) [71,72]. Bempegaldesleukin binds to heterodimeric IL-2R $\beta\gamma$ (CD122), which preferentially activates effector cytotoxic T lymphocytes and NK cells in the peripheral blood and tumour microenvironment. In contrast, pegylated IL-2 has a low affinity towards the receptor for IL-2 built of alpha, beta, and gamma subunits (IL-2R $\alpha\beta\gamma$, CD25), which is mainly found on Treg cells [71–73]. Due to these properties, bempegaldesleukin does not activate T lymphocytes and NK cells immediately after infusion and only transiently activates Treg cells, resulting in a higher safety profile compared to that of aldesleukin [71–73].

Clinical studies on the use of recombinant IL-15 have also been undertaken. However, this molecule was quickly replaced by an IL-15 superagonist (ALT-803), which consists of a modified IL-15 molecule with an introduced N72D mutation, a modified receptor for IL-15 (IL-15R), and an Fc fragment of IgG1 class antibody linking everything [74–77]. The IL-15 molecule is supposed to bind to IL-2R $\beta\gamma$ in order to stimulate cytotoxic T lymphocytes and NK cells. The modified IL-15R ensures specific binding of ALT-803 to IL-2R $\beta\gamma$, rather than to IL-2R $\alpha\beta\gamma$, which is found on Treg cells, while the Fc fragment of the antibody prolongs the half-life of the complex and attracts NK cells [74–78].

Bempegaldesleukin and ALT-803 have been used in combination with nivolumab and atezolizumab in patients with various types of cancer (including hematologic) in phase I clinical trials with promising results and satisfactory safety [79,80]. In turn, therapies in which anti-PD-1, anti-PD-L1, or anti-CTLA-4 antibodies were combined with therapies aimed at reducing the activity of immunosuppressive cytokines such as TGF- β (tumour growth factor beta), M-CSF (macrophage-colony stimulating factor), and IL-10 seem to be less effective [81,82]. Addition of drugs blocking IL-10 or TGF- β function to classical immunotherapy increased the risk of adverse effects in the form of autoimmune reactions [83–86]. Nevertheless, clinical trials are underway to investigate the effectiveness of M7824—a fusion protein consisting of a human IgG1 monoclonal antibody against PD-L1 fused to the extracellular domain of the receptor for TGF- β , which captures TGF- β in the tumour environment [86]. This drug may have great potential in the treatment of cancer patients in combination with other immunotherapy. It has selective effects in PD-L1 positive tumours and has fewer side effects than other anti-TGF- β agents [86].

The tumour microenvironment has a very adverse effect on the immune system functioning therein [83]. An unfavourable tumour microenvironment results in exclusion of immune response outside the tumour. Two substances play a special role here. One of them is adenosine [87,88]. Adenosine and ATP are present at exceptionally low concentrations in extracellular fluids. However, inflammation, ischemia, or the cancer process can lead to the release of ATP through transport channels in cell membranes, active exocytosis, and directly from damaged cells [89-91]. Extracellular ATP acts as a danger-associated molecular pattern (DAMP) to promote the immune response. However, during inflammation, extracellular ATP is progressively dephosphorylated by ectonucleotidases (mainly CD39 and CD73), resulting in the formation of adenosine. Adenosine binds to its receptors A1, A2a, A2b, and A3. Stimulation of the A2a receptor inhibits the cytotoxic T cell activity and promotes the Treg cell activity by increasing FoxP3 expression. Under the influence of this stimulation, the expression of immune checkpoints including PD-1, CTLA-4, and LAG-3 increases on effector lymphocytes. Therefore, it is not surprising that molecules that block adenosine binding to the A2a receptor and molecules that inhibit the activity of the CD39 and CD73 enzymes have been developed and used in combination with anti-PD-1 or anti-CTLA-4 antibodies in early phase clinical trials in cancer patients [89-91].

Another substance that causes elimination of tumour cells from the tumour area is indoleamine 2,3-dioxygenase (IDO) [92–95]. This enzyme metabolizes tryptophan to kynurenine. The production of IDO by tumour cells reduces tryptophan levels in the tumour. Tryptophan, i.e., an exogenous amino acid, is essential for normal lymphocyte function. Its absence in the tumour environment prevents T lymphocytes from entering the tumour [92–94]. Studies on the possibility of combining IDO inhibitors (e.g., epacadostat) with classical ICIs in NSCLC and melanoma patients have been conducted for several years. However, phase III trials failed to demonstrate the effectiveness of such therapy, which resulted in the lack of registration of epacadostat in combination with pembrolizumab for the treatment of melanoma and NSCLC patients [92,93].

4. Conclusions

Standard anti-cancer therapies, such as radiotherapy or chemotherapy, destabilize tumour cell function, contribute to the release of tumour antigens and the formation of neoantigens, and affect the production of cytokines, chemokines, and other substances that stimulate immune cell activity. As a result, tumours with low immunogenicity ("cold") could be transformed into tumours with high immunogenicity ("hot," "inflammatory"), abundant with infiltrates of activated specific lymphocytes [29,30]. This breaks down the mechanism by which tumour cells escape from immune surveillance. The addition of immunotherapy targeting immune checkpoints to chemotherapy or chemoradiotherapy further enhances the antitumor effects of cytotoxic T lymphocytes.

On the other hand, combining two different immunotherapy methods in cancer patients may be as effective as chemoimmunotherapy or chemoradiotherapy in cancer therapy. The combination of two immunotherapy methods is based on the idea of stimulating or inhibiting different immune cells at different levels of their activity with two different immune point activators or inhibitors, or using conventional ICIs in combination with non-specific immunostimulatory agents or agents that modify the tumour microenvironment. However, patients should be very well suited to this type of treatment. At present, there are no conclusively proven predictors for combination therapies, but the selection of patients should be based on clinical factors, such as the performance status of the patients, the presence of comorbidities, and the availability to a multidisciplinary cancer centre, which is extremely important for the proper management of patients.

Currently, scientists have a wide range of possibilities to investigate and combine different therapeutic approaches. The treatment method based on the use of specific genetically modified CAR-T cells (chimeric antigen receptor) is developing dynamically. Attempts are underway to combine classical immunotherapy targeting immune checkpoints with treatment using modified oncolytic viruses. Already, the median survival

of patients with advanced non-small-cell lung cancer has increased significantly. The development of modern personalized treatments, including immunotherapies, enables many patients to act in good functional status for 3 years and beyond. In the near future, it is expected that many patients will live with cancer just as patients with cardiovascular or infectious diseases (e.g., AIDS and hepatitis C) are currently living in near-complete comfort.

In conclusion, combination immunotherapies will be used in cancer patients, not only those with lung cancer. Therefore, is seems extremely important to understand the mechanisms of action of combined immunotherapy, firstly to understand how these therapies work in the patient's body and, secondly, to be able to quickly recognize the side effects and properly secure the patients.

Author Contributions: Resources, K.W.-K., P.K. and M.G.; Validation, K.W.-K.; Visualization, P.K. and M.S.; Writing—original draft, K.W.-K., P.K. and M.G.; Writing—review and editing, M.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing not applicable. No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Chen, R.; Tao, Y.; Xu, X.; Shan, L.; Jiang, H.; Yin, Q.; Pei, L.; Cai, F.; Ma, L.; Yu, Y. The efficacy and safety of nivolumab, pembrolizumab, and atezolizumab in treatment of advanced non-small cell lung cancer. *Discov. Med.* **2018**, *26*, 155–166. [PubMed]
- Hargadon, K.M.; Johnson, C.E.; Williams, C.J. Immune checkpoint blockade therapy for cancer: An overview of FDA-approved immune checkpoint inhibitors. *Int. Immunopharmacol.* 2018, 62, 29–39. [CrossRef]
- 3. Korman, A.J.; Peggs, K.S.; Allison, J.P. Checkpoint blockade in cancer immunotherapy. Adv. Immunol. 2006, 90, 297–339.
- 4. Valecha, G.K.; Vennepureddy, A.; Ibrahim, U.; Safa, F.; Samra, B.; Atallah, J.P. Anti-PD-1/PD-L1 antibodies in non-small cell lung cancer: The era of immunotherapy. *Expert Rev. Anticancer Ther.* **2017**, *17*, 47–59. [CrossRef]
- Fehrenbacher, L.; Spira, A.; Ballinger, M.; Kowanetz, M.; Vansteenkiste, J.; Mazieres, J.; Park, K.; Smith, D.; Artal-Cortes, A.; Lewanski, C.; et al. Atezolizumab versus docetaxel for patients with previously treated non-small-cell lung cancer (POPLAR): A multicentre, open-label, phase 2 randomised controlled trial. *Lancet* 2016, 387, 1837–1846. [CrossRef]
- Mok, T.S.K.; Wu, Y.L.; Kudaba, I.; Kowalski, D.M.; Cho, B.C.; Turna, H.Z.; Castro, G., Jr.; Srimuninnimit, V.; Laktionov, K.K.; Bondarenko, I.; et al. Pembrolizumab versus chemotherapy for previously untreated, PD-L1-expressing, locally advanced or metastatic non-small-cell lung cancer (KEYNOTE-042): A randomised, open-label, controlled, phase 3 trial. *Lancet* 2019, 393, 1819–1830. [CrossRef]
- Reck, M.; Rodríguez-Abreu, D.; Robinson, A.G.; Hui, R.; Csőszi, T.; Fülöp, A.; Gottfried, M.; Peled, N.; Tafreshi, A.; Cuffe, S.; et al. Updated analysis of Keynote-024: Pembrolizumab versus platinum-based chemotherapy for advanced non-small-cell lung cancer with PD-L1 tumor proportion score of 50% or greater. J. Clin. Oncol. 2019, 37, 537–546. [CrossRef]
- Ancevski Hunter, K.; Socinski, M.A.; Villaruz, L.C. PD-L1 testing in guiding patient selection for PD-1/PD-L1 inhibitor therapy in lung cancer. *Mol. Diagn. Ther.* 2018, 22, 1–10. [CrossRef]
- Spencer, K.R.; Wang, J.; Silk, A.W.; Ganesan, S.; Kaufman, H.L.; Mehnert, J.M. Biomarkers for immunotherapy: Current developments and challenges. *Am. Soc. Clin. Oncol. Educ. Book* 2016, 35, e493. [CrossRef]
- 10. Weber, J.S. Biomarkers for checkpoint inhibition. Am. Soc. Clin. Oncol. Educ. Book 2017, 37, 205–209. [CrossRef]
- 11. Zappasodi, R.; Merghoub, T.; Wolchok, J.D. Emerging concepts for immune checkpoint blockade-based combination therapies. *Cancer Cell* **2018**, *33*, 581–598. [CrossRef]
- Das, R.; Verma, R.; Sznol, M.; Boddupalli, C.S.; Gettinger, S.N.; Kluger, H.; Callahan, M.; Wolchok, J.D.; Halaban, R.; Dhodapkar, M.V.; et al. Combination therapy with anti-CTLA-4 and anti-PD-1 leads to distinct immunologic changes in vivo. *J. Immunol.* 2015, 194, 950–959. [CrossRef]
- Gide, T.N.; Quek, C.; Menzies, A.M.; Tasker, A.T.; Shang, P.; Holst, J.; Madore, J.; Lim, S.Y.; Velickovic, R.; Wongchenko, M.; et al. Distinct immune cell populations define response to anti-PD-1 monotherapy and anti-PD-1/anti-CTLA-4 combined therapy. *Cancer Cell* 2019, 35, 238–255. [CrossRef]
- 14. Wei, S.C.; Duffy, C.R.; Allison, J.P. Fundamental mechanisms of immune checkpoint blockade therapy. *Cancer Discov.* **2018**, *8*, 1069–1086. [CrossRef] [PubMed]

- Wei, S.C.; Levine, J.H.; Cogdill, A.P.; Zhao, Y.; Anang, N.A.S.; Andrews, M.C.; Sharma, P.; Wang, J.; Wargo, J.A.; Pe'er, D.; et al. Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* 2017, 170, 1120–1133.e17. [CrossRef] [PubMed]
- Paz-Ares, L.; Ciuleanu, T.E.; Cobo, M.; Schenker, M.; Zurawski, B.; Menezes, J.; Richardet, E.; Bennouna, J.; Felip, E.; Juan-Vidal, O.; et al. First-line nivolumab plus ipilimumab combined with two cycles of chemotherapy in patients with non-small-cell lung cancer (CheckMate 9LA): An international, randomised, open-label, phase 3 trial. *Lancet* 2021, 22, 198–211. [CrossRef]
- 17. Ramalingam, S.S.; Ciuleanu, T.E.; Pluzanski, A.; Lee, J.S.; Schenker, M.; Caro, R.B.; Lee, K.H.; Zurawski, B.; Audigier-Valette, C.; Provencio, M.; et al. Nivolumab + ipilimumab versus platinum-doublet chemotherapy as first-line treatment for advanced non-small cell lung cancer: Three-year update from CheckMate 227 Part 1. J. Clin. Oncol. **2020**, *38*, 9500. [CrossRef]
- Rodriguez-Abreu, D.; Johnson, M.L.; Hussein, M.; Cobo, M.; Patel, A.J.; Secen, N.M.; Lee, K.H.; Massuti, B.; Hiret, S.; Yang, J.C.; et al. Primary analysis of a randomized, double-blind, phase II study of the anti-TIGIT antibody tiragolumab (tira) plus atezolizumab (atezo) versus placebo plus atezo as first-line (1L) treatment in patients with PD-L1-selected NSCLC (CITYSCAPE). *J. Clin. Oncol.* 2020, *38*, 9503. [CrossRef]
- Kooshkaki, O.; Derakhshani, A.; Hosseinkhani, N.; Torabi, M.; Safaei, S.; Brunetti, O.; Racanelli, V.; Silvestris, N.; Baradaran, B. Combination of ipilimumab and nivolumab in cancers: From clinical practice to ongoing clinical trials. *Int. J. Mol. Sci.* 2020, 21, 4427. [CrossRef]
- 20. Seidel, J.A.; Otsuka, A.; Kabashima, K. Anti-PD-1 and Anti-CTLA-4 therapies in cancer: Mechanisms of action, efficacy, and limitations. *Front. Oncol.* **2018**, *8*, 1–14. [CrossRef]
- Wei, S.C.; Anang, N.A.S.; Sharma, R.; Andrews, M.C.; Reuben, A.; Levine, J.H.; Cogdill, A.; Mancuso, J.J.; Wargo, J.A.; Pe'er, D.; et al. Combination anti–CTLA-4 plus anti-PD-1 checkpoint blockade utilizes cellular mechanisms partially distinct from monotherapies. *Proc. Natl. Acad. Sci. USA* 2019, *116*, 22699–22709. [CrossRef]
- 22. Hellmann, M.D.; Paz-Ares, L.; Bernabe Caro, R.; Zurawski, B.; Kim, S.W.; Carcereny Costa, E.; Park, K.; Alexandru, A.; Lupinacci, L.; de la Mora Jimenez, E.; et al. Nivolumab plus ipilimumab in advanced non–small-cell lung cancer. *N. Engl. J. Med.* **2019**, *381*, 2020–2031. [CrossRef] [PubMed]
- Reck, M.; Schenker, M.; Lee, K.H.; Provencio, M.; Nishio, M.; Lesniewski-Kmak, K.; Sangha, R.; Ahmed, S.; Raimbourg, J.; Feeney, K.; et al. Nivolumab plus ipilimumab versus chemotherapy as first-line treatment in advanced non-small-cell lung cancer with high tumour mutational burden: Patient-reported outcomes results from the randomised, open-label, phase III CheckMate 227 trial. *Eur. J. Cancer* 2019, *116*, 137–147. [CrossRef] [PubMed]
- 24. Reck, M.; Ciuleanu, T.E.; Cobo Dols, M.; Schenker, M.; Zurawski, B.; Menezes, J.; Richardet, E.; Bennouna, J.; Felip, E.; Juan-Vidal, O.; et al. Nivolumab plus ipilimumab plus 2 cycles of platinum doublet chemotherapy vs 4 cycles of chemo as first-line treatment for stage IV/recurrent non-small cell lung cancer: CheckMate 9LA. *ASCO* 2020, 9501. [CrossRef]
- 25. Boyer, M.; Şendur, M.A.N.; Rodríguez-Abreu, D.; Park, K.; Lee, D.H.; Çiçin, I.; Yumuk, P.F.; Orlandi, F.J.; Leal, T.A.; Molinier, O.; et al. Pembrolizumab plus ipilimumab or placebo for metastatic non-small-cell lung cancer with PD-L1 tumor proportion score ≥ 50%: Randomized, double-blind phase III KEYNOTE-598 Study. J. Clin. Oncol. **2021**. [CrossRef]
- 26. Rizvi, N.A.; Cho, B.C.; Reinmuth, N.; Lee, K.H.; Luft, A.; Ahn, M.J.; van den Heuvel, M.M.; Cobo, M.; Vicente, D.; Smolin, A.; et al. Durvalumab with or without tremelimumab vs standard chemotherapy in first-line treatment of metastatic non-small cell lung cancer: The MYSTIC phase 3 randomized clinical trial JAMA. *Oncology* 2020, *6*, 661–674.
- 27. Bodor, J.N.; Boumber, Y.; Borghaei, H. Biomarkers for immune checkpoint inhibition in non-small cell lung cancer (NSCLC). *Cancer* 2020, *126*, 260–270. [CrossRef]
- 28. Krieger, T.; Pearson, I.; Bell, J.; Doherty, J.; Robbins, P. Targeted literature review on use of tumor mutational burden status and programmed cell death ligand 1 expression to predict outcomes of checkpoint inhibitor treatment. *Diagn. Pathol.* **2020**, *15*, 6. [CrossRef]
- Lizotte, P.H.; Ivanova, E.V.; Awad, M.M.; Jones, R.E.; Keogh, L.; Liu, H.; Dries, R.; Almonte, C.; Herter-Sprie, G.S.; Santos, A.; et al. Multiparametric profiling of non-small-cell lung cancers reveals distinct immunophenotypes. *JCI Insight* 2016, 1, e89014. [CrossRef]
- Bremnes, R.M.; Busund, L.T.; Kilvær, T.L.; Andersen, S.; Richardsen, E.; Paulsen, E.E.; Hald, S.; Khanehkenari, M.R.; Cooper, W.A.; Kao, S.C.; et al. The role of tumor-infiltrating lymphocytes in development, progression, and prognosis of non-small cell lung cancer. J. Thorac. Oncol. 2016, 11, 789–800. [CrossRef]
- Socinski, M.A.; Jotte, R.M.; Cappuzzo, F.; Orlandi, F.; Stroyakovskiy, D.; Nogami, N.; Rodríguez-Abreu, D.; Moro-Sibilot, D.; Thomas, C.A.; Barlesi, F.; et al. Atezolizumab for first-line treatment of metastatic nonsquamous NSCLC. *N. Engl. J. Med.* 2018, 378, 2288–2301. [CrossRef]
- 32. Curran, M.A.; Montalvo, W.; Yagita, H.; Allison, J.P. PD-1 and CTLA-4 combination blockade expands infiltrating T cells and reduces regulatory T and myeloid cells within B16 melanoma tumors. *Proc. Natl. Acad. Sci. USA* **2010**, 107, 4275–4280. [CrossRef]
- 33. De Sousa Linhares, A.; Leitner, J.; Grabmeier-Pfistershammer, K.; Steinberger, P. Not all immune checkpoints are created equal. *Front. Immunol.* **2018**, *9*, 1909. [CrossRef]
- 34. Hayashi, H.; Nakagawa, K. Combination therapy with PD-1 or PD-L1 inhibitors for cancer. *Int. J. Clin. Oncol.* **2020**, *25*, 818–830. [CrossRef]
- 35. Pardoll, D.M. The blockade of immune checkpoints in cancer immunotherapy. Nat. Rev. Cancer 2012, 12, 252–264. [CrossRef]

- 36. Parry, R.V.; Chemnitz, J.M.; Frauwirth, K.A.; Lanfranco, A.R.; Braunstein, I.; Kobayashi, S.V.; Linsley, P.S.; Thompson, C.B.; Riley, J.L. CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms. *Mol. Cell. Biol.* 2005, *25*, 9543–9553. [CrossRef]
- Hui, E.; Cheung, J.; Zhu, J.; Su, X.; Taylor, M.J.; Wallweber, H.A.; Sasmal, D.K.; Huang, J.; Kim, J.M.; Mellman, I.; et al. T cell costimulatory receptor CD28 is a primary target for PD-1-mediated inhibition. *Science* 2017, 355, 1428–1433. [CrossRef]
- Blackburn, S.D.; Shin, H.; Freeman, G.J.; Wherry, E.J. Selective expansion of a subset of exhausted CD8 T cells by alphaPD-L1 blockade. *Proc. Natl. Acad. Sci. USA* 2008, 105, 15016–15021. [CrossRef]
- 39. Carlino, M.S.; Long, G.V. Ipilimumab combined with nivolumab: A standard of care for the treatment of advanced melanoma? *Clin. Cancer Res.* **2016**, *22*, 3992–3998. [CrossRef]
- 40. Larkin, J.; Chiarion-Sileni, V.; Gonzalez, R.; Grob, J.J.; Cowey, C.L.; Lao, C.D.; Schadendorf, D.; Dummer, R.; Smylie, M.; Rutkowski, P.; et al. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N. Engl. J. Med.* **2015**, *373*, 23–34. [CrossRef]
- Gestermann, N.; Saugy, D.; Martignier, C.; Tillé, L.; Fuertes Marraco, S.A.; Zettl, M.; Tirapu, I.; Speiser, D.E.; Verdeil, G. LAG-3 and PD-1+LAG-3 inhibition promote anti-tumor immune responses in human autologous melanoma/T cell co-cultures. Oncoimmunology 2020, 9, 1736792. [CrossRef] [PubMed]
- 42. Pesce, S.; Trabanelli, S.; Di Vito, C.; Greppi, M.; Obino, V.; Guolo, F.; Minetto, P.; Bozzo, M.; Calvi, M.; Zaghi, E.; et al. Cancer immunotherapy by blocking immune checkpoints on innate lymphocytes. *Cancers* **2020**, *12*, 3504. [CrossRef] [PubMed]
- 43. Anderson, A.C.; Joller, N.; Kuchroo, V.K. Lag-3, Tim-3, and TIGIT co-inhibitory receptors with specialized functions in immune regulation. *Immunity* **2016**, *44*, 989–1004. [CrossRef] [PubMed]
- 44. Fan, X.; Quezada, S.A.; Sepulveda, M.A.; Sharma, P.; Allison, J.P. Engagement of the ICOS pathway markedly enhances efficacy of CTLA-4 blockade in cancer immunotherapy. *J. Exp. Med.* **2014**, *211*, 715–725. [CrossRef]
- 45. Qin, S.; Xu, L.; Yi, M.; Yu, S.; Wu, K.; Luo, S. Novel immune checkpoint targets: Moving beyond PD-1 and CTLA-4. *Mol. Cancer* **2019**, *18*, 155. [CrossRef]
- 46. Solinas, C.; Gu-Trantien, C.; Willard-Gallo, K. The rationale behind targeting the ICOS-ICOS ligand costimulatory pathway in cancer immunotherapy. *ESMO Open* **2020**, *5*, e000544. [CrossRef]
- 47. Solomon, B.L.; Garrido-Laguna, I. TIGIT: A novel immunotherapy target moving from bench to bedside. *Cancer Immunol. Immunother.* **2018**, *67*, 1659–1667. [CrossRef]
- Chauvin, J.M.; Pagliano, O.; Fourcade, J.; Sun, Z.; Wang, H.; Sander, C.; Kirkwood, J.M.; Chen, T.H.; Maurer, M.; Korman, A.J.; et al. TIGIT and PD-1 impair tumor antigen-specific CD8+ T cells in melanoma patients. *J. Clin. Investig.* 2015, 125, 2046–2058. [CrossRef]
- 49. Targeted Oncology. Available online: https://www.targetedonc.com/view/fda-grants-breakthrough-therapy-to-first-anti-tigit-therapy-in-nsclc-with-high-pd-l1 (accessed on 26 February 2021).
- 50. Bendell, J.C.; Bedard, P.; Bang, Y.J.; LoRusso, P.; Hodi, S.; Gordon, M.; D'Angelo, S.; Desai, J.; Garralda, E.; Italiano, A.; et al. Phase Ia/Ib dose-escalation study of the anti-TIGIT antibody tiragolumab as a single agent and in combination with atezolizumab in patients with advanced solid tumors. In Proceedings of the AACR Virtual Annual Meeting II 2020, Philadelphia, PA, USA, 27–28 April, 22–24 June 2020. Abstract number CT302.
- Johnston, R.J.; Comps-Agrar, L.; Hackney, J.; Yu, X.; Huseni, M.; Yang, Y.; Park, S.; Javinal, V.; Chiu, H.; Irving, B.; et al. The immunoreceptor TIGIT regulates antitumor and antiviral CD8+T cell effector function. *Cancer Cell* 2014, 26, 923–937. [CrossRef]
- 52. Kurtulus, S.; Sakuishi, K.; Ngiow, S.F.; Joller, N.; Tan, D.J.; Teng, M.W.; Smyth, M.J.; Kuchroo, V.K.; Anderson, A.C. TIGIT predominantly regulates the immune response via regulatory T cells. *J. Clin. Investig.* **2015**, *125*, 4053–4062. [CrossRef]
- 53. A Study of Tiragolumab in Combination with Atezolizumab Compared with Placebo in Combination with Atezolizumab in Patients with Previously Untreated Locally Advanced Unresectable or Metastatic pd-l1-Selected Non-Small Cell Lung Cancer (SKYSCRAPER-01). Available online: www.clinicaltrial.gov (accessed on 26 February 2021).
- 54. Chen, L.; Flies, D.B. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat. Rev. Immunol.* **2013**, *13*, 227–242. [CrossRef]
- Ko, K.; Yamazaki, S.; Nakamura, K.; Nishioka, T.; Hirota, K.; Yamaguchi, T.; Shimizu, J.; Nomura, T.; Chiba, T.; Sakaguchi, S. Treatment of advanced tumors with agonistic anti-GITR mAb and its effects on tumor-infiltrating Foxp3 + CD25 + CD4 + regulatory T cells. J. Exp. Med. 2005, 202, 885–891. [CrossRef]
- Mitsui, J.; Nishikawa, H.; Muraoka, D.; Wang, L.; Noguchi, T.; Sato, E.; Kondo, S.; Allison, J.P.; Sakaguchi, S.; Old, L.J.; et al. Two distinct mechanisms of augmented antitumor activity by modulation of immunostimulatory/inhibitory signals. *Clin. Cancer Res.* 2010, 16, 2781–2791. [CrossRef]
- Valzasina, B.; Guiducci, C.; Dislich, H.; Killeen, N.; Weinberg, A.D.; Colombo, M.P. Triggering of OX40 (CD134) on CD4+CD25+T cells blocks their inhibitory activity: A novel regulatory role for OX40 and its comparison with GITR. *Blood* 2005, 105, 2845–2851.
 [CrossRef]
- 58. Zappasodi, R.; Sirard, C.; Li, Y.; Budhu, S.; Abu-Akeel, M.; Liu, C.; Yang, X.; Zhong, H.; Newman, W.; Qi, J.; et al. Rational design of anti-GITR-based combination immunotherapy. *Nat. Med.* **2019**, *25*, 759–766. [CrossRef]
- Sanmamed, M.F.; Pastor, F.; Rodriguez, A.; Perez-Gracia, J.L.; Rodriguez-Ruiz, M.E.; Jure-Kunkel, M.; Melero, I. Agonists of co-stimulation in cancer immunotherapy directed against CD137, OX40, GITR, CD27, CD28, and ICOS. *Semin. Oncol.* 2015, 42, 640–655. [CrossRef]

- 60. Starzer, A.M.; Berghoff, A.S. New emerging targets in cancer immunotherapy: CD27 (TNFRSF7). *ESMO Open* **2019**, *4*, e000629. [CrossRef]
- 61. Van de Ven, K.; Borst, J. Targeting the T-cell co-stimulatory CD27/CD70 pathway in cancer immunotherapy: Rationale and potential. *Immunotherapy* **2015**, *7*, 655–667. [CrossRef]
- 62. Wong, H.Y.; Schwarz, H. CD137/CD137 ligand signalling regulates the immune balance: A potential target for novel immunotherapy of autoimmune diseases. J. Autoimmun. 2020, 112, 102499. [CrossRef]
- 63. Buchan, S.; Manzo, T.; Flutter, B.; Rogel, A.; Edwards, N.; Zhang, L.; Sivakumaran, S.; Ghorashian, S.; Carpenter, B.; Bennett, C.; et al. OX40- and CD27-mediated costimulation synergizes with anti-PD-L1 blockade by forcing exhausted CD8+ T cells to exit quiescence. *J. Immunol.* **2015**, *194*, 125–133. [CrossRef]
- 64. Buchan, S.L.; Fallatah, M.; Thirdborough, S.M.; Taraban, V.Y.; Rogel, A.; Thomas, L.J.; Penfold, C.A.; He, L.Z.; Curran, M.A.; Keler, T.; et al. PD-1 blockade and CD27 stimulation activate distinct transcriptional programs that synergize for CD8+ T-cell-driven antitumor immunity. *Clin. Cancer Res.* **2018**, *24*, 2383–2394. [CrossRef]
- Etxeberria, I.; Glez-Vaz, J.; Teijeira, Á.; Melero, I. New emerging targets in cancer immunotherapy: CD137/4-1BB costimulatory axis. ESMO Open 2019, 4, e000733. [CrossRef]
- Buchan, S.L.; Rogel, A.; Al-Shamkhani, A. The immunobiology of CD27 and OX40 and their potential as targets for cancer immunotherapy. *Blood* 2018, 131, 39–48. [CrossRef]
- Ansell, S.M.; Flinn, I.; Taylor, M.H.; Sikic, B.I.; Brody, J.; Nemunaitis, J.; Feldman, A.; Hawthorne, T.R.; Rawls, T.; Keler, T.; et al. Safety and activity of varlilumab, a novel and first-in-class agonist anti-CD27 antibody, for hematologic malignancies. *Blood Adv.* 2020, 4, 1917–1926. [CrossRef]
- Sanborn, R.E.; Pishvaian, M.; Callahan, M.; Weise, A.; Sikic, B.; Rahma, O.; Cho, D.; Rizvi, N.; Bitting, R.; Starodub, A.; et al. Anti-CD27 agonist antibody varlilumab (varli) with nivolumab (nivo) for colorectal (CRC) and ovarian (OVA) cancer: Phase (Ph) 1/2 clinical trial results. *J. Clin. Oncol.* 2018, *36*, 3001. [CrossRef]
- 69. Barroso-Sousa, R.; Ott, P.A. Transformation of old concepts for a new era of cancer immunotherapy: Cytokine therapy and cancer vaccines as combination partners of PD1/PD-L1 Inhibitors. *Curr. Oncol. Rep.* **2018**, *21*, 1. [CrossRef]
- 70. Golay, J.; Andrea, A.E. Combined anti-cancer strategies based on anti-checkpoint inhibitor antibodies. *Antibodies* **2020**, *9*, 17. [CrossRef]
- 71. Doberstein, S.K. Bempegaldesleukin (NKTR-214): A CD-122-biased IL-2 receptor agonist for cancer immunotherapy. *Expert Opin. Biol. Ther.* **2019**, *19*, 1223–1228. [CrossRef]
- 72. Rouanne, M.; Zitvogel, L.; Marabelle, A. Pegylated engineered IL2 plus anti–PD-1 monoclonal antibody: The nectar comes from the combination. *Cancer Discov.* **2020**, *10*, 1097–1099. [CrossRef] [PubMed]
- Khushalani, N.I.; Diab, A.; Ascierto, P.A.; Larkin, J.; Sandhu, S.; Sznol, M.; Koon, H.B.; Jarkowski, A.; Zhou, M.; Statkevich, P.; et al. Bempegaldesleukin plus nivolumab in untreated, unresectable or metastatic melanoma: Phase III PIVOT IO 001 study design. *Future Oncol.* 2020, *16*, 2165–2175. [CrossRef]
- Fujii, R.; Jochems, C.; Tritsch, S.R.; Wong, H.C.; Schlom, J.; Hodge, J.W. An IL-15 superagonist/IL-15Ralpha fusion complex protects and rescues NK cell-cytotoxic function from TGF-beta1-mediated immunosuppression. *Cancer Immunol. Immunother.* 2018, 67, 675–689. [CrossRef] [PubMed]
- 75. Knudson, K.M.; Hodge, J.W.; Schlom, J.; Gameiro, S.R. Rationale for IL-15 superagonists in cancer immunotherapy. *Expert Opin. Biol. Ther.* **2020**, *20*, 7. [CrossRef] [PubMed]
- 76. Rosario, M.; Liu, B.; Kong, L.; Collins, L.I.; Schneider, S.E.; Chen, X.; Han, K.; Jeng, E.K.; Rhode, P.R.; Leong, J.W.; et al. The IL-15-based ALT-803 complex enhances FcgammaRIIIa-triggered NK cell responses and in vivo clearance of B cell lymphomas. *Clin. Cancer Res.* 2016, 22, 596–608. [CrossRef] [PubMed]
- 77. Waldmann, T.A. The shared and contrasting roles of IL2 and IL15 in the life and death of normal and neoplastic lymphocytes: Implications for cancer therapy. *Cancer Immunol. Res.* **2015**, *3*, 219–227. [CrossRef]
- 78. Kim, P.S.; Kwilas, A.R.; Xu, W.; Alter, S.; Jeng, E.K.; Wong, H.C.; Schlom, J.; Hodge, J.W. IL-15 superagonist/IL-15RalphaSushi-Fc fusion complex (IL-15SA/IL-15RalphaSu-Fc; ALT-803) markedly enhances specific subpopulations of NK and memory CD8+ T cells, and mediates potent anti-tumor activity against murine breast and colon carcinomas. *Oncotarget* 2016, 7, 16130–16145. [CrossRef]
- 79. Knudson, K.M.; Hicks, K.C.; Alter, S.; Schlom, J.; Gameiro, S.R. Mechanisms involved in IL-15 superagonist enhancement of anti-PD-L1 therapy. *J. Immunother. Cancer* 2019, 7, 82. [CrossRef]
- 80. Wrangle, J.M.; Velcheti, V.; Patel, M.R.; Garrett-Mayer, E.; Hill, E.G.; Ravenel, J.G.; Miller, J.S.; Farhad, M.; Anderton, K.; Lindsey, K.; et al. ALT-803, an IL-15 superagonist, in combination with nivolumab in patients with metastatic non-small cell lung cancer: A non-randomised, open-label, phase 1b trial. *Lancet Oncol.* 2018, 19, 694–704. [CrossRef]
- 81. Liu, S.; Ren, J.; Ten Dijke, P. Targeting TGFβ signal transduction for cancer therapy. *Signal Transduct. Target. Ther.* **2021**, *6*, 8. [CrossRef]
- Van den Bulk, J.; de Miranda, N.F.C.C.; Ten Dijke, P. Therapeutic targeting of TGF-β in cancer: Hacking a master switch of immune suppression. *Clin. Sci.* 2021, 135, 35–52. [CrossRef]
- 83. Armitage, J.D.; Newnes, H.V.; McDonnell, A.; Bosco, A.; Waithman, J. Fine-tuning the tumour microenvironment: Current perspectives on the mechanisms of tumour immunosuppression. *Cells* **2021**, *10*, 56. [CrossRef]

- 84. Lee, H.J. Recent advances in the development of TGF-β signaling inhibitors for anticancer therapy. *J. Cancer Prev.* **2020**, *25*, 213–222. [CrossRef]
- Ni, G.; Zhang, L.; Yang, X.; Li, H.; Ma, B.; Walton, S.; Wu, X.; Yuan, J.; Wang, T.; Liu, X. Targeting interleukin-10 signalling for cancer immunotherapy, a promising and complicated task. *Hum. Vaccines Immunother.* 2020, 16, 2328–2332. [CrossRef]
- 86. Strauss, J.; Heery, C.R.; Schlom, J.; Madan, R.A.; Cao, L.; Kang, Z.; Lamping, E.; Marté, J.L.; Donahue, R.N.; Grenga, I.; et al. Phase I trial of M7824 (MSB0011359C), a bifunctional fusion protein targeting PD-L1 and-TGF-β, in advanced solid tumors. *Clin. Cancer Res.* 2018, 24, 1288–1295. [CrossRef]
- 87. Leone, R.D.; Emens, L.A. Targeting adenosine for cancer immunotherapy. J. Immunother. Cancer 2018, 6, 57. [CrossRef]
- Vijayan, D.; Young, A.; Teng, M.W.L.; Smyth, M.J. Targeting immunosuppressive adenosine in cancer. *Nat. Rev. Cancer* 2017, 17, 709–724. [CrossRef]
- 89. Helms, R.S.; Powell, J.D. Rethinking the adenosine-A2AR checkpoint: Implications for enhancing anti-tumor immunotherapy. *Curr. Opin. Pharmacol.* **2020**, *53*, 77–83. [CrossRef]
- 90. Vigano, S.; Alatzoglou, D.; Irving, M.; Ménétrier-Caux, C.; Caux, C.; Romero, P.; Coukos, G. Targeting adenosine in cancer immunotherapy to enhance t-cell function. *Front. Immunol.* **2019**, *10*, 925. [CrossRef]
- 91. Zhang, J.; Yan, W.; Duan, W.; Wüthrich, K.; Cheng, J. Tumor immunotherapy using A2A adenosine receptor antagonists. *Pharmaceuticals* **2020**, *13*, 237. [CrossRef]
- 92. Cheong, J.E.; Sun, L. Targeting the IDO1/TDO2-KYN-AhR pathway for cancer immunotherapy—challenges and opportunities. *Trends Pharmacol. Sci.* **2018**, *39*, 307–325. [CrossRef]
- Labadie, B.W.; Bao, R.; Luke, J.J. Reimagining IDO pathway inhibition in cancer immunotherapy via downstream focus on the tryptophan-kynurenine-aryl hydrocarbon axis. *Clin. Cancer Res.* 2019, 25, 1462–1471. [CrossRef]
- Zhai, L.; Bell, A.; Ladomersky, E.; Lauing, K.L.; Bollu, L.; Sosman, J.A.; Zhang, B.; Wu, J.D.; Miller, S.D.; Meeks, J.J.; et al. Immunosuppressive IDO in cancer: Mechanisms of action, animal models, and targeting strategies. *Front. Immunol.* 2020, 11, 1185. [CrossRef]
- Zhai, L.; Ladomersky, E.; Lenzen, A.; Nguyen, B.; Patel, R.; Lauing, K.L.; Wu, M.; Wainwright, D.A. IDO1 in cancer: A gemini of immune checkpoints. *Cell. Mol. Immunol.* 2018, 15, 447–457. [CrossRef]