



Modulated Gene Expression of *Toxoplasma gondii* Infected Retinal Pigment Epithelial Cell Line (ARPE-19) via PI3K/Akt or mTOR Signal Pathway

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Abstract: Due to the critical location and physiological activities of the retinal pigment epithelial (RPE) cell, it is constantly subjected to contact with various infectious agents and inflammatory mediators. However, little is known about the signaling events in RPE involved in *Toxoplasma gondii* infection and development. The aim of the study is to screen the host mRNA transcriptional change of 3 inflammation-related gene categories, PI3K/Akt pathway regulatory components, blood vessel development factors and ROS regulators, to prove that PI3K/Akt or mTOR signaling pathway play an essential role in regulating the selected inflammation-related genes. The selected genes include PH domain and leucine-rich-repeat protein phosphatases (PHLPP), casein kinase2 (CK2), vascular endothelial growth factor (VEGF), pigment epithelium-derived factor (PEDF), glutamate-cysteine ligase (GCL), glutathione S-transferase (GST), and NAD(P)H: quinone oxidoreductase (NQO1). Using reverse transcription polymerase chain reaction (RT-PCR) and quantitative real-time reverse transcription polymerase chain reaction (qRT-PCR), we found that *T. gondii* up-regulates PHLPP2, CK2 β , VEGF, GCL, GST, and NQO1 gene expression levels, but down-regulates PHLPP1 and PEDF mRNA transcription levels. PI3K inhibition and mTOR inhibition by specific inhibitors showed that most of these host gene expression patterns were due to activation of PI3K/Akt or mTOR pathways with some exceptional cases. Taken together, our results reveal a new molecular mechanism of these gene expression change dependent on PI3K/Akt or mTOR pathways and highlight more systematic insight of how an intracellular *T. gondii* can manipulate host genes to avoid host defense.

Key words: *Toxoplasma gondii*, reactive oxygen species, PI3K/Akt pathway, mTOR, ARPE-19 cell

INTRODUCTION

Toxoplasma gondii is one of the most widespread zoonotic pathogens in the world that can infect almost any nucleated vertebrate cell. Most of toxoplasmosis patients are asymptomatic in immunocompetent individuals, but ocular lesions may be present in up to 20% of infected patients [1]. Clinical examination revealed free tachyzoites and cysts in the Retinal pigment epithelial (RPE) cell and the retina of *T. gondii*-induced retinochoroiditis [2]. RPE cell is an integral part of the neuroretina in the posterior segment of the eye, which normal

functions is phagocytosis and the recycling of components of the outer segments of the rods and cones of the retina during physiological photoreceptor recycling. RPE cell also forms a blood-retinal barrier between the highly vascularized choroid and the retina with a complex architecture of neuronal cells [3]. Hence, studies of the influences of *T. gondii* replication in RPE cells and the responses of the host cells to parasite invasion would be valuable in understanding the immunopathological basis of ocular toxoplasmosis.

Following invasion, *T. gondii* resides and replicates within a parasitophorous vacuole (PV) that resists host endosomal acidification and lysosomal fusion. Once the *T. gondii* begins growing and dividing within the PV, it must be deprived nutrients such as glucose, lipids and amino acids and nucleotides from the host cell [4]. *T. gondii* can secrete multiple effector molecules into host cell, and these effectors are known to interfere with host cell signaling pathways and alter host defenses al-

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though the molecular details are unclear [5]. Previous data demonstrated that *T. gondii* can recognize and interfere with host signaling pathways to subvert innate immunity to establish a life-long chronic infection [6]. One of the initial reactions of host innate immunity is the generation of reactive oxygen species (ROS) in reaction to parasites. Invasion and expulsion of parasites [7]. It has been reported that upon infection with *T. gondii*, host phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt) pathway was activated, and this activation was required for host cell resistance to apoptosis [8], and Akt/mTOR activation was also required for *T. gondii* to coordinate control of host centrosome position, organelle distribution, and migratory response [9]. However, most of the mechanisms and pathways involved are still not clearly understood.

T. gondii-induced phenotypic changes are characterized by marked changes in gene expression [10]. There is a report that host mRNAs display altered abundance increased 15% relative to uninfected cells after 24 hr post-infection [11]. The eye is a distinctive immunologic environment that aimed at reducing inflammation. RPE cells are constantly subjected to contact with numerous infectious agents and inflammatory. After parasite infection, changes of various host gene expressions are observed, which can be deadly or beneficial to the parasite. Since the combination of 3 individual host signals or molecules would determine the fate of the infected parasite, identification of the match between specific signal pathways and downstream target genes would be essential for establishing a strategy to control the infected parasite. Therefore, we categorized selected host genes into 3 categories, signal modulation, vascular formation, and ROS regulation, and to determine which are under control of PI3K/Akt or mTOR pathways.

PHLPP has been identified to serve as a negative regulator of Akt by dephosphorylation of Akt "Ser-473" directly. Two highly related isoforms in this family, PHLPP1 and PHLPP2, both dephosphorylate the same residue on Akt, however, they differentially terminate Akt signaling by regulating distinct Akt isoforms. PHLPP1 specifically modulates the phosphorylation of Akt2, nevertheless PHLPP2 purposely dephosphorylates Akt3 [12]. A number of studies have clearly demonstrated that PTEN antagonizes PI3K activity and negatively regulates its downstream target, the serine/threonine kinase Akt [13]. CK2 is a serine/threonine kinase that consists of 2 catalytic subunits (α , α') and the regulatory subunit (β) [14]. Recent reports have shown that CK2 can induce PTEN phosphorylation to inhibit PTEN activity and is also essential to regulate PI3K/Akt

signaling pathway [15].

Choroidal neovascularization (CNV) is a severe complication of ocular toxoplasmosis [16]. Among the cytokines involved in pathological angiogenesis, vascular endothelial growth factor (VEGF) and pigment epithelial derived factor (PEDF) serve as the most potent angiogenic stimulators and angiogenic inhibitors, respectively [17,18]. Counterbalancing VEGF angiogenic stimulation and PEDF inhibition is essential to maintain the angiogenic homeostasis.

Recent studies provide evidence that *T. gondii* can diminish ROS production in the host cell, but the exact mechanism is unclear [19]. It is well known that a family of proteins called "Phase II genes" can protect against oxidative stress by catalyzing a wide variety of reactions that neutralize reactive oxygen species, toxic electrophiles and carcinogens when they are up-regulated [20]. However, the gene expression levels of selected Phase II enzymes such as: γ -glutamylcysteine ligase and glutathione synthetase, that regulates the key steps in glutathione (GSH) biosynthesis; NQO-1, a reducing agent that plays a crucial role in antioxidant defenses with the cofactor NADH or NADPH were not reported in *T. gondii* infection. GSH is a well-studied tri-peptide and has numerous roles in protecting cells from oxidants and maintaining the cellular thiol redox status. The rate-limiting enzyme in the de novo synthesis of GSH is GCL. GCL consists of a catalytic heavy subunit (GCLC) and a modulatory light subunit (GCLM) [21]. Alpha-class glutathione S-transferases (GSTA) can protect cells against oxidative stress [22]. GSTA4 enzyme, the alpha subunit that exhibits the highest activity against 4-hydroxynonenal (4-HNE) also efficiently protects against oxidative damage mediated by this cytotoxic product of lipid peroxidation generated by ROS overproduction [23]. Glutathione S-transferase pi 1 (GSTP1) is a subgroup of GST family, which provides cellular protection against free radical and carcinogenic compounds due to its detoxifying function [24].

Our results indicate that RPE cells might have a set of genes which can be regulated by *T. gondii*-induced PI3K/Akt pathway and the sum of individual gene expression changes (PI3K-dependent or -independent) will contribute to the PI3K/Akt positive feedback, vascular formation, and ROS down-regulation. Eventually, these gene expression change patterns will be beneficial for *T. gondii* growth. This study may provide new potential drug targets for the control of RPE-infected *T. gondii* in cases of ocular toxoplasmosis.

MATERIALS AND METHODS

Parasites

Tachyzoites of the virulent *Toxoplasma gondii* RH strain were used. Parasite were maintained by 3 days passage on human retinal pigment epithelium (ARPE-19 cells) cultured in a 1:1 mixture of Dulbecco's modified Eagle's medium (DMEM) with F12 (DMEM/F12) supplemented with 10% heat-inactivated fetal bovine serum (FBS), and antibiotic-antimycotics (all from Gibco, Grand Island, New York, USA).

Cell culture

ARPE-19 cell line was obtained from American Type Culture Collection (ATCC, Manassas, Virginia, USA) and used at passages 15 to 20 in all experiments. Cultures of ARPE-19 cells were maintained at 37°C, DMEM-F12 medium supplemented with 10% FBS in a humidified atmosphere containing 5% CO₂.

Western blot

ARPE-19 cells were cultured in 6-well dishes and infected with RH tachyzoites for 24 hr and then treated for 1 hr with specific PI3-kinase inhibitors LY294002 (LY 0.1, 1, 10 μM),

Wortmannin (WM 5, 50, 500 ng/ml), or mTOR inhibitor Rapamycin (Rap 5, 50, 500 ng/ml). The protein samples were separated by SDS-PAGE and transferred onto polyvinylidene difluoride membranes and probed with the relevant antibodies. The blots were developed using an enhanced ECL chemiluminescence detection kit (GE Healthcare, Little Chalfont, UK). The primary antibodies used were rabbit anti-phospho-Akt (Thr308), phospho-Akt (Ser473), Akt, phospho-mTOR, mTOR, phospho-p70 S6 kinase and p70 S6 kinase (Cell Signaling Technology, Danvers, Massachusetts, USA).

Profiling of host gene expression analysis by reverse transcription-PCR (RT-PCR)

ARPE-19 cells were cultured in 6-well dishes and infected with RH tachyzoites for 24 hr, and then treated for 1 hr with specific PI3-kinase inhibitors LY294002 (LY 10 μM), wortmannin (WM 500 ng/ml), or mTOR inhibitor rapamycin (Rap, 500 ng/ml). Total RNA was extracted using Trizol Reagent (Invitrogen Life Technologies, Carlsbad, California, USA). The cDNA was synthesized using a Moloney murine leukemia virus cDNA synthesis kit (Enzymomics, Daejeon, Korea) with 2 μg of total RNA. PCR was performed with the Maxime PCR

Table 1. Primer sequences used for PCR and qRT-PCR in this study

Gene name	Primer sequence (5' to 3')		Products size (bp)
	RT-PCR	qRT-PCR	
PHLPP1	F-ACACCGTGATTGCTCACTCC R-TTCCAGTCAGGTCTAGCTCC	F-ACACCGTGATTGCTCACTCC R-TTCCAGTCAGGTCTAGCTCC	156/156
PHLPP2	F-AGGTTCTGAGCATCTCTTC R-GTTCAGGCCCTTCAGTTGAG	F-AGGTTCTGAGCATCTCTTC R-GTTCAGGCCCTTCAGTTGAG	128/128
CK2α	F-ATTGAATTAGATCCAGTTT R-GGTTGGCAGCAGCAATCACT	F-CCCAACATCATCACACTGGC R-CCAGGGCCTTCAGAATCTCA	346/159
CK2β	F-ACTGTGAGAACAGCCAATG R-CTTGAAGTTGCTGGCGGCTT	F-TCAAGACACCATCACACGGA R-CTGGGCTCTTGAAGTTGCTG	287/187
VEGF	F-GAAGTGGTGAAGTTCATGGATGTC R-CGATCGTTCTGTATCAGTCTTTCC	F-CTGCTTTGGGTGCATTGGAG R-ACCAGGGTCTCGATTGGATG	165/121/164
PEDF	F-CAGAAGAACCCTCAAGAGTGCC R-CTTCATCCAAGTAGAAATCCTC	F-AGAACCTCAAGAGTGCCCTCC R-GAGCTTCCCTTTCATCTGCG	310/176
GCLC	F-GAGGCTATGTGTACAGACATTGATTGTCG R-GTGTACTCTCTGCAGCGAGCTCCGTG	F-CAGGTGACATTCCAAGCCTG R-CGGTAAAAGGGAGATGCAGC	688/120
GCLM	F-GTCCAOCACAGCGAGGAGCTTCATG R-GATCATTGTGAGTCAACAGCTGTATG	F-CTGTATCAGTGGGCACAGGT R-GTGCCTTGAATGTCAGGAA	542/198
GSTP1	F-GCTGCGCGGCCCTGCGCATGCTG R-GCAGGTTGTAGTC AGCGAAGGAG	F-GGACTTGCTGCTGATCCATG R-ATTGATGGGGAGGTTACCGT	429/145
GSTA4	F-GGATGAAGTTGGTACACCCGAAG R-GAGGCTTCTCTTGTGCTGCCAGGTTCAAG	F-CAACAAGTGCCCATGGTTGA R-CTGGGCCATGTTAACCAT	435/223
NQO1	F-ATGGTCGGCAGAAGAGCACTGATCG R-TTTTCTAGCTTTGATCTGGTTGTCAGTTGGG	F-ACTCTCTGCAAGGGATCCAC R-TCTCCAGGCGTTTCTCCAT	708/174
GAPDH	F-ACCACAGTCCATGCCATCAC R-TCCACCACCCTGTTGCTGT	F-GAAGGTGAAGGTCGGAGTCA R-CCTGGAAGATGGTATGGGA	452/131

premix kit (Takara Bio, Otsu, Japan). All PCR reactions were performed with a MyCycler instrument (Bio-Rad, Richmond, California, USA) for 25 cycles. The PHLPP1 and PHLPP2 primer sequences for RT-PCR were described by Hirano et al. [25]. VEGF and PEDF primer sequences for RT-PCR were described previously [26,27]. The GCLC, GCLM, GSTP1, GSTA4, and NQO1 primer sequences used were described by Saravananakumar et al. [28]. Amplified products were electrophoresed in a 2% agarose gel and visualized with ethidium bromide. The relative intensity of bands for each mRNA was related to the intensity of the autoradiogram band of the internal control, glyceraldehyde-3-phosphate dehydrogenase (GAPDH). Quantification of mRNA was performed using an imaging densitometer (Bio-Rad Laboratories, Otsu, Japan).

Profiling of host gene expression analysis by quantitative real-time reverse transcription-PCR (qRT-PCR)

After isolation total mRNA with Trizol Reagent, the cDNA was synthesized using the Primer Script RT reagent kit (Takara) with 1 µg of total mRNA. Real-time PCR was performed using the SYBR Premix Ex Taq™ II (2×) mix (Takara) and specific primer sets (Table 1) with the Rotor-Gene Q Real Time PCR Systems (Qiagen, Hilden, Germany). The thermal cycle parameters were 95°C for 10 min and 40 cycles of each for 5 sec at 95°C and 40 sec at 60°C. Ct values for specific genes were corrected by the Ct value for the GAPDH housekeeping gene and expresses as Δ Ct. Data represent mean \pm SD, values of Δ Ct.

Statistical analysis

The unpaired Student's *t*-test was used for statistical analysis. Three independent experimental data are presented as means \pm SD. *P*-values less than 0.05 are considered significant.

RESULTS

T. gondii induced PI3K/Akt and mTOR/S6 signaling activity

It is well known that *T. gondii* trigger PI3-kinase signaling to activate Akt pathway upon infection in vivo and in vitro [11]. To investigate the impact of *T. gondii*-infection on the PI3K/Akt or mTOR pathways, ARPE-19 cells were infected with *T. gondii* at multiplicity of infection (MOI) 10 for 24 hr and then treated with specific PI3K inhibitors LY294002 (LY 0.1, 1, 10 µM), wortmannin (WM 5, 50, 500 ng/ml) or mTOR inhibitor rapamycin (Rap 5, 50, 500 ng/ml) for 1 hr and the activity of each signaling protein were measured by western blot. As

shown in Fig. 1A, the phospholation levels of Akt (Thr308), Akt (Ser473), mTOR and S6 were elevated remarkably by *T. gondii* infection. However, *T. gondii*-induced increased activity Akt and mTOR were markedly attenuated by the LY294002, wortmannin and rapamycin. Furthermore, the densitometric analysis of western blot also confirmed that phosphorylation protein levels of Akt (Thr308), Akt (Ser473), mTOR and S6 were significantly increased by *T. gondii*, but LY294002, wortmannin and rapamycin significantly prevented the *T. gondii*-induced activity of Akt and mTOR, respectively (Fig. 1A-E). These observations together suggest that *T. gondii*-infection both activates the PI3K/Akt and mTOR pathways.

Differential regulation of PHLPP1, PHLPP2, and CK2β by *T. gondii*-infection through PI3K signal pathway

To directly address the consequences of activation of PI3K pathway induced by *T. gondii*-infection on modulation of host gene expression, we first analyzed mRNA levels of PHLPP1, PHLPP2, CK2α, and CK2β. RT-PCR results showed that *T. gondii*-infection suppressed PHLPP1 mRNA expression. Interestingly, the PI3 kinase inhibitor, LY294002, was able to repress the effect of *T. gondii* to reverse the PHLPP1 mRNA level of host cell to normal level, but not wortmannin. The mTOR inhibitor rapamycin also had no effect on *T. gondii*-induced PHLPP1 gene expression. Surprisingly, PHLPP2 mRNA expression level was elevated with *T. gondii* infection, and all 3 inhibitors restrained the *T. gondii*-dependent PHLPP2 mRNA induction. *T. gondii*-infection had no effect on CK2α gene expression, but significantly up-regulated CK2β mRNA level. The induction of CK2β in response to *T. gondii* infection was partially increased by rapamycin (Fig. 2A). Similarly, *T. gondii*-induced PHLPP1, PHLPP2, CK2α, and CK2β mRNA expression levels and effect of LY294002, wortmannin or rapamycin on these gene expression patterns were also confirmed by qRT-PCR method (Fig. 2B-E).

These results suggest that PHLPPs are under control of PI3 kinase pathway; however, CK2β expression was insensitive to PI3K inhibitors but was sensitive to rapamycin which suggests that CK2β might be under control of mTOR. Their sensitivity difference to LY294002, wortmannin or rapamycin implied the presence of micro-regulation system for these enzymes and signaling pathways.

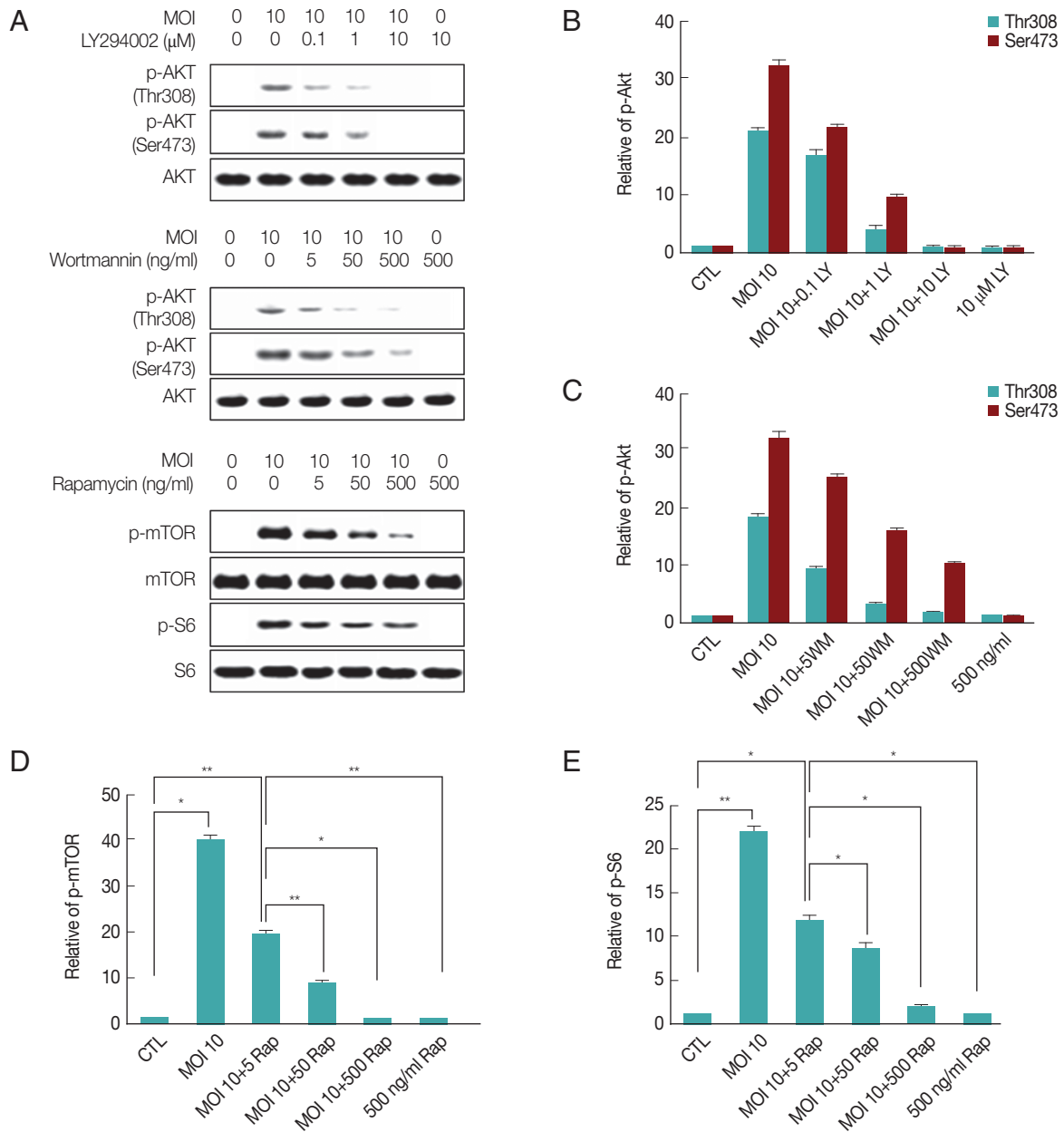


Fig. 1. *T. gondii* infection induces PI3K/Akt and mTOR/S6 signaling activity, and the activity was suppressed by PI3K/Akt or mTOR inhibitors. ARPE-19 cells were infected with *T. gondii* at MOI 10 for 24 hr and then treated with specific PI3K inhibitors LY294002 (LY 0.1, 1, 10 μM), wortmannin (WM 5, 50, 500 ng/ml) or mTOR inhibitor rapamycin (Rap 5, 50, 500 ng/ml) for 1 hr. The activity of each signaling protein levels were measured by western blot (A). Bar plot depicting the phospho-Akt (Thr308)/Total Akt, phospho-Akt (Ser473)/Total Akt, phospho-mTOR/Total mTOR, phospho-S6/Total S6 (B-E) ratios as determined by densitometric analysis of western blot and expressed as fold change compared with control ARPE-19 cells. For all panels, data are presented as the mean ± SD. **P* < 0.05, ***P* < 0.01 compared with control or MOI 10 of *T. gondii*-infected groups. All data shown are representative of 3 independent experiments.

T. gondii infection up-regulated pro-angiogenic VEGF and down-regulated anti-angiogenic PEDF in ARPE-19 cells

Choroidal neovascularization is a severe complication of ocular toxoplasmic retinochoroiditis [16]. Increased expression

of VEGF, in addition to compromise in Bruch's membrane and inflammation secondary to infection with *T. gondii*, may contribute to neovascular disease in ocular toxoplasmosis [29]. Therefore, we examined whether the *T. gondii* infection can change the mRNA expression levels of VEGF and PEDE, well-

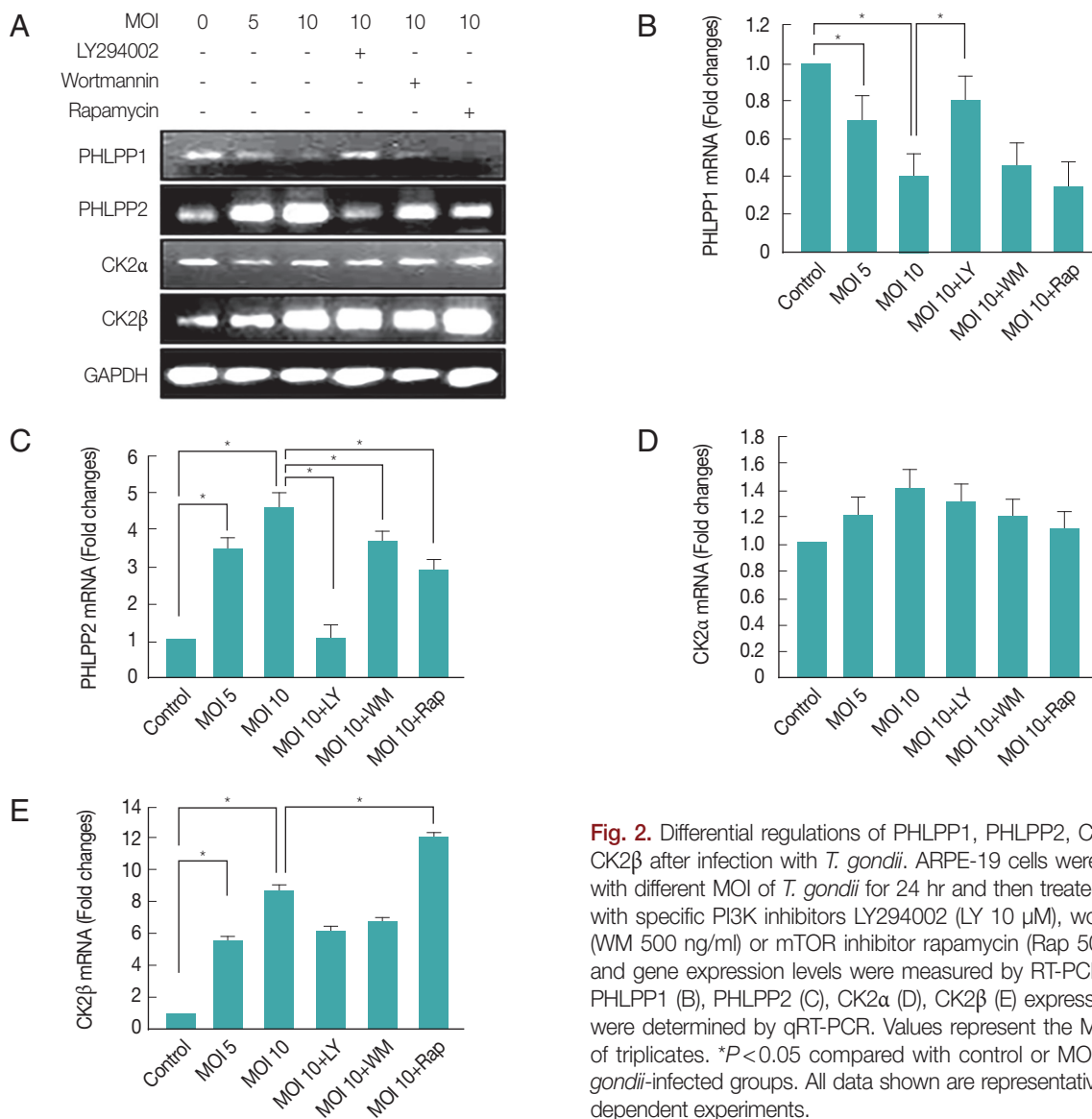


Fig. 2. Differential regulations of PHLPP1, PHLPP2, CK2α, and CK2β after infection with *T. gondii*. ARPE-19 cells were infected with different MOI of *T. gondii* for 24 hr and then treated for 1 hr with specific PI3K inhibitors LY294002 (LY 10 μM), wortmannin (WM 500 ng/ml) or mTOR inhibitor rapamycin (Rap 500 ng/ml) and gene expression levels were measured by RT-PCR (A) and PHLPP1 (B), PHLPP2 (C), CK2α (D), CK2β (E) expression levels were determined by qRT-PCR. Values represent the Mean ± SD of triplicates. **P* < 0.05 compared with control or MOI 10 of *T. gondii*-infected groups. All data shown are representative of 3 independent experiments.

known factors for neovascularization regulation in PI3K-Akt signaling dependent manner. As shown in Fig. 3A and 3B, *T. gondii* infection increased VEGF mRNA level of host cell in a MOI (parasite burden)-dependent manner, but post-infection treatment of the inhibitors of PI3K or mTOR did not suppress *T. gondii* induced VEGF isoform transcription (VEGF₁₆₅ and VEGF₁₂₁) gene expression. These results suggest that *T. gondii* can up-regulate VEGF expression in ARPE-19 cells, but this effect is not under the control of PI3K signaling pathway.

However, PEDF mRNA level was 4-5 times down-regulated by *T. gondii* in a MOI (parasite burden)-dependent manner, and treatment of LY294002 was able to restore the reduced PEDF level to basal level. Wortmannin and rapamycin also

showed partial suppression of *T. gondii* effect on PEDF expression (Fig. 3A, C). These results suggest *T. gondii* induced down-regulation of PEDF expression is dependent on PI3K/Akt and partially on mTOR signaling pathways.

Effects of *T. gondii*-infection on GCLC, GCLM, GSTP1, GSTA4, and NQO1 mRNA expressions

We further investigated *T. gondii* modulated Phase II enzymes related gene expression levels via PI3K/Akt or mTOR pathway. As shown in Fig. 4A and 4B, *T. gondii*-infection significantly increased GCLC expression level as compared with uninfected control cell, and treatment of infected cell with LY294002, wortmannin or rapamycin significantly reduced its expression

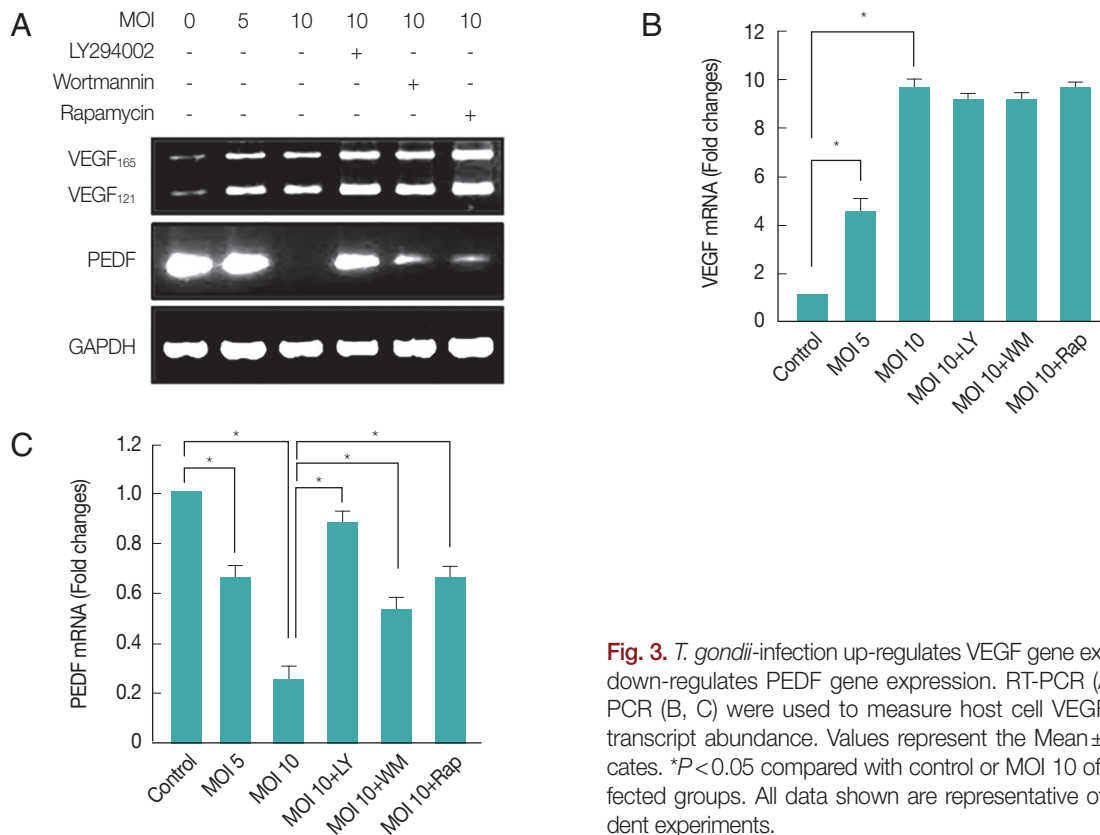


Fig. 3. *T. gondii*-infection up-regulates VEGF gene expression but down-regulates PEDF gene expression. RT-PCR (A) and qRT-PCR (B, C) were used to measure host cell VEGF and PEDF transcript abundance. Values represent the Mean \pm SD of triplicates. * $P < 0.05$ compared with control or MOI 10 of *T. gondii*-infected groups. All data shown are representative of 3 independent experiments.

levels based on RT-PCR and qRT-PCR. Surprisingly, *T. gondii*-infection dramatically increased GCLM mRNA levels, and treatment of infected cells with LY294002 or wortmannin had no effect on *T. gondii*-induced up-regulation of GCLM gene expression, but treatment with rapamycin significantly suppressed *T. gondii*-induced GCLM mRNA expression (Fig. 4A, C). These results show that mTOR but not PI3K was important for *T. gondii* to induce GCLM transcription regulation.

Furthermore, *T. gondii*-infection significantly induced both GSTP1 and GSTA4 expression in host cells, when *T. gondii*-infected cells were treated with PI3K or mTOR inhibitors, these inhibitors had no effect on *T. gondii* induced GSTP1 gene expression. In contrast, LY294002, wortmannin or rapamycin remarkably suppressed *T. gondii*-induced GSTA4 mRNA transcription to basal level (Fig. 4D, E). Indicating that GSTP1 expression induced by *T. gondii* is insensitive to PI3K or mTOR pathway, but GSTA4 expression is dependent on PI3K and mTOR signaling.

We focused on another oxidoreductase, NQO1, which uses either NADH or NADPH as a reducing cofactor to catalyze the obligate 2-electron reduction of quinones to hydroquinones

[30]. *T. gondii*-infection significantly increased (about 1.7 folds) NQO1 mRNA expression and LY294002 was significantly inhibited the *T. gondii*-induced NQO1 gene expression (Fig. 4A, F). These results provide strong evidence that *T. gondii* may cause Phase II enzyme regulation through the PI3K/Akt pathway.

DISCUSSION

This is the first study to investigate the host PI3 kinase-Akt pathway affiliated genes upon to *T. gondii* infection in human RPE cell. Since *T. gondii* is an intracellular parasite, the retina sustains the major damage and primary insult. We found that *T. gondii* replication is associated with expression level changes of multiple known and novel genes involved in signaling and ROS regulation in ARPE-19 cells on host PI3K/Akt or mTOR pathway activation.

T. gondii have devised strategies to extend the life of infected host cells by targeting key signal transduction pathways such as PI3 kinase-Akt or mTOR pathways [31,32]. In vitro experiments revealed that the activation of PKB in host cell infected

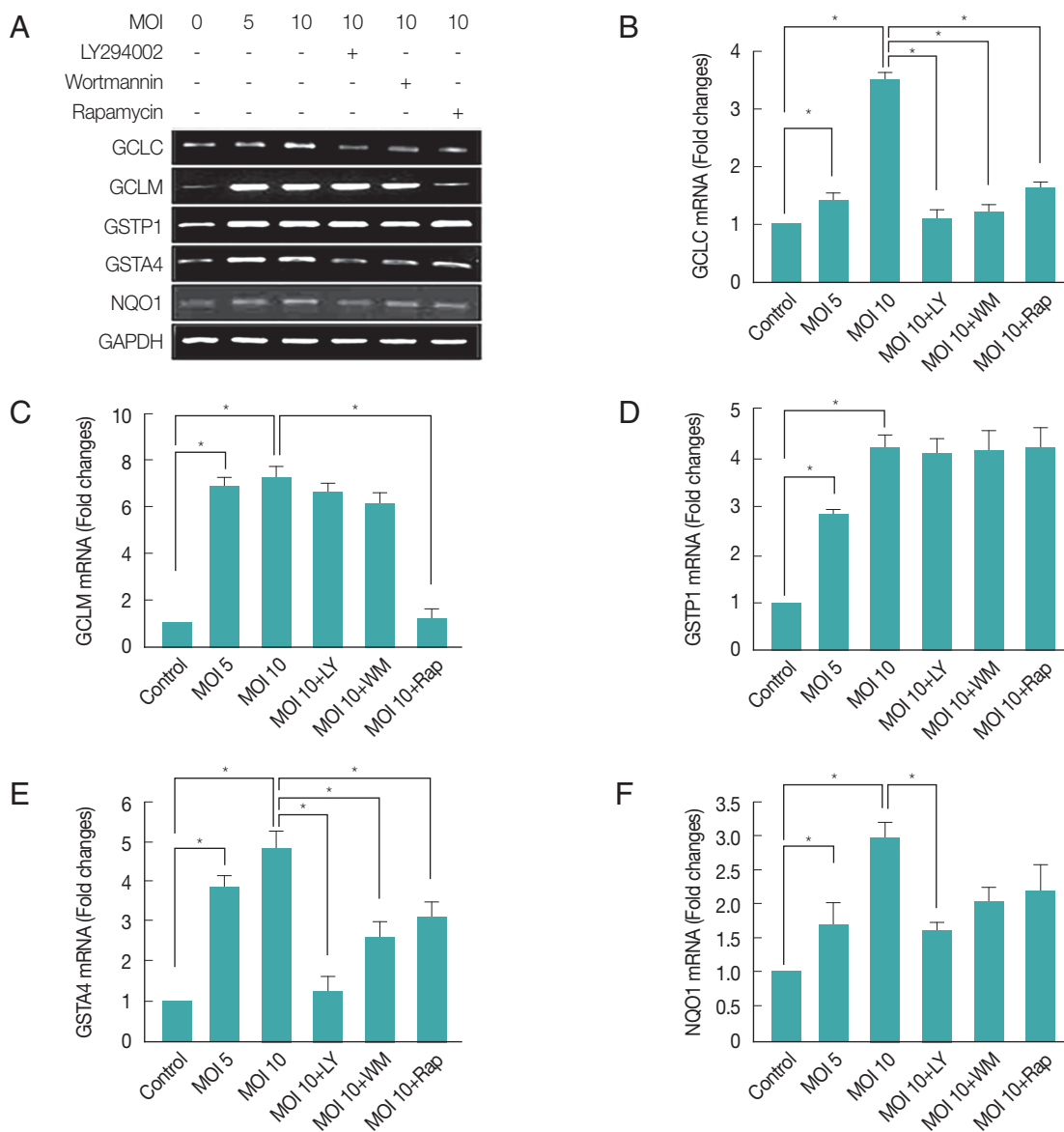


Fig. 4. *T. gondii*-induced regulation of “Antioxidants and Phase II Enzymes” in ARPE-19 cell. The mRNA levels of both the catalytic (GCLC) and modulatory (GCLM) subunits of GCL as well as 4 other phases 2 detoxification genes, GSTP1, GSTA4, and NQO1 were measured by RT-PCR (A) and qRT-PCR (B, C, D, E, F) after *T. gondii* infection and inhibitors treatment. Values represent the Mean ± SD of triplicates. * $P < 0.05$ compared with control or MOI 10 of *T. gondii*-infected groups. All data shown are representative of 3 independent experiments.

with *T. gondii* was dependent upon the host PI3K and Gi protein-coupled trans-membrane receptors (GiPCR) [8]. Gene silencing studies reveal that PHLPP1 knockdown or genetic deletion increase Akt phosphorylation at Ser473 which results in significant increase in Akt catalytic activity [33].

In this study, PHLPP1 mRNA expression was significantly reduced after *T. gondii* infection which can contribute to maintain high levels of activated Akt that may prevent host cell death. But for PHLPP2, it is observed that *T. gondii* infection induces

its gene expression of host cell. This phenomenon could be due to a compensation role of PHLPP2 for the loss of PHLPP1. Indeed, in human cells, it has been reported that knockdown of PHLPP1 resulted in greater expression of PHLPP2, and vice versa [34]. Treatment with PI3K inhibitor, LY294002, inverted *T. gondii*-induced the phosphatases mRNA expression pattern that PHLPP1 increased however PHLPP2 decreased. The *T. gondii*-induced reduction of Akt’s negative regulator, PHLPP1, provides a novel mechanism used by *T. gondii* to elongate the

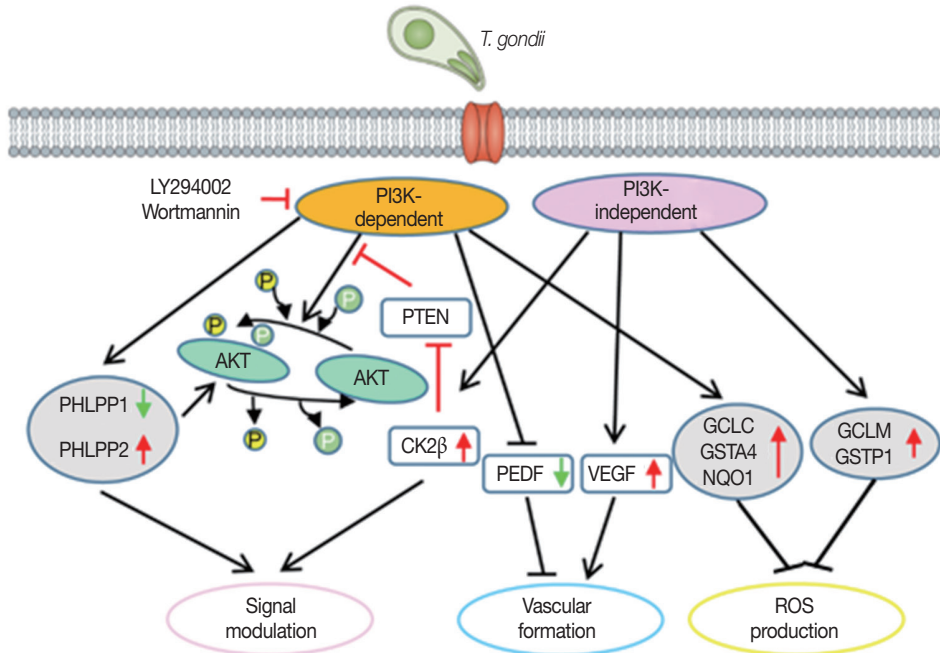


Fig. 5. The model of *T. gondii*-modulated PI3K/Akt or mTOR pathways on regulating 3 different cellular events to generate parasite-favorable host environment.

Akt activation in RPE cells.

It has been reported that CK2 kinase mediates phosphorylation of PTEN, and its phosphorylation inhibits PTEN function [15]. Gene silencing studies of CK2 indicated that CK2 β is essential for cell viability [35], based on the reports, it was wondered that the main regulated subunit by *T. gondii* is CK2 β but not CK2 α . A very interesting result of the present study is the relationship between CK2 β and the PI3K pathway, CK2 β showed a significant positive correlation with Akt and a nearly dramatic negative correlation with PTEN immunoprecipitation [36].

In the present study, CK2 α mRNA level was not sensitive to *T. gondii* infection, but CK2 β mRNA level increased highly in a moi-dependent manner. Inhibitor experiment showed that only rapamycin leads to additional up-regulation of CK2 β by *T. gondii*. These results suggest that endogenous PTEN level can be regulated by *T. gondii*-controlled CK2 β level, and mTOR functions as a negative regulating component in CK2 β gene expression.

Clinical experiments show that intravitreal anti-VEGF agents therapy plus anti-*T. gondii* medicines have potentials to be efficacious in treatment of ocular neovascularization secondary to ocular toxoplasmosis [29]. Recent studies have identified that PEDF inhibits angiogenesis in the eye and as a therapeutic

agent for retinal and choroidal diseases triggered by photoreceptor degenerations and abnormal neovascularization has been explored in animal models and is being evaluated in a clinical trial [36]. The counterbalance of VEGF and PEDF is supported by the previous demonstrations that either inhibition of the VEGF system or over-expression of PEDF inhibits choroidal neovascularization [37]. Our experimental results showed that *T. gondii* down-regulates PEDF gene expression, and causes the over-expression of VEGF, which will bring the disruption of the balance between VEGF and PEDF. This event might be a valuable target for therapeutic studies about ocular toxoplasmosis.

Inflammatory ROS is believed to play critical roles in various ocular diseases. Several studies have indicated that RPE cells are capable of producing ROS under certain conditions [38]. It has been reported that proliferation of *T. gondii* can diminish oxygen radical production in the macrophage cell in vivo, but most of the mechanisms and pathways involved are not still understood [19]. Recent studies have shown that the PI3K/Akt pathway is essential in regulating the Nrf2-ARE-dependent protection against oxidative stress in the RPE [39]. Our results have shown that *T. gondii* can induce the expression of the phase II detoxification genes, increase GLC, GST, and NQO1 genes expression, and confirmed that these gene

changes were mediated by PI3K/Akt or mTOR pathway. In accordance with our data, it was reported that protein levels of GSTA4 and activity of GST were elevated with the increase of Insulin signaling pathway [40]. However, it also has been reported that, in Human foreskin fibroblasts, *T. gondii* infection reduced GSTA2 levels [41]. The different reaction between GSTA2 and GSTA4 expression against *T. gondii* may imply various considerable factors; different cell context based on intracellular ROS level or nutrition conditions, possible mutual independent gene expression stimulation mechanisms, or even compensation mechanism based on other intracellular GST levels to sustain homeostasis of host cell. Further studies are required to solve the mystery of this observation.

In conclusion, we found that many host genes of ARPE-19 cells involved in Akt signal regulation, vascular formation and ROS suppression can be modulated by *T. gondii*-induced PI3K/Akt pathway. Based on their biological functions, we suggest that the compilation of these PI3K/Akt dependent and certain mTOR dependent gene expression change will promote the establishing of host environment which is favorable for infected parasite (Fig. 5). Characterization of PI3K/Akt or mTOR pathway regulation in response to *T. gondii* infection will provide a better understanding of the pathogenic mechanisms of ocular toxoplasmosis and may lead to more innovative methods for developing appropriate therapeutic therapy in correlation with metabolism disease such as diabetes mellitus or age-dependent vision loss.

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CONFLICT OF INTEREST

The authors declare no conflict of interest related to this study.

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