



Complete Genome Sequence of *Streptococcus mutans* Strain LAB761, Which Harbors Several Bacteriocin Loci, Isolated from a Caries-Active Child in Canada

Abdelahhad Barbour, a Richard Yi He, a Siew-Ging Gong, a DCéline M. Lévesque

^aFaculty of Dentistry, University of Toronto, Toronto, Ontario, Canada

ABSTRACT Streptococcus mutans LAB761 has been isolated from dental plaque collected from a child with severe caries. We report here the complete genome sequence of *S. mutans* strain LAB761, which has a chromosome of 2.0 Mb. The genome sequence reported herein contains several loci encoding double-glycine-motif peptides and lantibiotic and nonlantibiotic bacteriocins.

Itreptococcus mutans is an important human pathogen. It resides in the oral biofilm (dental plaque), and is one of the main causative agents of dental caries, the most prevalent streptococcal disease (1). The transmission of S. mutans usually occurs from mother to child via salivary transfer. However, detection of genotypes that are not shared with any household members suggest both horizontal and vertical routes of transmission (2). S. mutans possesses several virulence factors that contribute to its pathogenicity. These factors include its ability to adhere to the tooth surface as part of the multispecies biofilm community, to both produce and withstand highly acidic conditions, and to compete within the biofilm through the production of antimicrobial peptides called bacteriocins or mutacins (3, 4). S. mutans strain LAB761 was isolated from the supragingival plaque collected from facial and lingual smooth surfaces of primary maxillary incisors of a severe early childhood caries child age 3 years 4 months (REB protocol reference 32740). The plaque sample was plated on Mitis-Salivarius-Bacitracin agar supplemented with 20% sucrose using a spiral plater. Isolate LAB761 was verified as S. mutans by PCR (5) and 16S rRNA sequencing (6). Here, we present the complete genome sequence of this strain.

Strain LAB761 was cultivated in a 50-ml volume of Todd-Hewitt broth supplemented with 0.3% yeast extract at 37°C in air with 5% CO₂ for 18 h without agitation. Genomic DNA was extracted using an in-house protocol. Briefly, cells were lysed with lysozyme (50 mg/ml at 37°C for 1 h), and proteins were digested with proteinase K (20 mg/ml at 37°C for 15 min) and precipitated with cold potassium acetate. The DNA was fished out using a sterile glass pipette and treated with RNase. DNA was quantified using the Quant-iT PicoGreen double-stranded DNA (dsDNA) assay kit (Thermo Fisher). Wholegenome sequencing was performed using the PacBio sequencing technology. The DNA library was prepared following the Pacific Biosciences 20-kb template preparation using the BluePippin size-selection system protocol. Qualified genomic DNA was fragmented using the Covaris g-TUBE device and then end-repaired to prepare SMRTbell DNA template libraries (with a fragment size of 15 kb to 50 kb) selected using a BluePippin system. Sequencing was performed using a Pacific Biosciences RSII sequencer using the MagBead one cell per well (OCPW) protocol at the Génome Québec Innovation Centre and Canadian Centre for Computational Genomics (McGill University, Québec, Canada). PacBio sequencing using 1 single-molecule real-time (SMRT) cell generated a total of 42,960 raw subreads of average length 12,149 bp, with an N_{50} value of 26,099 bp. Contig assembly was done using the HGAP workflow using default settings (7). The

Citation Barbour A, He RY, Gong S-G, Lévesque CM. 2019. Complete genome sequence of *Streptococcus mutans* strain LAB761, which harbors several bacteriocin loci, isolated from a caries-active child in Canada. Microbiol Resour Announc 8:e01483-18. https://doi.org/10.1128/ MRA.01483-18.

Editor J. Cameron Thrash, Louisiana State University

Copyright © 2019 Barbour et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Céline M. Lévesque, celine.levesque@dentistry.utoronto.ca.

Received 29 October 2018 Accepted 3 December 2018 Published 10 January 2019 assembled genome had 246× genome coverage. The genome with a total size of 2,076,490 bp is composed of a single contig with a G+C content of 37.8%. Circularization of the contig at the overlapping ends was demonstrated using the Gepard software version 1.4 (8). Gene prediction and annotation were performed using RAST (9) and BLASTp (10). The genome annotation consisted of 1,841 protein-coding genes (CDSs), 64 tRNAs, and 5 rRNAs. The genomic information was analyzed to predict putative bacteriocin gene clusters and biosynthesis genes using the BAGEL4 (11) and antiSMASH 3.0 (12) Web servers with default search options. These pipelines predicted several genes for bacteriocin production and genes encoding uncharacterized double-glycine motif-containing peptides, in particular, genes related to mutacin Smb, mutacin IV, mutacin V, and mutacin VI. We also identified a locus of nine genes related to the production of the lantibiotic bacteriocin B-Ny266. Sequencing of multiple complete bacterial genomes is critical for evolutionary genomics. A comparative genomics approach will most likely be useful for a better characterization of the bacteriocin repertoire in *S. mutans*.

Data availability. The complete genome sequence has been deposited in GenBank under the accession number CP033199. Raw sequencing reads were deposited in the NCBI Sequence Read Archive (SRA) under accession number SRR8245044 and Bio-Project number PRJNA497888.

ACKNOWLEDGMENTS

This research was supported by the Canadian Institutes of Health Research (CIHR) grant CMA-151711. R.Y.H. was supported by a CIHR Undergraduate Award. C.M.L. is a recipient of a Canada Research Chair.

REFERENCES

- Loesche WJ. 1986. Role of *Streptococcus mutans* in human dental decay. Microbiol Rev 50:353–380.
- Momeni SS, Whiddon J, Cheon K, Ghazal T, Moser SA, Childers NK. 2016. Genetic diversity and evidence for transmission of *Streptococcus mutans* by DiversiLab rep-PCR. J Microbiol Methods 128:108–117. https://doi .org/10.1016/j.mimet.2016.07.010.
- Nicolas GG, Lavoie MC, LaPointe G. 2007. Molecular genetics, genomics and biochemistry of mutacins. G3 1:193–208.
- Merritt J, Qi F. 2012. The mutacins of *Streptococcus mutans*: regulation and ecology. Mol Oral Microbiol 27:57–69. https://doi.org/10.1111/j .2041-1014.2011.00634.x.
- Chen Z, Saxena D, Caufield PW, Ge Y, Wang M, Li Y. 2007. Development of species-specific primers for detection of *Streptococcus mutans* in mixed bacterial samples. FEMS Microbiol Lett 272:154–162. https://doi .org/10.1111/j.1574-6968.2007.00756.x.
- Edwards U, Rogall T, Blöcker H, Emde M, Böttger EC. 1989. Isolation and direct complete nucleotide determination of entire genes. Characterization of a gene coding for 16S ribosomal RNA. Nucleic Acids Res 17: 7843–7853.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT

sequencing data. Nat Methods 10:563-569. https://doi.org/10.1038/ nmeth.2474.

- Krumsiek J, Arnold R, Rattei T. 2007. Gepard: a rapid and sensitive tool for creating dotplots on genome scale. Bioinformatics 23:1026–1028. https://doi.org/10.1093/bioinformatics/btm039.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/gkt1226.
- Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res 25:3389–3402. https://doi .org/10.1093/nar/25.17.3389.
- van Heel AJ, de Song A, Song C, Viel JH, Kok J, Kuipers OP. 2018. BAGEL4: a user-friendly web server to thoroughly mine RIPPs and bacteriocins. Nucleic Acids Res 46:W278–W281. https://doi.org/10.1093/nar/gky383.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0–a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–D243. https://doi.org/10.1093/nar/gkv437.