

Why Are Children With Bronchiolitis At Risk Of Urinary Tract Infections?

This article was published in the following Dove Press journal:
Risk Management and Healthcare Policy

Mohamed A Hendaus^{1,2}

¹Department of Pediatrics, Section of Academic General Pediatrics, Sidra Medicine, Doha, Qatar; ²Department of Clinical Pediatrics, Weill-Cornell Medicine, Doha, Qatar

Abstract: Viral respiratory infections are frequently eliminated from human bodies without any sequelae. Secondary serious bacterial infection (SBI) in children with acute bronchiolitis has been an apprehension expressed by health care providers. Several published studies have shown an association between acute bronchiolitis and secondary bacterial infection, including urinary tract infections (UTI). However, the proposed mechanism by which a virus can induce UTIs is not yet known. The aim of this commentary is to update the current evidence of risk of UTI in children with bronchiolitis. We present several clinical studies related to the topic as well as a brief review of the potential pathophysiology of secondary infections that could present with viral respiratory illness.

Keywords: bronchiolitis, infection, urine

Review Of The Literature

Viral respiratory infections are frequently eliminated from human bodies without any sequelae. Nevertheless, in some occasions viruses can evade the immune reaction of the airways, leading to austere respiratory diseases.¹ Potent mechanical and immunosuppressive methods protect the lungs against external infections, but a solitary respiratory tract infection can change immunity and pathology.² Secondary serious bacterial infection (SBI) in children with acute bronchiolitis has been an apprehension expressed by health care providers.³ Several published studies have shown an association between acute bronchiolitis and secondary bacterial infection, including urinary tract infections (UTI).⁴⁻¹³ However, the proposed mechanism by which a virus can induce UTIs is not yet known.

In a review of the literature, the percentage of patients with fever with positive urine cultures ranged from 4.2% to 20.0% in infants <3 months of age and 0% to 7.4% in older children (3 to 36 months of age).¹⁴ Ralston et al³ conducted a systematic review delineating the risk of occult SBI in young febrile infants presenting with either “clinical bronchiolitis” or “proven RSV infection”. The review included 11 studies.^{4-7,9-13,15,16} The rate of urinary tract infections in the 11 studies analyzed was 3.3% (95% confidence interval, 1.9–5.7%). The authors concluded the rate of urine cultures positive for bacteria was noteworthy, though asymptomatic bacteriuria may have muddled the results. Recently, McDaniel et al¹⁷ conducted a systematic review and meta-analysis exploring the prevalence of UTI in infants and young children with bronchiolitis when positive urinalysis (UA) results being incorporated into the UTI definition. The investigators included 18 studies,^{4-7,9,11-13,15,16,18-25} seven of which had UA information.^{4,11,16,18,20-22} The definition of positive UA varied among the studies. Some considered positive UA as

Correspondence: Mohamed A Hendaus
Department of Pediatrics, Sidra Medicine,
Doha 26999, Qatar
Tel +974-4003-6559
Email mhendaus@yahoo.com

having more than 5 white blood cells per high-powered field, while others considered the UA positive by having bacteriuria, positive leukocyte esterase, or positive nitrite. The prevalence of UTI in bronchiolitis in the 18 studies was 3.1% (95% CI, 1.8–4.6%). The authors further analyzed the data of the 7 studies where the presence of pyuria or nitrites was a diagnostic criterion to define UTI and the prevalence of UTI was 0.8% (95% CI, 0.3–1.4%).

However, the above studies did not sub-categorize the prevalence of UTI in bronchiolitis per specific respiratory virus as a trigger. Hendaus et al⁸ studied the prevalence of urinary tract infection in infants and children with bronchiolitis. The study included 835 pediatric patients with acute bronchiolitis. The mean (\pm SD) age at diagnosis was 3.47 \pm 2.99 months. There were 325 (39%) girls and 510 (61%) boys. Participants were divided into three groups: group 1 comprised of children hospitalized with bronchiolitis and a positive diagnosis for respiratory syncytial virus (RSV) bronchiolitis; group 2 comprised of children hospitalized with clinical bronchiolitis with no virus detected; and group 3 comprised of children hospitalized with clinical with bronchiolitis and a positive diagnosis respiratory virus other than RSV. After applying inclusion and exclusion criteria, RSV was notorious in 352 (45.7%) patients; respiratory viruses other than RSV were identified in 275 (35.7%) patients and 142 (18.4%) were studied but had no viruses detected. Non-RSV viruses comprised of rhinovirus (n=85 [31%]), parainfluenza virus type 4 (n=40 [14%]), adenovirus (n=40 [14%]), human metapneumovirus (HMPV) (n=27 [10%]), bocavirus (n=27 [10%]), coronavirus (n=20 [7%]), parainfluenza virus (hPIV) type 1 (n=9 [3.4%]), hPIV type 2 (n=9 [3.4%]), hPIV type 3 (n=9 [3.4%]), and H1N1pdm09 (n=9 [3.4%]). The definition of UTI was adopted from the American Academy of Pediatrics as

clinicians should require both urinalysis results that suggest infection (pyuria and/or bacteriuria) and the presence of \geq 50,000 colony-forming units (CFUs) per milliliter of a uropathogen cultured from a urine specimen obtained through catheterization or suprapubic aspirate.²⁶

The overall prevalence of UTI was 10%, and was most common in group 3 (13.4%) trailed by group 2 (9.7%), and was least common in group 1 (6%) (P=0.030). The most reasonable explanation of why the rate of UTI was higher in the study conducted by Hendaus et al⁸ was because it was the first published study that sub-

categorized the prevalence of UTI in bronchiolitis per specific respiratory virus as a trigger.

So What Could Be The Potential Pathophysiology?

The epithelium is usually protected by a layer of mucus that functions as a border.^{27–29} Viruses can inflict impairment on host epithelial cells, and mammalian cells are susceptible to bacterial attachment during a viral sickness.³⁰ Moreover, viruses can incapacitate the mucociliary clearance arrangement, causing increased attachment of bacteria to mucins and colonization.³¹ Viruses like the influenza and RSV might injure ciliated cells, causing ciliostasis, and hence worsening of mucociliary clearance.^{30,32,33} Furthermore, virus-induced cell demise weakens the mechanical elimination of the close pathogens and exhibits new receptors for bacterial adherence.³⁴ The respiratory virus-infected epithelia enable the attraction of inflammatory cells, including natural killer cells, neutrophils, macrophages, and eosinophils from the bloodstream into the infected area.³⁵

The epithelium identifies microorganisms through pattern recognition receptors, such as nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), retinoic acid-inducible gene (RIG)-like helicases,^{36,37} and Toll-like receptors (TLRs).²⁹ NLRs and RIG-like helicases activate innate immune reactions through cytosolic detection of viral and bacterial components,^{37,38} while TLRs are single, noncatalytic, membrane-spanning receptor proteins utilized by the innate immune system.³⁹ Post-viral continued desensitization of lung sentinel cells to TLR signals might contribute to secondary bacterial infection. For instance, TLR4 and TLR5 pathways are modified after influenza virus infection, leading to decreased neutrophil attraction, hence resulting in increased attachment of bacteria.³⁸

Several epithelial cells can also express the classical antiviral interferons (INFs), especially IFN- α and IFN- β .^{40,41} The link between host cells and microorganisms during sickness prompts immune reaction that comprise the generation of pro-inflammatory molecules. In spite of their important role as a bactericidal, pro-inflammatory cytokines such as TNF- α produced in response to infection could be injurious to the host cells.⁴² Viruses can also have an impact in modulating many molecules such as intercellular adhesion molecule 1 (ICAM-1), carcinoembryonic antigen-related cellular adhesion 1 (CEACAM-1), and platelet-activating factor receptor (PAF-r),⁴³ resulting in a risk of bacterial adherence.⁴⁴

Throughout a viral episode, TLR and RIG-I-like receptor activation prompts fabrication of type I IFNs, which can then boost the inflammatory response to TLR ligands including lipopolysaccharide (LPS).^{45,46} Interface between type I IFNs and Nod1/Nod2 signalling results in bacterial recognition, and causes damaging effects in the virally infected host.⁴⁷

Conclusions And Recommendations

1. Published studies do not robustly support the idea that viral bronchiolitis can lead to bacterial UTI.
2. Despite the fact that a viral infection can lead to secondary bacterial infection, the mechanism of acquiring UTI after viral bronchiolitis is not very well known.
3. Large randomized studies are required to tackle the possible causation/association (bronchiolitis vs UTI).
4. Performing urine microscopy and urine culture on a febrile young child with bronchiolitis should be individualized per patient's condition.

Acknowledgment

Publication of this manuscript has been funded by Qatar National Library.

Disclosure

The author reports no conflicts of interest in this work.

References

1. Varelle M, Kieninger E, Edwards MR, Regamey N. The airway epithelium: soldier in the fight against respiratory viruses. *Clin Microbiol Rev*. 2011;24(1):210–229. doi:10.1128/CMR.00014-10
2. Walzl G, Tafuro S, Moss P, Openshaw PJ, Hussell T. Influenza virus lung infection protects from respiratory syncytial virus-induced immunopathology. *J Exp Med*. 2000;192(9):1317–1326. doi:10.1084/jem.192.9.1317
3. Ralston S, Hill V, Waters A. Occult serious bacterial infection in infants younger than 60 to 90 days with bronchiolitis: a systematic review. *Arch Pediatr Adolesc Med*. 2011;165:951–956. doi:10.1001/archpediatrics.2011.155
4. Melendez E, Harper MB. Utility of sepsis evaluation in infants 90 days of age or younger with fever and clinical bronchiolitis. *Pediatr Infect Dis J*. 2003;22:1053–1056. doi:10.1097/01.inf.0000101296.68993.4d
5. Levine DA, Platt SL, Dayan PS, et al. Multicenter RSV-SBI Study Group of the Pediatric Emergency Medicine Collaborative Research Committee of the American Academy of Pediatrics Risk of serious bacterial infection in young febrile infants with respiratory syncytial virus infections. *Pediatrics*. 2004;113:1728–1734. doi:10.1542/peds.113.6.1728
6. Purcell K, Fergie J. Concurrent serious bacterial infections in 2396 infants and children hospitalized with respiratory syncytial virus lower respiratory tract infections. *Arch Pediatr Adolesc Med*. 2002;156:322–324. doi:10.1001/archpedi.156.4.322
7. Titus MO, Wright SW. Prevalence of serious bacterial infections in febrile infants with respiratory syncytial virus infection. *Pediatrics*. 2003;112:282–284. doi:10.1542/peds.112.2.282
8. Hendaus MA, Alhammadi AH, Khalifa MS, Muneer E, Chandra P. Risk of urinary tract infection in infants and children with acute bronchiolitis. *Paediatr Child Health*. 2015;20(5):e25–e29. doi:10.1093/pch/20.5.e25
9. Oray-Schrom P, Phoenix C, St Martin D, Amoateng-Adjepong Y. Sepsis workup in febrile infants 0–90 days of age with respiratory syncytial virus infection. *Pediatr Emerg Care*. 2003;19:314–319. doi:10.1097/01.pec.0000092576.40174.28
10. Byington CL, Enriquez FR, Hoff C, et al. Serious bacterial infections in febrile infants 1 to 90 days old with and without viral infections. *Pediatrics*. 2004;113:1662–1666. doi:10.1542/peds.113.6.1662
11. Antonow JA, Hansen K, McKinstry CA, Byington CL. Sepsis evaluations in hospitalized infants with bronchiolitis. *Pediatr Infect Dis J*. 1998;17:231–236. doi:10.1097/00006454-199803000-00011
12. Bilavsky E, Shouval DS, Yarden-Bilavsky H, Fisch N, Ashkenazi S, Amir J. A prospective study of the risk for serious bacterial infections in hospitalized febrile infants with or without bronchiolitis. *Pediatr Infect Dis J*. 2008;27:269–270. doi:10.1097/INF.0b013e31815e85b1
13. Kuppermann N, Bank DE, Walton EA, Senac MO Jr, McCaslin I. Risks for bacteremia and urinary tract infections in young febrile children with bronchiolitis. *Arch Pediatr Adolesc Med*. 1997;151:1207–1214. doi:10.1001/archpedi.1997.02170490033006
14. Baraff LJ, Bass JW, Fleisher GR, et al. Practice guideline for the management of infants and children 0 to 36 months of age with fever without source. *Pediatrics*. 1993;92:1–12.
15. Liebelt EL, Qi K, Harvey K. Diagnostic testing for serious bacterial infections in infants aged 90 days or younger with bronchiolitis. *Arch Pediatr Adolesc Med*. 1999;153:525–530. doi:10.1001/archpedi.153.5.525
16. Luginbuhl LM, Newman TB, Pantell RH, Finch SA, Wasserman RC. Office-based treatment and outcomes for febrile infants with clinically diagnosed bronchiolitis. *Pediatrics*. 2008;122:947–954. doi:10.1542/peds.2007-3206
17. McDaniel CE, Ralston S, Lucas B, Schroeder AR. Association of diagnostic criteria with urinary tract infection prevalence in bronchiolitis: a systematic review and meta-analysis. *JAMA Pediatr*. 2019. doi:10.1001/jamapediatrics.2018.5091.
18. Elkhunovich MA, Wang VJ. Assessing the utility of urine testing in febrile infants aged 2 to 12 months with bronchiolitis. *Pediatr Emerg Care*. 2015;31(9):616–620. doi:10.1097/PEC.0000000000000359
19. Purcell K, Fergie J. Concurrent serious bacterial infections in 912 infants and children hospitalized for treatment of respiratory syncytial virus lower respiratory tract infection. *Pediatr Infect Dis J*. 2004;23(3):267–269. doi:10.1097/01.inf.0000116759.21252.29
20. Kaluarachchi D, Kaldas V, Roques E, Nunez R, Mendez M. Comparison of urinary tract infection rates among 2- to 12-month-old febrile infants with RSV infections using 1999 and 2011 AAP diagnostic criteria. *Clin Pediatr (Phila)*. 2014;53(8):742–746. doi:10.1177/0009922814529015
21. Randolph AG, Reder L, Englund JA. Risk of bacterial infection in previously healthy respiratory syncytial virus-infected young children admitted to the intensive care unit. *Pediatr Infect Dis J*. 2004;23(11):990–994. doi:10.1097/01.inf.0000143647.88873.66
22. Schlechter Salinas AK, Hains DS, Jones T, Harrell C, Meredith M. Testing for urinary tract infection in the influenza/respiratory syncytial virus-positive febrile infant aged 2 to 12 months. *Pediatr Emerg Care*. 2017. doi:10.1097/PEC.0000000000001073
23. Yarden-Bilavsky H, Ashkenazi-Hoffnung L, Livni G, Amir J, Bilavsky E. Month-by-month age analysis of the risk for serious bacterial infections in febrile infants with bronchiolitis. *Clin Pediatr (Phila)*. 2011;50(11):1052–1056. doi:10.1177/0009922811412949
24. Garcia CG, Bhoire R, Soriano-Fallas A, et al. Risk factors in children hospitalized with RSV bronchiolitis versus non-RSV bronchiolitis. *Pediatrics*. 2010;126(6):e1453–e1460. doi:10.1542/peds.2010-0507
25. Librizzi J, McCulloh R, Koehn K, Alverson B. Appropriateness of testing for serious bacterial infection in children hospitalized with bronchiolitis. *Hosp Pediatr*. 2014;4(1):33–38. doi:10.1542/hpeds.2013-0073

26. Roberts KB, American Academy of Pediatrics, Subcommittee on Urinary Tract Infection, Steering Committee on Quality Improvement and Management. Urinary tract infection: clinical practice guideline for diagnosis and management of the initial UTI in febrile infants and children 2 to 24 months. *Pediatrics*. 2011;128:595–610.
27. Voynow JA, Rubin BK. Mucins, mucus, and sputum. *Chest*. 2009;135(2):505–512. doi:10.1378/chest.08-0412
28. Thornton DJ, Rousseau K, McGuckin MA. Structure and function of the polymeric mucins in airways mucus. *Annu Rev Physiol*. 2008;70:459–486. doi:10.1146/annurev.physiol.70.113006.100702
29. Rose MC, Voynow JA. Respiratory tract mucin genes and mucin glycoproteins in health and disease. *Physiol Rev*. 2006;86(1):245–278. doi:10.1152/physrev.00010.2005
30. Pittet LA, Hall-Stoodley L, Rutkowski MR, Harmsen AG. Influenza virus infection decrease stracheal mucociliary velocity and clearance of *Streptococcus pneumoniae*. *Am J Respir Cell Mol Biol*. 2010;42(4):450–460. doi:10.1165/rcmb.2007-0417OC
31. Wilson R, Dowling RB, Jackson AD. The biology of bacterial-colonization and invasion of the respiratory mucosa. *Eur Respir J*. 1996;9(7):1523–1530. doi:10.1183/09031936.96.09071523
32. Tristram DA, Hicks W Jr, Hard R. Respiratory syncytial virus and human bronchial epithelium. *Arch Otolaryngol Head Neck Surg*. 1998;124:777–783. doi:10.1001/archotol.124.7.777
33. McCullers JA, Iverson AR, McKeon R, Murray PJ. The platelet activating factor receptor is not required for exacerbation of bacterial pneumonia following influenza. *Scand J Infect Dis*. 2008;40(1):11–17. doi:10.1080/00365540701477568
34. Bragonzi A, Copreni E, de Bentzmann S, Ulrich M, Conese M. Airway epithelial cell-pathogen interactions. *J Cyst Fibros*. 2004;3(suppl 2):197–201. doi:10.1016/j.jcf.2004.05.041
35. Katze MG, He Y, Gale M Jr. Viruses and interferon: a fight for supremacy. *Nat Rev Immunol*. 2002;2(9):675–687. doi:10.1038/nri888
36. Akira S. Pathogen recognition by innate immunity and its signaling. *Proc Jpn Acad Ser B Phys Biol Sci*. 2009;85(4):143–156. doi:10.2183/ptjab.85.143
37. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006;124:783–801. doi:10.1016/j.cell.2006.02.015
38. Didierlaurent A, Goulding J, Patel S, et al. Sustained desensitization to bacterial Toll-like receptor ligands after resolution of respiratory influenza infection. *J Exp Med*. 2008;205(2):323–329. doi:10.1084/jem.20070891
39. Kanneganti TD, Lamkanfi M, Nunez G. Intracellular NOD-like receptors in host defense and disease. *Immunity* 2007;27:549–559. doi:10.1016/j.immuni.2007.10.002
40. Inohara N, Ogura Y, Fontalba A, et al. Host recognition of bacterial muramyl dipeptide mediated through NOD2. Implications for Crohn's disease. *J Biol Chem*. 2003;278:5509–5512.
41. Basler CF, Garcia-Sastre A. Viruses and the type I interferon antiviral system: induction and evasion. *Int Rev Immunol*. 2002;21(4–5):305–337. doi:10.1080/08830180213277
42. Cook DN, Pisetsky DS, Schwartz DA. Toll-like receptors in the pathogenesis of human disease. *Nat Immunol*. 2004;5:975–979. doi:10.1038/ni1116
43. Message SD, Johnston SL. Host defense function of the airway epithelium in health and disease: clinical background. *J Leukoc Biol*. 2004;75(1):5–17. doi:10.1189/jlb.0703315
44. Ishizuka S, Yamaya M, Suzuki T, et al. Effects of rhinovirus infection on the adherence of *Streptococcus pneumoniae* to cultured human airway epithelial cells. *J Infect Dis*. 2003;188(12):1928–1939. doi:10.1086/jid.2003.188.issue-12
45. Nansen A, Randrup Thomsen A. Viral infection causes rapid sensitization to lipopolysaccharide: central role of IFN-alpha beta. *J Immunol*. 2001;166:982–988. doi:10.4049/jimmunol.166.2.982
46. Doughty L, Nguyen K, Durbin J, Biron C. A role for IFN-alpha beta in virus infection-induced sensitization to endotoxin. *J Immunol*. 2001;166:2658–2664. doi:10.4049/jimmunol.166.4.2658
47. Kim YG, Park JH, Reimer T, et al. Viral infection augments Nod1/2 signaling to potentiate lethality. *Cell Host Microbe*. 2011;9(6):496–507. doi:10.1016/j.chom.2011.05.006

Risk Management and Healthcare Policy

Dovepress

Publish your work in this journal

Risk Management and Healthcare Policy is an international, peer-reviewed, open access journal focusing on all aspects of public health, policy, and preventative measures to promote good health and improve morbidity and mortality in the population. The journal welcomes submitted papers covering original research, basic science, clinical & epidemiological studies, reviews and evaluations,

guidelines, expert opinion and commentary, case reports and extended reports. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/risk-management-and-healthcare-policy-journal>