

1 **Suboptimal response to COVID-19 mRNA vaccines in hematologic malignancies patients**

2 Mounzer Agha¹, Maggie Blake¹, Charles Chilleo³, Alan Wells², and Ghady Haidar⁴

3 ¹Hillman Cancer Center, UPMC, Pittsburgh, PA, USA

4 ²Department of Pathology, University of Pittsburgh School of Medicine, Pittsburgh, PA, USA

5 ³UPMC Clinical Laboratories, Pittsburgh, PA, USA

6 ⁴Division of Infectious Diseases, University of Pittsburgh School of Medicine, Pittsburgh, PA,
7 USA

8

9 **Abstract:**

10 Studies describing SARS-CoV-2 immune responses following mRNA vaccination in hematology
11 malignancy (HM) patients are virtually non-existent. We measured SARS-CoV-2 IgG production
12 in 67 HM patients who received 2 mRNA vaccine doses. We found that 46% of HM patients did
13 not produce antibodies and were therefore vaccine non-responders. Patients with B-cell CLL
14 were at a particularly high risk, as only 23% had detectable antibodies despite the fact that
15 nearly 70% of these patients were not undergoing cancer therapy. HM patients should be
16 counseled about the ongoing risk of COVID-19 despite vaccination. Routine measurement of
17 post-vaccine antibodies in HM patients should be considered. Novel strategies are needed to
18 prevent COVID-19 in these individuals.

19 Patients with hematologic malignancies are at high risk for coronavirus disease 2019
20 (COVID-19)-related complications, with mortality rates exceeding 30%¹⁻³. These patients have
21 also been shown to develop prolonged shedding of infectious severe acute respiratory
22 syndrome coronavirus-2 (SARS-CoV-2), often lasting several months, and have been implicated
23 in being sources of novel SARS-CoV-2 variants⁴⁻⁷. Such patients should be therefore be
24 prioritized for primary prevention of COVID-19 via vaccination⁸. However, the performance of
25 COVID-19 mRNA vaccines in hematological malignancy patients is unknown, as these
26 individuals were excluded from COVID-19 vaccine clinical trials^{9,10}.

27 To address these knowledge gaps, we measured SARS-CoV-2 antibody responses in
28 patients with hematological malignancies seen at UPMC Hillman Cancer Center who have
29 received two doses of either the mRNA-1273 (Moderna) or the BNT162b2 (Pfizer) vaccine.
30 Patients with prior COVID-19 were excluded. Antibody assays were performed at the UPMC
31 clinical laboratories using the semi-quantitative Beckman Coulter SARS-CoV-2 platform, which
32 detects IgG against the Spike protein receptor-binding domain (RBD). These results are
33 expressed as extinction coefficient (signal/cutoff) ratios and are interpreted as positive (≥ 1.00),
34 equivocal (> 0.80 to < 1.00), or non-reactive (≤ 0.80)¹¹. Reactive results are confirmed by the
35 Siemens SARS-CoV-2 Total Ig Assay, which detects both IgM and IgG antibodies against RBD
36 of the S1 subunit of the Spike protein¹². For analysis, reactive results were defined as positive,
37 and equivocal or non-reactive results were defined as negative. We calculated the proportion of
38 patients with a positive versus negative result (vaccine responders versus non-responders,
39 respectively) with 95% Coppler-Pearson exact confidence intervals and used χ^2 or Wilcoxon
40 Rank Sum testing for comparisons as appropriate. Analyses were performed using Stata
41 version 16.1 (StataCorp) and GraphPad Prism 8.3.1. Institutional Board Review Approval was
42 obtained.

43 Sixty-seven patients were included. Median age was 71 (interquartile range (IQR) 65 -
44 77), and 47.8% (32/67) percent were female. Underlying malignancies were B-cell chronic
45 lymphocytic leukemia (CLL, 19.4% (13/67)), lymphomas (31.3%, 21/67), multiple myeloma
46 (43.3%, 29/67), and other myeloid malignancies (5.97%, 4/68) (**Table 1**). Thirty patients (44.8%)
47 were undergoing therapy for their cancers, whereas 37 (55.2%) were under observation. Among
48 the 62 patients whose vaccine type was available, 50.8% (34/67) and 41.8% (28/67) had
49 received the BNT162b2 or mRNA-1273 vaccines, respectively. Median duration from the 2nd
50 vaccine dose to the antibody test was 23 days (IQR 16 - 31 days).

51 In total, 31/67 patients (46.3%, 95% CI 35.4%– 60.3%) had a negative antibody result
52 after vaccination and were therefore considered to be vaccine non-responders. Older patients

53 were more likely to be vaccine non-responders than younger patients (**Table 1**). Sex,
54 immunoglobulin G (IgG) levels, number of days between 2nd vaccine dose and antibody
55 measurement, and cancer therapy status did not differ among vaccine responders versus non-
56 responders. However, patients with CLL were significantly less likely to develop SARS-CoV-2
57 antibodies compared to patients with other hematological malignancies (23.1% (3/13) versus
58 61.1% (33/54), respectively, $p = 0.01$), even though 69.2% (9/13) of CLL patients were not
59 actively undergoing cancer therapy. There was no difference between age or IgG level between
60 CLL and non-CLL patients.

61 We further analyzed SARS-CoV-2 IgG extinction coefficient (signal/cutoff) ratios in order
62 to quantify antibody responses. These ratios were obtained from the Beckman assay, with
63 higher values generally indicating more robust antibody responses. Among vaccine responders,
64 there was no difference in the extinction coefficient ratios between the different hematological
65 malignancies (median ratio among CLL versus non-CLL patients = 7.88 (range 1.42 – 20.19)
66 versus 15.44 (range 1.05 – 38.6), respectively, $p = 0.39$) (**Figure 1A**). Among vaccine non-
67 responders however, patients with CLL had significantly lower extinction coefficient ratios
68 compared to those without CLL (median ratio 0.02 (range 0.02 – 0.06) versus 0.15 (range 0.02
69 – 0.91), respectively, $p < 0.001$) (**Figure 1B**). It should be noted that values below 0.10 are
70 suggestive of no antibody response, whereas values closer 1.00 may suggest evolving or
71 declining responses¹².

72 Our data show that nearly half of patients with hematological malignancies do not
73 generate antibodies after completing their COVID-19 vaccine series, which is in stark contrast
74 with the results of phase 1 mRNA vaccine immunogenicity trials, in which robust antibody
75 responses were seen in essentially 100% of participants^{13,14}. This lack of response was
76 particularly pronounced among patients with CLL, in whom the results of qualitative testing
77 demonstrated significantly lower antibody signals compared to patients without CLL, suggesting
78 that patients with CLL are unable to develop any antibody response after COVID-19
79 vaccination. These findings cannot be explained by age, cancer therapy, or IgG levels, and are
80 therefore likely a result of the humoral defects that are characteristic of CLL¹⁵.

81 Our findings underscore the importance of adherence to non-pharmaceutical
82 interventions to prevent COVID-19 in hematological malignancy patients, particularly in the
83 context of the limited arsenal of SARS-CoV-2 antiviral therapies, the high mortality rates of
84 cancer patients with COVID-19¹⁻³, and the emerging risk of prolonged SARS-CoV-2 replication
85 and variant generation in cancer patients⁴⁻⁷. Indeed, as the March 2021 CDC guidance has
86 been modified to allow for unmasked gatherings between vaccinated individuals and low-risk

87 unvaccinated individuals¹⁶, clinicians caring for patients with hematological malignancies and
88 other immunocompromising conditions should be aware of the possibility of COVID-19 vaccine
89 failure. Although immunological correlates of vaccine protection may be more complex than the
90 presence or absence of antibody responses¹⁷, these patients should be advised to wear masks
91 and observe social distancing regardless of vaccination status.

92 Limitations of this study include a small sample size, lack of serial measurements, and
93 lack of a control group. In addition, we did not determine whether antibodies from vaccine
94 responders are able to neutralize SARS-CoV-2. Nonetheless, these early findings suggest that
95 COVID-19 vaccine responses in hematological malignancy patient are suboptimal, and that
96 patients with CLL are at a very high risk for vaccine failure. Future studies should focus on post-
97 vaccine antibody durability, B-cell and T-cell responses after vaccination, and novel strategies of
98 COVID-19 prevention in hematological malignancy patients, such as administration of additional
99 vaccine doses or the use of monoclonal antibodies for primary prophylaxis¹⁸. Routine
100 measurement of SARS-CoV-2 antibody responses in immunocompromised patients should be
101 considered.

102

103

104 **Table 1. Comparison of hematological malignancy patients with positive versus negative**
 105 **SARS-CoV-2 antibody results after administration of two doses of an mRNA COVID-19**
 106 **vaccine.**

	SARS-CoV-2 antibody result		P-value
	Positive (N=36)	Negative (N=31)*	
Age (media, IQR)	70 (62.5 – 73.5)	74 (68 – 79)	0.009
Sex (N, %)			
Male	19 (54.3%)	16 (45.7%)	0.92
Female	17 (53.1%)	15 (46.9%)	
Vaccine type (N, %)			
BNT162b2	15 (44.1%)	19 (55.9%)	0.31
mRNA-1273	16 (57.1%)	12 (42.9%)	
Days between 2 nd dose of vaccine and antibody level (median, IQR)	23 (14-33)	25 (16-31)	0.93
IgG level (mg/dL) (median, IQR) [†]	723.5 (510-1045)	549 (472-939)	0.22
Therapy (N, %)			
Active treatment	15 (50%)	15 (50%)	0.58
Observation	21 (56.8%)	16 (43.2%)	
Cancer type (N, %)			
CLL	3 (23.1%)	10 (76.9%)	0.01 [§]
Non-CLL	33 (61.1%)	21 (38.9%)	
Lymphomas	11 (52.4%)	10 (47.6%)	
Multiple myeloma	19 (65.5%)	10 (34.5%)	
Other [‡]	3 (75.0%)	1 (25.0%)	

107

108 CLL, chronic lymphocytic leukemia; IgG, immunoglobulin G; IQR, interquartile range

109 *Includes 31 patients with non-reactive tests and 1 patient with an equivocal test.

110 †Represents lowest IgG level obtained within 90 days of the SARS-CoV-2 antibody. IgG levels
 111 available for 55 patients. Only 2 patients had received intravenous immunoglobulin during this
 112 time period.

113 ‡Includes 2 patients with acute myelogenous leukemia (1 of whom had undergone a
 114 hematopoietic cell transplant 10 years prior) and with 2 chronic myeloid leukemia.

115 §Comparison between CLL versus non-CLL patients

116

117

137

138 **Contribution:** M.A. and G.H. designed the research and wrote the first draft of the paper. M.A.
139 and M.B. collected the data. G.H. analyzed the data. A.W. and C.C. performed the experiments.
140 All authors have reviewed the paper.

141 **Conflicts of interest:** None

142 **Disclosures:** “Research reported in this publication was supported by the National Institute Of
143 Allergy And Infectious Diseases of the National Institutes of Health under Award Number
144 K23AI154546 awarded to G.H. The content is solely the responsibility of the authors and does
145 not necessarily represent the official views of the National Institutes of Health.”

146 **Correspondence:** Ghady Haidar, MD. 3601 Fifth Ave, Falk Medical BLDG, Suite 5B.
147 Pittsburgh, PA, 15213, USA. +1-412-648-6601. haidarg@upmc.edu

148

149

150 **References:**

151

152 1. Sharma A, Bhatt NS, St Martin A, et al. Clinical characteristics and outcomes of COVID-
153 19 in haematopoietic stem-cell transplantation recipients: an observational cohort study. *Lancet*
154 *Haematol.* 2021.

155 2. Vijenthira A, Gong IY, Fox TA, et al. Outcomes of patients with hematologic
156 malignancies and COVID-19: a systematic review and meta-analysis of 3377 patients. *Blood.*
157 2020;136(25):2881-2892.

158 3. Fung M, Babik JM. COVID-19 in Immunocompromised Hosts: What We Know So Far.
159 *Clin Infect Dis.* 2021;72(2):340-350.

160 4. Avanzato VA, Matson MJ, Seifert SN, et al. Case Study: Prolonged Infectious SARS-
161 CoV-2 Shedding from an Asymptomatic Immunocompromised Individual with Cancer. *Cell.*
162 2020;183(7):1901-1912 e1909.

163 5. Aydillo T, Gonzalez-Reiche AS, Aslam S, et al. Shedding of Viable SARS-CoV-2 after
164 Immunosuppressive Therapy for Cancer. *N Engl J Med.* 2020;383(26):2586-2588.

165 6. Hensley MK, Bain WG, Jacobs J, et al. Intractable COVID-19 and Prolonged SARS-
166 CoV-2 Replication in a CAR-T-cell Therapy Recipient: A Case Study. *Clin Infect Dis.* 2021.

167 7. Kemp SA, Collier DA, Datir RP, et al. SARS-CoV-2 evolution during treatment of chronic
168 infection. *Nature.* 2021.

169 8. Ribas A, Sengupta R, Locke T, et al. Priority COVID-19 Vaccination for Patients with
170 Cancer while Vaccine Supply Is Limited. *Cancer Discov.* 2021;11(2):233-236.

171 9. Baden LR, El Sahly HM, Essink B, et al. Efficacy and Safety of the mRNA-1273 SARS-
172 CoV-2 Vaccine. *N Engl J Med.* 2021;384(5):403-416.

173 10. Polack FP, Thomas SJ, Kitchin N, et al. Safety and Efficacy of the BNT162b2 mRNA
174 Covid-19 Vaccine. *N Engl J Med.* 2020;383(27):2603-2615.

175 11. Beckman Coulter Access Immunoassay Systems Instructions for Use. FDA Emergency
176 Use Authorization.. <https://www.fda.gov/media/139627/download>. Accessed April 2, 2021.

177

178 12. Zilla M, Wheeler BJ, Keetch C, et al. Variable Performance in 6 Commercial SARS-CoV-
179 2 Antibody Assays May Affect Convalescent Plasma and Seroprevalence Screening. *Am J Clin*
180 *Pathol.* 2021;155(3):343-353.

- 181 13. Jackson LA, Anderson EJ, Roupael NG, et al. An mRNA Vaccine against SARS-CoV-2
182 - Preliminary Report. *N Engl J Med*. 2020;383(20):1920-1931.
- 183 14. Mulligan MJ, Lyke KE, Kitchin N, et al. Phase I/II study of COVID-19 RNA vaccine
184 BNT162b1 in adults. *Nature*. 2020;586(7830):589-593.
- 185 15. Forconi F, Moss P. Perturbation of the normal immune system in patients with CLL.
186 *Blood*. 2015;126(5):573-581.
- 187 16. When You've Been Fully Vaccinated. Centers for Disease Control and Prevention.
188 <https://www.cdc.gov/coronavirus/2019-ncov/vaccines/fully-vaccinated.html>. Accessed March 26,
189 2021.
- 190 17. Jin P, Li J, Pan H, Wu Y, Zhu F. Immunological surrogate endpoints of COVID-2019
191 vaccines: the evidence we have versus the evidence we need. *Signal Transduct Target Ther*.
192 2021;6(1):48.
- 193 18. Lilly's neutralizing antibody bamlanivimab (LY-CoV555) prevented COVID-19 at nursing
194 homes in the BLAZE-2 trial, reducing risk by up to 80 percent for residents.
195 [https://investor.lilly.com/news-releases/news-release-details/lillys-neutralizing-antibody-](https://investor.lilly.com/news-releases/news-release-details/lillys-neutralizing-antibody-bamlanivimab-ly-cov555-prevented)
196 [bamlanivimab-ly-cov555-prevented](https://investor.lilly.com/news-releases/news-release-details/lillys-neutralizing-antibody-bamlanivimab-ly-cov555-prevented). Accessed February 18, 2021.

197