

Tailor-made PEG-DA-CuS nanoparticles enriched in tumor with the aid of retro Diels–Alder reaction triggered by their intrinsic photothermal property

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Introduction: In recent years, near-infrared laser-induced photothermal therapy is being considered as a promising approach to kill tumors owing to its noninvasive nature and excellent antitumor efficiency. However, the lack of ideal photothermal agents hinders further development of this technology.

Materials and methods: Aiming at solving this long-standing obstacle, we report here about the polyethylene glycol (PEG)-DA modified copper sulfide (CuS) nanoparticles (NPs) (PEG-DA-CuS NPs), a kind of semiconductor photothermal agents that show excellent photothermal stability and high heat conversion efficiency.

Results and discussion: Owing to the surrounding PEG, the water solubility of CuS NPs was significantly improved when circulating in blood in the body. When the NPs reached the tumors and were irradiated by a 1,064 nm laser (1 W/cm², 10 minutes), the local temperature increased above 90°C, triggering the retro Diels–Alder reaction. After the release of PEG chain, CuS NPs soon formed aggregates and enriched the tumor via the enhanced permeability and retention effect, promoting the efficacy of photothermal therapy.

Conclusion: Therefore, we believe PEG-DA-CuS NPs are able to serve as a kind of cytotoxic and efficient photothermal agent to kill cancer.

Keywords: CuS, PEG, retro Diels–Alder reaction, excellent photothermal stability, high heat conversion efficiency, aggregation in tumor

Introduction

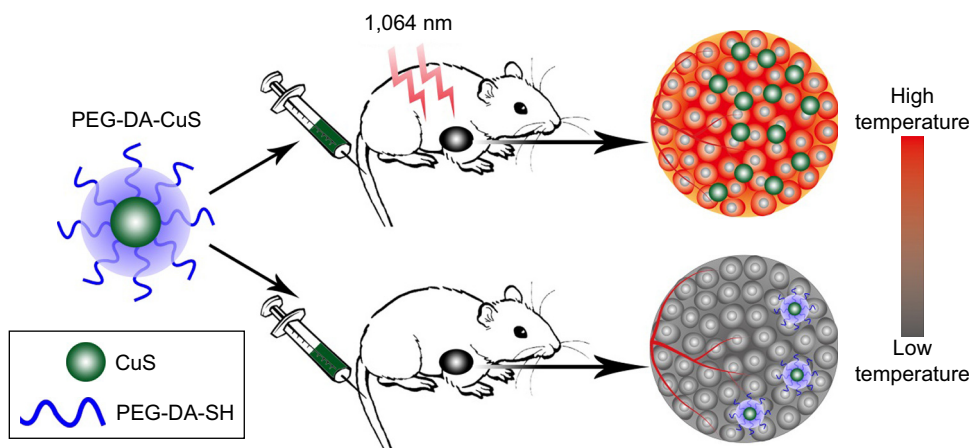
With the accumulating fundamental study of tumor, we realized that tumor is not only a mass formed by malignant cells, but also a highly heterogeneous multi-cell aggregation. Tumor microenvironment plays a decisive role in the occurrence, development, and treatment of tumor tissues.^{1,2} The terrible extracellular environment including metabolic stress, lack of oxygen, nutrients and growth factors, lack of blood supply insufficiency, and antitumor treatment ensures tumor cells the ability to resist tumor apoptosis, thus greatly improving the ability to survive adversity.^{3,4} Traditional cancer treatments include surgery, radiation, and chemotherapy. However, they all have obvious flaws. The surgery cannot remove the small lesions that the human eye cannot see. Chemotherapy often causes tumor cell resistance and tumor recurrence after treatment.⁵ Radiotherapy faces the risk of carcinogenesis in normal tissues.⁶ Therefore, it is of great therapeutic and fundamental significance to develop new strategies for tumor treatment.

Near-infrared (NIR) laser-induced photothermal therapy (PTT) has attracted significant attention due to the noninvasive nature and safe biocompatibility of NIR laser

irradiation in normal tissues.⁷ In the development of PTT, obtaining ideal photothermal agents is the key step, since they can convert absorbed optical energy into heat, which enables the ablation of tumors while preventing the surrounding tissues from being affected.^{8–18} So far, a variety of functional nanoparticles (NPs) have been examined, including polymer NPs, noble metal nanomaterials, carbon-based nanomaterials, and semiconductor nanomaterials, due to their strong absorbency under NIR.^{19–26} Among them, semiconductor copper sulfide (CuS) NPs possess excellent physical properties and high heat conversion efficiency, which triggers our interest in exploring their clinical applications in photothermal ablation of tumors. Compared with noble metal nanomaterials such as golden nanorods, CuS NPs exhibit higher heat tolerance. Club-shaped golden nanorods are easily transformed to spherical shape at a high temperature.²⁷ On the contrary, uniformly spherical CuS NPs have a stable conformation, which does not change after the hyperthermia process, ensuring their high heat conversion efficiency after repeated laser irradiation. In addition, their absorption peaks are positioned at 1,060 nm, longer than that of golden nanorods, which means they can efficiently absorb laser of longer wavelength and lower energy, causing smaller side effects to normal tissues. Unfortunately, considering their fast metabolic rate, CuS NPs can hardly accumulate into tumor cells, which is bound to limit the efficacy of PTT. In order to overcome this obstacle, various approaches have been explored. Meng's group used thermosensitive MEO₂MA nanogel as a nanocarrier to deliver CuS NPs.²⁸ Zhang et al's article introduced an activatable Cy5.5-hyaluronic acid nanoparticles/CuS (HANPC) nanocomposite to achieve

optical/photoacoustic image-guided PTT.²⁹ However, these methodologies capable of loading and delivering CuS NPs all relied on a complicated vehicle and an immediate outside stimulus to obtain a good delivery outcome. As such, these systems failed in taking advantage of the excellent photothermal qualities of CuS NPs.

Here, we successfully synthesized polyethylene glycol (PEG)-linked Diels–Alder cycloadducts-modified CuS NPs (PEG-DA-CuS NPs). The Diels–Alder reaction is a reversible reaction, and the forward reaction proceeds at 60°C while the backward reaction proceeds at 90°C.³⁰ In the forward reaction process, the water solubility of CuS NPs was improved thanks to the presence of PEG on their surfaces. When the PEG-DA-modified CuS NPs were injected into the body, they were stable and had prolonged circulation life in the blood. After they reached the tumor area, we used 1,064 nm laser to irradiate the lesion to obtain a local high temperature of >90°C. Then, the backward reaction (retro Diels–Alder reaction) happened, releasing PEG chains from the CuS NPs. After laser irradiation, the aggregation and enrichment of CuS NPs in the tumor via enhanced permeability and retention effect were rigorously demonstrated by the *in vivo* biodistribution of CuS NPs. *In vitro* and *in vivo* tests showed that PTT rendered by the PEG-DA-CuS NPs under 1,064 nm laser irradiation maintained a wonderful effect on cancer treatment and effectively erased the tumor. The whole process is illustrated in Scheme 1. We believe these tailor-made PEG-DA-CuS NPs will open a new avenue for tumor treatment and throw light on their clinical applications in cancer.



Scheme 1 When the PEG-DA-modified CuS nanoparticles were injected in the body, they were stable and had prolonged circulation life in the blood. After they reached the tumor area, 1,064 nm laser was employed to irradiate the lesion to obtain a local high temperature of >90°C; then the backward reaction (retro Diels–Alder reaction) happened, releasing the PEG chains from the CuS nanoparticles. After laser irradiation, the aggregation and enrichment of CuS nanoparticles displayed a superior photothermal therapy.

Abbreviation: PEG, polyethylene glycol.

Materials and methods

CuCl₂·2H₂O, sodium hydroxide (NaOH), and thioacetamide were all purchased from Aladdin (Shanghai, China). m-PEG-maleimide (molecular weight 20,000 Da) was bought from J&K Chemicals (Beijing, China). Calcein-acetoxymethyl (calcein-AM), dimethylformamide, tris (2-carboxyethyl) phosphine hydrochloride, ethidium homodimer-1 (EthD-1), furfuryl disulfide, and PBS (pH ~7.4) were all bought from Jinpan biotech (Nantong, China). Human umbilical vein endothelial cells (HUVECs) and human cervical carcinoma (HeLa) cell line were both ordered from Servicebio Company (Wuhan, China). Nude mice (female, 4–6 weeks) were bought from Comparative Medicine Center of Yangzhou University. Eight mice per cage were housed under controlled environmental conditions (22°C±3°C, 45%–70% relative humidity; 12-hour light/dark cycle). All animal experiments were approved by the Care Committee of Nanjing University (including guidelines for animal care & use, and guidelines for euthanasia of mice, Protocol #: 20170322-016).

Preparation of PEG-DA-CuS NPs

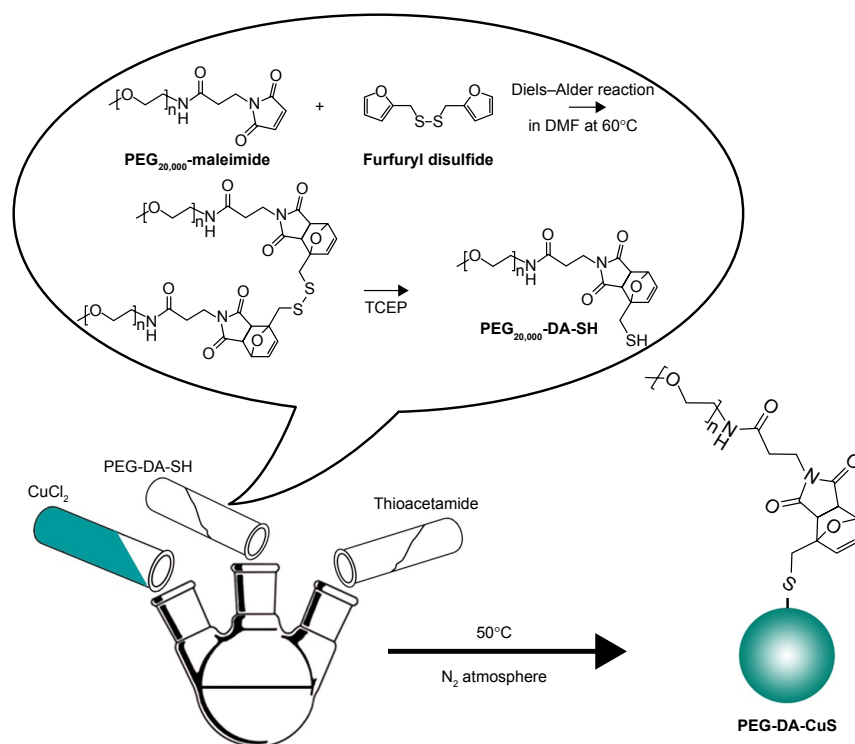
PEG-DA-CuS NPs were synthesized as illustrated in Scheme 2. First, PEG-DA-SH was prepared according to previously reported procedures.³⁰ Then, an aqueous solution

(40 mL) of the as-obtained PEG-DA-SH (0.1 mmol) was mixed with an aqueous solution (40 mL) of CuCl₂·2H₂O (0.1 mmol) and made to react for 10 minutes. Afterward, an aqueous solution (10 mL) of NaOH (0.1 mmol) was added dropwise to adjust the solution pH to 9. Then, the whole solution was transferred to a three-necked flask and an aqueous solution (10 mL) of thioacetamide (0.1 mmol) was added. The solution was under constant stirring at 60°C for 1.5 hours, and the process was protected by N₂ atmosphere.

Characterization

The morphologies of the obtained NPs were observed by transmission electron microscopy (TEM; JEOL, Tokyo, Japan), and the ultraviolet (UV)–Vis absorption spectra were recorded on a spectrophotometer (UV3100; Shimadzu Corporation, Kyoto, Japan). The size distribution was confirmed by dynamic light scattering method (BI-9000AT; Brookhaven, MS, USA). To study the photothermal effects, a 1,064 nm diode laser (LE-LS-1064-3000TFC-D; LaserGlow Technologies, Toronto, Canada) was used. The photothermal conversion efficiency (η) was derived from the following formula:^{10,27}

$$\eta (\%) = \frac{cmt}{wt} \times 100\% = \frac{4.2 \times 10^3 \times 1.43 \times 10^{-3} t}{1 \times 600} \times 100\%$$



Scheme 2 Synthesis of PEG-DA-CuS NPs.

Abbreviations: DMF, dimethylformamide; NP, nanoparticle; PEG, polyethylene glycol; TCEP, tris (2-carboxyethyl) phosphine hydrochloride.

where w represents the laser power, 1 W; m represents the mass of PEG-DA-CuS NP solution, 1.43×10^{-3} kg; t represents the laser irradiation time, 600 seconds; c represents the specific heat capacity of PEG-DA-CuS NP solution, 4.2×10^3 J/(kg·K); and Δt represents the temperature increase of PEG-DA-CuS NP solution. Release of PEG chains from PEG-DA-CuS NPs was quantified by the barium iodine method.³⁰

In vitro cytotoxicity assay

The Annexin-V-fluorescein isothiocyanate/propidium iodide staining method was used to evaluate the cytotoxicity of PEG-DA-CuS NPs to HUVECs. At first, HUVECs were seeded into 96-well plates (1×10^4 cells, 100 mL medium, per well) and incubated in 5% CO₂ atmosphere at 37°C for 1 day. Then, the cells were washed twice with PBS, followed by incubating them with PEG-DA-CuS NPs of various concentrations (0.1, 0.2, 0.4, and 1 mg/mL). Afterward, the cells were cultured with fresh culture medium at 37°C for 4 hours. Finally, the cells were collected and resuspended in a binding buffer (500 μ L). Annexin V-fluorescein isothiocyanate and propidium iodide were added following the manufacturer's guidance. After incubation in darkness at room temperature for 20 minutes, samples were analyzed using flow cytometry (FACS verse; BD Biosciences, Franklin Lake, NJ, USA) and relevant FlowJo software.

Biodistribution of PEG-DA-CuS NPs

An aqueous solution (0.1 mL) of PEG-DA-CuS NPs (100 mg) was intravenously (iv) administered into the HeLa-xenograft-bearing mice with the mice divided into two groups. One group received no laser irradiation, while the other group was irradiated by 1,064 nm laser 0.5 hour after the injection. The mice were sacrificed (six mice per time point) at 0.5, 1, 3, 8, 12, and 24 hours after the injection. The tumor, liver, heart, spleen, kidney, lung, and blood were collected, weighed, completely lysed in aqua regia, and examined by an inductively coupled plasma-mass spectrometer (NexION 300 D; PerkinElmer Inc., Waltham, MA, USA).

In vitro antitumor effect

To assess the in vitro PTT rendered by PEG-DA-CuS NPs, HeLa cells were incubated in the confocal dish for a day at 37°C. Then, the HeLa cells were cultured with PEG-DA-CuS NPs (200 μ g/mL) and incubated for 5 hours. The media were discarded and the cells were washed with PBS thrice and exposed to 1,064 nm laser at a power of 1 W/cm² (10 minutes). The viability of HeLa cells posttreatment was

indicated by staining the cells with calcein-AM or EthD-1 accordingly. Finally, laser scanning confocal microscopy was used to determine the cells' viability with and without formalin fixation.

In vivo antitumor effect

Mice bearing HeLa tumors in the right flank, with average tumor sizes of 500 mm³, were used to assess the in vivo antineoplastic efficiency. The mice were randomized into four different groups (iv injection of saline [50 mg/kg], iv injection of PEG-DA-CuS [50 mg/kg] alone, iv injection of saline [50 mg/kg] plus 1,064 nm laser, iv injection of PEG-DA-CuS [50 mg/kg] plus 1,064 nm laser). Tumor size was evaluated by monitoring and recording the volume and body weight. After the mice were sacrificed 12 days after the treatments, the tumors were collected, weighed, and fixed in a 10% neutral buffered formalin solution after three washes with PBS. Immunohistochemistry for Ki67 staining tests and H&E staining tests were also conducted by Yili biotechnology Company (Wu Han, China) and observed via fluorescence microscopy (IX71; Olympus Corporation, Tokyo, Japan).

Pathology analysis

Organs (lung, heart, spleen, liver, and kidney) and skin tissues were collected from the mice on day 7, day 15, and day 30 after the PEG-DA-CuS NPs-based PTT treatment to observe histological changes in mice. These organs were embedded in paraffin after immobilization in 4% paraformaldehyde at 4°C for 4 hours. At last, the sections were stained with H&E and representative images were collected using a light microscope.

Hematology and biochemical assay

Hematology and biochemical assays were conducted by collecting mouse blood on day 7, day 15, and day 30 after the PEG-DA-CuS NPs-based PTT treatment or from untreated normal mice. In order to assess the biocompatibility and in vivo toxicity of PEG-DA-CuS NPs, we analyzed kidney function and spleen function by monitoring blood urea nitrogen (BUN), platelet (PLT) level, and urea creatinine (CRE). ALT, total protein, and total bilirubin level were determined to study serological liver function. In addition, blood CRE and BUN were used to detect kidney function, and the PLT level was correlated with spleen function. Neutrophils, red blood cells, monocytes, lymphocytes, and white blood cells were counted by flow cytometry assay, and levels of hemoglobin were tested to evaluate the potential cytotoxic effect and immune response.

Results and discussion

Characterization of PEG-DA-CuS NPs

The morphology of the as-obtained PEG-DA-CuS NPs was depicted by TEM as indicated in Figure 1A. From the micrograph, we could find that the as-acquired PEG-DA-CuS NPs have the morphology of homogeneously dispersed sphere and a mean diameter of about 10 nm. UV-Vis spectra (Figure 1B) of PEG-DA-CuS displayed a strong absorption peak at 1,065 nm, suggesting the superior potency of PEG-DA-CuS

NPs for converting the absorbed optical energy into heat. After a 10-minute irradiation by 1,064 nm laser, there was no apparent absorption change in the UV spectra of PEG-DA-CuS NPs, indicating the wonderful photothermal stability. The profile of temperature elevation was investigated and is recorded in Figure 1C. Aqueous solution (1 mL) containing 1 mg PEG-DA-CuS NPs was put in a quartz cell and subjected to laser irradiation (1,064 nm, 1 W/cm²). A thermographic camera was used to track the increasing

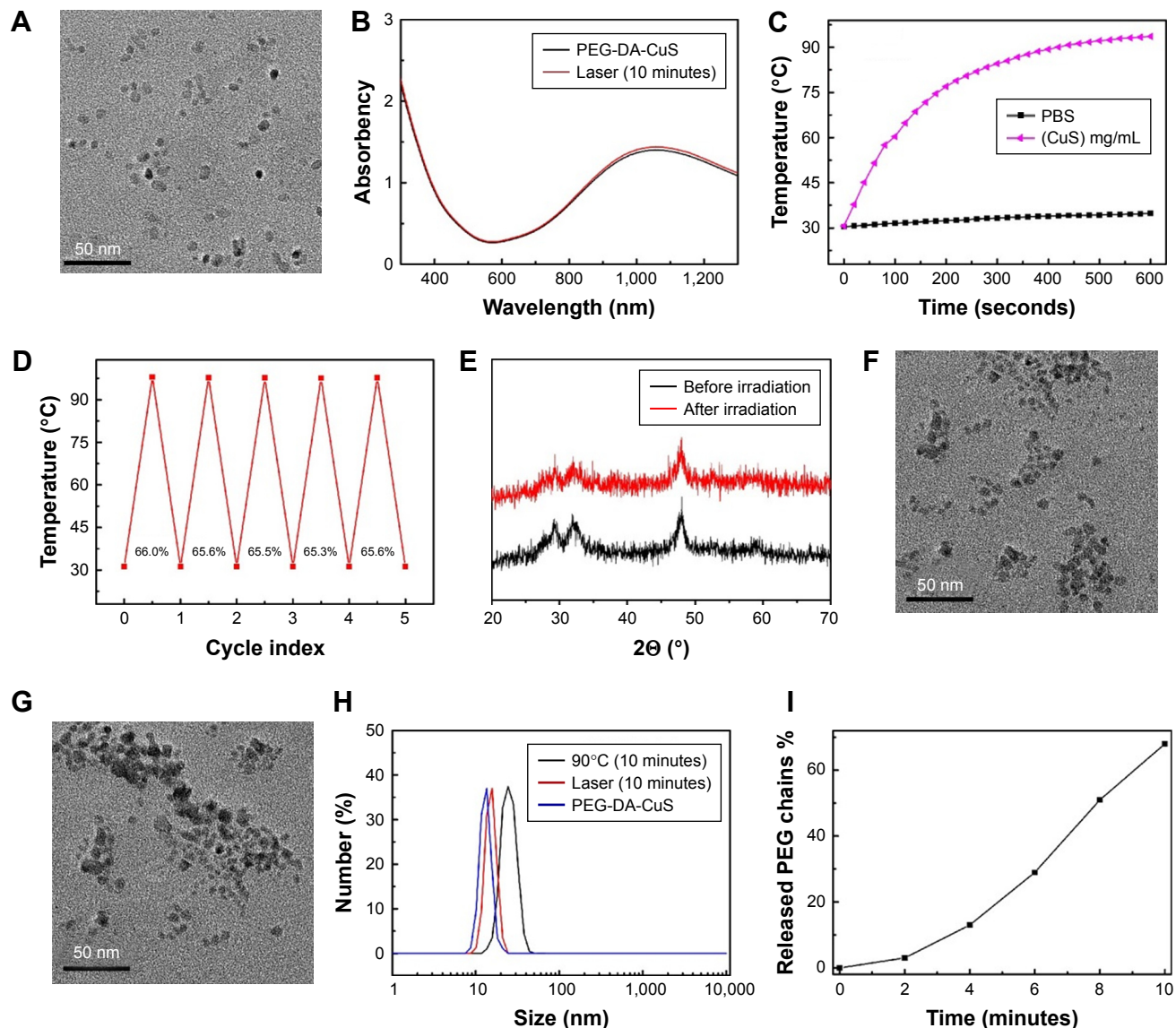


Figure 1 Characterization of PEG-DA-CuS NPs.

Notes: (A) TEM image of PEG-DA-CuS NPs. (B) UV-Vis spectrum of PEG-DA-CuS NPs before and after NIR laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes). (C) Temperature-time curve of PBS and PEG-DA-CuS NPs (1 mg/mL) dissolved in distilled water. (D) Temperature of PEG-DA-CuS NPs solution (1.43 g, 1.4 mL) after five cycles of NIR laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes)/cooling down (1 hour) process. (E) XRD of PEG-DA-CuS NPs before and after NIR laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes). (F) TEM image of PEG-DA-CuS NPs after NIR laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes). (G) TEM image of PEG-DA-CuS NPs after oil bath heating (90°C, 10 minutes). (H) Particle size distribution of PEG-DA-CuS NPs before NIR laser irradiation, after NIR laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes), and after oil bath heating (90°C, 10 minutes). (I) Release of PEG chains from the PEG-DA-CuS after irradiation with 1,064 nm laser (1 W/cm²) for 10 minutes.

Abbreviations: NIR, near infrared; NP, nanoparticle; PEG, polyethylene glycol; TEM, transmission electron microscopy; UV, ultraviolet; XRD, X-ray powder diffraction.

temperature. As indicated in Figure 1C, the temperature curve of PEG-DA-CuS NPs solution exhibited significant growth (increased from 30.2°C to 91.5°C) 10 minutes after 1,064 nm laser irradiation; on the contrary, the control solution only increased from 30.2°C to 32°C. These data prove that the PEG-DA-CuS NPs featured such impressive photothermal properties that ensured the success of retro Diels–Alder reaction. According to Figure 1D, the conversion efficiency was estimated to be around 65.6%, and this value did not decrease significantly after repeated laser irradiation, which confirmed that PEG-DA-CuS NPs represented excellent stability as photothermal agents. X-ray power diffraction measurements of PEG-DA-CuS NPs (Figure 1E) exhibited typical diffraction peaks of CuS (JCPDS no 06-0464). There was no detectable change in these NPs after irradiation with 1,064 nm laser, again showing the stability of PEG-DA-CuS NPs. Since the temperature of PEG-DA-CuS NPs when exposed to 1,064 nm laser for 10 minutes was high enough to induce retro Diels–Alder reaction and release the PEG chain, we next investigated the morphology change and size change of PEG-DA-CuS NPs after laser irradiation via TEM and dynamic light scattering. In Figure 1F–H, we could

see the aggregate morphology and enhanced hydrodynamic diameter regardless of 10 minutes irradiation with 1,064 nm laser or heating using oil at 90°C. All these taken together, we conclude that the as-obtained PEG-DA-CuS NPs had two major properties. On one hand, they maintained good photothermal and structural stability. On the other hand, their superior photothermal effect made it possible that the PEG chain would be released from PEG-DA-CuS NPs once the retro Diels–Alder reaction was triggered by 1,064 nm laser irradiation. Next, we conducted the iodine–barium assay and analyzed the quantified release of PEG chains from the PEG-DA-CuS NPs. In Figure 1I, it could be seen that little PEG chain was released in the first 5 minutes owing to the low temperature. However, as the temperature rose, more and more PEG was released and ~68% of the modified PEG chain was detected in the PEG-DA-CuS solution after irradiation with 1,064 nm laser (1 W/cm²) for 10 minutes.

In vitro cytotoxicity

The in vitro cytotoxicity of PEG-DA-CuS NPs was assessed in normal HUVECs via flow cytometry analysis. As displayed in Figure 2A–C, no obvious loss of cell viability (<7%) existed

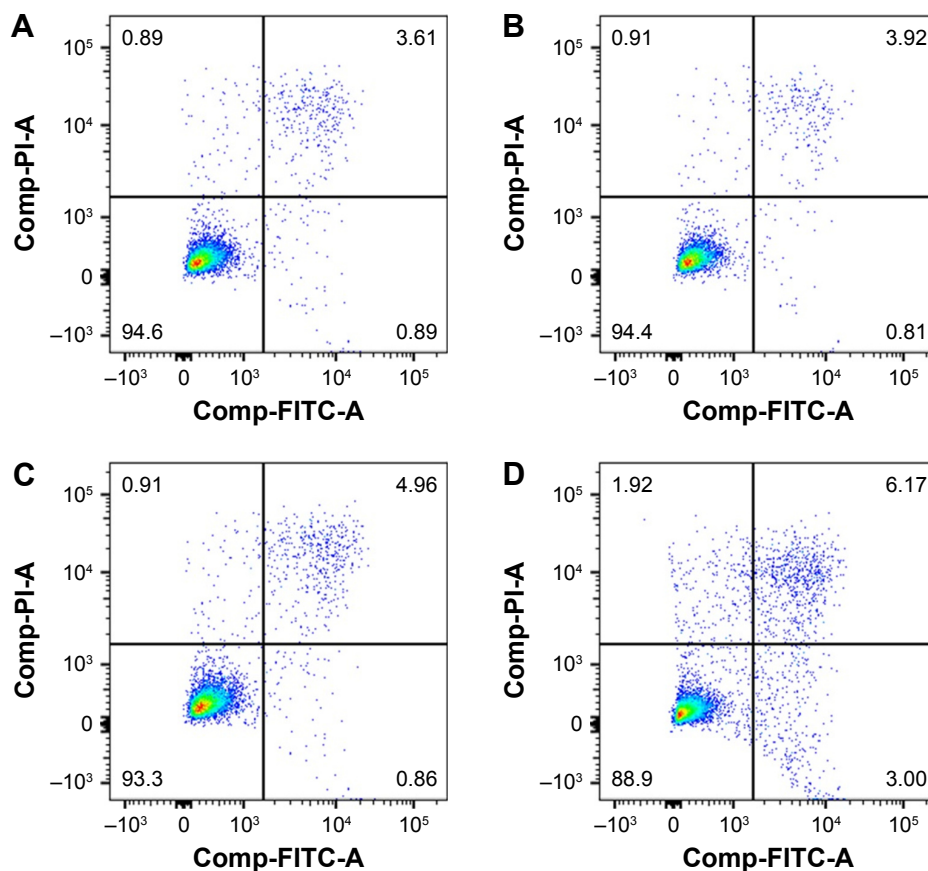


Figure 2 Flow cytometry analysis of HUVECs cultured with PEG-DA-CuS NPs at various concentrations: (A) 0.1 mg/mL; (B) 0.2 mg/mL; (C) 0.4 mg/mL; and (D) 1 mg/mL. **Abbreviations:** FITC, fluorescein isothiocyanate; HUVECs, human umbilical vein endothelial cells; NPs, nanoparticles; PEG, polyethylene glycol; PI, propidium iodide.

when the concentration was below 0.5 mg/mL, indicating the cells demonstrate excellent tolerance to PEG-DA-CuS NPs of low concentration. In addition, even if the drug concentration was raised to 1 mg/mL (Figure 2D), over 88% viability could still be reached. These data are good proof of the compatibility of PEG-DA-CuS NPs for practical applications.

Biodistribution of PEG-DA-CuS NPs

PEG was reported to have the function of prolonging the blood circulation of NPs, and we analyzed the blood retention profiles of PEG-DA-CuS NPs postinjection. As displayed in Figure 3A, the half-life of PEG-DA-CuS NPs in blood is 252 minutes, which proves the long circulation time. As such, we speculated that the removal of PEG from PEG-DA-CuS NPs at the tumor site would shorten the blood circulation of CuS NPs and further cause the aggregation of CuS around the tumor, leading to the passive enrichment of CuS in the tumor. To demonstrate this hypothesis, the biodistribution of CuS NPs was monitored on HeLa-xenograft-bearing mice treated with 10-minute 1,064 nm laser at the power of

1 W/cm² at 0.5 hour after the iv injection or with nothing after the injection. We noticed that, as a result of the aggregation of CuS around the tumor (Figure 3B), a large proportion of the iv-injected CuS NPs was enriched in the tumor tissue through the enhanced permeability and retention effect. By contrast, only a small amount of the CuS NPs was enriched in the tumor due to the long blood circulation time of PEG (Figure 3C).

In vitro antitumor efficacy

Live/dead double-staining method (Figure 4) was applied to verify the therapeutic outcome of PTT in cells in vitro under different treatments. Calcein-AM stained live cells with green fluorescence, while EthD-1 showed dead cells with red fluorescence. No red color could be detected in the control group (HeLa cells treated with nothing) and two negative groups (HeLa cells irradiated by 1,064 nm laser (10 minutes, 1 W/cm²) or incubated with PEG-DA-CuS NPs alone), indicating negligible cytotoxic damage to the HeLa cells was exerted by these treatments. On the contrary, a mass

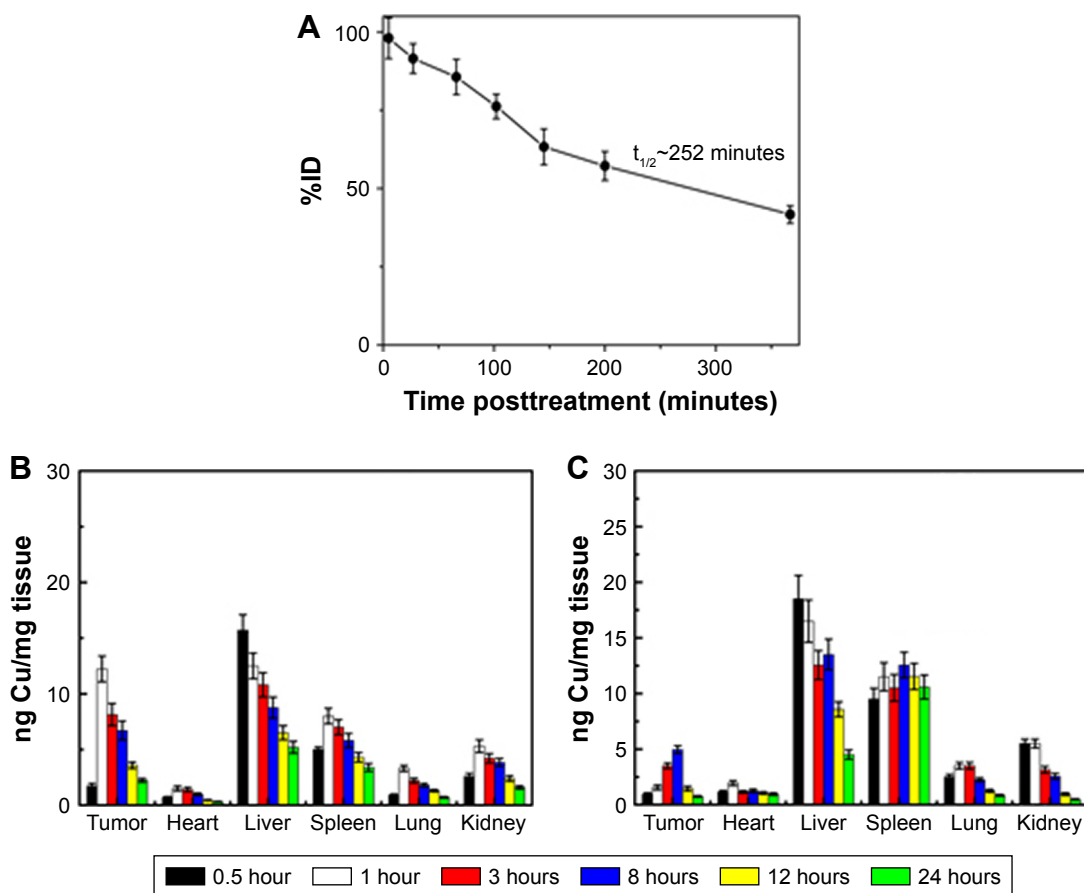


Figure 3 (A) In vitro distribution profiles of CuS NPs in blood. (B) Biodistribution analysis of CuS NPs without laser irradiation. (C) Biodistribution analysis of CuS NPs with laser irradiation (1 W/cm², 1,064 nm laser, 10 minutes).

Abbreviation: NP, nanoparticle.

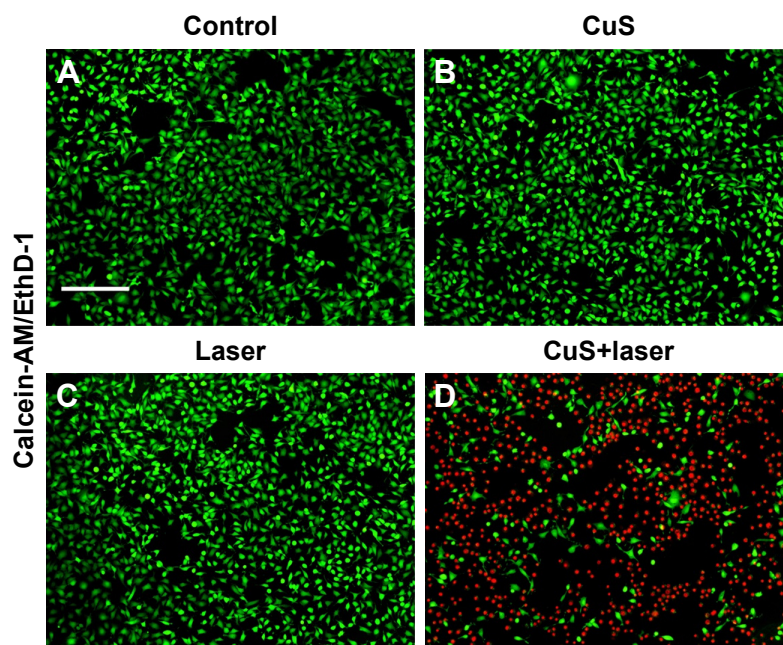


Figure 4 Fluorescence images of HeLa cells.

Notes: (A) The cells were incubated with nothing. (B) The cells were incubated with PEG-DA-CuS NPs (200 $\mu\text{g}/\text{mL}$) alone. (C) The cells were exposed to laser irradiation (1 W/cm^2 , 1,064 nm laser, 10 minutes) alone. (D) The cells were incubated with PEG-DA-CuS NPs (200 $\mu\text{g}/\text{mL}$) and exposed to laser irradiation (1 W/cm^2 , 1,064 nm laser, 10 minutes). Calcein-AM (green fluorescence representing live cells) and EthD-1 (red fluorescence representing dead cells) were used as indicators. The scale bar is 50 μm . Magnification is 10 \times .

Abbreviations: AM, acetoxymethyl; EthD-1, ethidium homodimer-1; NPs, nanoparticles; PEG, polyethylene glycol.

of red fluorescence was observed when the cells received the PTT treatment rendered by PEG-DA-CuS NPs, which confirmed the wonderful therapeutic efficacy of PEG-DA-CuS NPs against HeLa cells.

In vivo antitumor efficacy

Subsequently, the in vivo antitumor efficacy of PTT rendered by PEG-DA-CuS NPs on mice bearing HeLa tumors was investigated. As represented in Figure 5A, the tumor from the control group (saline group) and other two negative groups (PEG-DA-CuS NPs alone and 1,064 nm laser alone) showed faster growth rates, accompanied by an ~ 7 -fold increase in volume (from day 12 to day 0). On the contrary, the PEG-DA-CuS NPs treatment significantly ablated the tumor volume. Compared to day 0, the tumor volume of the PEG-DA-CuS NPs plus laser group reached 1.2-fold on day 12, showing dramatic photothermal effect in the elimination of cancer simultaneously. According to the recorded body weight curve of mice depicted in Figure 5B, no abnormal weight changes were detected throughout the whole process, confirming the PEG-DA-CuS NPs-based PTT would not cause apparent acute toxicity in mice bearing HeLa tumor.

H&E staining (Figure 5C) and Ki67 staining (Figure 5D) were used to evaluate the therapeutic effect after different treatments. Tumor tissues from mice treated with PTT rendered by PEG-DA-CuS NPs presented remarkable

necrosis and apoptosis. Therefore, we concluded that the PTT rendered by PEG-DA-CuS NPs offered an excellent therapeutic efficacy in cancer treatment.

Safety evaluation

Major organs (heart, lung, spleen, kidney, and liver) of mice receiving PTT rendered by PEG-DA-CuS NPs were collected and weighed to assess the in vivo cytotoxicity. Compared with the control group, H&E staining pictures obtained from the experimental group displayed in Figure 6A represented negligible histological abnormalities, necrosis, inflammation lesions, and pulmonary fibrosis, demonstrating excellent in vivo biocompatibility of PEG-DA-CuS NPs.

We also selected four nude mice and picked the skin tissues in the same location of the mice body to conduct safety evaluation via histological analysis. The selected skin tissues were treated in different ways (treated with nothing as the control group, injected with PEG-DA-CuS NPs alone, laser irradiation alone, and injected with PEG-DA-CuS NPs and irradiated with 1,064 nm laser), and the H&E results are shown in Figure 6B. Compared with the control group, the skin tissues of mice treated with PEG-DA-CuS NPs alone or laser irradiation alone represented no apparent histological changes. On the contrary, the skin structure of those receiving PEG-DA-CuS NPs treatment under laser irradiation was destroyed, leaving only two to three layers of stratum corneum.

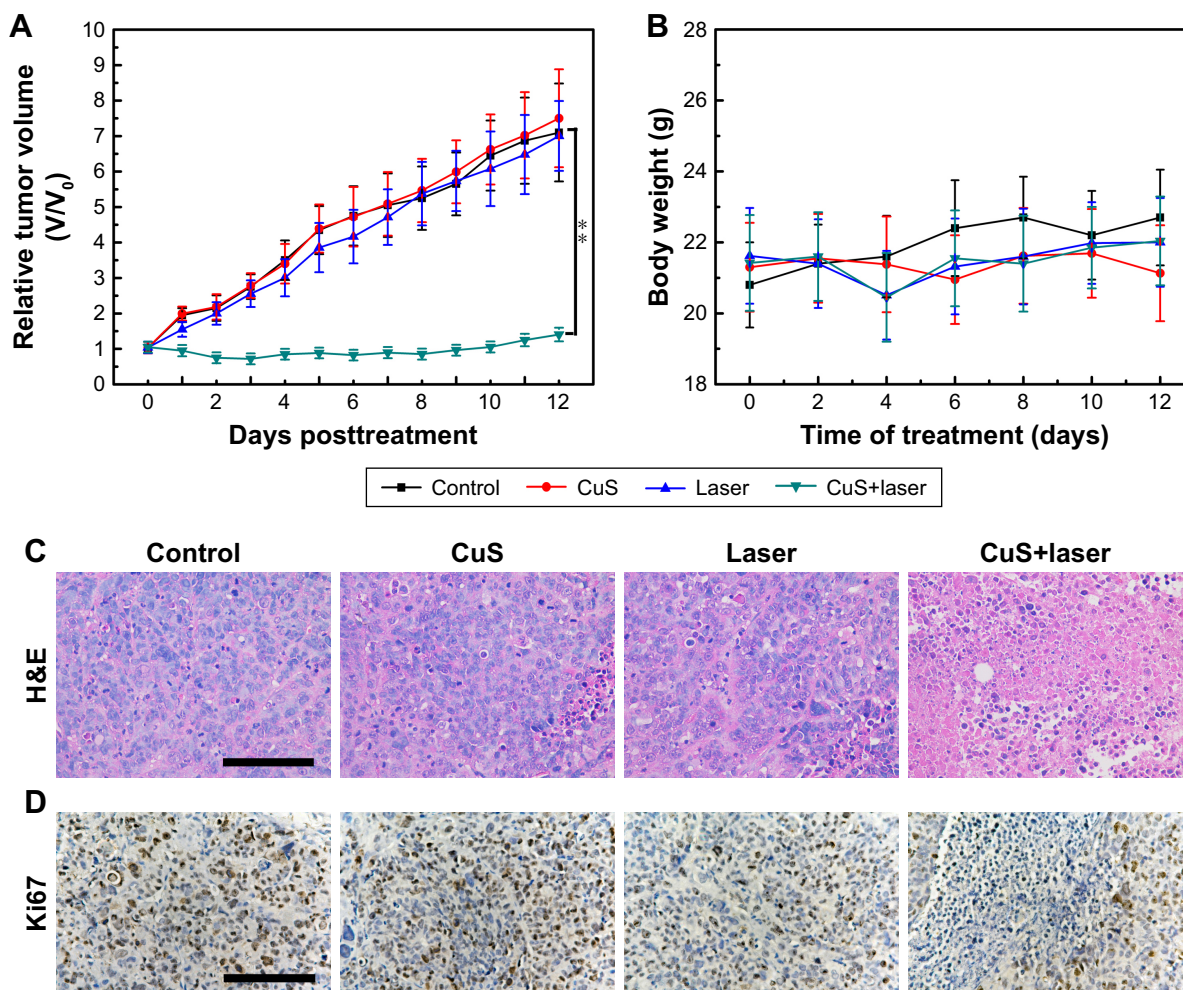


Figure 5 (A) Tumor growth curves of HeLa tumor-bearing mice subjected to different treatments as displayed. The data are expressed as the mean \pm SD (n=8). ** P <0.01. (B) Body weight of HeLa tumor-bearing mice subjected to different treatments. (C) IHC staining images of HeLa tumor sections from mice subjected to different treatments, obtained with H&E staining. (D) Representative IHC for Ki67 staining images of HeLa tumor sections obtained from mice subjected to different treatments. The scale bar is 50 μ m. **Abbreviation:** IHC, immunohistochemistry.

These data proved that damages on the normal tissues occurred only in NP-enriched location with the help of laser irradiation. Therefore, guiding the laser to the lesions (such as tumors) directly without penetrating the normal tissues will be a solution. We believe, with the development of modern medical techniques, approaches to direct the laser to the lesions (such as tumors) without penetrating the skin will be discovered. In addition, serum biochemical study and hematology assay were conducted to further investigate the potential cytotoxicity of PEG-DA-CuS NPs. No obvious changes in statistical significance were found in total bilirubin, ALT, total protein, CRE, BUN, and PLT when compared with the control group (untreated mice), indicating liver, kidney, and spleen functions were normal and their metabolism was not disrupted after PEG-DA-CuS NPs treatment (Figure 7A and B). The clinical fate of NPs was decided by hepatotoxicity. As shown in Figure 7C, hematopoietic and aerobic capacity reflected by

the counts of hemoglobin, white blood cells, and red blood cells was similar compared with the control group (untreated mice). There were no detectable changes in peripheral blood monocytes, lymphocytes, and neutrophil counts (Figure 7D) when compared with the control group, suggesting no potential immune responses occurred throughout the treatment. Taken together, all these data offer solid evidence that PEG-DA-CuS NPs have promising clinical applications.

Conclusion

We reported a kind of ideal photothermal agent, PEG-DA-CuS NPs, for NIR laser-induced PTT. Success of this methodology relies on the occurrence of retro Diels–Alder reaction triggered by the intrinsic photothermal property of CuS NPs, allowing the aggregation of photothermal agents in the tumor. In addition, based on the results of in vitro experiments, we noticed that PEG-DA-CuS NPs show excellent photothermal stability

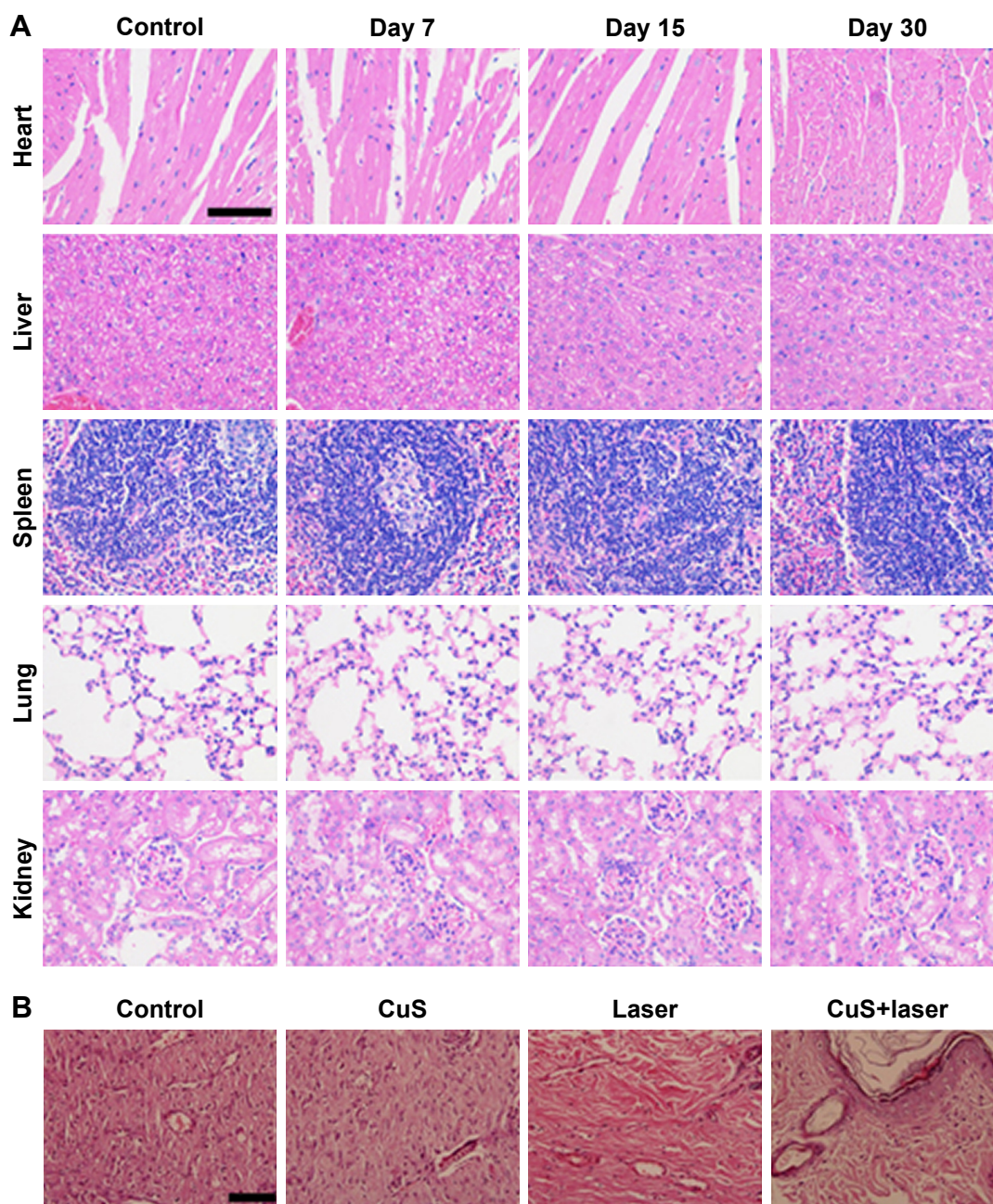


Figure 6 (A) Histological images of the HE-stained heart, liver, spleen, lung, and kidney harvested from mice after treatment with PTT rendered by PEG-DA-CuS NPs or from mice treated with nothing as the control group. **(B)** Histological images of the H&E-stained skin harvested from mice after different treatments (treated with nothing as the control group, injected with PEG-DA-CuS NPs alone, laser irradiation alone, and injected with PEG-DA-CuS NPs and irradiated with 1,064 nm laser). We selected four nude mice and picked the skin tissues in the same location of the mice body to conduct the histological analysis. The scale bar is 50 μ m.

Abbreviations: NPs, nanoparticles; PEG, polyethylene glycol; PTT, photothermal therapy.

and high heat conversion efficiency. Moreover, the *in vivo* experiments indicated the tumor size of mice receiving PTT rendered by PEG-DA-CuS NPs was significantly reduced, prolonging the lives of mice, and further confirmed that PEG-DA-CuS NPs exhibited low cytotoxicity. Therefore, we are quite sure that PEG-DA-CuS NPs can be used as promising photothermal agents in the ablation of tumors.

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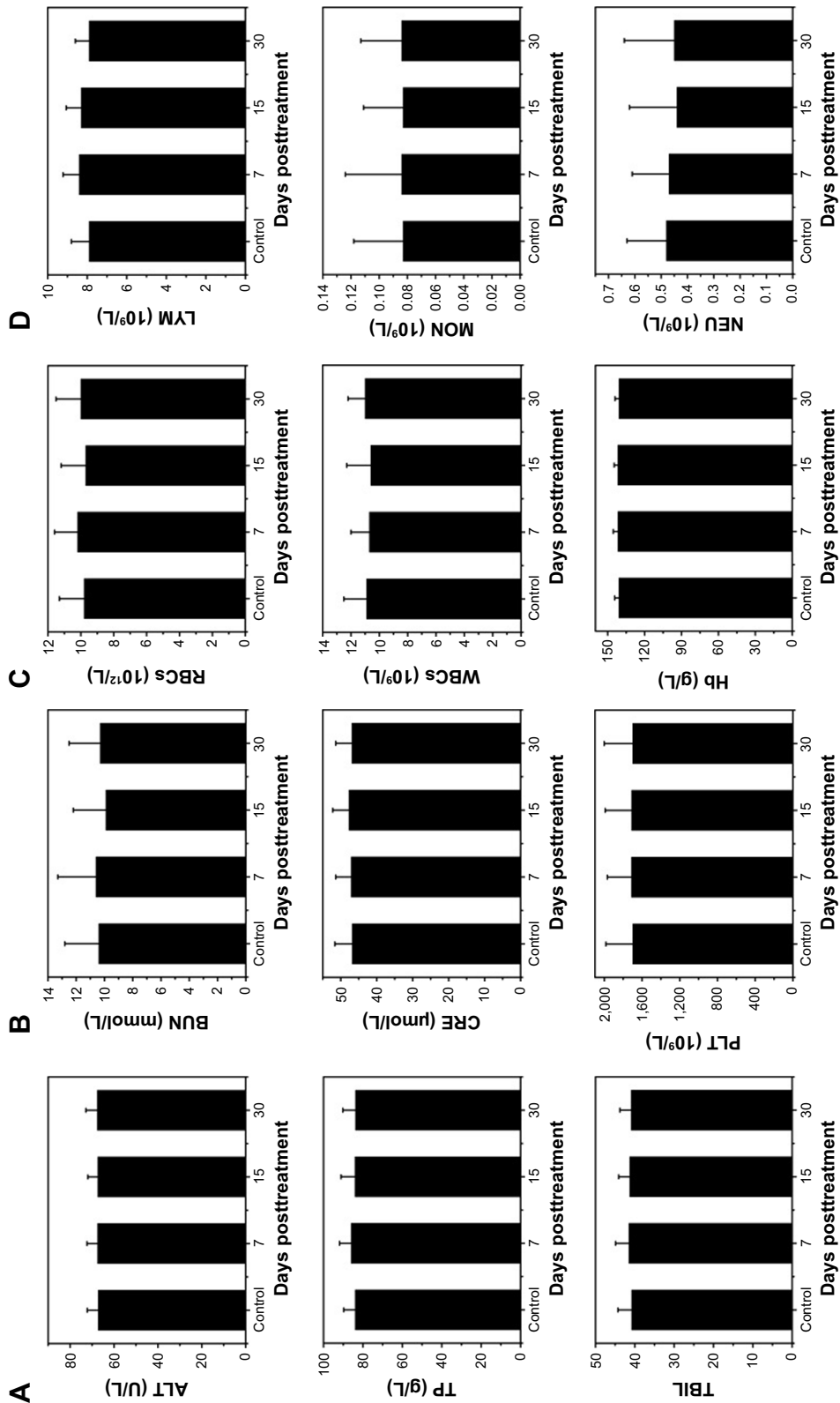


Figure 7 Serum biochemical study and hematology assay conducted in mice at 7, 15, and 30 days after PTT rendered by PEG-DA-CuS NPs and in mice treated with nothing as the control group.

Notes: (A) Serum biochemical test of ALT, TBIL, TP. (B) CRE, BUN, and hematology assay of PLT. (C) RBCs, Hb, and WBCs. (D) MON, LYM, and NEU.

Abbreviations: BUN, blood urea nitrogen; CRE, creatinine; Hb, hemoglobin; LYM, lymphocyte; MON, monocyte; NEU, neutrophil; NPs, nanoparticles; PEG, polyethylene glycol; PLT, platelet; PTT, photothermal therapy; RBC, red blood cell; TBIL, total bilirubin; TP, total protein; WBC, white blood cell; ALT, alkaline phosphatase.

Author contributions

All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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