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Anticonvulsant effects of *antiaris toxicaria* aqueous extract: investigation using animal models of temporal lobe epilepsy

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Abstract

Background: *Antiaris toxicaria* has previously shown anticonvulsant activity in acute animal models of epilepsy. The aqueous extract (AAE) was further investigated for activity in kindling with pentylenetetrazole and administration of pilocarpine and kainic acid which mimic temporal lobe epilepsy in various animal species.

Results: ICR mice and Sprague–Dawley rats were pre-treated with AAE (200–800 mg kg⁻¹) and convulsive episodes induced using pentylenetetrazole, pilocarpine and kainic acid. The potential of AAE to prevent or delay onset and alter duration of seizures were measured. In addition, damage to hippocampal cells was assessed in kainic acid-induced *status epilepticus* test. 800 mg kg⁻¹ of the extract suppressed the kindled seizure significantly ($P < 0.05$) as did diazepam. AAE also produced significant effect ($P < 0.01$) on latency to first myoclonic jerks and on total duration of seizures. The latency to onset of wet dog shakes was increased significantly ($P < 0.05$) by AAE on kainic acid administration. Carbamazepine and Nifedipine (30 mg kg⁻¹) also delayed the onset. Histopathological examination of brain sections showed no protective effect on hippocampal cells by AAE and nifedipine. Carbamazepine offered better preservation of hippocampal cells in the CA1, CA2 and CA3 regions.

Conclusion: *Antiaris toxicaria* may be effective in controlling temporal lobe seizures in rodents.

Keywords: Hippocampus, Kainic acid, Pentylenetetrazole, Pilocarpine, Kindling

Background

Epilepsy is a common neurological disorder which may be due to an imbalance between excitatory and inhibitory arms of the central nervous system—produced by a decrease in GABAergic and/or an increase in glutamatergic transmission [1].

Kindling and Status Epilepticus are the two most commonly used animal models of Temporal Lobe Epilepsy (TLE). Both models provide a dependable induction of a persistent, epileptic-like condition, despite their unique characteristics. Kindling is a simple phenomenon in which repeated induction of focal seizure discharge produces a progressive, highly reliable, increase in epileptic

response to the inducing agent, usually electrical stimulation [2]. However, the use of chemical inducing agents, such as pentylenetetrazole has been shown to be equally effective [3, 4]. Acute administration of a high dose of pilocarpine in rodents is widely used to study the pathophysiology of seizures. It was first described by Turski et al. in 1983 [5, 6]. Pilocarpine-induced seizures reveal behavioural and electroencephalographic features that are similar to those of human temporal lobe epilepsy. Kainic acid, like pilocarpine, can also be used to induced a similar TLE or status epilepticus state in a variety of species using either systemic, intrahippocampal or intramygdaloid administrations [7].

Temporal lobe epilepsy is the most common form of complex partial seizures accounting for approximately 60% of all patients with epilepsy. Medial temporal lobe epilepsy which is the commonest temporal lobe epilepsy is also frequently resistant to medications and associated

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with hippocampal sclerosis. Management is challenging and often surgery has to be resorted to [8].

The plant *Antiaris toxicaria* (family Moraceae) is a common plant in Ghanaian forests. It has been employed traditionally as an analgesic and anticonvulsant [9]. Previous studies have shown that *Antiaris* possesses anticonvulsant activity in various acute murine models [10]. *Antiaris toxicaria* in this present study was evaluated to determine its properties in kindling models and post-status models of temporal lobe epilepsy. This investigation sought to determine if the extract possessed potential as an antiepileptogenic agent as well as efficacy in the management of temporal lobe epilepsy.

Methods

Plant material

Stem bark of *A. toxicaria* was harvested from the KNUST campus, Kumasi and identified by a staff member of the Pharmacognosy Department where a voucher specimen (KNUST/HM1/011/S007) has been retained in the herbarium.

Preparation of *Antiaris toxicaria* aqueous extract

The dry stem bark was powdered using a commercial grinder. The coarse powder (431 g) was extracted by cold maceration with distilled water as solvent at room temperature for 5 days. The resultant filtrate was oven-dried to obtain 23.40% w/w of *A. toxicaria* aqueous extract (AAE).

Animals

Naïve male ICR mice (20–25 g) and Sprague–Dawley Rats (120–145 g) were obtained from Noguchi Memorial Institute for Medical Research, Accra, Ghana and kept in the departmental Animal House. Animals were maintained under laboratory conditions (room temperature; 12-h light–12-h dark cycle) in stainless steel cages (34 × 47 × 18 cm³) with wood shavings as bedding and allowed free access to water and food ad libitum. They were fed with normal commercial diet (GAFCO Ltd). Animals were tested in groups of eight. Groups were assigned randomly. Sample size was calculated using method of power analysis using the G-power software version 3.0.5. Experiments were carried out during the day. All animals were handled in accordance the Guide for the Care and Use of Laboratory Animals [11] and experiments were approved by the Faculty of Pharmacy and Pharmaceutical Sciences Ethics Committee, KNUST.

Drugs and chemicals

Diazepam (DZP), pentylenetetrazole (PTZ) pilocarpine (PILO) and kainic acid (KA) were purchased from Sigma-Aldrich Inc., St. Louis, MO, USA.

Kindling induction

PTZ kindling was initiated using a subconvulsive dose of PTZ 40 mg kg⁻¹ body weight injected into the soft skin fold of the neck on every 2nd day (i.e. Day 1, Day 3, Day 5...). The PTZ injections were stopped when the control animals showed adequate kindling, i.e. Racine score of 5. After each PTZ injection, the convulsive behaviour of the rodent was observed for 30 min in an observation chamber. The resultant seizures were scored as follows: Stage 0 (no response); Stage 1 (hyperactivity, restlessness and vibrissae twitching); Stage 2 (head nodding, head clonus and myoclonic jerks); Stage 3 (unilateral or bilateral limb clonus); Stage 4 (forelimb clonic seizures); Stage 5 (generalized clonic seizures with loss of postural control). AAE was tested at doses of 200, 400 and 800 mg kg⁻¹ body weight orally and diazepam (0.1, 0.3 and 1 mg kg⁻¹, i.p). PTZ was injected 30 min after administration of test drugs. Control animals received 3 ml kg⁻¹ of distilled water. Seven groups of eight animals each were used. Group 1 = distilled water-treated control group; groups 2–4 = AAE-treated groups and groups 5–7 = diazepam-treated group.

Pilocarpine-induced Status epilepticus

Seizures were induced by an i.p. injection of pilocarpine (PILO) (300 mg kg⁻¹, i.p.) into drug or vehicle-treated male rats. Rats were pre-treated with AAE (100–1000 mg kg⁻¹, *p.o.*) or diazepam (0.3–3.0 mg kg⁻¹, i.p.) for 30 or 15 min, respectively, before PILO injection. To reduce peripheral autonomic effects produced by PILO, the animals were pre-treated with *n-butyl*-bromide hyoscine (1 mg kg⁻¹, i.p.) 30 min before PILO administration. Animals were placed in observation cages and observed via video recordings. Latency to and duration of seizures were scored.

Rat kainate model

Animals were pre-treated with the plant extract 30 min as above before administration of kainic acid (10 mg kg⁻¹, i.p.). Other animals were treated with carbamazepine (30 mg kg⁻¹, *p.o.*) and nifedipine (30 mg kg⁻¹, *p.o.*) 30 min before induction of convulsions. Animals were observed for wet dog shakes over a 1 h period [12]. Brains were harvested for histopathological examination after an hour. Tissues were fixed in 10% buffered formalin (pH 7.2). Dehydration was done with a series of ethanolic solutions, embedded in paraffin wax and processed for histological analysis. Coronal sections (2 μm thick) were cut and stained with haematoxylin-eosin for examination. The stained tissues were observed through an Olympus microscope (BX-51) and photographed by a chare-couple device (CCD) camera.

Data analysis

Data were presented as mean ± S.E.M and significant differences between means determined by one-way analysis of variance (ANOVA) followed by Newman–Keuls’ post hoc test. Statistical analyses were carried out with Graph Pad Prism® Version 5.0 (GraphPad Software, San Diego, CA, USA) and SigmaPlot® Version 11.0 (Systat Software, Inc.). Data from 5 to 8 animals in each group were included in the analyses. $P < 0.05$ was considered significant in all cases. None were excluded.

Results

Effects in kindling

In PTZ + vehicle-treated group, repeated administration of subconvulsive dose of PTZ (40 mg kg⁻¹) on every alternate day for 20 days resulted in increasing convulsive activity leading to generalized clonic seizures (Racine score of 5). Administration of AAE in the dose of 200 and 400 mg kg⁻¹ did not modify the course of kindling induced by PTZ significantly. However, a higher dose of 800 mg kg⁻¹ suppressed the kindled seizure significantly ($P < 0.05$; Fig. 1a, b) as the group could not achieve a

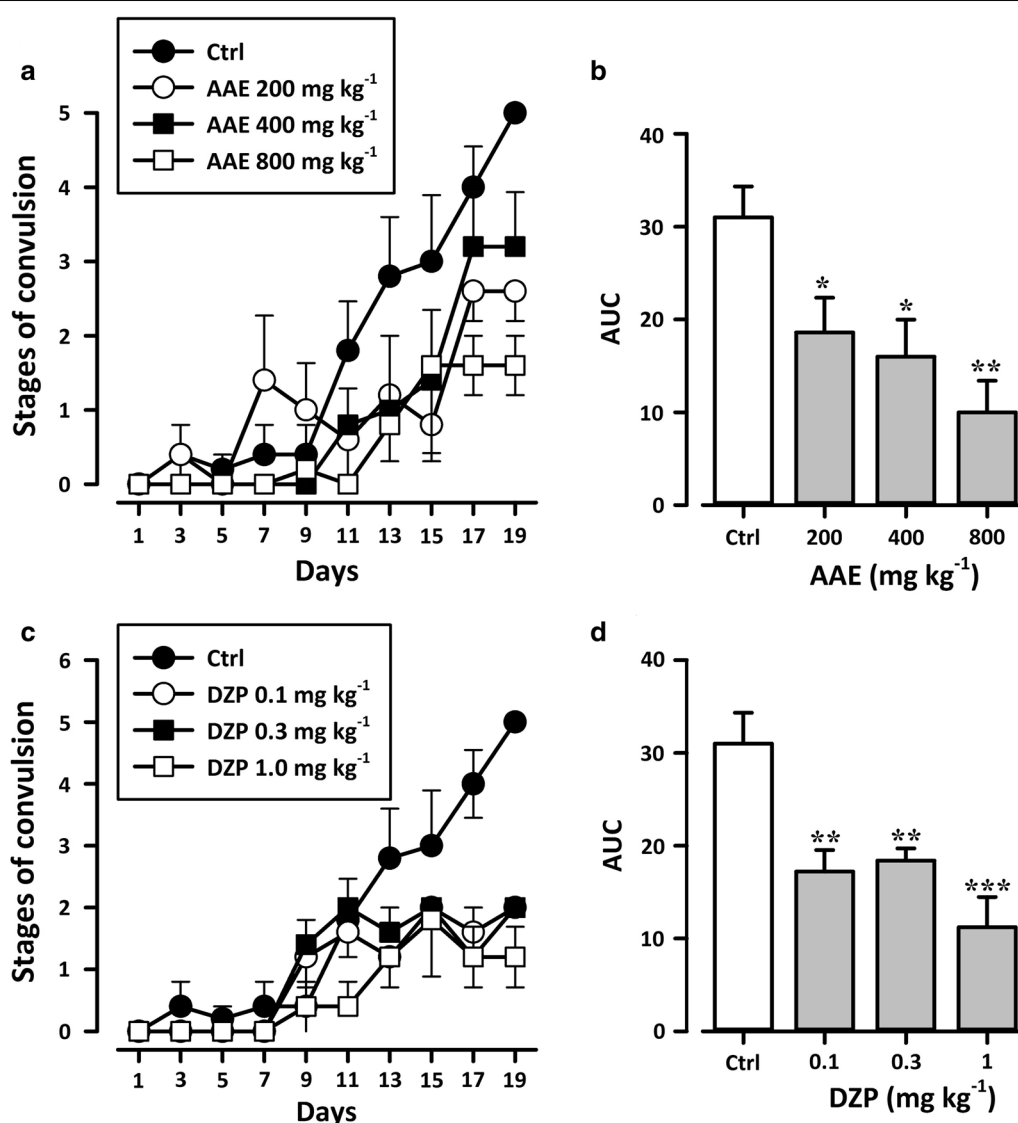


Fig. 1 Effects of AAE (200, 400 and 800 mgkg⁻¹, p. o.; **a** and **b**) and diazepam (0.1, 0.3 and 1 mgkg⁻¹, i.p.; **c** and **d**) on the stages of convulsion attained in PTZ-induced kindling. Data are presented as group mean ± SEM (n = 8). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared with vehicle treated group (One-way analysis of variance followed by Newman–Keuls post hoc test)

mean score of 5. The standard anticonvulsant diazepam significantly ($P < 0.01$; Fig. 1c, d) modified the course of kindling at all three dose levels compared to the control. ED_{50} obtained for the extract was $276.70 \text{ mg kg}^{-1}$ compared to 0.05 mg kg^{-1} for diazepam. The extract was however more efficacious than diazepam achieving an E_{max} of 88.83% compared to 60.36% for diazepam (Fig. 2).

Pilocarpine-induced status epilepticus

Pilocarpine induced behavioural changes including hypoactivity, tremor and myoclonic movements of the limbs progressing to recurrent myoclonic convulsions with rearing, falling, and status epilepticus. AAE produced significant effect ($P < 0.01$, Fig. 3a) on the latency to first myoclonic jerks as compared to control at the highest dose only. It had a similar effect on the total duration of seizures (Fig. 3c). Diazepam was used as the reference drug and it also significantly reduced the total duration of seizures ($P < 0.01$, Fig. 3d) and latency ($P < 0.001$, Fig. 3b) at 1 and 3 mg kg^{-1} . Diazepam was more potent than the extract in increasing the % latency with an ED_{50} of 0.66 mg kg^{-1} as against $424.50 \text{ mg kg}^{-1}$ for the extract (Fig. 4a). Diazepam was also more efficacious achieving an E_{max} of 108.90% compared to 100% for the extract. Likewise, for the % duration AAE produced $ED_{50} = 80.06 \text{ mg kg}^{-1}$ and $E_{\text{max}} = 100\%$ while the standard diazepam achieved $ED_{50} = 1.67 \text{ mg kg}^{-1}$ and $E_{\text{max}} = 100\%$ (Fig. 4b).

Effects in rat kainate model

Kainic acid (10 mg kg^{-1} , i. p) produced wet dog shakes in all animals. AAE (400 mg kg^{-1}) produced a significant

($P < 0.05$) increase in time taken to the onset of wet dog shakes (Fig. 5a). Carbamazepine (30 mg kg^{-1}) and Nifedipine (30 mg kg^{-1}) also delayed the onset. Histopathological examination of the coronal section of the brain showed no protective effect on hippocampal cells by AAE and nifedipine. Carbamazepine offered better preservation of hippocampal cells in the CA1, CA2 and CA3 regions (Fig. 6). The brain to body ratio decreased significantly ($P < 0.001$; Fig. 5b) with all three treatments.

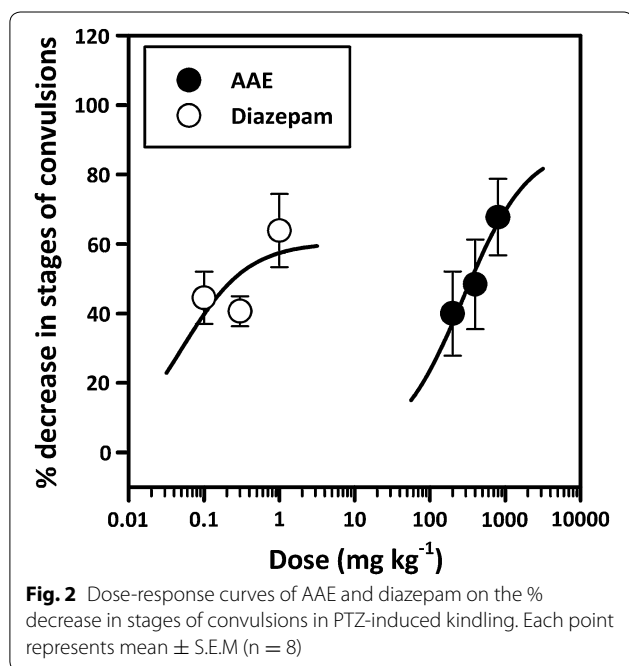
Discussion

Kindling is a chronic model of epilepsy and epileptogenesis. Repeated administration of a subconvulsive dose of PTZ (a blocker of the $GABA_A$ receptor) results in the progressive intensification of convulsant activity, culminating in a generalized seizure [3, 4]. The highest dose of AAE (800 mg kg^{-1}) significantly delayed progression of convulsion similarly to diazepam.

Many substances interacting with GABA receptors have been shown to produce potent anticonvulsant effects on seizures in previously kindled animals [2, 13]. It has been shown that AAE produces anticonvulsant effects by interacting with the $GABA_A$ receptor. The fact that it acts via GABAergic mechanisms may be a possible explanation for anticonvulsant effects being exhibited in the kindling model.

There is some evidence that free radicals are actively involved in physiological processes during oxidative stress induced by administration of convulsants [14]. Of all the free oxygen radicals that occur in vivo, the hydroxyl-free radicals (OH^-) are considered to be most hazardous [15–17]. Different mechanisms may lead to the increase of the free radicals in PTZ-induced convulsions. It may be assumed that further reason exist for the increased formation of OH^- in kindled animals during PTZ seizure, such as reduced activity of superoxide dismutase (SOD), a major defence system for counteracting the toxic effects of reactive oxygen species such as O^{2-} . However, antioxidant activity of AAE has not been firmly established.

AAE exhibited anticonvulsant effects against pilocarpine-induced seizures. Pilocarpine is a cholinergic agonist, widely used experimentally to induce limbic seizures in structures containing a high concentration of muscarinic receptors such as the cerebrum [18–20]. Status Epilepticus produces significant decreases in M_1 , M_2 , and GABAergic receptor densities [21] and hence neurotransmission. Freitas et al. have also reported in 2004 on increased levels of superoxide dismutase and catalase and reductions in acetylcholinesterase enzymatic activities in the rat frontal cortex and hippocampus. During pilocarpine-induced seizures and SE in adult rats, lipid peroxidation processes are increased [21, 22] suggesting



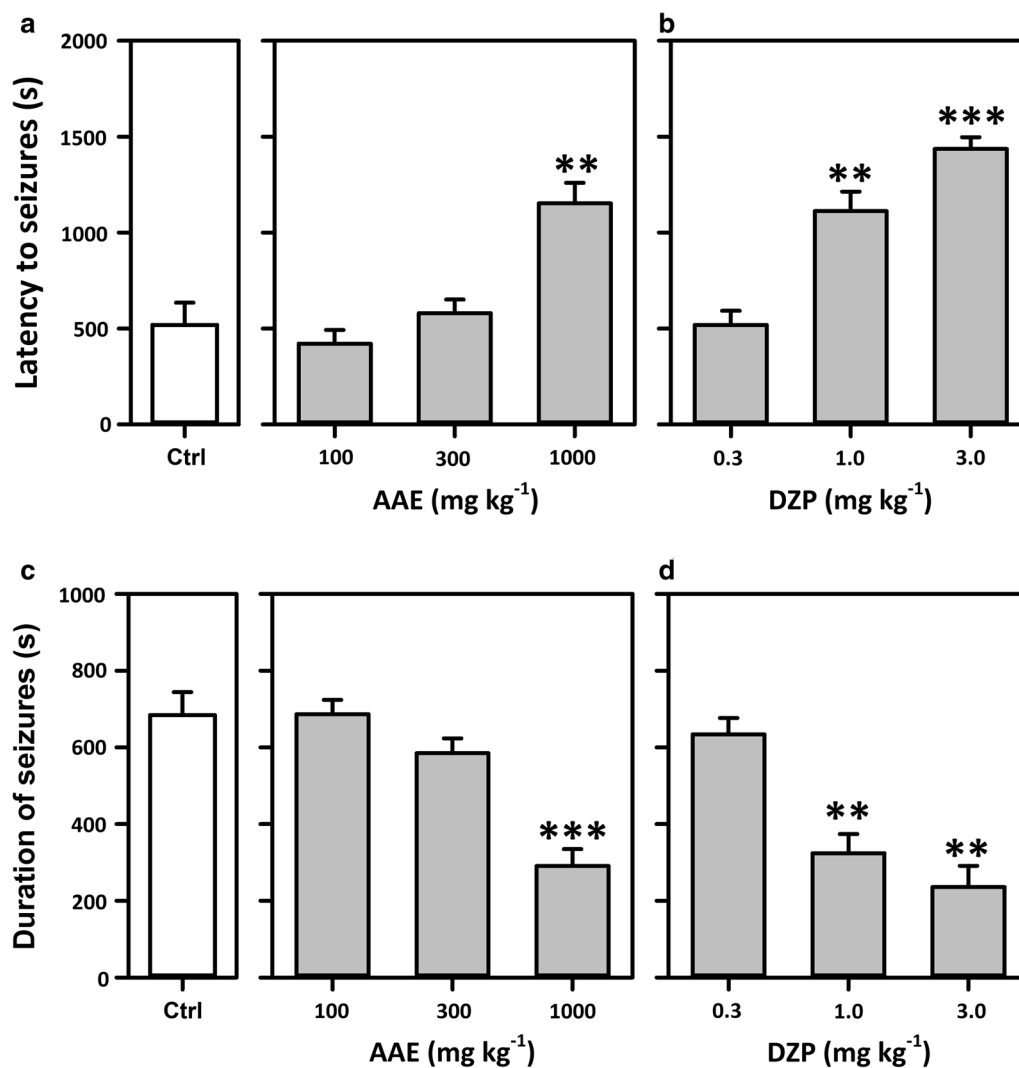
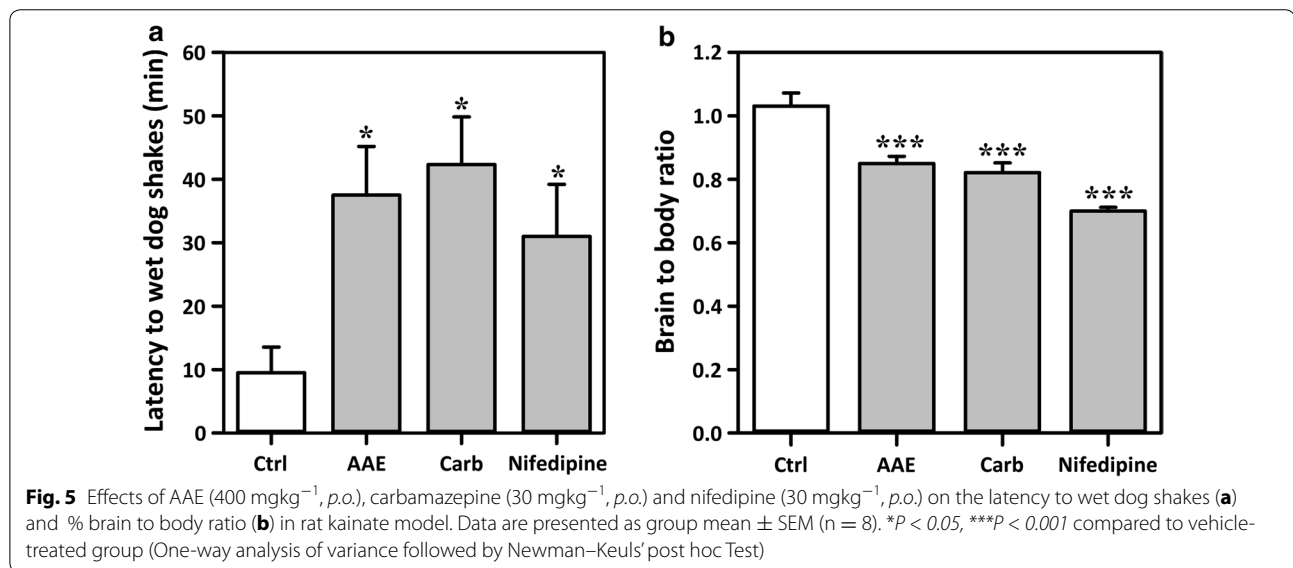
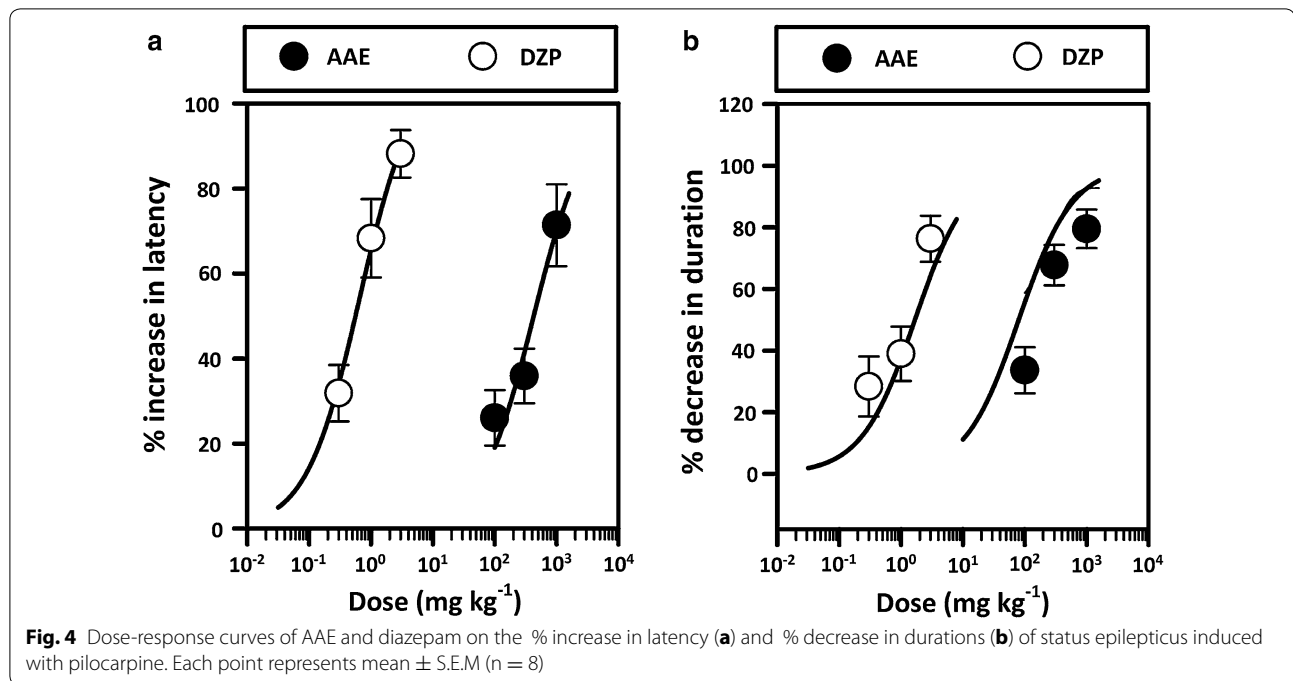


Fig. 3 Effect of AAE (100–1000 mg kg⁻¹, *p.o.*) and diazepam (0.3–3 mg kg⁻¹, *i.p.*) on the latency to (a and b) and total duration of seizures (c and d) induced by PILO. Each column represents the mean \pm SEM (n = 8). **P < 0.01, ***P < 0.001 compared to vehicle-treated group (One-way ANOVA followed by Newman-Keuls post hoc test)

free radical involvement in the pilocarpine-induced brain damage. Certain antioxidants, such as ascorbic acid, have therefore been shown to possess anticonvulsant activity against pilocarpine-induced SE [22, 23]. Muscarinic receptor stimulation is alleged to be responsible for the onset of pilocarpine-induced seizures, while glutamate acting on NMDA receptors sustains seizure activity [18]. Analysis of the brain morphology after pilocarpine administration demonstrates that the CA1 hippocampal neurones and the hilus of dentate gyrus are predominantly susceptible to neuronal cell loss [5]. Neuronal cell death during SE occurs largely by excitotoxic injury caused by the activation of glutamatergic pathways [6, 24]. Thus, the ability of AAE to attenuate seizures

induced by pilocarpine could be attributed to cholinergic antagonism at the M₁ or M₂ receptors, increase in GABA, and/or its receptor densities, decrease in glutamate levels or through antioxidant pathways. Activation of potassium ion conductance can also contribute as it results in inhibition of the release of glutamate [25, 26]. AAE may therefore have potential in the management of *status epilepticus*.

Kainic acid is a neuro excitotoxic analogue of glutamate used in studies of epilepsy to model experimentally induced limbic seizures [27, 28]. Kainate-treated rats may respond differently. Some may produce wet dog shakes (equivalent to a class III seizure on the Racine scale) or more severe seizures [29]. Previous studies have shown



pattern of neurodegeneration in the hippocampus with high concentration of high affinity KA binding sites (CA3 pyramidal cells of the hippocampus) [7, 30]. The dentate gyrus from kainate-treated rats has shown the presence of mossy fibre sprouting in the inner molecular layer [7, 31]. Examination of the hippocampus after seizures revealed hippocampal damage, especially in the CA3 and CA2 regions as shown in the photomicrographs. The extract showed no significant protection against such damage even though it significantly delayed the latency

to wet dog shakes. This implies that the extract possesses general anticonvulsant properties but offers no protection against morphological changes. The kainate-treated rat model is used to study temporal lobe epilepsy. However, similarity of seizure occurrence to human temporal lobe epilepsy has not been studied comprehensively. But there are several characteristics of the seizures that resemble temporal lobe epilepsy in humans. For instance, some of the animals produce a few observed motor seizures (even with several months of observation) after a

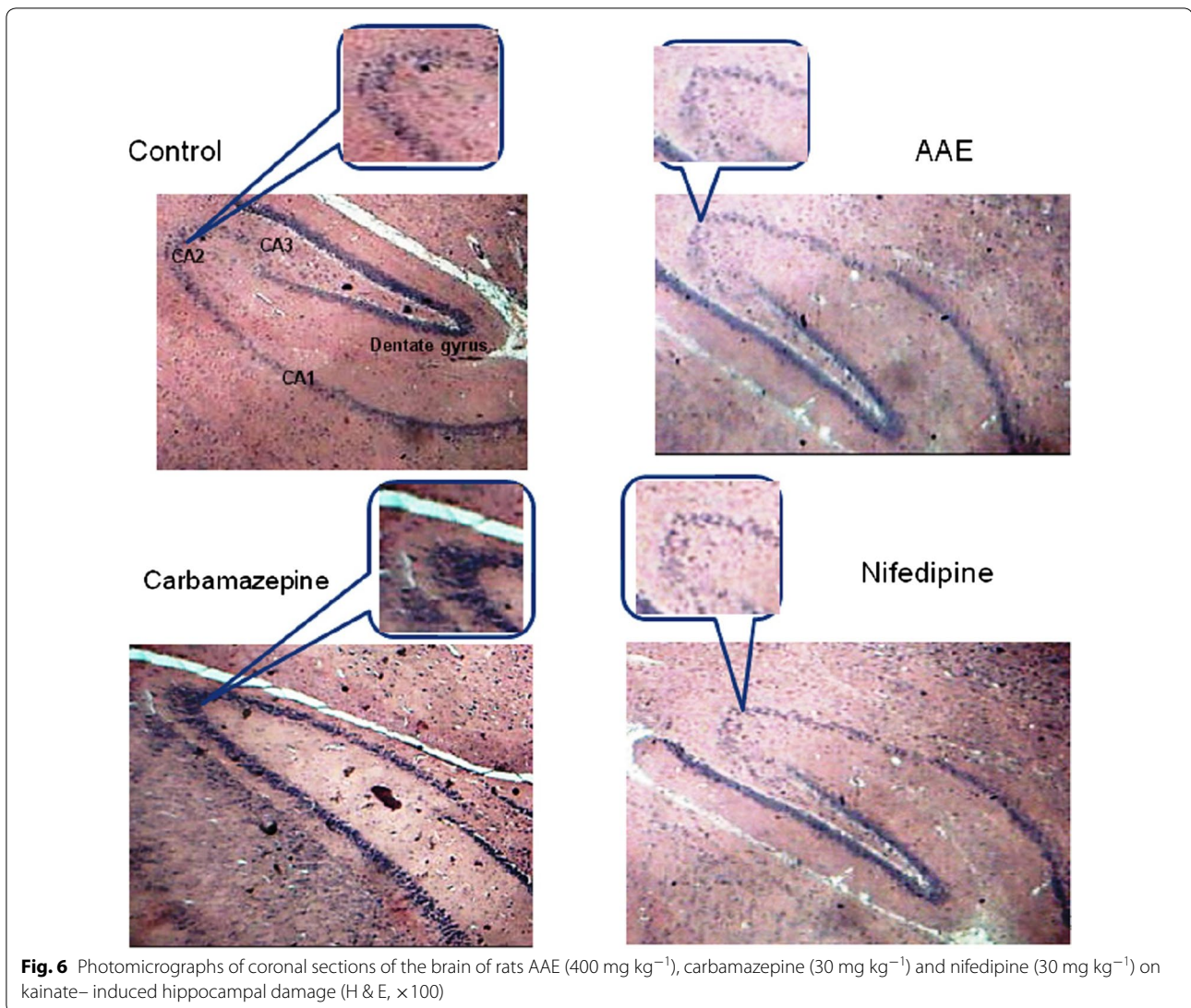


Fig. 6 Photomicrographs of coronal sections of the brain of rats AAE (400 mg kg^{-1}), carbamazepine (30 mg kg^{-1}) and nifedipine (30 mg kg^{-1}) on kainate-induced hippocampal damage (H & E, $\times 100$)

latent period, while other animals have seizures at a frequency as high as 1–2 hz which is reminiscent of epilepsy in the human population [8]. The rats often demonstrate confusion (e.g. hyperactive exploration of their cage) after a seizure, resembling the post-ictal confusion in most humans with temporal lobe epilepsy [32, 33]. Rats treated with the extract exhibited fewer seizures as compared to the control implying a possibility that it might be effective in the treatment of temporal lobe epilepsy. The CA1 and CA3 regions of the hippocampus are also known to possess one of the highest densities of the dihydropyridine receptors in the rat brain [34, 35]. The results of many experimental studies have shown that calcium channel blockers are effective against several different types of seizures [35–37]. Hence, nifedipine proved its effect in this model. As much as these experiments

model human temporal lobe epilepsy, caution has to be exercised in translating the results directly to man without the needed clinical trials. The results however further lend scientific credence to the traditional use of *A. toxicaria* as an antiepileptic.

Conclusion

Antiaris toxicaria possesses anticonvulsant properties in kindling and status epilepticus murine models and may be antiepileptogenic and a candidate for the management of temporal lobe epilepsy (Additional file 1).

Additional file

[Additional file 1.](#) Raw data.

Authors' contributions

PKM: Was involved in the conception and design, acquisition of all data as well as the analysis and interpretation of data. She was also involved in drafting and revising the manuscript. DWA: Was involved in the acquisition of data, analysis and interpretation of data in addition to drafting and revising the manuscript. EW: Was involved in the conception and design, analysis and interpretation of data as well as drafting and revising the manuscript. All the authors read and approved the final manuscript.

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Not applicable.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data generated or analysed during this study are included in this published article as supplementary information files.

Ethics approval and consent to participate

All animals were handled in accordance the Guide for the Care and Use of Laboratory Animals [11] and experiments were approved by the Faculty of Pharmacy and Pharmaceutical Sciences Ethics Committee, KNUST.

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References

- Meldrum BS. Antiepileptic drugs potentiating GABA. *Electroencephalogr Clin Neurophysiol Suppl.* 1999;50:450–7.
- Morimoto K, Fahnestock M, Racine RJ. Kindling and status epilepticus models of epilepsy: rewiring the brain. *Prog Neurobiol.* 2004;73(1):1–60.
- Corda MG, Giorgi O, Orlandi M, Longoni B, Biggio G. Chronic administration of negative modulators produces chemical kindling and GABAA receptor down-regulation. *Adv Biochem Psychopharmacol.* 1990;46:153–66.
- Dhir A. Pentylenetetrazol (PTZ) kindling model of epilepsy. *Curr Protoc Neurosci.* 2012. doi:10.1002/0471142301.ns0937s58.
- Turski WA, Cavalheiro EA, Schwarz M, Czuczwar SJ, Kleinrok Z, Turski L. Limbic seizures produced by pilocarpine in rats: behavioural, electroencephalographic and neuropathological study. *Behav Brain Res.* 1983;9(3):315–35.
- Lopes MW, Lopes SC, Costa AP, Gonçalves FM, Rieger DK, Peres TV, et al. Region-specific alterations of AMPA receptor phosphorylation and signaling pathways in the pilocarpine model of epilepsy. *Neurochem Int.* 2015;87:22–33.
- Levesque M, Avoli M. The kainic acid model of temporal lobe epilepsy. *Neurosci Biobehav Rev.* 2013;37(10 Pt 2):2887–99.
- French J, Williamson P, Thadani V, Darcey T, Mattson R, Spencer S, et al. Characteristics of medial temporal lobe epilepsy: i. results of history and physical examination. *Ann Neurol.* 1993;34(6):774–80.
- Mshana RN, Abbiw DK, Addae-Mensah I, Adjanouhou E, Ahyi MRA, Ekpere JA, et al. Traditional medicine and pharmacopoeia; Contribution to the revision of ethnobotanical and floristic studies in Ghana. *Organization of African Unity/Scientific, Technical & Research Commission*; 2000.
- Mante PK, Adongo DW, Woode E, Kukuia KK, Ameyaw EO. Anticonvulsant effect of *Antiaris toxicaria* (Pers.) Lesch. (Moraceae) aqueous extract in rodents. *ISRN Pharmacol.* 2013;2013:9.
- National Research Council. *Guide for the care and use of laboratory animals.* Washington D.C.: The National Academies Press; 1996.
- Cilio M, Bolanos A, Liu Z, Schmid R, Yang Y, Stafstrom C, et al. Anticonvulsant action and long-term effects of gabapentin in the immature brain. *Neuropharmacology.* 2001;40(1):139–47.
- Bittencourt S, Dubiela FP, Queiroz C, Covolan L, Andrade D, Lozano A, et al. Microinjection of GABAergic agents into the anterior nucleus of the thalamus modulates pilocarpine-induced seizures and status epilepticus. *Seizure.* 2010;19(4):242–6.
- Coyle JT, Puttfarcken P. Oxidative stress, glutamate, and neurodegenerative disorders. *Science.* 1993;262(5134):689–95.
- Halliwell B. Reactive oxygen species and the central nervous system. *J Neurochem.* 1992;59(5):1609–23.
- Halliwell B. Oxidative stress and neurodegeneration: where are we now? *J Neurochem.* 2006;97(6):1634–58.
- Nowak JZ. Oxidative stress, polyunsaturated fatty acids derived oxidation products and bisretinoids as potential inducers of CNS diseases: focus on age-related macular degeneration. *Pharmacol Rep.* 2013;65(2):288–304.
- Turski L, Ikonomidou C, Turski WA, Bortolotto ZA, Cavalheiro EA. Review: cholinergic mechanisms and epileptogenesis. The seizures induced by pilocarpine: a novel experimental model of intractable epilepsy. *Synapse.* 1989;3(2):154–71.
- Clifford DB, Olney JW, Maniotis A, Collins RC, Zorumski CF. The functional anatomy and pathology of lithium-pilocarpine and high-dose pilocarpine seizures. *Neuroscience.* 1987;23(3):953–68.
- Gao F, Liu Y, Li X, Wang Y, Wei D, Jiang W. Fingolimod (FTY720) inhibits neuroinflammation and attenuates spontaneous convulsions in lithium-pilocarpine induced status epilepticus in rat model. *Pharmacol Biochem Behav.* 2012;103(2):187–96.
- Freitas RM, Sousa FC, Vasconcelos SM, Viana GS, Fonteles MM. Pilocarpine-induced status epilepticus in rats: lipid peroxidation level, nitrite formation, GABAergic and glutamatergic receptor alterations in the hippocampus, striatum and frontal cortex. *Pharmacol Biochem Behav.* 2004;78(2):327–32.
- Xavier SM, Barbosa CO, Barros DO, Silva RF, Oliveira AA, Freitas RM. Vitamin C antioxidant effects in hippocampus of adult Wistar rats after seizures and status epilepticus induced by pilocarpine. *Neurosci Lett.* 2007;420(1):76–9.
- Tejada S, Sureda A, Roca C, Gamundi A, Esteban S. Antioxidant response and oxidative damage in brain cortex after high dose of pilocarpine. *Brain Res Bull.* 2007;71(4):372–5.
- Cavalheiro EA, Naffah-Mazzacoratti MG, Mello LE, Leite JP. The pilocarpine model of seizures. *Models of seizures and epilepsy.* New York: Elsevier; 2006. p. 433–448.
- Morales-Villagrán A, Tapia R. Preferential stimulation of glutamate release by 4-aminopyridine in rat striatum in vivo. *Neurochem Int.* 1996;28(1):35–40.
- Maljevic S, Lerche H. Potassium channels: a review of broadening therapeutic possibilities for neurological diseases. *J Neurol.* 2013;260(9):2201–11.
- Ben-Ari Y, Cossart R. Kainate, a double agent that generates seizures: two decades of progress. *Trends Neurosci.* 2000;23(11):580–7.
- Biziere K, Slevin J, Zaczek R, Collins J. Kainic acid neurotoxicity and receptor in CNS pharmacology neuropeptides. In: *Proceedings of the 8th international congress of pharmacology*, Tokyo, 1981. Elsevier; 2013.
- Hellier JL, Dudek FE. Chemoconvulsant model of chronic spontaneous seizures. *Curr Protoc Neurosci.* Chapter 9: Unit 9.19.
- Fisher RS. Animal models of the epilepsies. *Brain Res Rev.* 1989;14(3):245–78.
- Buckmaster PS, Dudek FE. Neuron loss, granule cell axon reorganization, and functional changes in the dentate gyrus of epileptic kainate-treated rats. *J Comp Neurol.* 1997;385(3):385–404.
- Fenwick P. *Psychiatric disorders and epilepsy.* In: *Epilepsy*, Hopkins A, Shorvon S, Cascino G, editors. London: Chapman and Hall Medical; 1995. p. 453–502.
- Liu A, Bryant A, Jefferson A, Friedman D, Minhas P, Barnard S, et al. Exploring the efficacy of a 5-day course of transcranial direct current stimulation (TDCS) on depression and memory function in patients with well-controlled temporal lobe epilepsy. *Epilepsy Behav.* 2016;55:11–20.

34. Meyer JH, Gruol DL. Dehydroepiandrosterone sulfate alters synaptic potentials in area CA1 of the hippocampal slice. *Brain Res.* 1994;633(1):253–61.
35. Koskimäki J, Matsui N, Umemori J, Rantamäki T, Castrén E. Nimodipine activates TrkB neurotrophin receptors and induces neuroplastic and neuroprotective signaling events in the mouse hippocampus and prefrontal cortex. *Cell Mol Neurobiol.* 2015;35(2):189–96.
36. van Lujtelaar G, Wiaderna D, Elants C, Scheenen W. Opposite effects of T- and L-type Ca²⁺ channels blockers in generalized absence epilepsy. *Eur J Pharmacol.* 2000;406(3):381–9.
37. Kriz J, Župan G, Simonić A. Differential effects of dihydropyridine calcium channel blockers in kainic acid-induced experimental seizures in rats. *Epilepsy Res.* 2003;52(3):215–25.

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