



Article Physical Properties, Chemical Analysis, and Evaluation of Antimicrobial Response of New Polylactide/Alginate/Copper Composite Materials

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Received: 8 November 2020; Accepted: 17 December 2020; Published: 21 December 2020



Abstract: In recent years, due to an expansion of antibiotic-resistant microorganisms, there has been growing interest in biodegradable and antibacterial polymers that can be used in selected biomedical applications. The present work describes the synthesis of antimicrobial polylactide-copper alginate (PLA-ALG- Cu^{2+}) composite fibers and their characterization. The composites were prepared by immersing PLA fibers in aqueous solution of sodium alginate, followed by ionic cross-linking of alginate chains within the polylactide fibers with Cu(II) ions to yield PLA–ALG–Cu²⁺ composite fibers. The composites, so prepared, were characterized by scanning electron microscopy (SEM), UV/VIS transmittance and attenuated total reflection Fourier-transform infrared spectroscopy ATR-FTIR, and by determination of their specific surface area (SSA), total/average pore volumes (through application of the 5-point Brunauer-Emmett-Teller method (BET)), and ability to block UV radiation (determination of the ultraviolet protection factor (UPF) of samples). The composites were also subjected to in vitro antimicrobial activity evaluation tests against colonies of Gram-negative (E. coli) and Gram-positive (S. aureus) bacteria and antifungal susceptibility tests against Aspergillus niger and Chaetomium globosum fungal mold species. All the results obtained in this work showed that the obtained composites were promising materials to be used as an antimicrobial wound dressing.

Keywords: alginic acid; poly(lactide); copper; polymers; nonwoven fabric; melt-blown; composite; biodegradable; antibacterial activity

1. Introduction

Poly(lactic acid) (PLA) presents a polyester-type polymer [1,2], which due to its physicochemical properties and technical parameters [3,4], biodegradability and bioavailability [3,5–11] is dedicated to biomedical applications [12–20] These pro-medical attributes of PLA can be utilized in the form of its antibacterial composites, prepared on the PLA surface-modified matrix with antimicrobial additivities [3,8]. Copper and its salts play a special role in inorganic antimicrobials; there were nearly 5300 documents on antibacterial copper abstracted by Scopus [21]; it is vitally essential for many biological processes [22,23] and is a cheap and antibacterially-efficient inorganic [23].

Since the effective antibacterial composite should exhibit prolonged antibacterial activity, stable surface deposition/attachment of copper to polymer presents the major problem. Due to low affinity of metallic cations to carboxylic ester bonds [23], PLA weakly binds copper ions, and its antibacterial PLA-Cu composites require an interface covering layer with high affinity to copper.

Such requirements are fulfilled by alginates, marine-originated [24], biodegradable [25–29] biopolymers applied as a basis for drug delivery, wound dressings, and tissue engineering [30–34].

The strong affinity of alginates to metal cations allows for their application as antibacterial hybrids (e.g., [35–44]), and also for the removal of heavy metal salts, e.g., copper, lead, and mercury (e.g., [45–50]). The role of alginate in antibacterial finishing of textiles has been reviewed recently by Li et al. [51].

We propose an application of alginate film covering the PLA matrix (PLA–ALG), which after addition of copper salts underwent cross-linking with formation of an outer-space coating, with strongly-fixed copper ions (Figure 1). Such PLA–ALG–Cu²⁺ nonwoven composites slowly release of copper ions providing long-termed antibacterial activity.

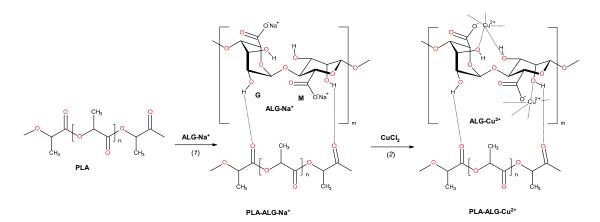


Figure 1. The reactions involved in the preparation of fibrous composite: PLA \rightarrow PLA–ALG–Na⁺ \rightarrow PLA–ALG–Cu²⁺. The structure of alginate is presented as a linear copolymer –[GM]_n– with homopolymeric blocks of (1–4)-linked β -D-mannuronate (M) and its C-5 epimer α -L-guluronate (G) residues.

Is worth noting that cellulose-based fibers [52–54] and also wool fabric functionalized with copper alginate were reported to exhibit antibacterial properties [55]. However, in spite of the huge number reports on alginate abstracted by Scopus (12,643 reports) [56], or those on alginate composites (3450 reports) [57] or alginate hybrids (1237 reports) [58], the application of PLA–ALG hybrids has been reported in only a few papers, e.g., as a tissue engineering material [59], as composite microcapsules for a single-shot vaccine [60], and as bio-polymer carriers which can be implanted in subcutaneous tissue for continuous monitoring of glucose [61].

As part of our investigations focused on biologically-active phosphonates [62–64] and fibrous materials functionalization [65–70] we present the preparation and biological and physico-chemical properties of polylactide/alginate/copper composite materials.

2. Results and Discussion

2.1. Preparation of PLA–ALG– Cu^{2+} Composites

Sodium alginate with abundant carboxylate and hydroxyl groups [71], reacts with divalent cations such as Cu(II) [72], and others [73–75], to form cross-linked hydrogels, existing in reticular structures called "egg box" structures [76,77]. In transition metal-alginate systems, the sol-gel transition is characterized by a complex formation in which only the carboxyl groups in both M and G residues are coordinated to the metal ions [78]. In these structures, ALG-metal metal ions are bonded so strongly that alginates are used as metal sorbents for removal from environmental wastes [77,79–82]. Therefore, ALG–Cu²⁺ complexes, and subsequently PLA–ALG–Cu²⁺ composites are stable with a prolonged application period.

The preparation of PLA–ALG–Cu²⁺ composites wase performed by treating of PLA nonwovens with aqueous solutions of sodium alginate, during which alginate underwent adhesion to the PLA surface. In the second stage of the procedure, solution of copper chloride was added and consequently copper ions substituted sodium ions, and in turn initiated a cross-linking reaction with subsequent sol

transition. The reactions involved in the fibrous composite preparation are depicted schematically in Figure 1.

2.2. Scanning Electron Microscopy

SEM micrographs of polylactide nonwoven (PLA), polylactide/sodium alginate (PLA–ALG–Na⁺), and PLA–ALG–Cu²⁺-2 composites are presented in Figures 2–4, respectively.

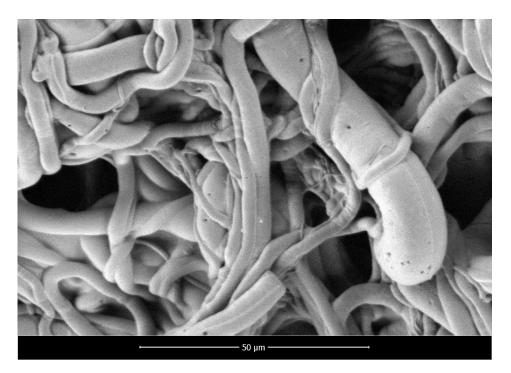


Figure 2. Scanning electron microscopy images of PLA nonwoven, magnification: 2000×.



Figure 3. Scanning electron microscopy images of PLA–ALG–Na⁺, magnification: 2000×.

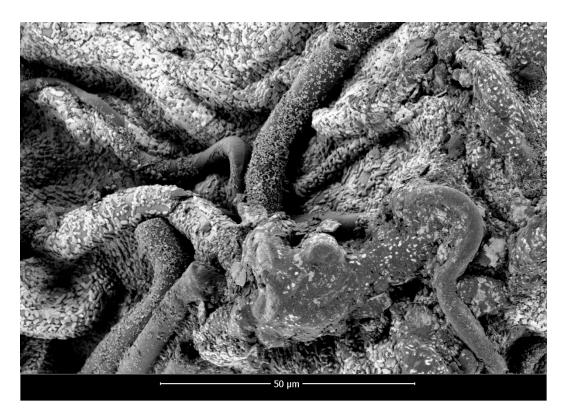


Figure 4. Scanning electron microscopy images of PLA–ALG–Cu²⁺-2, magnification: 2000×.

The presented SEM images illustrate the changes in the surface morphology of the investigated samples occurring due to the modification of PLA nonwovens with the solution of alginic acid sodium salt and CuCl₂. The SEM image of unmodified PLA nonwoven presents a mesh of randomly-oriented fibers, with interconnected pores and relatively-smooth surface (Figure 2). The diameters of the fibers vary significantly from approximately 0.7 μ m up to 14.0 μ m. Surface modification of PLA nonwovens with the alginate resulted in the occurrence of the coating on the surface of the randomly-oriented fibers (Figure 3). As a result, fewer pores are visible. In addition, the coating covers thinner fibers and only fibers with a diameter greater than ~3.5 μ m can be distinguished. Furthermore, the agglomerates of alginate may be noticed on the surface of the fibrous composite. Due to that, the surface of PLA–ALG–Na⁺ fibers is more coarse in comparison to unmodified PLA fibers.

Figure 4 illustrates the surface of the PLA–ALG–Cu²⁺-2 composite (charged with ~7.6% of Cu²⁺), with the agglomerates, presumably composed of ALG–Cu²⁺, visible over the entire surface of the sample. Similarly as in the case of the PLA–ALG–Na⁺ sample, only the fibers with the diameter higher than ~4. μ m are distinguishable and the majority of pores are covered by the coating. Moreover, the surface of the PLA–ALG–Cu²⁺ composite is less uniform and rougher due to the presence of numerous agglomerates.

Figures 5–7 show the exemplary EDS spectra of PLA–ALG–Na⁺ (Figure 5), PLA–ALG–Cu²⁺-1 (Cu: 1.4%) (Figure 6) and PLA–ALG–Cu²⁺-2 (Cu: 7.6%) (Figure 7) composites (the EDS data are presented as a graph with energy (keV) on the *x*–axis and peak intensity on the *y*–axis). Table 1 presents the chemical composition of the investigated samples obtained from the quantitative analysis of EDS results. The presented values of mean concentration of each element are calculated from six to eight spot measurements, on two different samples.

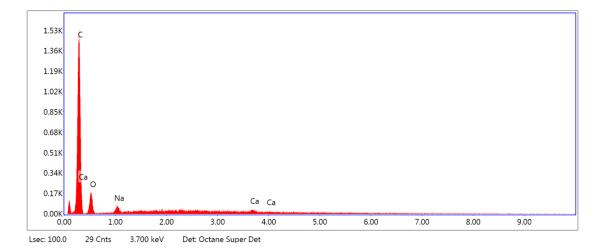


Figure 5. Example of the energy-dispersive X-ray Spectroscopy (EDS) spectrum of PLA–ALG–Na⁺.

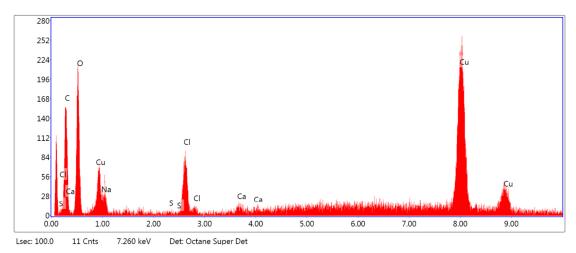


Figure 6. Example of the EDS spectrum of the PLA–ALG–Cu²⁺-1.

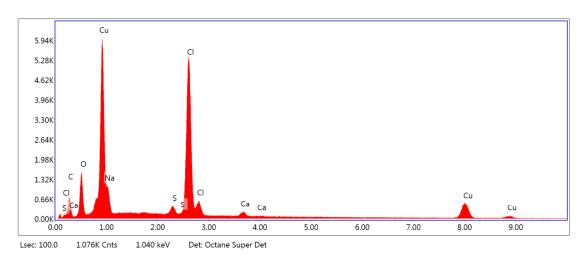


Figure 7. Example of the EDS spectrum of PLA–ALG–Cu²⁺-2.

	PLA						
Atom	С	0					
At. %	51.70	48.33	-				
Std. dev.	0.11	0.11					
	P	LA-ALG-Na	a ⁺				
Atom	С	0	Na	Ca			
At. %	71.41	28.42	0.10	0.08			
Std. dev.	9.74	9.89	0.14	0.17			
			PLA-ALC	G-Cu ²⁺ -1			
Atom	С	0	Cu	Na	S	C1	Ca
At. %	55.26	40.95	0.92	0.78	0.02	1.93	0.15
Std. dev.	1.26	2.59	1.26	1.10	0.04	0.65	0.21
			PLA-ALO	G–Cu ²⁺ -2			
Atom	С	0	Cu	Na	S	Cl	Ca
At. %	35.16	19.96	15.06	5.87	1.18	21.44	1.30
Std. dev.	8.66	2.67	1.53	0.89	0.59	5.23	0.75

Table 1. Quantitative results of EDS analysis of PLA, PLA–ALG–Na⁺, PLA–ALG–Cu²⁺-1 and PLA–ALG–Cu²⁺-2.

As can be observed from Figure 5, the PLA–ALG–Na⁺ sample was composed primarily from carbon, oxygen, and sodium. This is consistent with the chemical composition of PLA, which is built of carbon, oxygen, and hydrogen. The occurrence of a sodium peak may be attributed to the presence of alginate, which apart from carbon, oxygen, and hydrogen also contains sodium. In the case of the PLA–ALG–Cu²⁺-1 and PLA–ALG–Cu²⁺-2 composites, additional peaks associated with copper and chloride were detected. This confirms the presence of NaCl (ALG–Na⁺ + Cu²⁺ + 2Cl⁻ \rightarrow ALG–Cu²⁺ + 2Na⁺ + 2Cl⁻) in those samples. Apart from that, the calcium peak was observed for all the samples. Moreover, in the case of fibrous composites containing NaCl a peak related to sulfur was also noticed. The relative intensity of the sulfur peak increased with the NaCl concentration in the surface modifier.

Quantitative EDS analysis (Table 1) exhibited that the modification of PLA with alginate led to an increase in carbon concentration (from 51.70 at. % to 71.41 at. %). Simultaneously, a decrease in the oxygen content (from 48.33 at. % to 28.42 at. %) was observed. Additionally, for the PLA-ALG-Na⁺ composite a small concentration of sodium was detected (0.10 at. %). Furthermore, the obtained results indicate that the content of Cu and Cl significantly increased when the concentration of CuCl₂ in the surface modifier changed from 5% to 10%. In the case of the PLA–ALG–Cu²⁺-1 sample, the content of copper was 0.92 at. %, while the chloride concentration was 1.93 at. % (Figure 5). For the PLA-ALG-Cu²⁺-2 composite the concentrations of copper and chloride rose to 15.06 at. % and 21.44 at. %, respectively (Figure 6). At the same time, with the increase of $CuCl_2$ concentration from 5% to 10%, the carbon content dropped to 55.25 at. % and 35.16 at. %, while sodium content increased to 0.78 at. % and 5.87 at. %, accordingly. In the case of oxygen concentration, at first an increase of up to 40.95 at. % was observed for the PLA–ALG–Cu²⁺-1 sample, followed by a decrease to 19.96 at. % for the PLA-ALG-Cu²⁺-2 composite. As far as calcium and sulfur contents were concerned, both rose with the increase in the CuCl₂ concentration, however, the observed values did not exceed 1.5 at. % and thus, may be regarded as a contamination. The observed high standard deviation values indicates that the distribution of chemical components was not uniform and homogeneous.

2.3. FAAS

The copper content in the PLA–ALG–Cu²⁺ composites was assessed by the flame atomic absorption spectrometry (FAAS) method, after prior composite degradation, as shown in Figure 8. The results are given in Table 2 [83].

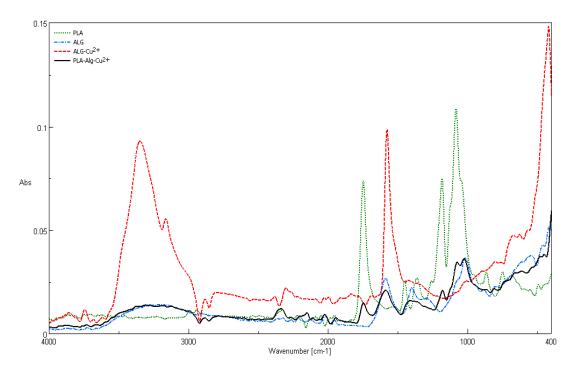


Figure 8. Comparison of attenuated total reflection Fourier–Transform infrared spectroscopy ATR–FTIR spectra of PLA nonwovens, alginic acid sodium salt (ALG–Na⁺), ALG–Cu²⁺ complex, and PLA–ALG–Cu²⁺-2 composite.

Cu Concentration (g/kg)
0.004
0.003
13.56
73.91

Table 2. Copper concentration in polylactide/alginate/copper composite materials samples.

The results have been measured in triplicate and presented as a mean value.

The results of FAAS analysis showed that copper content in the fibrous composites samples depends on the concentation of water solution of copper(II) chloride in the second step of a dip-coating modification. The higher coating concentration of the copper(II) chloride modifier (10%) gave the higher content of the Cu^{2+} on PLA sample (73.91 g/kg).

2.4. ATR-FTIR Spectra

The recorded ATR–FTIR spectra for polylactide nonwoven, alginic acid sodium salt (ALG–Na⁺), ALG–Cu²⁺ complex, and PLA–ALG–Cu²⁺-2 composite are presented in Figure 8. Characteristic FTIR signals of the composite and its components (PLA, ALG–Na⁺, and ALG–Cu²) are summarized in Table 3 [84–92].

Table 3. The Characteristic FTIR band frequencies determined for polylactide nonwoven, alginic acid
sodium salt (ALG–Na ⁺), ALG–Cu ²⁺ complex, and PLA–ALG–Cu ²⁺ -2 composite [84–92].

					1 .					
	PLA				ALG-Na ⁺		ALG-0	Cu ²⁺	PLA-ALC	G–Cu ²
(v/cm ⁻¹)	Int. ^{/a}	Assign. [84]	(v/cm ⁻¹)	Int. ^{/a}	Lit. bands (v/cm ⁻¹) [85–91]	Assign. /[Ref.][88]	(v/cm ⁻¹)	Int. ^{/a}	(v/cm ⁻¹)	Int./a
					3350 ± 350	ν_s O–H	3750	0.013 ^{/c}		
							3345	0.09 ^{/c}		
			3200	0.14 ^{/b}		ν _s Ο–Η			3200	0.01
							3164	0.05		
2997	0.01	$\nu_{as}CH_3$								
2947	0.01	ν_sCH_3								
					2926 ± 1	ν_s C–H			2890	0.01
			2897	0.01			2886	0.02	2886	0.01
							2389	0.02		
									2341	0.01
			2322	0.01			2300	0.02	2337	0.01
									2265	0.01
			2216	0.01					2270 2205	0.01
			2134	0.01					2203	0.00
			2134	0.01					2132	0.01
			2021	0.01					2025	0.01
			1957	0.01					1946	0.01
							1833	0.02		
1760	0.07	ν C=O							1755	0.01
					1650 [86]	ν_{as} COO ⁻	1690	0.02		
					1614 [85,86]	ν_{as} COO ⁻				
			1588	0.03			1581	0.09	1588	0.02
1452	0.02	$\delta_{as}CH_3$								
					1417 [85,86]	δ C–H; ν _s COO ⁻			1411	0.02
			1400	0.02						
1388-1368	0.02	$\delta_s CH_3$								
1368-1360	0.02	$\overset{\delta_1}{CH+\delta_sCH_3}$								
1270	0.02	δCH+νCOC								
			1295	0.02	1301 (ms)	δ C-H				
1130	0.07	r _{as} CH3			1124 (s) [86]	ν C–O, ν C–C, δ C–C–C,			1178	0.02
						$\nu_{as}COC^{/d}$				
1100-1090	0.11	$\nu_{s}COC$			1096 (s) [86]	ν C-O, ν C-C, δ C-C-O;			1073	0.03
1045	0.08	ν C-CH3			1034 (vs) [86]	ν _{as} C–O–C ^{/d} , ν C–O ^{/d} ν C–C				
			1017	0.04			1013	0.03	1020	0.03
875-860	0.03	νC-COO							866	0.02
					826 (ms) [86]	δ C-O-C ^{/d} ,δ C-C-C, δ C-C-O ^{/d} δ C-C-H, δω O-H	810	0.03	810	0.02
760–740	0.03	δC=Ο			776 (w)	rb, δ C-C-H, δ C-C-O				
715-695	0.03	γC=O			703(ms)	rb				
			555	0.04		δω Ο–Η			555	0.03

^a Band assignment: v, stretching vibration; δ , deformation; sh, shoulder; s, symmetric; as, asymmetric; r, rocking; γ , out-of-plane bending mode; ω , wagging; rb, ring breathing. ^b Band intensity: The bands with absorbance values ≥ 0.005 are listed, and subsequently approximated to a second decimal place. In the literature data: s, strong; vs, very strong; ms, medium strong; w, weak. ^c Derived from water hydrogen–bonded O–H stretching vibrations [92]. ^d Glycosidic linkage.

The infrared vibrations spectrum of PLA–ALG–Cu²⁺ composite contains the major bands derived from the composite components, namely PLA (1760 cm⁻¹ (ν C=O), 1073 cm⁻¹ (ν _s C–O–C), and 866 cm⁻¹ (ν C–CO₂⁻)), ALG–Na⁺ /ALG–Cu₂+ (3200 cm⁻¹ (ν _s O–H), 2890 cm⁻¹ (ν _s C–H), 2337 cm⁻¹, 2152 cm⁻¹, 1158 cm⁻¹, 1417 cm⁻¹ (δ C–H, ν C–CO₂⁻), 1073 cm⁻¹, 1020 cm⁻¹, 801 cm⁻¹, and 555 cm⁻¹ (δ_{ω} O–H)).

Some bands characteristic for PLA or ALG–Na⁺ disappeared, due to an overlapping effect or a band broadening caused by influence of cupric ion, for instance in PLA spectrum (2997 cm⁻¹ (ν_{as} CH₃), 2947 cm⁻¹ (ν_{s} CH₃), 1452 cm⁻¹ (δ_{as} CH₃), 1388–1368 cm⁻¹ (δ CH⁺, δ_{s} CH₃), 1270 cm⁻¹ (δ CH⁺, ν C–O–C), 1045 cm⁻¹ (ν C–CH₃), 875–860 cm⁻¹ (ν C–CO₂⁻), 760–740 cm⁻¹ (δ C=O), and 715–695 cm⁻¹ (γ C=O)).

2.5. Specific Surface Area, Total Pore Volume, and Average Pore Diameter Measurement

The specific surface area, total pore volume, and average pore diameter of the PLA–ALG–Cu²⁺ composites samples are presented in Table 4.

Sample Name	Specific Surface Area S _{BET} (m²/g)	Total Pore Volume V (cm ³ /g)	Average Pore Diameter D (nm)
PLA	0.2405 ± 0.0220	$9.084 \cdot 10^{-4}$	13.85
PLA-ALG-Na ⁺	0.5548 ± 0.0452	$1.581 \cdot 10^{-3}$	10.54
PLA-ALG-Cu ²⁺ -1	0.8429 ± 0.0109	$3.450 \cdot 10^{-3}$	16.59
PLA-ALG-Cu ²⁺ -2	1.4280 ± 0.0220	$4.661 \cdot 10^{-3}$	13.26

Table 4. Specific surface area, total pore volume, and average pore diameter of the examined samples.

The specific surface area of the unmodified PLA nonwoven was equal to 0.2405 m²/g. The modification of the PLA nonwoven with the solution of alginic acid sodium salt resulted in ~130% increase in the value of specific surface area (up to 0.5548 m²/g). At the same time, the addition of 5% and 10% of CuCl₂ caused even further growth of the specific surface area up to 0.8429 and 1.4280 m²/g, respectively. When compared to the PLA–ALG–Na⁺ sample, the specific surface area of PLA–ALG–Cu²⁺-1 sample rose by ~150%, while in the case of the PLA–ALG–Cu²⁺-2 sample an ~260% increase in the specific surface area value was observed. Thus, it can be concluded that the modification of PLA nonwovens with alginate and CuCl₂ leads to a significant growth of the specific surface area.

The increase in the specific surface area observed for the modified nonwovens may be associated with the higher mesoporosity as indicated by the larger hysteresis loops for the adsorption–desorption isotherms of modified samples. This is confirmed by the total pore volume, which significantly rose from $9.084 \cdot 10^{-4}$ cm³/g for the PLA nonwoven up to $1.581 \cdot 10^{-3}$ cm³/g (170%), $3.450 \cdot 10^{-3}$ cm³/g (380%) and $4.661 \cdot 10^{-3}$ cm³/g (510%) for the PLA–ALG–Na⁺, PLA–ALG–Cu²⁺-1, and PLA–ALG–Cu²⁺-2 samples, respectively. It is worth highlighting, that despite the observed covering of pores visible in the SEM images of the modified samples, the mesoporosity of those samples was greater.

At the same time, there was no correlation between the specific surface area or total pore volume and the average pore diameter. The estimated average pore diameter of the investigated samples varied in the range of 10.54–16.59 nm and did not explicitly depend on the modification and CuCl₂ concentration.

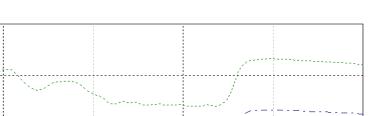
2.6. UV-VIS Analysis and Determination of the Protective Properties against UV Radiation

Figure 9 presents spectrophotometric transmittance spectrum in the wavelength $\lambda = 200-800$ nm of PLA nonwoven and PLA-ALG-Na⁺ and PLA-ALG-Cu²⁺ (PLA-ALG-Cu²⁺-1 and PLA-ALG-Cu²⁺-2) composites.

PLA PLA-ALG PLA-ALG-Cu²⁺⁻¹ PLA-ALG-Cu²⁺⁻²

25

20



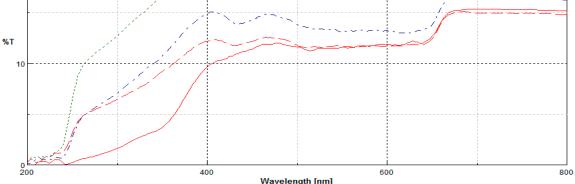


Figure 9. Comparison of transmittance spectra (%T) of PLA nonwoven and PLA–ALG–Na⁺ and PLA–ALG–Cu²⁺ (PLA–ALG–Cu²⁺-1 and PLA–ALG–Cu²⁺-2) composites, recorded in the range: $\lambda = 200-800$ nm.

The transmittance (%T) spectra in the range $\lambda = 200-800$ nm of the PLA and its composites (PLA-ALG-Na⁺ and PLA-ALG-Cu²⁺) revealed changes in their macrostructures in comparison to PLA nonwoven, expressed by a decrease in transmittance in all ranges of measurement. The unmodified (PLA) and modified samples (PLA-ALG-Na⁺, PLA-ALG-Cu²⁺ (PLA-ALG-Cu²⁺-1 and PLA-ALG-Cu²⁺-2) had similar spectral characteristics across the entire spectral range. The reduction in spectral transmission (PLA-ALG-Na⁺, PLA-ALG-Cu²⁺ (PLA-ALG-Cu²⁺-1 and PLA-ALG-Cu²⁺-2) was caused by an additional layer of alginate coating on the surface of the samples. There was a noticeable influence of the Cu content in a modified sample (PLA-ALG-Cu²⁺ (PLA-ALG-Cu²⁺)) on a transmission level, especially in the range of a 250-500 nm-decrease of transmission level in the sample PLA-ALG-Cu²⁺-2 vs. PLA-ALG-Cu²⁺-1.

Table 5 compare average transmittance (T%) and calculated UPF values of modified samples PLA–ALG, PLA–ALG–Cu²⁺ (PLA–ALG–Cu²⁺-1 and PLA–ALG–Cu²⁺-2) with those of non–modified samples (PLA and PLA–ALG–Na⁺).

	PLA	PLA–ALG–Na ⁺	PLA-LG-Cu ²⁺ -1	PLA-ALG-Cu ²⁺ -2
UPF	7.23	12.35	13.97	43.28
average %T, $\lambda = 290-400 \text{ nm}$	16.46	10.63	9.03	4.43

Table 5. UPF values of modified nonwoven fabric samples.

The results have been measured in triplicate and presented as a mean value with ± deviation approximately 6%.

Very good barrier properties against UV radiation were obtained for PLA–ALG–Cu²⁺-2 (UPF = 43.28). This result indicates that the modification performed imparts proper barrier properties against UV radiation according to EN 13758-1:2002 [93].

3. Antimicrobial Properties

3.1. Antibacterial Activity

The polylactide/alginate/copper (PLA–ALG–Cu²⁺) composites were subjected to antimicrobial activity tests against Gram-negative *Escherichia coli* (ATCC11229) and Gram-positive

Staphylococcus aureus (ATCC 6538) bacteria. Results of biological activity tests are listed in Table 6 and illustrated in Figures 10 and 11.

Table 6. Results of tests on the antibacterial activity of polylactide/alginate/copper (PLA–ALG–Cu²⁺) on the basis of standards EN–ISO 20645:2006 [94].

Samula Nama	Bacterial Average Inhibition Zone (mm)			
Sample Name	Escherichia coli	Staphylococcus aureus		
PLA	0	0		
PLA-ALG-Na ⁺	0	0		
PLA-ALG-Cu ²⁺ -1	3	2		
PLA-ALG-Cu ²⁺ -2	3	4		

Concentration of inoculum (bacterial suspension). Amount of live bacteria: *Escherichia coli*: CFU/mL = 1.8×10^8 and *Staphylococcus aureus*: CFU/mL = 1.6×10^8 .

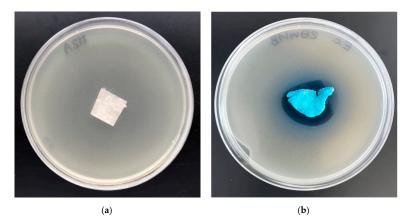


Figure 10. The polylactide/alginate/copper (PLA–ALG– Cu^{2+}) composites' antimicrobial activity tests. Inhibition zones of *Escherichia Coli* bacterial growth on Petri dishes, sample: (**a**) PLA and (**b**) PLA–ALGC u^{2+} –2.

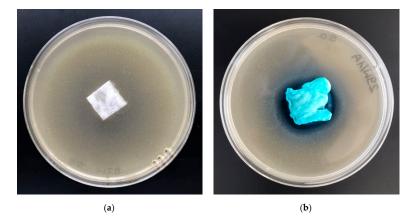


Figure 11. The polylactide/alginate/copper (PLA–ALG– Cu^{2+}) composites' antimicrobial activity tests. Inhibition zones of *Staphylococcus aureus* bacterial growth on Petri dishes, sample: (**a**) PLA and (**b**) PLA–ALG– Cu^{2+} –2.

Results of these studies prove antimicrobial protection against different bacterial microorganisms of functionalized materials (PLA–ALG–Cu²⁺) for *Escherichia coli* and *Staphylococcus aureus* (Table 6), expressed by strong, visible inhibition zones of bacterial growth on Petri dishes (Figures 10b and 11b). A higher ability to inhibit the bacterial growth of *Staphylococcus aureus* was exhibited by the PLA–ALG–Cu²⁺-2 samples. Inhibition zones of bacterial growth of 3–4 mm and no bacterial growth in the samples tested confirmed this observation.

3.2. Antifungal Activity

Results of antifungal activity tests in accordance with the PN EN 14119: 2005 point 10.5 (B2) against a colony of *Aspergillus niger* and *Chaetomium globosum* (ATCC 6205) of PLA fabrics and PLA–ALG–Cu²⁺ composites are illustrated in Figures 12 and 13 and listed in Table 7 [95].

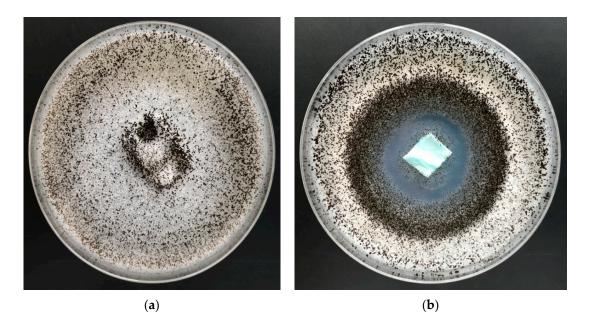


Figure 12. The PLA–ALG– Cu^{2+} composites antimicrobial activity tests against *Aspergillus niger* (ATCC 6275). Inhibition properties of fungal growth on Petri dishes: (a) PLA and (b) PLA–ALG– Cu^{2+} –2.

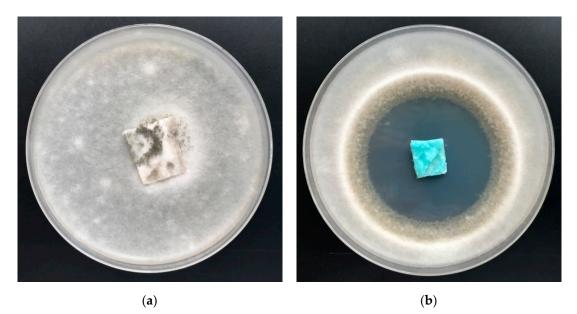


Figure 13. The PLA–ALG– Cu^{2+} composites' antimicrobial activity tests against *Chaetomium globosum*. Inhibition properties of fungal growth on Petri dishes: (a) PLA and (b) PLA–ALG– Cu^{2+} –2.

	Fungal Average Inhibition Zone (mm)					
Sample Name	Aspergillus niger	Chaetomium globosum	Visual Evaluation (Magnification 50×)			
PLA	0	0	Visible growth			
PLA-ALG-Na ⁺	0	0	on sample surface			
PLA-ALG-Cu ²⁺ -1	3	3	No visible growth			
PLA-ALG-Cu ²⁺ -2	3	3	on sample surface			

Table 7. The antifungal activities and growth inhibition effects of PLA–ALG– $Cu^{2+}-1/2$ composites.

Concentration of inoculum, number of fungal spores in 1 mL: *Aspergillus niger*: CFU/mL = 2.9×10^6 , *Chaetomium globosum*: CFU/mL = 2.3×10^6 .

Non–modified samples of polylactide nonwoven (PLA) as well as alginate–modified polylactide nonwoven (PLA–ALG–Na⁺) did not inhibit the growth of *Aspergillus niger* or of *Chaetomium globosum* fungus, either around the samples or in the contact zone with the culture media. In addition, strong fungal growth covering of the entire surface of the samples was visible (Figures 12a and 13a).

Antifungal activity was demonstrated by the PLA–ALG–Cu²⁺ composites samples and the results revealed strong visible inhibition zones of fungal growth on Petri dishes (Figures 12b and 13b), and therefore proved antifungal protection against *Aspergillus niger* and *Chaetomium globosum*.

4. Conclusions

Medicine, especially at present, is focused on the search for new and more effective methods of combating microbials, especially multi-drug-resistant pathogens (viruses, bacteria, and fungi). In recent years there has been growing interest in biodegradable, "eco–friendly" and multifunctional polymers that can be used in selected biomedical applications. This paper presents a method for charging of poly(lactide) nonwoven fabrics with copper salt, and in this way making equipment with antibacterial activity. This was achieved by a two–step procedure of coating of PLA fibers with, prepared in situ, copper alginate (PLA \rightarrow PLA–ALG–Cu²⁺).

The structural properties of these new products were characterized by scanning electron microscopy (SEM), energy-dispersive spectroscopy (EDS), attenuated total reflectance Fourier-transform infrared (ATFR-IR) and ultraviolet-visible spectrophotometry (UV-Vis), and specific surface area, total pore volume and average pore diameter measurement. Copper content in the composites was determined by FAAS.

The results revealed that PLA–ALG– Cu^{2+-2} with ca. 7.4% copper in the composite, presents less uniform and rougher nonwoven in comparison with PLA, with diameter range of 4 to 6 μ m, and a majority of pores covered by the coating (PLA–ALG– Cu^{2+-2} vs. PLA), and spotted with the agglomerates, presumably also ALG– Cu^{2+} .

Specific surface area, total pore volume, and average pore diameter of the examined samples presented ~6-fold increase, ~2-fold decrease and similar average pore diameters for PLA–ALG–Cu²⁺-2 vs. PLA, respectively.

The transmittance (%T) spectra in the range $\lambda = 200-800$ nm of the PLA nonwoven, PLA-ALG-Na⁺ and PLA-ALG-Cu²⁺ composites revealed a decrease in transmittance in all range of measurement and very good barrier properties against UV radiation were obtained for PLA-ALG-Cu²⁺-2 (UPF = 43.28).

The ATR–FTIR spectra of PLA–ALG–Cu²⁺ composites contained the major bands derived from the composite components, namely PLA (1760 cm⁻¹ (ν C=O), 1073 cm⁻¹ (ν s C–O–C) and 866 cm⁻¹ (ν C–CO₂⁻)), ALG–Na⁺/ALG–Cu²⁺ (3200 cm⁻¹ (ν s O–H), 2890 cm-1 (ν s C–H), 2337cm⁻¹, 2152cm⁻¹, 1158cm⁻¹, 1417cm⁻¹ (δ C–H, ν C–CO₂⁻), 1073 cm⁻¹, 1020 cm⁻¹, 801 cm⁻¹, and 555 cm⁻¹ (δ ω O–H)).

The application benefits of the polylactide/alginate/copper materials were confirmed by antimicrobial tests against *Chaetomium globosum*, *Aspergillus niger* fungus species, and representative Gram–negative (*E. coli*) and Gram–positive (*S. aureus*) bacteria.

In summary, PLA–ALG–Cu²⁺ is composed of two easily–biodegradable composites (PLA and ALG) and the entire procedure fulfills the requirements of eco–friendly standards, due to the exhibited technical properties and significant antimicrobial action that can find an application in different fields of medical or healthcare industry (e.g., air filtration, bioadhesives, and tissue engineering). The performed study allows us to state that modification of poly(lactide) nonwoven fabric using alginic acid and copper (II) chloride offers the possibility of additional functions and the creation of new added-value materials. Further studies will focus on the determination of mechanical and cytotoxicity properties of PLA-alginate composites.

5. Materials and Methods

5.1. Materials

- Poly(lactic acid) (PLA) type Ingeo[™] Biopolymer 3251D, in the form of granulates was purchased from NatureWorks LLC (Minnetonka, MN, USA), T_{mp} = 160–170 °C, MFR = 30–40 g/10min (190 °C/2.16 kg) and was used for the forming of samples of nonwoven fabrics;
- Alginic acid sodium salt (CAS Number 9005-38-3, the molecular weight: 120,000–190,000 g/moL, mannuronic acid to guluronic acid–M/G ratio: 1.56) from Millipore Sigma (St. Louis, MO, USA), was used for surface modification of polymer nonwovens;
- Copper(II) chloride, CuCl₂, 97% (CAS Number: 7447-39-4) from Millipore Sigma (St. Louis, MO, USA) was used for surface modification of nonwoven composite;
- Bacterial strains: *Escherichia coli* (ATCC 25922) and *Staphylococcus aureus* (ATCC 6538) were purchased from Microbiologics (St. Cloud, MN, USA).
- Fungal strains: *Aspergillus niger* van Tieghem (ATCC 6275) and *Chaetomium globosum* (ATCC 6205) were purchased from Microbiologics (St. Cloud, MN, USA).

5.2. Methods

5.2.1. Nonwoven Fabrics

Nonwovens were fabricated by the melt-blown technique using a one-screw laboratory extruder (Axon, Limmared, Sweden) with a head with 30 holes of 0.25 mm diameter each, compressed air heater and collecting drum. The melt-blown process conditions applied for preparation of PLA nonwovens are presented in Table 8.

Parameter	
Polymer yields	5 g/min
Mass per unit area of nonwovens	160 g/m^2
Air flow rate	7–8 m ³ /h
Temperature of the extruder: zone 1	195 °C
Temperature of the extruder: zone 2	245 °C
Temperature of the extruder: zone 3	260 °C
Head temperature	260 °C
Air heater temperature	260 °C

Table 8. Melt-blown process conditions applied for preparation of polylactide (PLA) nonwovens.

5.2.2. Dip-Coating Modification

Nonwoven samples ($10 \text{ cm} \times 10 \text{ cm}$; $2.00 \pm 0.05 \text{ g}$) were modified by the dip-coating, two-step method, impregnated in the solution of sodium alginate (step 1) and immersed in the solution of copper(II) chloride (step 2). An aqueous solution of sodium alginate (0.5%, 5 g/L) was homogeneously dispersed and had appropriate viscosity (about 50–70 dPas). An additional two different water solutions of copper(II) chloride was prepared—5 and 10%. The nonwoven fabric samples were impregnated in the polysaccharide solution for 1 min, then the samples were immediately transferred

to one of the two variants of the aqueous copper(II) chloride solution (5 or 10%) and immersed in the solution for 1 min. Then the samples were squeezed and dried for 5 h at 50 °C (to a constant weight of 2.5 ± 0.05 g). After this modification, the formed nonwoven composites, further referred to as PLA–ALG–Cu²⁺-1 (5% CuCl₂) or PLA–ALG–Cu²⁺-2 (10% CuCl₂), respectively, or PLA–ALG–Na⁺ for a composite of PLA and coating paste without CuCl₂, presented a visually–uniform, homogeneous structure. The components of the modifier used and the corresponding modifier abbreviations are given in Table 9.

	Coating Components (%)				
Assignments for Composites and Their Components	Sodium Alginate Solution (ALG–Na ⁺)	Copper(II) Chlorid Solutions (CuCl ₂)			
components	0, 5%	5%	10%		
PLA	_	_	_		
PLA-ALG-Na ⁺	+	_	_		
PLA-ALG-Cu ²⁺ -1	+	+	-		
PLA-ALG-Cu ²⁺ -2	+	-	+		

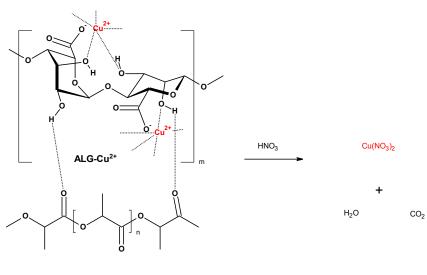
Table 9. Assignments of samples and composition of the coating components of poly(lactide) nonwoven samples surface modifier (%).

5.2.3. Scanning Electron Microscopy/Energy–Dispersive X–ray Spectroscopy (SEM/EDS)

The microscopic analysis of fibers was performed on a Tescan Vega 3 scanning electron microscope (Brno, Czech Republic). Magnification was from 20,000×.

5.2.4. Flame Atomic Absorption Spectrometry (FAAS)

Determination of copper content in PLA–ALG–Cu²⁺ composites was assessed by prior sample mineralization (Figure 14), using single-module Magnum II microwave mineralizer from Ertec (Wroclaw, Poland), as described earlier [70].



PLA-ALG-Cu²⁺

Figure 14. Mineralization of PLA–ALG–Cu²⁺.

Determination of copper (II) ions was performed by atomic absorption spectrometry with flame excitation using Thermo Scientific Thermo Solar M6 (LabWrench, Midland, Canada) spectrometer equipped with a 100-mm titanium burner, coded lamps with a single-element hollow cathode, background correction: D2 deuterium lamp, as described earlier [65].

The total copper content M (mg/kg; ppm) in the PLA–ALG–Cu²⁺ composite sample was calculated according to the Equation (1) [83]:

$$M = \frac{C \cdot V}{m} \tag{1}$$

where:

C—metal concentration in the mineralized PLA–ALG–Cu²⁺ sample solution (mg/L);

m—mass of the mineralized sample of PLA–ALG– Cu^{2+} composites (g);

V—volume of the sample solution (mL).

5.2.5. Attenuated Total Reflection Fourier-Transform Infrared Spectroscopy (ATR-FTIR)

An analysis of the chemical surface structure of the of PLA–ALG– Cu^{2+} fibrous composite samples was performed by ATR–FTIR spectroscopy using a Jasco 4200 spectrometer (Tokyo, Japan) with an Pike Gladi ATR attachment (Cottonwood, AZ, USA) in the range of 400–4000 cm⁻¹.

5.2.6. Specific Surface Area, Total Pore Volume, and Average Pore Diameter Measurement

The specific surface area of the investigated samples was measured using the Autosorb-1 (Quantachrome Instruments, Boynton Beach, FL, USA) apparatus. The analysis was performed using the physisorption method with nitrogen used as a sorption agent. The measurements were carried out at 77 K. For each experiment, about 1–2 g of a given sample was weighed and used. Prior to the analysis, the samples were dried in 105 °C for 24 h and degassed overnight at room temperature.

In order to determine the specific surface area, the 5-point Brunauer–Emmett–Teller (BET) method was applied. The specific surface area was calculated twice for each sample, using the 5-point adsorption isotherm (P/P₀ in the range of 0.10–0.30) and the 39-point adsorption-desorption isotherm. The total pore volume and average pore diameter were determined from the 39-point adsorption-desorption isotherm (P/P₀ in the range of 0.05–1.00). For that purpose, a single point at P/P₀ \approx 1.00 was analyzed.

5.2.7. UV-VIS Analysis and Determination of the Protective Properties against UV Radiation

Changes of the physical properties as transmittance (%T) of samples occurring during modifications were assessed using double beam Jasco V-550 UV-VIS spectrophotometer (Tokyo, Japan) with integrating sphere attachment in the range of 200–800 nm, analogously as we described earlier [65]. The same apparatus was used to determine the ultraviolet protection factor (UPF) of samples.

The UPF value of the samples was determined, according to EN 13758-1:2002 standard [93], as the arithmetic mean of the UPF values (Equation (2)) for each of the samples (a confidence interval of 95%), analogously, as described earlier [67].

$$UPF = \frac{\int_{290}^{400} E(\lambda)\varepsilon(\lambda)d(\lambda)}{\int_{290}^{400} E(\lambda)\varepsilon(\lambda)T(\lambda)d(\lambda)}$$
(2)

where:

 $E(\lambda)$ —the solar irradiance;

 $\varepsilon(\lambda)$ —the erythema action spectrum (measure of the harmfulness of UV radiation for human skin); $\Delta\lambda$ —the wavelength interval of the measurements;

 $T(\lambda)$ —the spectral transmittance at wavelength λ .

5.2.8. Antibacterial Activity

The PLA–ALG–Cu²⁺ fabrics' anti-bacterial activity was tested according to standard: PN-EN ISO 20645:2006 [94], against a colony of Gram-positive bacteria, *Staphylococcus aureus* (ATCC 6538) and Gram-negative bacteria, *Escherichia coli* (ATCC 25922), analogously as in polypropylene nonwovens [65].

5.2.9. Antifungal Activity

The antifungal activity of resulting samples was tested according to PN-EN 14119:2005 [95]. This standard indicates tests of antifungal activity on a *Chaetomium globosum* (ATCC 6205), analogously, as in PP nonwovens [65].

Author Contributions: M.H.K. developed the concept, designed and performed the experiments, analyzed the data, and wrote the paper; M.B. developed the concept and designed the experiments and participated in the publication preparation; Z.M. and A.K. performed the experiments, analyzed the data, and participated in the publication preparation. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Polish Ministry of Science and Higher Education within statutory research work carried out at the Lukasiewicz Research Network-Textile Research Institute, Lodz, Poland.

Acknowledgments: This work was partly supported by National Science Centre, Poland, via Grant: *Miniatura* 2, No. 2018/02/X/ST8/01775. The authors would like to thank Grzegorz Celichowski (UŁ) for performing SEM and EDS analyses.

Conflicts of Interest: The authors declare no conflict of interest.

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