

Similarity of salivary microbiome in parents and adult children

Kati Sundström^{1,2,3,4}, Pashupati P. Mishra^{2,3,4}, Mikko J. Pyysalo^{5,6,7,8},
Terho Lehtimäki^{2,3,4}, Pekka J. Karhunen^{1,2,3,4} and Tanja Pessi^{1,8,9}

¹ Department of Forensic Medicine, Faculty of Medicine and Life Sciences, University of Tampere, Tampere, Finland

² Department of Clinical Chemistry, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

³ Finnish Cardiovascular Research Center-Tampere, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

⁴ Department of Clinical Chemistry, Fimlab Laboratories Ltd, Tampere, Finland

⁵ Department of Otorhinolaryngology, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

⁶ Department of Oral and Maxillofacial Diseases, Tampere University Hospital, Tampere, Finland

⁷ Oral Health Services, City of Tampere, Tampere, Finland

⁸ Department of Molecule Microbiology, Faculty of Medicine and Health Technology, Tampere University, Tampere, Finland

⁹ Science Centre, Tampere, Finland

ABSTRACT

Background: Human saliva contains approximately 700 bacterial species. It has been reported that the salivary microbiome of a large family of closely related individuals consisting of multiple households is similar but the relatedness of salivary bacteria between generations of parents and their children has not yet been investigated. The objectives were to investigate the entirety of salivary bacterial DNA profiles and whether and how families share these profiles and also compare these communities between grandparents and their first daughter generations (F1) using 16S rRNA gene amplicon sequencing.

Results: The most abundant phyla in two separate families were *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Fusobacteria* and *Actinobacteria*. Family ties explained 13% of the variance between individuals' bacterial communities ($R^2 = 0.13$; $P = 0.001$). Mothers shared more OTUs with adult children compared to fathers, but this linkage seemed to be weaker in the nuclear family with older adult children. We identified 29 differentially abundant genus level OTUs ($FDR < 0.05$) between families, which accounted for 31% of the total identified genus level OTUs.

Conclusions: Our results indicate that adult family members share bacterial communities and adult children were more similar to mothers than fathers. The observed similarity in oral microbiome between parent-child pairs seemed to weaken over time. We suggest that our analysis approach is suitable for relatedness study of multigenerational salivary bacteria microbiome.

Submitted 4 March 2019
Accepted 25 February 2020
Published 9 April 2020

Corresponding author
Kati Sundström,
kati.sundstrom@tuni.fi

Academic editor
Tatiana Tatarinova

Additional Information and
Declarations can be found on
page 10

DOI 10.7717/peerj.8799

© Copyright
2020 Sundström et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Subjects Bioinformatics, Genetics, Microbiology, Molecular Biology

Keywords Metagenomics, Next generation sequencing, Saliva, Microbiota, Similarity, 16S rRNA gene

BACKGROUND

The human body is considered as holobiont, meaning an organism consisting of the host and all its associated microbiota (*Rosenberg & Zilber-Rosenberg, 2016*), which consists of approximately 3.0×10^{13} human cells and 3.8×10^{13} bacterial cells (*Sender, Fuchs & Milo, 2016*). Bacteria are transmitted to the host in two ways: horizontal transmission occurs between unrelated individuals by contact or by respiratory, oral or fecal spread. Vertical transmission occurs directly from parents to offspring, for example, via ovum, placenta, vagina, milk or saliva (*Van Winkelhoff & Boutaga, 2005; Matsumiya et al., 2002*). According to the holobiont theory, where humans are a combination of host and microbial cells, birth is more than just an origin of a new individual; it is an origin of a new community, that is, a new holobiont (*Gilbert, 2014*).

The holobiont theory is supported by studies on meconium, the first stool of a mammalian infant, which is secreted during foetal time and shown to contain bacteria (*Gosalbes et al., 2013*). *Jimenez et al. (2008)* showed in mice studies that labelled *Enterococcus faecium* was found in the pup's meconium after an aseptic caesarean section in those pregnant mice whose diet contained the same bacteria. Infants acquire their mothers' microbiota from multiple anatomic sites after birth. Previous studies have demonstrated a high degree of similarity of oral bacterial compositions between mothers and their children; the two may share as much as 94% of their oral bacterial spectra, including cariogenic species (*Li et al., 2007*).

The oral cavity is a major gateway for bacteria to enter the human body and a natural route for passage to respiratory and gastrointestinal tracts. The oral cavity consists of a diverse and complex community containing hundreds of different bacterial species. Saliva is a good candidate to study human microbiota since the sampling is non-invasive and fast. Salivary microbiota can also be distinguished from other oral microbiomes, such as gingival or tongue microbiome (*Segata et al., 2012*). It contains approximately 700 different bacterial species (*Aas et al., 2005*) at an average density of 1.4×10^8 organisms per millilitre (*Lazarevic et al., 2011*). Due to the abundance of bacteria and its' distinguished characteristics, it is easy to build up individual bacterial profiles. Moreover, the microbiome in the mouth is considered more stable than in the gastrointestinal tract and other microbial sites of the body (*Costello et al., 2009*). A longitudinal twin study showed that there is a core oral microbiome that does not change over time, but also that there is no difference between monozygotic and dizygotic twins, indicating that genetics do not affect oral microbiome composition (*Stahringer et al., 2012*). However, the similarity of the oral bacterial microbiome among adult family members has been reported (*Shaw et al., 2017*), but it is not known whether this bacterial microbiome profile characterizes families over generations.

Our aim was to do a feasibility study on the relatedness of oral microbiome by amplifying the 16S rRNA gene from salivary samples and to evaluate similarity of salivary bacterial profile between generations of parents and their children.

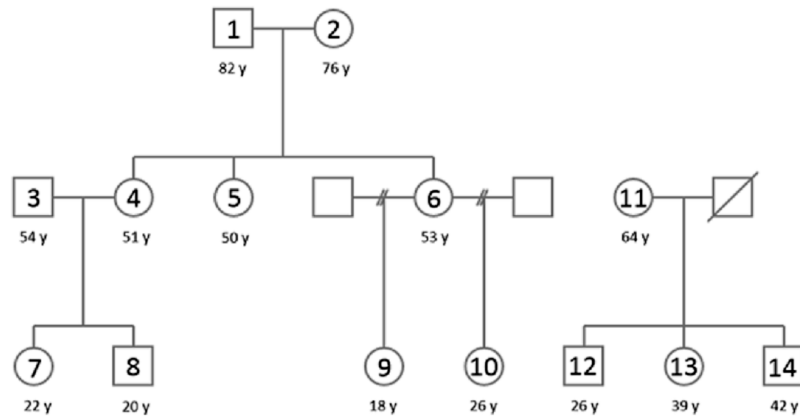


Figure 1 A pedigree of the population used in this study. Family 1 (subjects 1–10) is located on the left and family 2 (subjects 11–14) on the right. The squares denote males and circles females, sample numbers are marked inside and ages (y) below the circles/squares. Subject 11 was excluded for pathogenic bacteria dominance in sequencing results and subject 13 was excluded due to smoking. Therefore core Family 1 included subjects 1–10 and core family 2 included subjects 12 and 14.

Full-size [DOI: 10.7717/peerj.8799/fig-1](https://doi.org/10.7717/peerj.8799/fig-1)

MATERIALS AND METHODS

Study population

The study subjects were a family of three generations including ten adults, and an unrelated family of two generations including four adults (Fig. 1) (ethical approval by the Regional Ethics Committee of the Expert Responsibility area of Tampere University Hospital, reference number: R12217, and oral consent). Subjects were asked using a questionnaire about their general health, smoking habits and living conditions. Relatedness was based on a self-reported pedigree and not confirmed by DNA-testing since household sharing is the proposed main driver of salivary microbiome similarity. All adult children have shared household with their parents at least until the age of 18 years. Both families live in the same area in Southern Finland in an urban or suburban setting. All sampled subjects were used to study the entirety and total bacterial genera of oral microbiota using NGS.

Collection of saliva samples

Unstimulated saliva samples were collected into sterile plastic vials (Sarstedt AG & Co., Nümbrecht, Germany). Samples were stored at -20°C and analysed within 18 h. The subjects were asked to not eat or drink for 2 h prior to sampling. Subject 13 was excluded due to being the only smoker, thus smoking is not a confounding factor in this study.

DNA extraction and sequencing of the 16S rRNA gene

DNA was extracted from a maximum volume of two ml saliva according to the PureLink microbiome DNA purification kit (Thermo Fisher Scientific, Waltham, MA, USA). All samples were amplified in triplicates using universal primers targeting the V3–V4

regions on 16S rRNA gene: the forward primer with adapter was 341F TCG TCG GCA GCG TCA GAT GTG TAT AAG AGA CAG CCT ACG GGA GGC AGC AG (Muyzer, De Waal & Uitterlinden, 1993) and the reverse primer with adapter was R806 GTC TCG TGG GCT CGG AGA TGT GTA TAA GAG ACA GGG ACT ACH VGG GTW TCT AAT (Caporaso et al., 2011). The reaction mixture (25 µL) contained 2.5 µL genomic DNA, 2× KAPA HiFi HotStart ReadyMix (Kapa Biosystems, Wilmington, MA, USA), and 0.2 mM forward and 0.5 mM reverse primer. The PCR reaction conditions for amplification of DNA were as follows: Initial denaturation at 95 °C for 3 min, followed by 35 cycles of denaturation at 95 °C for 20 s, annealing beginning at 65 °C and ending at 55 °C for 15 s and extension at 72 °C for 30 s. The annealing temperature was lowered 1 °C every cycle until reaching 55 °C, which was used for the remaining cycles. Final elongation was for 5 min at 72 °C. Negative controls were included in triplicates during amplification. Magnetic bead purification (Beckman Coulter, Brea, CA, USA), second PCR, normalization and pooling were performed according to Illumina's 16S metagenomic sequencing library preparation protocol (Illumina Ltd., San Diego, CA, USA). MiSeq® Reagent Kit v3 for 600 sequencing cycles (Illumina Ltd., San Diego, CA, USA) was used for MiSeq library with a final concentration of 4 pM and with 10% PhiX control. The DNA pool included a commercial *Streptococcus mitis* strain (ATCC® 49456™, LGC Standards, Teddington, Middlesex, UK) as mock community (Pyysalo et al., 2019).

Data analysis

The analyses were performed with Quantitative insight to microbial ecology (QIIME) (Caporaso et al., 2010) and Mothur software (Schloss et al., 2009). Low quality sequences were trimmed with minimum average PHRED quality score threshold of 20 (Q20) using Trimmomatic version 0.33 (Bolger, Lohse & Usadel, 2014). Sequences shorter than 200 nucleotide bases were dropped out. Chimeric sequences were identified using usearch61 (Edgar, 2010) method in de novo mode via identify_chimeric_seqs.py script in QIIME 1.9.1. Contaminants, that is, *Archaea*, *Eukarya*, mitochondrial and unknown sequences were filtered out with remove.lineage command in Mothur (version 1.38.1.). Taxonomies that were different among replicates were considered as bacterial contamination and were removed separately from each sample.

OTU picking was done with QIIME (version 1.9.1.) with UCLUST (Edgar, 2010) in de novo mode via the pick_otu.py script. Default parameters were used and clusters were generated with 97% similarity threshold but we focused our report to genus level based on assigned taxonomy to OTUs. Taxonomy assignment was done to the representative sequences for each of the OTUs via assign_taxonomy.py script against SILVA database (123 release) (Pruesse et al., 2007) as well as Human oral microbiome database (HOMD) (version 14.51) (Chen et al., 2010) using default parameter settings. Similarly, alignment of the representative sequences, filtration of the gaps present in all sequences in the alignment and building of a phylogenetic tree using the alignment were accomplished using the scripts in QIIME 1.9.1 software suite. Picked OTUs were converted to an OTU table in BIOM format for subsequent analysis using make_otu_table.py script.

The OTU table was normalized using cumulative sum scaling (Paulson et al., 2013) via `normalize_table.py` script in QIIME 1.9.1.

Beta diversity was calculated using unweighted UniFrac method via `beta_diversity.py` script. Adonis test (Anderson, 2001) was performed to assess the difference in bacterial community between the two families. Homogeneity of dispersion among groups and the validity of Adonis was tested using PERMDISP method (Anderson, 2006) via `compare_categories.py` script in QIIME 1.9.1 software suite. Differential abundance analysis was done using DESeq2 method via `differential_abundance.py` script in QIIME 1.9.1. Two nuclear families from our cohort were chosen to investigate the difference in OTUs shared between mother or father with adult children using Venn diagrams in R. Significance of the overlaps were estimated with hypergeometric test. Nuclear family A consists of parents and three daughters (subjects 1, 2, 4, 5 and 6) and nuclear family B consists of parents, one son and one daughter (subjects 3, 4, 7 and 8) (Figs. S1–S3). Figures S1 and S2 represent core A dataset. Figure S3 represent core B dataset. Two plots were generated for core A dataset because there are five groups in it and venn diagram can be clearly plotted with maximum of four groups.

RESULTS

The quality control rate, Q30%, for the run was 75.7%. The total number of sequences obtained in one single run from the analyzed samples was 5,293,569, the average number of reads per sample was 182,536, and the average Shannon species diversity was 2.997 (SD = 0.108). Family transmission study was conducted in Family 1 for subjects 1–8 because only core families with both father, mother and all adult children present can be used for family studies. Subjects 9 and 10 from family 1 were not used for transmission study because they had two different fathers and comparison could not be made. Family 2 was not used for transmission study due to lack of father. Moreover, even if dental health was not examined in detail before sampling, no subjects claimed oral disease except for subject 11. In case of this subject, the saliva sample was mixed with a significant amount of blood and sequencing results showed dominance in pathogenic, mostly periodontitis bacterial sequences. For this reason, we excluded subject 11.

Differential abundance analysis of SILVA based taxonomy yielded 69 oral taxa. The analysis was repeated with HOMD based taxonomy, which yielded 91 taxa. According to SILVA, the major phyla were *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Fusobacteria* and *Actinobacteria* (38% of the total identified phyla). The most common genera were *Streptococcus* spp., *Veillonella* spp., *Prevotella* spp., *Neisseria* spp. and *Leptotrichia* spp. (3.7% of the total identified genera). The most significant abundances were observed in bacterial taxa like unclassified *Synergistaceae*, *Atopobium* spp., Human oral bacterium BD1-5, *Lactobacillus* spp. and *Butyrivibrio* spp. (Table 1).

Differences in oral microbiome between and within families

The R^2 value obtained from Adonis test indicates that approximately 13% of the variances in the distances is explained by grouping based on families ($R^2 = 0.13$; $p = 0.001$).

Table 1. Five most abundant taxa and their differences compared between the two families and obtained with SILVA. Stat is DESeq2 Wald statistics that represents difference in abundance of taxa between the compared families. The p -value (adjusted) represents the probability that the observed difference (Wald statistics) is observed even though there is no real difference (by chance).

Operational taxonomic unit	P_{adjusted}	Stat
<i>Synergistaceae</i> unclassified	1.66×10^{-11}	-7.381
<i>Atopobium</i> spp.	6.28×10^{-9}	6.441
Human oral bacteria BD1-5	8.21×10^{-8}	-5.973
<i>Lactobacillus</i> spp.	1.84×10^{-7}	5.792
<i>Butyrivibrio</i> spp.	1.07×10^{-4}	-4.492

Significant difference in dispersion was indicated between the two families by PERMDISP test (F -value = 9.17, $p = 0.006$).

Of the 69 oral taxa detected by Differential abundance analysis in SILVA based taxonomy, 29 were significantly different in two families (FDR < 0.05, [Table S1](#)). Of the 91 taxa detected with HOMD based taxonomy, 39 were found to be significantly different (FDR < 0.05, [Table S2](#)). Of all observed taxa, 22 were common to both databases. Major differences were observed in unclassified taxa ($n = 6$, [Tables S1](#) and [S2](#)).

Shared OTUs between parents and adult children were mapped in Venn diagrams ([Figs. S1–S3](#)). Overlaps with hypergeometric p -value were analyzed, indicating adult children share more OTUs with mothers as compared to fathers but the difference in shared OTUs decreased over time with the child aging ([Fig. 2; Table 2](#)).

DISCUSSION

Saliva is one of the most studied oral microbiomes in humans due to the ease of collection ([Cameron et al., 2015](#)). We investigated the entirety of salivary bacterial DNA profiles and whether and how families share these profiles and also compare these communities between grandparents and their first daughter generation (F1) using 16S rRNA gene amplicon sequencing. Our results indicate that adult family members share bacterial communities and adult children were more similar to mothers than fathers. The observed similarity in oral microbiome between parent–child pairs seemed to weaken over time.

We used next generation sequencing and two databases (SILVA, HOMD) to analyze the entirety and vertical transmission of bacterial community in saliva in two families. We used two databases for comparison to increase reliability of our results due to small sample size. In our cohort results were better seen and presented with SILVA (123 release), thus we chose to use this database for presenting results. Databases are constantly updated and for this reason new versions may give different results.

Of the two databases, SILVA is older, and for long considered as the gold standard. SILVA provides updated data sets of aligned small (16S/18S) and large subunit (23S/28S) sequences for all three domains of life (*Bacteria*, *Archaea* and *Eukarya*) ([Quast et al., 2013](#)), whereas HOMD is a relatively new database, but has lately been used a lot in oral microbiome related articles ([Timby et al., 2017](#); [Vesty et al., 2017](#); [Adams et al., 2017](#)). HOMD is a smaller database, since the human oral cavity only consists of approximately

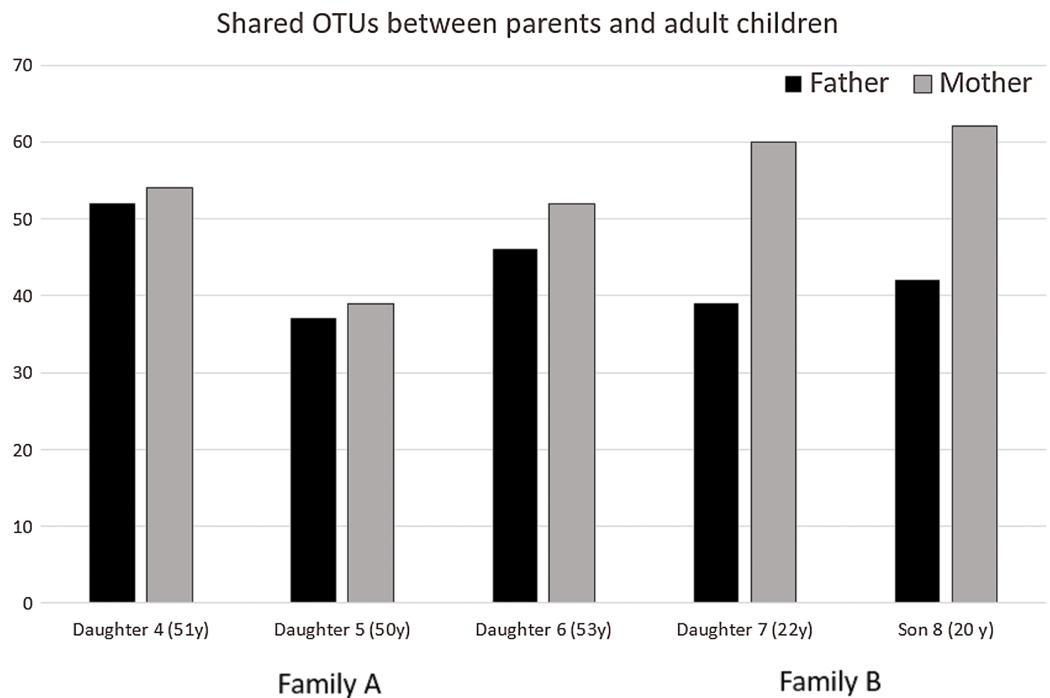


Figure 2 Shared OTUs between parents and adult children according to Fig. 1.

Full-size DOI: 10.7717/peerj.8799/fig-2

700 species, whereof 400 are currently listed in HOMD. It is possible to go down to species level with this phylogeny-curated database ([Dewhurst et al., 2010](#)). The variety between results from databases can partly be explained by the biases for each database, where certain bacteria genera or phyla are often overrepresented, but also since assigning down to genera level from 16S rRNA gene sequences can hide micro-heterogeneity and thus falsify OTU results ([Poretsky et al., 2014](#)). A set of universal primers was used for amplification of bacterial DNA, however, no primer pair is actually universal, and thus there is a possibility of DNA sequence dropout due to the primers, which are not amplifying all sequences.

The most significantly abundant taxa according to SILVA, *Synergistaceae* unclassified, was not recognized by HOMD. Among all significant taxa, 22 same taxa were observed in both databases, among the 10 most significantly abundant taxa *Peptostreptococcaceae*, *Megasphaera* spp., *Capnocytophaga* spp. and *Slackia* spp. were recognized in both databases. Overall, the taxa recognized by the databases were relatively similar, but their RFD-values were for the most part not consistent. According to SILVA, we found Firmicutes to be the most abundant phyla. Our results are in line with those by [Shaw et al. \(2017\)](#) who established the same result with a significantly larger sample size and by using the database HOMD. Our results indicating Streptococcus, Veillonella, Prevotella and Neisseria being the most abundant genera. Similarly, [Shaw et al. \(2017\)](#) found that the most abundant genera were Streptococcus, Rothia, Neisseria and Prevotella.

The main reason for similarity of microbiota between newborns and mothers is considered bacteria that relocate from the birth canal during labor and from breast milk in

Table 2 The parent-child-pairs OTU overlap and hypergeometric *p*-value. The samples with replicates were summarized by taking average (avg) of the replicates (Figs. S1–S3). The change in sample name after summarizing is mapped to its original pedigree number.

	Pedigree number	Age of child (y)	Age of parent (y)	Overlap (number of OTUs)	Hypergeometric <i>p</i> -value
Nuclear family A					
CoreA_Mother–coreA_daughter1_avg (Fig. S1)	2 4	51	76	54	0.006811284
CoreA_Mother–coreA_daughter2 (Fig. S1)	2 5	50	76	39	NS
CoreA_Mother–coreA_daughter3 (Fig. S2)	2 6	53	76	52	4.593393e-05
CoreA_father–coreA_daughter1_avg (Fig. S1)	1 4	51	82	52	3.138095e-05
CoreA_father_avg–coreA_daughter2 (Fig. S1)	1 5	50	82	37	NS
CoreA_father_avg–coreA_Daughter3_avg (Fig. S2)	1 6	53	82	46	0.0001144497
Nuclear family B					
CoreB_mother_avg–coreB_daughter_avg (Fig. S3)	4 7	22	51	60	2.649257e-06
CoreB_mother_avg–coreB_Son (Fig. S3)	4 8	20	51	62	NS
CoreB_father_avg–coreB_daughter_avg (Fig. S3)	3 7	22	54	39	0.0005486819
CoreB_father_avg–coreB_Son (Fig. S3)	3 8	20	54	42	NS

infancy (Verdu, Galipeau & Jabri, 2015). From the birth canal, the child obtains the mother's vaginal and faecal bacteria (Rosenberg & Zilber-Rosenberg, 2016) and bacteria from milk during breastfeeding (Fernandez *et al.*, 2013). It has been shown that the diversity of a new-born's gut microbiome changes gradually over time, reflecting changes in diet (Koenig *et al.*, 2011). The earliest colonizers in the child's oral cavity depend on both surrounding microbes and antibodies inherited from the mother.

Thus, the greater similarity of maternal bacteria dates back to childhood and a close physical contact between mother and infant. In contrast, to our knowledge, the stability of bacterial transfer has not been studied in adulthood between adult mother and child pairs over generations or by comparing oral microbial profiles of the adult child to the father's microbiome. Previous studies (Li *et al.*, 2007; Könönen, 2000; Milani *et al.*, 2015) have focused on the development of the microbiome in children and adolescents, but not on the resemblance of the microbiome in adulthood and of three generations, as we have now done.

A study based on microbiome analysis of twin-pairs concluded that environmental factors provide the greatest influence on oral microbial composition (Stahringer *et al.*, 2012; Shaw *et al.*, 2017). Kort *et al.* (2014) however, showed that intimate kissing increases

similarity between oral microbiomes of couples only temporarily, suggesting that an adult's microbiota is stable. In our study, oral bacterial profile comparison between parents and younger adult children show a higher resemblance compared to elderly parents and their older adult children. Younger adult children, seven (22 years) and eight (20 years), still live with their parents, and this could partly explain the larger amount of shared OTUs. Older adult children four (51 years), five (50 years) and six (53 years) have lived in their own households for at least two decades.

It has been suggested that a large part of the oral microbiome is similar around the world (*Shaw et al., 2017; Nasidze et al., 2009*). Our PERMDISP results suggest that there are certain differences between families, and those differences might be due to the difference in dispersion instead of center. Moreover, the Adonis result reporting 13% variance due to family ties and rest due to environmental bacteria in the mouth is consistent with studies by *Kort et al. (2014)* and *Nasidze et al. (2009)*.

The major weakness of this study is, however, the sample size. The small sample size interferes especially the heterogeneity calculated using PERMDISP and due the fact that we had only a few parents-adult children pairs to draw Venn diagrams and to calculate hypergeometric *p*-values. Thus, we conclude that larger cohorts are needed to confirm our preliminary results.

CONCLUSION

In conclusion, our exploratory study making use of 16S rRNA amplicon sequencing followed by extensive data analysis suggests that even if mothers could be closer to their adult children compared to fathers in early adulthood, this similarity may change over time. Our study suggests that even though the oral cavity is very prone to inhabit environmental bacteria, the mother still has a role in her offspring's oral microbiota in the adulthood. This research setting can serve as a foundation for further research with larger sample sizes and better defined families.

LIST OF ABBREVIATIONS

NGS	Next generation sequencing
OTU	Operational Taxonomic Unit FDR
QIIME	Quantitative insights to microbial ecology
Q20	Quality score threshold of 20
HOMD	Human oral microbiome database
SD	Standard deviation
16S/18S	Small subunit of rRNA
23S/28S	Large subunit of rRNA

ACKNOWLEDGEMENTS

We want to thank the volunteers for participating in this study by providing saliva samples.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by Jane and Aatos Erkko Foundation, Finnish Clinical Chemistry Foundation, State research funding for Tampere University Hospital Region, Pirkanmaa Regional Fund, Terveystutkimuksen Toimikunta (No. 286284), Finnish Foundation for Cardiovascular Research, Yrjö Jahnssoon Foundation, Tampereen Tuberkuloosisäätiö, EU Horizon 2020 Framework Programme (NO. 755320 and Aition grant 8484146), Finnish Academy grant no 322098, Signe and Ane Gyllenberg Foundation, Diabetestutkimussäätiö, Swedish Cultural Foundation in Finland, Päivikki and Sakari Sohlberg Foundation, Laboratoriolääketieteen edistämssäätiö. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Finnish Clinical Chemistry Foundation.

Tampere University Hospital Region, Pirkanmaa Regional Fund, Terveystutkimuksen Toimikunta: 286284.

Finnish Foundation for Cardiovascular Research, Yrjö Jahnssoon Foundation, Tampereen Tuberkuloosisäätiö, EU Horizon 2020 Framework Programme: 755320 and Aition grant 8484146.

Finnish Academy: 322098.

Signe and Ane Gyllenberg Foundation, Diabetestutkimussäätiö, Swedish Cultural Foundation in Finland, Päivikki, Sakari Sohlberg Foundation, Laboratoriolääketieteen edistämssäätiö.

Competing Interests

Terho Lehtimäki, Pekka Karhunen, Kati Sundström and Pashupati Mishra are employees of Fimlab Laboratories Ltd. The authors declare there are no competing interests concerning this study.

Author Contributions

- Kati Sundström conceived and designed the experiments, performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Pashupati P. Mishra conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Mikko J. Pyysalo conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Terho Lehtimäki conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

- Pekka J. Karhunen conceived and designed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Tanja Pessi conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.

Human Ethics

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

The Pirkanmaa Hospital District granted ethical approval to carry out this study (ethical application ref: ETL R12217).

Data Availability

The following information was supplied regarding data availability:

Raw sequence data and code are available at Zenodo: Sundström Kati, Mishra Pashupati P, Pyysalo Mikko J, Karhunen Pekka J, Lehtimäki Terho & Pessi Tanja. (2019). Similarity of salivary microbiome in parents and adult children. (Data set). Zenodo. DOI [10.5281/zenodo.2582046](https://doi.org/10.5281/zenodo.2582046).

Additional data (pre-processed data run through a database to identify a bacterial genera for each sequence) and codes are available in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.8799#supplemental-information>.

REFERENCES

- Aas JA, Paster BJ, Stokes LN, Olsen I, Dewhirst FE. 2005. Defining the normal bacterial flora of the oral cavity. *Journal of Clinical Microbiology* **43**(11):5721–5732 DOI [10.1128/JCM.43.11.5721-5732.2005](https://doi.org/10.1128/JCM.43.11.5721-5732.2005).
- Adams SE, Arnold D, Murphy B, Carroll P, Green AK, Smith AM, Marsh PD, Chen T, Marriott RE, Brading MG. 2017. A randomised clinical study to determine the effect of a toothpaste containing enzymes and proteins on plaque oral microbiome ecology. *Scientific Reports* **7**(1):43344 DOI [10.1038/srep43344](https://doi.org/10.1038/srep43344).
- Anderson MJ. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecology* **26**:32–46.
- Anderson MJ. 2006. Distance-based tests for homogeneity of multivariate dispersions. *Biometrics* **62**(1):245–253 DOI [10.1111/j.1541-0420.2005.00440.x](https://doi.org/10.1111/j.1541-0420.2005.00440.x).
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**(15):2114–2120 DOI [10.1093/bioinformatics/btu170](https://doi.org/10.1093/bioinformatics/btu170).
- Cameron SJ, Huws SA, Hegarty MJ, Smith DP, Mur LA. 2015. The human salivary microbiome exhibits temporal stability in bacterial diversity. *FEMS Microbiology Ecology* **91**(9):fiv091 DOI [10.1093/femsec/fiv091](https://doi.org/10.1093/femsec/fiv091).
- Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JI, Huttley GA, Kelley ST, Knights D, Koenig JE, Ley RE, Lozupone CA, McDonald D, Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Turnbaugh PJ, Walters WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R. 2010. QIIME allows analysis

- of high-throughput community sequencing data. *Nature Methods* 7(5):335–336
DOI 10.1038/nmeth.f.303.
- Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Lozupone CA, Turnbaugh PJ, Fierer N, Knight R. 2011. Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *Proceedings of the National Academy of Sciences of the United States of America* 108(Suppl. 1):4516–4522 DOI 10.1073/pnas.1000080107.
- Chen T, Yu WH, Izard J, Baranova OV, Lakshmanan A, Dewhirst FE. 2010. The human oral microbiome database: a web accessible resource for investigating oral microbe taxonomic and genomic information. *Database (Oxford)* 2010:baq013
DOI 10.1093/database/baq013.
- Costello EK, Lauber CL, Hamady M, Fierer N, Gordon JI, Knight R. 2009. Bacterial community variation in human body habitats across space and time. *Science* 326(5960):1694–1697
DOI 10.1126/science.1177486.
- Dewhirst FE, Chen T, Izard J, Paster BJ, Tanner ACR, Yu W-H, Lakshmanan A, Wade WG. 2010. The human oral microbiome. *Journal of Bacteriology* 192(19):5002–5017
DOI 10.1128/JB.00542-10.
- Edgar RC. 2010. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* 26(19):2460–2461 DOI 10.1093/bioinformatics/btq461.
- Fernandez L, Langa S, Martin V, Maldonado A, Jimenez E, Martin R, Rodríguez JM. 2013. The human milk microbiota: origin and potential roles in health and disease. *Pharmacological Research* 69(1):1–10 DOI 10.1016/j.phrs.2012.09.001.
- Gilbert SF. 2014. A holobiont birth narrative: the epigenetic transmission of the human microbiome. *Frontiers in Genetics* 5:282 DOI 10.3389/fgene.2014.00282.
- Gosalbes MJ, Llop S, Vallès Y, Moya A, Ballester F, Francino MP. 2013. Meconium microbiota types dominated by lactic acid or enteric bacteria are differentially associated with maternal eczema and respiratory problems in infants. *Clinical & Experimental Allergy* 43(2):198–211
DOI 10.1111/cea.12063.
- Jimenez E, Marin ML, Martin R, Odriozola JM, Olivares M, Xaus J, Fernández L, Rodríguez JM. 2008. Is meconium from healthy newborns actually sterile? *Research in Microbiology* 159(3):187–193 DOI 10.1016/j.resmic.2007.12.007.
- Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, Angenent LT, Ley RE. 2011. Succession of microbial consortia in the developing infant gut microbiome. *Proceedings of the National Academy of Sciences of the United States of America* 108(Suppl. 1):4578–4585
DOI 10.1073/pnas.1000081107.
- Kort R, Caspers M, Van de Graaf A, Van Egmond W, Keijser B, Roeselers G. 2014. Shaping the oral microbiota through intimate kissing. *Microbiome* 2:41.
- Könönen E. 2000. Development of oral bacterial flora in young children. *Annals of Medicine* 32(2):107–112 DOI 10.3109/07853890009011759.
- Lazarevic V, Whiteson K, François P, Schrenzel J. 2011. The salivary microbiome assessed by a high-throughput and culture-independent approach. *Journal of Integrated OMICS* 1(1):2–35
DOI 10.5584/jiomics.v1i1.43.
- Li Y, Ismail AI, Ge Y, Tellez M, Sohn W. 2007. Similarity of bacterial populations in saliva from African-American mother-child dyads. *Journal of Clinical Microbiology* 45(9):3082–3085
DOI 10.1128/JCM.00771-07.
- Matsumiya Y, Kato N, Watanabe K, Kato H. 2002. Molecular epidemiological study of vertical transmission of vaginal *Lactobacillus* species from mothers to newborn infants in Japanese, by

- arbitrarily primed polymerase chain reaction. *Journal of Infection and Chemotherapy* **8**(1):43–49 DOI [10.1007/s101560200005](https://doi.org/10.1007/s101560200005).
- Milani C, Mancabelli L, Lugli GA, Duranti S, Turrone F, Ferrario C, Mangifesta M, Viappiani A, Ferretti P, Gorfer V, Tett A, Segata N, Van Sinderen D, Ventura M. 2015.** Exploring vertical transmission of bifidobacteria from mother to child. *Applied and Environmental Microbiology* **81**(20):7078–7087 DOI [10.1128/AEM.02037-15](https://doi.org/10.1128/AEM.02037-15).
- Muyzer G, De Waal EC, Uitterlinden AG. 1993.** Profiling of complex microbial populations by denaturing gradient gel electrophoresis analysis of polymerase chain reaction-amplified genes coding for 16S rRNA. *Applied and Environmental Microbiology* **59**(3):695–700 DOI [10.1128/AEM.59.3.695-700.1993](https://doi.org/10.1128/AEM.59.3.695-700.1993).
- Nasidze I, Li J, Quinque D, Tang K, Stoneking M. 2009.** Global diversity in the human salivary microbiome. *Genome Research* **19**(4):636–643 DOI [10.1101/gr.084616.108](https://doi.org/10.1101/gr.084616.108).
- Paulson JN, Stine OC, Bravo HC, Pop M. 2013.** Differential abundance analysis for microbial marker-gene surveys. *Nature Methods* **10**(12):1200–1202 DOI [10.1038/nmeth.2658](https://doi.org/10.1038/nmeth.2658).
- Poretzky R, Rodriguez-R LM, Luo C, Tsementzi D, Konstantinidis KT. 2014.** Strengths and limitations of 16S rRNA gene amplicon sequencing in revealing temporal microbial community dynamics. *PLOS ONE* **9**(4):e93827 DOI [10.1371/journal.pone.0093827](https://doi.org/10.1371/journal.pone.0093827).
- Pruesse E, Quast C, Knittel K, Fuchs BM, Ludwig W, Peplies J, Glöckner FO. 2007.** SILVA: a comprehensive online resource for quality checked and aligned ribosomal RNA sequence data compatible with ARB. *Nucleic Acids Research* **35**(21):7188–7196 DOI [10.1093/nar/gkm864](https://doi.org/10.1093/nar/gkm864).
- Pyysalo MJ, Mishra PP, Sundström K, Lehtimäki T, Karhunen PJ, Pessi T. 2019.** Increased tooth brushing frequency is associated with reduced gingival pocket bacterial diversity in patients with intracranial aneurysms. *PeerJ* **7**(Suppl. 1):e6316 DOI [10.7717/peerj.6316](https://doi.org/10.7717/peerj.6316).
- Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, Peplies J, Glöckner FO. 2013.** The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic Acids Research* **41**:D590–D596 DOI [10.1093/nar/gks1219](https://doi.org/10.1093/nar/gks1219).
- Rosenberg E, Zilber-Rosenberg I. 2016.** Microbes drive evolution of animals and plants: the hologenome concept. *mBio* **7**(2):1395 DOI [10.1128/mBio.01395-15](https://doi.org/10.1128/mBio.01395-15).
- Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ, Sahl JW, Stres B, Thallinger GG, Van Horn DJ, Weber CF. 2009.** Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Applied and Environmental Microbiology* **75**(23):7537–7541 DOI [10.1128/AEM.01541-09](https://doi.org/10.1128/AEM.01541-09).
- Segata N, Haake SK, Mannon P, Lemon KP, Waldron L, Gevers D, Huttenhower C, Izard J. 2012.** Composition of the adult digestive tract bacterial microbiome based on seven mouth surfaces, tonsils, throat and stool samples. *Genome Biology* **13**(6):R42 DOI [10.1186/gb-2012-13-6-r42](https://doi.org/10.1186/gb-2012-13-6-r42).
- Sender R, Fuchs S, Milo R. 2016.** Revised estimates for the number of human and bacteria cells in the body. *PLOS Biology* **14**(8):e1002533 DOI [10.1371/journal.pbio.1002533](https://doi.org/10.1371/journal.pbio.1002533).
- Shaw L, Ribeiro ALR, Levine AP, Pontikos N, Balloux F, Segal AW, Roberts AP, Smith AM. 2017.** The human salivary microbiome is shaped by shared environment rather than genetics: evidence from a large family of closely related individuals. *mBio* **8**(5):e0123717 DOI [10.1128/mBio.01237-17](https://doi.org/10.1128/mBio.01237-17).
- Stahringer SS, Clemente JC, Corley RP, Hewitt J, Knights D, Walters WA, Knight R, Krauter KS. 2012.** Nurture trumps nature in a longitudinal survey of salivary bacterial

communities in twins from early adolescence to early adulthood. *Genome Research* **22(11)**:2146–2152 DOI [10.1101/gr.140608.112](https://doi.org/10.1101/gr.140608.112).

Timby N, Domellöf M, Holgerson PL, West CE, Lönnerdal B, Hernell O, Johansson I.. 2017.

Oral microbiota in infants fed a formula supplemented with bovine milk fat globule membranes—a randomized controlled trial. *PLOS ONE* **12(1)**:e0169831

DOI [10.1371/journal.pone.0169831](https://doi.org/10.1371/journal.pone.0169831).

Van Winkelhoff AJ, Boutaga K. 2005. Transmission of periodontal bacteria and models of infection. *Journal of Clinical Periodontology* **32(Suppl. 6)**:16–27

DOI [10.1111/j.1600-051X.2005.00805.x](https://doi.org/10.1111/j.1600-051X.2005.00805.x).

Verdu EF, Galipeau HJ, Jabri B. 2015. Novel players in coeliac disease pathogenesis: role of the gut microbiota. *Nature Reviews Gastroenterology & Hepatology* **12(9)**:497–506

DOI [10.1038/nrgastro.2015.90](https://doi.org/10.1038/nrgastro.2015.90).

Vesty A, Biswas K, Taylor MW, Gear K, Douglas RG. 2017. Evaluating the impact of DNA extraction method on the representation of human oral bacterial and fungal communities. *PLOS ONE* **12(1)**:e0169877 DOI [10.1371/journal.pone.0169877](https://doi.org/10.1371/journal.pone.0169877).