Integrated High-Throughput Bioinformatics (Microarray, RNA-Seq, and RNA Interaction) and qRT-PCR Investigation of *BMPR1B* Axis as a Potential Diagnostic Biomarker of Isfahan Breast Cancer

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Abstract

Background: According to the bioinformatics analyses and previous studies, bone morphogenetic protein receptor type 1B (*BMPR1B*) dysregulation could remarkably affect breast cancer (BC) status as a potential biomarker and tumor suppressor. Therefore, the analysis of the expression level of *BMPR1B* and other relevant biological factors such as microRNAs, long non-coding RNAs, downstream proteins in the relevant signaling pathways, and finding the accurate biological mechanism of *BMPR1B* could be helpful for a better understanding of BC pathogenicity and discovering the new treatment methods and drugs.

Materials and Methods: R Studio software (4.0.2) was used for microarray data analyses. GSE31448 dataset was downloaded by GEOquery package and analyzed by limma package. STRING and miRWalk online databases and Cytoscape software were used for interaction analyses. Quantitative measurement of *BMPR1B* expression level was performed by qRT-PCR experiment.

Result: Microarray and real-time PCR analysis revealed that *BMPR1B* has a significant downregulation in the transforming growth factor (TGF)-beta and bone morphogenic protein (BMP) signaling pathways in BC samples. *BMPR1B* is a potential diagnostic biomarker, regulated by hsa-miR-181a-5p. Also, *BMPR1B* regulates the function of BMP2, BMP6, SMAD4, SMAD5, and SMAD6 proteins.

Discussion: *BMPR1B* have a significant role in the development of BC by regulating the potential proteins' function, playing the diagnostic biomarker role, and regulation of TGF-beta and BMP signaling pathways. The high amount of *BMPR1B* protein helps in increasing the survival rate of the patients.

Keywords: Bioinformatics, bone morphogenetic protein receptor, breast neoplasms, microarray analysis, MicroRNAs

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INTRODUCTION

During the last 50 years, the bulk of cancer research has concentrated on discovering how tumor cells vary from normal cells in gene expression.^[1] The occurrence of abnormality in the gene expression levels could provide various human diseases, including Alzheimer's,^[2] multiple sclerosis,^[3] hepatitis,^[4] diabetes,^[5] and several cancer



types, including retinoblastoma,^[6] colorectal cancer,^[7] lung cancer,^[8] head and neck cancer,^[9] hepatocellular carcinoma,^[10] and breast cancer (BC).^[11-14] An in-depth examination of gene expression patterns in a number of pathogenic conditions, such as BC, can aid in therapy and disease prediction.

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The leading cause of cancer-related deaths among women globally is BC.^[15-18] According to Cancer Statistics 2018, BC is the most frequent female malignancy and the main cause of cancer mortality in women, with more than 2.1 million females diagnosed yearly and more than 62,000 fatalities. More than 60% of BC fatalities occur in poorer nations.^[19] Despite improvements in important outcomes for BC patients because of early identification and recent developments in anti-cancer medication, the disease's recurrence rate remains significant.^[20-22] Overall, measuring the expression of genes linked to BC, identifying diagnostic and prognostic biomarkers, and understanding gene expression patterns in different clinical and pathological situations linked to BC could provide useful information about the disease and aid in its prevention.

Scientists have developed a number of approaches for measuring gene expression. Two solid tools for evaluating gene expression are real-time PCR and microarray. Gene expression profiling and genome-wide gene expression analysis using DNA microarray might offer information on the amount of expression and relative expression of genes and RNAs in various groups, such as "tumor" and "normal," or "treated" and "untreated."^[23]

The bone morphogenic protein (BMP) receptor family of transmembrane serine/threonine kinases includes bone morphogenetic protein receptor type 1B (*BMPR1B*), which is a member of the transforming growth factor (TGF) superfamily, whose members are dynamically expressed in the endometrium during menstruation, pregnancy, and endometriosis.^[24,25] According to recent research, BMP signaling appears to be a critical regulator during embryonic, cardiac, and brain development.^[26] BMP members and their receptors are essential for the development and proper reproductive function of the ovaries as well as tumor-suppressors against ovarian cancer.^[27]

Therefore, in this research, we aimed to find the expression level of *BMPR1B* in the Isfahan BC samples as a potential prognostic biomarker. Furthermore, demonstrating the biological RNA and protein interactions with the downstream genes, the correlation of the *BMPR1B* expression with the survival rate of BC patients, and relevant signaling pathways are the main goals of this study.

MATERIALS AND METHODS

Tissue collection and ethics statement

All research methods in this study involving human samples were authorized by the Ethics Committee of Al-Zahra Hospital, Isfahan University of Medical Sciences, and all patients signed written consent forms. The survey protocol was authorized via the ethical committee of Islamic Azad Rasht Branch University (the bioethical code number: IR.IAU. RASHT.REC.1398.055). In case–control research, the BC and surrounding normal breast tissue samples were examined from 50 individuals with BC. None of the patients had radiation or chemotherapy experiences in the past. Tissue samples were washed in distilled water before being immersed in RNA later solution (Invitrogen, USA) and frozen in liquid nitrogen for pathologist review. Table 1 lists the clinicopathological features of BC patients.

Real-time PCR experiment

BC tissue samples and their normal breast tissue equivalents from the same people were obtained, and their total RNA content was extracted using an RNA extraction kit in accordance with the manufacturer's instructions (GeneAll, Seoul, Korea). The first-strand cDNA synthesis kit (Thermo Fisher Scientific, Waltham, MA, USA) was employed to produce cDNA in accordance with the manufacturer's instructions. To analyze the expression of *BMPR1B* and *GAPDH* as the reference gene, the cDNA products were kept at -20° C. Magnetic induction cycler was used to conduct the qRT-PCR experiment (Bio molecular Systems, Australia). The program used the device to multiply cDNA in three different temperatures. Initial 95°C for 15 min, the secondary 95°C for 15 s, a 60°C for 20 s, and a 72°C for 20 s. It should be mentioned that from the secondary 95°C onwards, the number of cycles has been 40.

Microarray analysis

Microarray analysis was performed by R Studio software (4.0.2). The GEOquery,^[28] limma,^[29] ggplot2,

Table 1: Clinicopathological features of samples		
Characteristic	Status	Number of patients (%)
Stage	Ι	8 (16)
	II	15 (30)
	III	12 (24)
	IV	9 (18)
	Unknown	6 (12)
Age	<45	20 (40)
	>45	24 (48)
	Unknown	6 (12)
Lymph node metastasis	Yes	17 (34)
	No	26 (52)
	Unknown	7 (14)
Tumor size (TS)	<2 cm	15 (30)
	5>TS>2	5 (10)
	>5 cm	29 (58)
	Unknown	1 (2)
Menopausal status	Yes	12 (24)
	No	15 (30)
	Unknown	23 (46)
ER receptor	Positive	14 (28)
	Negative	13 (26%)
	Unknown	23 (46%)
PR receptor	Positive	19 (38%)
	Negative	21 (42%)
	Unknown	10 (20%)
HER2/neu receptor	Positive	12 (24%)
	Negative	21 (42%)
	Unknown	17 (34%)

ER, PR: Estrogen receptor, Progesterone receptor

and pheatmap packages were used to get the data from the GEO database, statistical analysis of microarray data, and drawing plots and heatmap. Three GSE31448by GPL570 ([HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array) were analyzed to find the significantly dysregulated genes. Quantile normalization method was performed to normalization of the raw microarray expression data. The Bioconductor and CRAN databases were used for downloading the R packages. About 353 BC samples and four control samples were analyzed.

Bioinformatics analyses

The miRNA interaction analysis was performed by miRWalk 2 online software. The RNA interaction network was visualized by the Cytoscape (3.8.0) software. Pathway enrichment analysis was performed by KEGG and Reactome online databases.

Statistical analysis

Statistical analysis was performed by GraphPad Prism 8 software. The Student's *t*-test, Kolmogorov–Smirnov test, Welch's *t*-test, one-way ANOVA, and Tukey's multiple comparisons test were performed on the expression data to get the significance level of the experiment.

RESULTS

Microarray analysis

Our microarray data analysis on three GSE31448 dataset revealed that *BMPR1B* could have a significant downregulation in BC patients compared to normal individuals and could be considered as a potential biomarker for BC [Figure 1]. P value and adjusted P value were considered as the statistical significance parameters. For both parameters, 0.05 was considered as the significance level.

Bioinformatics results

After choosing the *BMPR1B* as the target gene, the KEGG database was used for the analysis of molecular pathways.

Pathway enrichment analysis by the KEGG database revealed that *BMPR1B* is involved in several biological pathways, such as the TGF-beta signaling pathway. Furthermore, the alternative splicing pathway and pathway in cancer are the most relevant pathways to *BMPR1B*. According to KEGG result, *BMPR1B* gene family is one of the gene families involved in the pathways leading to cell tumor. Based on the Reactome pathway analysis, *BMPR1B* presents in signaling by BMP pathway [Figure 2].

Analysis of miRNA–mRNA interaction represents that the most regulatory effect of hsa-miR-181a-5p is on the target genes that have a crucial role in tumorigenesis pathways, such as escape from apoptosis and aberrant reproduction. In addition, *BMPR1B* is one of these target genes that have an important role in these pathways. Survival analysis based on the GEPIA2 result revealed that the low expression of *BMPR1B* was correlated to the less survival rate in BC samples.

Furthermore, the mRNA–miRNA interaction analysis showed that hsa-miR-181a-5p could have various biological interactions with some other genes, such as RAD9-HUS1-RAD1 interacting nuclear orphan 1 (*RHNO1*), sterol O-acyltransferase 1 (*SOAT1*), cell division cycle 5 like (*CDC5L*), UDP-glucuronate decarboxylase 1 (*UXS1*), and TNFAIP3 interacting protein 1 (*TNIP1*). This hub gene could be remarkable biomarkers for BC disease [Figure 3].

The protein–protein interaction analysis was performed using STRING online software. This analysis revealed that *BMPR1B* could have correlations with some other mRNAs, such as bone morphogenetic protein 2 (*BMP2*), bone morphogenetic protein 6, SMAD Family Member 4 (*SMAD4*), SMAD Family Member 5 (*SMAD5*), and SMAD Family Member 6 (*SMAD6*). These hub genes could have some important effect on the expression of *BMPR1B* and hsa-miR-181a-5p in various human diseases, including BC [Figure 4].

Survival analysis of *BMPR1B* expression revealed that the high expression of *BMPR1B* has a not-significant relation with low survival rate of patients [Figure 5].



Figure 1: The related plots of microarray data analysis. (a) Volcano plot is indicating the distribution of each gene analyzed in the GSE31448 microarray dataset, based on logFC and –log 10 (adj. *P.* value). The red color indicates upregulated genes and the green color indicates downregulated genes. *BMPR1B* is indicating in the plot by a black point as a significantly downregulated gene in the dataset (b) heatmap of top 200 up differentially expressed genes in the dataset



Figure 2: Reactome pathway analysis. Based on pathway enrichment analysis, *BMPR1B* has a significant role in signaling by bone morphogenic protein pathway. The signaling pathway picture provided by Reactome online database

Real-time PCR

BMPR1B is downregulated in BC

The expression of *BMPR1B* was evaluated by GEPIA2 database and real-time PCR experiment. GEPIA2 [Figure 6] and our experiment [Figure 7a] revealed that *BMPR1B* had a significantly low expression in BC patients (logFC: -2.335, *P* value = 0.0062). Furthermore, ROC (Receiver operating characteristic) analysis was performed on the expression data of *BMPR1B*. The result of ROC analysis revealed that *BMPR1B* could be a suitable diagnostic biomarker for BC patients (AUC (Area Under Curve) = 0.7, *P* value = 0.003, Figure 7b).

Clinicopathological analysis

Analysis of clinicopathological features of BC patients was performed on expression data of *BMPR1B*. Different stages, age, tumor size, lymph node metastasis, menopausal, ER (Estrogen receptor), PR (Progesterone receptor), and Her2 receptor status were analyzed, based on the expression level of gene and miRNA. Our analysis revealed that the *BMPR1B* expression did not correlate with different pathological statuses in BC.

DISCUSSION

To design a reasonable experiment and have a logical hypothesis, we performed an integrated high-throughput bioinformatics analysis to obtain BC's hub potential prognostic and diagnostic biomarker. Furthermore, a network analysis was performed to find the correlated transcript factors and the possible effect of coding and non-coding RNAs on each other. Finally, we evaluated the expression level of *BMPR1B* as the potential prognostic biomarkers that could have a relative regulatory effect on each one by binding to the has-miR-181a-5p in an RNA interaction network. The correlation of the *BMPR1B*



Figure 3: The mRNA–miRNA interaction analysis. *BMPR1B* is indicated by a red node in the network



Figure 5: Survival analysis of *BMPR1B*, based on GEPIA2 online database. The *x* axis of this plot represents the time (month) and the *y* axis represents the precent survival of the patients. Red color indicates the patients with high expression level and the blue color indicates the patients with low expression level. According to this analysis, there is a non-significant correlation between the low expression of *BMPR1B* and low survival rate of the patients

expression with the BC patients' survival rate was also evaluated.

In summary, we demonstrated that *BMPR1B* could be a significant prognostic biomarker in the BC samples of the Isfahan population, with a decrease in the expression level. ROC analysis revealed that *BMPR1B* could be considered as



Figure 4: The protein interaction analysis result of STRING online software



Figure 6: Expression analysis of *BMPR1B* by GEPIA2 revealed that *BMPR1B* has a significant low expression in the breast cancer samples, compared to control

a BC diagnostic biomarker in the population. Furthermore, the expression level of *BMPR1B* had a non-significant positive correlation with the survival rate of the patients.

Based on our bioinformatical analysis, we understand that some other mRNAs could be affected by has-miR-181a-5p, such as *ERBIN, SOAT1, RHNO1, CDC5L, THBS3*, and *TNIP1*. Also, during the mRNA–mRNA interaction analysis, we found that *BMPR1B* could have a remarkable interaction with some other genes, such as *SMAD4, SMAD6, SMAD7, SMAD9, BMP2*, and *ACVR2A*. Furthermore, we find that *SMAD2, SMAD3, SMAD6*, and *SMAD7* are involved in the TGF-beta signaling pathway; the same pathway that *BMPR1B* was involved in it. As a suggestion, future experiments could be performed on the expression level of these genes found. In addition, for a better understanding of the interactions and the effects of these biological factors on the expression level of each other, the correlation of these genes with each other and especially with *BMPR1B* could be examined.

Previous studies have revealed that *BMPR1B* could be affected by related proteins such as *SMAD4*.^[24] Based on this study,



Figure 7: qRT-PCR analysis of *BMPR1B* expression level. (a) Relative expression analysis of *BMPR1B* revealed that *BMPR1B* has a significant decrease in the breast cancer (BC) tissue's expression (b) ROC (Receiver operating characteristic) analysis revealed that *BMPR1B* could be a suitable prognostic biomarker for BC

luciferase and chip assay represented that *SMAD4* could enhance PII promoter activity of the ovine *BMPR1B* by interacting with the *SBE1* motif directly. BMPs, members of the TGF- β superfamily, bind to their receptors, including *BMPR1B*, resulting in phosphorylation of *SMAD1*/5/8, which activates the *SMAD*-dependent pathway.^[30] Our bioinformatics analysis of the signaling pathway of *BMPR1B* proved the previous studies about this gene and related pathways. About the role of different polymorphisms of *BMPR1B* and their role in the development of BC, Zheng *et al.* 2022^[31] revealed that BC susceptibility was shown to be elevated by the T allele of rs1434536 and rs1970801. Based on the study of Bokobza *et al.* in 2009,^[32] reducing the *BMPR1B* expression level has a significant correlation with poor prognosis and proliferation of BC cells. Our result supports mentioned result about the expression level of *BMPR1B*.

About hsa-miR-181a-5p, previous studies revealed that this miRNA could have a remarkable effect on cell proliferation in gastric cancer,^[29] reducing oxidation resistance in osteoarthritis,^[30] papillary thyroid cancer,^[31] and suppressing the proliferation and GC (Gastric cancer) cells migration.^[32]

It is highly recommended to perform the luciferase assay experiment to investigate the RNA interaction of hsa-miR-181a-5p and *BMPR1B*. Also, finding the novel relevant single nucleotide polymorphisms to the risk of BC in the *BMPR1B* and performing the HRM (High resolution melt) qRT-PCR experiment helps better understand the main mechanisms of *BMPR1B* as a diagnostic biomarker in BC patients. Pathway-specific studies also may help find the upstream and downstream factors regulated by *BMPR1B*.

Declaration of patient consent

The authors certify that they have obtained all appropriate patient consent forms. In the form the patient(s) has/have

given his/her/their consent for his/her/their images and other clinical information to be reported in the journal. The patients understand that their names and initials will not be published and due efforts will be made to conceal their identity, but anonymity cannot be guaranteed.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Zhang L, Zhou W, Velculescu VE, Kern SE, Hruban RH, Hamilton SR, et al. Gene expression profiles in normal and cancer cells. Science (80-) 1997;276:1268-72.
- Theuns J, Van Broeckhoven C. Transcriptional regulation of Alzheimer's disease genes: Implications for susceptibility. Hum Mol Genet2000;9:2383-94.
- Tajouri L, Fernandez F, Griffiths L. Gene expression studies in multiple sclerosis. Curr Genomics 2007;8:181-9.
- Asselah T, Bièche I, Sabbagh A, Bedossa P, Moreau R, Valla D, *et al.* Gene expression and hepatitis C virus infection. Gut 2009;58:846-58.
- Das UN, Rao AA. Gene expression profile in obesity and type 2 diabetes mellitus. Lipids Health Dis 2007;6:35.
- Kapatai G, Brundler MA, Jenkinson H, Kearns P, Parulekar M, Peet AC, *et al*. Gene expression profiling identifies different sub-types of retinoblastoma. Br J Cancer 2013;109:512-25.
- Kheirelseid EAH, Miller N, Chang KH, Nugent M, Kerin MJ. Clinical applications of gene expression in colorectal cancer. J Gastrointest Oncol 2013;4:144-57.
- Petty RD, Nicolson MC, Kerr KM, Collie-Duguid E, Murray GI. Gene expression profiling in non-small cell lung cancer: From molecular mechanisms to clinical application. Clin Cancer Res 2004;10:3237-48.
- Nagai MA. Genetic alterations in head and neck squamous cell carcinomas. Braz J Med Biol Res 1999;32:897-904.
- Chen X, Cheung ST, So S, Fan ST, Barry C, Higgins J, et al. Gene expression patterns in human liver cancers. Mol Biol Cell 2002;13:1929-39.

- Bao T, Davidson NE. Gene expression profiling of breast cancer. Adv Surg 2008;42:249-60.
- Arpino G, Generali D, Sapino A, Lucia DM, Frassoldati A, de Laurentis M, *et al.* Gene expression profiling in breast cancer: A clinical perspective. Breast 2013;22:109-20.
- Guler EN. Gene expression profiling in breast cancer and its effect on therapy selection in early-stage breast cancer. Eur J Breast Health 2017;13:168-74.
- Reis-Filho JS, Pusztai L. Gene expression profiling in breast cancer: Classification, prognostication, and prediction. Lancet 2011;378:1812-23.
- Azadeh M, Salehzadeh A, Ghaedi K, Sasani ST. Decreased expression level of long non-coding RNA CCAT1, was observed in breast cancer tissue of an Isfahanian population (Iran). Gene Rep 2021;23:101154.
- Assad Samani L, Javadirad SM, Parsafar S, Tabatabaeian H, Ghaedi K, Azadeh M. TP53 rs1625895 is related to breast cancer incidence and early death in Iranian population. Indian J Clin Biochem 2019;34:485-9.
- Rouigari M, Dehbashi M, Tabatabaeian H, Ghaedi K, Mohammadynejad P, Azadeh M. Evaluation of the expression level and hormone receptor association of miR-126 in breast cancer. Indian J Clin Biochem 2019;34:451-7.
- Azadeh M, Salehzadeh A, Ghaedi K, Talesh Sasani S. NEAT1 can be a diagnostic biomarker in the breast cancer and gastric cancer patients by targeting XIST, hsa-miR-612, and MTRNR2L8: Integrated RNA targetome interaction and experimental expression analysis. Genes Environ 2022;44:16.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018;68:394-424.
- Kim YA, Cho H, Lee N, Jung SY, Sim SH, Park IH, et al. Doxorubicin-induced heart failure in cancer patients: A cohort study based on the Korean National Health Insurance Database. Cancer Med 2018;7:6084-92.
- 21. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J

Clin 2020;70:7-30.

- 22. Li N, Deng Y, Zhou L, Tian T, Yang S, Wu Y, *et al.* Global burden of breast cancer and attributable risk factors in 195 countries and territories, from 1990 to 2017: Results from the Global Burden of Disease Study 2017. J Hematol Oncol 2019;12:1-12.
- Dufva M. Introduction to microarray technology. Methods Mol Biol 2009;529:1-22.
- Abdurahman A, Du X, Yao Y, Sulaiman Y, Aniwashi J, Li Q. Smad4 feedback enhances BMPR1B transcription in ovine granulosa cells. Int J Mol Sci 2019;20:2732.
- Yamaji N, Celeste AJ, Thies RS, Song JJ, Bernier SM, Goltzman D, et al. A mammalian serine/threonine kinase receptor specifically binds BMP-2 and BMP-4. Biochem Biophys Res Commun 1994;205:1944-51.
- Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. Growth Factors 2004;22:233-41.
- 27. Sætrom P, Biesinger J, Li SM, Smith D, Thomas LF, Majzoub K, et al. A risk variant in an miR-125b binding site in BMPR1B is associated with breast cancer pathogenesis. Cancer Res 2009;69:7459-65.
- Sean D, Meltzer PS. GEOquery: A bridge between the Gene Expression Omnibus (GEO) and BioConductor. Bioinformatics 2007;23:1846-7.
- Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. Limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res 2015;43:e47.
- Yoshino H, Yonezawa T, Yonemori M, Miyamoto K, Sakaguchi T, Sugita S, *et al.* Downregulation of microRNA-1274a induces cell apoptosis through regulation of BMPR1B in clear cell renal cell carcinoma. Oncol Rep 2018;39:173-81.
- 31. Zheng Y, Jiang X, Wang M, Yang S, Deng Y, Li Y, et al. BMPR1B polymorphisms (rs1434536 and rs1970801) are associated with breast cancer susceptibility in Northwest Chinese han females: A case-control study. Clin Breast Cancer 2022;22:e641-6.
- Bokobza SM, Ye L, Kynaston HE, Mansel RE, Jiang WG. Reduced expression of BMPR-IB correlates with poor prognosis and increased proliferation of breast cancer cells. Cancer Genomics Proteomics 2009;6:101-8.