Supplementary Figures

Long-term expansion of basal cells and the novel differentiation methods identify mechanisms for switching Claudin expression in normal epithelia

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The PDF file includes

Supplementary Figure 1 | Biological replicates with donor 2 and additional details for Figure 1. **Supplementary Figure 2** | Biological replicates with donor 2 and additional data for Figure 3.

Supplementary Figure 3 | Biological replicates with donor 2 and additional data for Figure 4.

Supplementary Figure 4 | Biological replicates with donor 2 and additional data for Figure 5.

Supplementary Figure 1. Biological replicates with donor 2 and additional details for Figure 1.

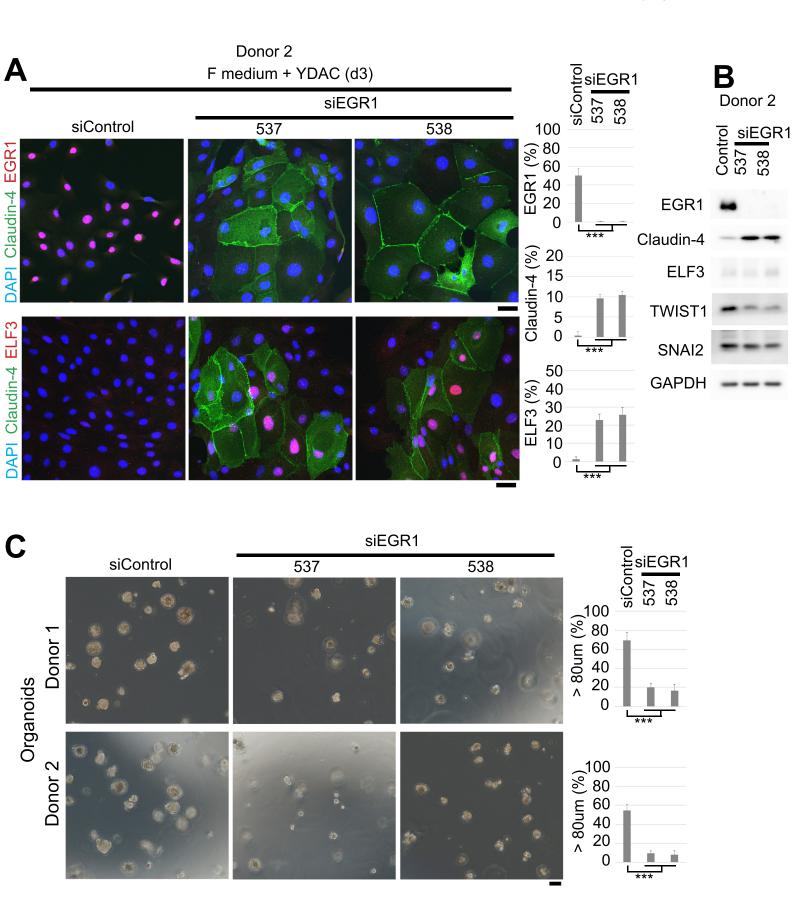
(A-D) Biological replicates of data in Figure 1, A, C, D, and E (donor 1) with independent donor 2. **(E)** Wide-field images of Figure 1 D plus an image of YDA-depleted cells extending the culture period up to 5 days. The extension did not improve the degree of differentiation. **(F)** P63 α antibody used for this research showed a single band at a molecular weight of 72 kDa in immunoblotting (donor 1), suggesting the recognition of Δ Np63 α as per a previous report [35]. Abbreviations of inhibitors: Y, 10 μ M Y-27632; D, 1 μ M DMH1; A, 1 μ M A-83-01; and C,1 μ M CHIR99021. Scale bars, 100 μ m, 10 μ m, and 10 μ m in B, C, and E, respectively.

Supplementary Figure 2. Biological replicates with donor 2 and additional data for Figure 3.

(A-E) Biological replicates of data in Figure 3 B–E (donor 1) with independent donor 2. (F) YDAC-maintained HMECs in Matrigel form well-polarized cystic structures on day 8 with partial expression of EGR1 and ELF3, suggesting heterogeneity in differentiation (immunostaining, arrows). Details are provided in the Materials and Methods section. (G) Phase-contrast micrograph of our new method for culturing mammary cells in an in vivo-like bilayer. The bilayer was assembled by culturing cells in the YDAC medium for approximately 3 days after they reached confluence. Cells on the first day of confluence (d1) were compressed and partially layered, and eventually bilayered (d2-3). Bilayering can be estimated based on the intersection of two types of cell-cell boundaries (d3-). See also Figure 3 H. Scale bars, 10 μm.

Supplementary Figure 3. Biological replicates with donor 2 and additional data for Figure 4.

(A and B) Biological replicates of data in Figure 4 A and B (donor 1) with independent donor 2. (C) Left, phase-contrast images of siEGR1-treated organoids from independent donors. Right, quantification of the organoids larger than 80 μ m in diameter. Fifty organoids were counted in each group (n=3). Scale bars, 10 μ m and 100 μ m, in A and C, respectively.



Supplementary Figure 4. Biological replicates with donor 2 and additional data for Figure 5.

(**A-C**) Biological replicates of Figure 5 A-C (donor 1) with independent donor 2. (**D**) Left, phase-contrast images of siELF3-treated organoids from independent donors. Right, quantification of the organoids with a bumpy shape. Fifty organoids were counted in each group (n=3). Scale bars, 10 μ m, 10 μ m, and 100 μ m, in A, C, and D, respectively.