© 2011 International Society of Nephrology

Open

Phosphate and FGF-23

Harald Jüppner¹

¹Endocrine Unit and Pediatric Nephrology Unit, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts, USA

Fibroblast growth factor (FGF)-23 is probably the most important regulator of serum phosphate and calcitriol (1,25(OH)₂D₃) levels. It is secreted by osteocytes and osteoblasts in response to oral phosphate loading or increased serum 1,25(OH)₂D₃ levels. In human chronic kidney disease (CKD), plasma FGF-23 appears to be a sensitive biomarker of abnormal renal phosphate handling, as FGF-23 levels increase during early stages of kidney malfunction. In humans and animals with CKD, elevated FGF-23 levels increase fractional phosphate excretion, reduce serum phosphate levels, and reduce 1a-hydroxylase activity, which reduces 1,25(OH)₂D₃ formation thereby increasing parathyroid hormone (PTH) secretion. FGF-23 thus has a key adaptive role in maintaining normophosphatemia. Plasma FGF-23 continues to increase as CKD progresses, increasing by orders of magnitude in end-stage renal disease. At the same time, responsiveness to FGF-23 declines as the number of intact nephrons declines, which is associated with reduced expression of Klotho, the co-receptor required for FGF-23 signaling. In late CKD, FGF-23 cannot reduce serum phosphate levels, and abnormally high plasma FGF-23 concentrations appear to exert unwarranted off-target effects, including left ventricular hypertrophy, faster CKD progression, and premature mortality. Lowering serum phosphate levels through the use of oral phosphate binders and/or long-acting PTH agents may reduce FGF-23 levels in early CKD stages, thereby limiting off-target effects, which may improve patient outcomes.

Kidney International (2011) **79** (Suppl 121), S24–S27; doi:10.1038/ki.2011.27; published online 23 February 2011

KEYWORDS: FGF-23; phosphate and calcium homeostasis; PTH

TO CITE THIS ARTICLE:

Jüppner H. Phosphate and FGF-23. Kidney Int 2011; 79 (Suppl 121): S24–S27.

Phosphate is essential for many cellular functions. It is a constituent of DNA, membrane lipids, high-energy phosphates, and second messengers, that is, inositol trisphosphate, cyclic adenosine monophosphate (cAMP), and cyclic guanosine monophosphate, and protein phosphorylation is an essential process, which helps regulate enzyme and receptor activities. Because phosphate is scarce in nature, vertebrate physiology has evolved to conserve phosphate through the efficient use of three sodium-dependent phosphate cotransporters (NaPi-IIa, NaPi-IIb, and NaPi-IIc, also referred to as NPT2a, NPT2b, and NPT2c) that are highly conserved throughout vertebrate taxa, from fish¹ to humans.² Additional molecules contributing to the regulation of phosphate homeostasis include Pit2 and the 1\alpha-hydroxylase, which allows the formation of calcitriol (1,25(OH)₂D₃). Expression of these proteins in the proximal renal tubules is regulated by serum phosphate concentration and different hormonal systems. For example, low serum phosphate levels induce NPT2b expression in the intestinal tract, thus enhancing absorption of this mineral from the diet. In addition, low serum phosphate levels induce NPT2a and NPT2c expression in the proximal tubules of the kidney, thus maximizing the reabsorption of phosphate and minimizing urinary losses of this mineral. The efficiency of this latter process was strikingly demonstrated in a 1976 study, in which healthy human volunteers on a low-phosphate diet (90 mg/day) reduced their phosphate excretion considerably, thereby avoiding profound hypophosphatemia.² In contrast to the efficient adaptation to hypophosphatemia, renal excretion of excess phosphate is a more difficult problem for human physiology to solve.

The principal hormones that regulate renal phosphate handling are parathyroid hormone (PTH), which is produced by the parathyroid gland, and fibroblast growth factor (FGF)-23, which is produced by osteocytes and osteoblasts in bone. In healthy individuals, increasing serum phosphate concentration induces secretion of PTH and FGF-23. These two phosphaturic hormones reduce expression of NPT2a and NPT2c in the proximal renal tubules, thereby diminishing phosphate reabsorption and increasing urinary phosphate excretion.^{3,4} PTH also increases the 1 α -hydroxylase in the kidney, thereby increasing 1,25(OH)₂D₃ production, thus enhancing intestinal calcium and phosphate absorption. Some evidence suggests that PTH induces expression and secretion of FGF-23; FGF-23 in turn decreases 1,25(OH)₂D₃ production, which is an inhibitor of PTH production.

Correspondence: Harald Jüppner, Endocrine Unit and Pediatric Nephrology Unit, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts, USA. E-mail: hjueppner@partners.org

In chronic kidney disease (CKD), increased FGF-23 production enhances the excretion of phosphate per nephron, thereby restoring normophosphatemia. However, it also reduces 1,25(OH)₂D₃ levels,⁵ contributing to an increase in PTH secretion, which appears to occur after FGF-23 levels increase. This process disrupts the bonekidney-parathyroid endocrine axis and eventually fails to prevent the development of hyperphosphatemia as CKD progresses. The resulting changes, constituting CKD-related mineral and bone disorder (CKD-MBD), are a reflection of the trade-offs postulated by Bricker's⁶ intact nephron hypothesis: 'If solute intake does not diminish as the number of excretory units diminishes, the adaptive increase in excretion rate per nephron may be accomplished only at the expense of one or more abnormalities of the uremic state.' This review will explore preclinical and clinical studies of the role of FGF-23 in phosphate metabolism in CKD and the contribution of FGF-23 to CKD-MBD.

RENAL REGULATION OF PHOSPHATE AND CALCIUM HOMEOSTASIS Role of PTH: lessons from familial pseudohypoparathyroidism

NPT2a and NPT2c facilitate the efficient reabsorption of phosphate in the proximal tubule; expression of both transporters is regulated by PTH and FGF-23. The acute PTH-dependent regulation of NPT2a and NPT2c expression is mediated predominantly through the cAMP and protein kinase A signaling pathway.⁴ The PTH-induced increase in urinary phosphate is coupled to cAMP excretion from proximal tubules. Cyclic AMP production requires $G_s \alpha$, which appears to be derived in this portion of the kidney predominantly, if not exclusively, from the maternal allele, thereby potentially limiting the maximal amount of $G_s \alpha$ protein that can be generated in this tissue. As a consequence of this parent-specific expression of $G_{s}\alpha$ in the proximal renal tubules, maternally inherited, heterozygous GNAS mutations cause PTH resistance, that is, pseudohyperparathyroidism.⁷ Inactivating GNAS mutations in those exons that encode $G_s \alpha$ cause pseudohypoparathyroidism type Ia, whereas microdeletions within or upstream of GNAS cause pseudohypoparathyroidism type Ib and are associated with GNAS methylation changes. The mechanism by which cAMP is excreted into the urine remains unknown, but is expected to involve a specific transporter that is expressed in the proximal tubule. In the distal tubule, PTH diminishes calcium excretion through cAMP/protein kinase A- and possibly also through inositol trisphosphate/protein kinase C-dependent actions on the function of calbindin-D and TRPV5. In contrast to the findings in proximal tubular cells, $G_s \alpha$ is expressed in the distal tubular cells from both parental alleles, and maternally inherited GNAS mutations consequently leave the PTH-induced regulation of calcium excretion intact.

Role of FGF-23: lessons from CKD

Similar to the actions of PTH, FGF-23 controls NPT2a and NPT2c expression. FGF-23 is secreted by bone cells in

response to 1,25(OH)₂D₃, which increases the mRNA levels encoding this phosphaturic hormone.⁸ FGF-23 mediates its action in the kidney through an FGF receptor (FGFR)/Klotho complex to downregulate NPT2a and NPT2c expression in the proximal tubules. However, it is uncertain whether FGF-23 signals initially through receptors in the distal convoluted tubule cells in which ERK phosphorylation occurs in response to FGF-23.9 These cells are adjacent to NPT2aexpressing proximal tubular cells and it has therefore been speculated that a paracrine signal from the distal to the proximal tubules is required for decreasing NPT2a expression. Even small changes in plasma FGF-23 levels are associated with significant changes in urinary phosphate excretion, as shown by the small increase in FGF-23 levels after unilateral nephrectomy in healthy kidney donors, which is associated with increased urinary phosphate excretion.¹⁰

Besides its effect on tubular phosphate handling, FGF-23 reduces PTH secretion, and it inhibits 1α -hydroxylase leading to a decrease in $1,25(OH)_2D_3$ production, which contributes to the development of hypocalcemia and leads to an increase in PTH production.¹¹ The $1,25(OH)_2D_3$ itself increases FGF-23 production.

FGF-23 AND PHOSPHATE IN CKD

FGF-23 appears to be an important biomarker for an abnormal regulation of phosphate homeostasis in CKD and is likely to be involved in CKD-MBD pathophysiology. As CKD progresses, plasma FGF-23 levels increase. This increase occurs earlier and to a greater extent than observed for serum phosphate; in late CKD, plasma FGF-23 levels can be elevated by several orders of magnitude.^{5,12} Findings in experimental renal disease suggest that the FGF-23 increase also precedes the increase in PTH levels.¹¹ In fact, bone biopsies of patients with CKD have shown increased expression of FGF-23 already by CKD stage 2, along with a marked increase in DMP1 protein (a negative regulator of FGF-23), which may be improperly processed and could thus be inactive.¹³ Furthermore, in a prospective study of patients with mildto-moderate CKD,¹⁴ higher plasma FGF-23 levels were shown to predict a more rapid progression toward end-stage renal disease (ESRD); similarly elevated FGF-23 levels predict CKD progression in patients with diabetic nephropathy.¹⁵ In addition, elevated FGF-23 levels are independently associated with left ventricular hypertrophy in patients with CKD,16 an important finding because CKD-MBD, and particularly hyperphosphatemia, worsen the cardiovascular prognosis in CKD. Aortic calcification, a major reason for cardiovascular morbidity, in hemodialysis recipients is independently predicted by plasma FGF-23.17

Phosphate is a major regulator of FGF-23 expression.¹⁸ Dietary phosphate loading increases FGF-23 expression, whereas phosphate depletion with binders decreases the circulating levels of this hormone.¹⁹ In mild CKD, FGF-23 seems to function as a protective factor, as it triggers adaptive changes that maintain normophosphatemia. For example, in animal models, FGF-23 protects against hyperphosphatemia



Figure 1 | Spectrum of serum fibroblast growth factor (FGF)-23 levels in early chronic kidney disease and end-stage renal disease (ESRD) as compared with the normal condition and with different disorders affecting FGF-23. ADHR, autosomal dominant hypophosphatemic rickets; ARHP, autosomal recessive hypophosphatemia; TIO, tumor-induced osteomalacia; XLH, X-linked hypophosphatemia. Adapted with permission from Isakova *et al.*²⁰

by increasing urinary phosphate excretion and reducing $1,25(OH)_2D_3$ production; diminished $1,25(OH)_2D_3$ levels lead to hypocalcemia and thus to an increase in PTH levels, which further enhances renal phosphate excretion.¹¹ The importance of FGF-23 in mediating these effects is shown by the effects of anti-FGF-23 antibodies in rats with experimental CKD.¹¹ Animals with early CKD that were treated with anti-FGF-23 antibodies showed an increase in $1,25(OH)_2D_3$ levels, a normalization of serum calcium levels, and a decrease in PTH levels. Furthermore, fractional phosphate excretion decreased because of inactivation of FGF-23 resulting in hyperphosphatemia.

As CKD progresses, elevated FGF-23 levels are no longer able to enhance urinary phosphate excretion, thus leading to the development of hyperphosphatemia. This may be partly related to declining Klotho expression and a reduction in functional nephrons. When patients require treatment by dialysis, the levels of immunoreactive FGF-23 can be markedly elevated (Figure 1).²⁰ However, unlike the appearance of large amounts of C-terminal PTH fragments, most of which are biologically inactive, practically all of the circulating FGF-23 present in patients with ESRD is intact and biologically active. This conclusion is based on observations with a reporter-cell assay (human embryonic kidney cells expressing Klotho and a luciferase reporter expressed under the control of the EGR-1 promoter) designed to quantify biologically active FGF-23, which demonstrated excellent correlation with immunoreactive measurement of FGF-23. The immunometric assays used for these measurements quantify the intact FGF-23 molecule alone (Kainos assay) or the intact protein along with a C-terminal fragment (Immutopics assay).^{21,22} Consistent with the excellent correlation between bioactive and immunoreactive FGF23, Western blot analyses of plasma FGF-23 from dialysis patients showed that most of the circulating FGF-23 is intact.²¹ Thus, with CKD progression, the excess of biologically active FGF-23, which is thought to occur in response to hyperphosphatemia, ceases to be protective and may lead to pathological off-target effects that potentially contribute to the increase in mortality as FGF-23 increases in patients with ESRD^{23,24} (Figure 2).

Interventions that lower serum phosphate, such as oral phosphate binders to prevent intestinal absorption or



Figure 2 | Mortality in patients with end-stage renal disease (ESRD) receiving hemodialysis in relationship to quartiles of serum fibroblast growth factor (FGF)-23 concentration at initiation of dialysis. R, reference. Reproduced with permission from Gutiérrez *et al.*²³ Copyright The Massachusetts Medical Society.



Figure 3 | 'Trade-off hypothesis' revisited. Involvement of fibroblast growth factor (FGF)-23 in end-stage renal disease (ESRD) pathology, reflecting potentially adverse effects of increased secretion of this phosphaturic hormone, which helps normalize phosphate homeostasis but contributes to the development of secondary hyperparathyroidism. PTH, parathyroid hormone.

possibly long-acting PTH analogs that reduce NPT2a or NPT2c expression, may help prevent CKD-related FGF-23 increases (Figure 3). For example, two recently reported, long-acting PTH analogs (M-PTH(1-28) and Trp1-M-PTH(1-28)) were found to efficiently reduce NPT2a expression in proximal tubules, leading to sustained hypophosphatemia in wild-type mice.²⁵ Targeting the proximal tubules with long-acting PTH agents in early CKD may add to the effects of phosphate binders in normalizing serum phosphate levels and may help avoid an increase in FGF-23, unless PTH has a role in the synthesis and/or secretion of FGF-23. Proactive interventions to avoid the development of transient hyperphosphatemia in CKD may thus prevent an increase in FGF-23 and the hormone's putative off-target effects in CKD-MBD. In support of this conclusion, a recent post hoc analysis of a randomized clinical trial has shown that treatment with phosphate binders can markedly decrease FGF-23 levels in patients undergoing hemodialysis.²⁶

CONCLUSIONS

FGF-23 is a bone-generated major regulator of phosphate homeostasis, which appears to be an important biomarker of phosphate homeostasis in patients with CKD. Plasma FGF-23 concentrations begin to increase early in CKD, with increases by orders of magnitude by the time patients reach ESRD. Increasing FGF-23 levels are independently associated with left ventricular hypertrophy, CKD progression, and mortality, possibly through off-target actions. Lowering serum phosphate levels through the use of phosphate binders may lower FGF-23 levels. Further research is needed to show whether lowering FGF-23 levels improves outcomes in patients with CKD.

DISCLOSURE

HJ has served as a paid consultant to Genzyme and Roche, and is a named inventor on the patent outlining the development of immunometric assays for the detection of FGF-23.

ACKNOWLEDGMENTS

This article was developed from the author's presentation and discussions at the '50 Years of Discovery Following the Intact Nephron Hypothesis' symposium in Munich, Germany, 24-25 June 2010. The author meets all International Council of Medical Journal Editors criteria and acknowledges the writing assistance of Kim Coleman Healy, PhD, of Envision Scientific Solutions. Publication of this supplement was supported by Genzyme Corporation.

REFERENCES

- 1. Coloso RM, King K, Fletcher JW et al. Dietary P regulates phosphate transporter expression, phosphatase activity, and effluent P partitioning in trout culture. J Comp Physiol B 2003; 173: 519-530.
- 2 Dominguez JH, Gray RW, Lemann Jr J. Dietary phosphate deprivation in women and men: effects on mineral and acid balances, parathyroid hormone and the metabolism of 25-OH-vitamin D. J Clin Endocrinol Metab 1976; 43: 1056-1068.
- 3 Gattineni J, Bates C, Twombley K et al. FGF23 decreases renal NaPi-2a and NaPi-2c expression and induces hypophosphatemia in vivo predominantly via FGF receptor 1. Am J Physiol Renal Physiol 2009; 297: F282-F291.
- Bergwitz C, Jüppner H. Regulation of phosphate homeostasis by PTH, 4. vitamin D, and FGF23. Annu Rev Med 2010; 61: 91-104.
- Gutierrez O, Isakova T, Rhee E et al. Fibroblast growth factor-23 mitigates hyperphosphatemia but accentuates calcitriol deficiency in chronic kidney disease. J Am Soc Nephrol 2005; 16: 2205-2215.
- 6. Bricker NS. On the pathogenesis of the uremic state. An exposition of the 'trade-off hypothesis'. N Engl J Med 1972; 286: 1093-1099.
- Jüppner H, Bastepe M. Different mutations within or upstream of the 7. GNAS locus cause distinct forms of pseudohypoparathyroidism. J Pediatr Endocrinol Metab 2006; 19(Suppl 2): 641-646.

- 8. Kolek OI, Hines ER, Jones MD et al. 1α,25-Dihydroxyvitamin D₃ upregulates FGF23 gene expression in bone: the final link in a renal-gastrointestinal-skeletal axis that controls phosphate transport. Am J Physiol Gastrointest Liver Physiol 2005; 289: G1036-G1042.
- 9. Farrow EG, Davis SI, Summers LJ et al. Initial FGF23-mediated signaling occurs in the distal convoluted tubule. J Am Soc Nephrol 2009; 20: 955-960.
- Westerberg PA, Ljunggren O, Larsson TE et al. Fibroblast growth factor-23 10. and mineral metabolism after unilateral nephrectomy. Nephrol Dial Transplant 2010; 25: 4068-4071.
- Hasegawa H, Nagano N, Urakawa I et al. Direct evidence for a causative 11. role of FGF23 in the abnormal renal phosphate handling and vitamin D metabolism in rats with early-stage chronic kidney disease. Kidney Int 2010; 78: 975-980.
- 12. van Husen M, Fischer AK, Lehnhardt A et al. Fibroblast growth factor 23 and bone metabolism in children with chronic kidney disease. Kidney Int 2010; 78: 200-206.
- 13. Pereira RC, Jüppner H, Azucena-Serrano CE et al. Patterns of FGF-23, DMP1, and MEPE expression in patients with chronic kidney disease. Bone 2009; 45: 1161-1168.
- 14. Fliser D, Kollerits B, Neyer U et al. Fibroblast growth factor 23 (FGF23) predicts progression of chronic kidney disease: the Mild to Moderate Kidney Disease (MMKD) Study. J Am Soc Nephrol 2007; 18: 2600-2608.
- 15. Titan SM, Zatz R, Jorgetti V et al. FGF-23 as a predictor of renal outcome in diabetic nephropathy. (Renal week 2009 abstract F-PO1872). J Am Soc Nephrol 2009; 20: 540A.
- 16. Gutiérrez OM, Januzzi JL, Isakova T et al. Fibroblast growth factor 23 and left ventricular hypertrophy in chronic kidney disease. Circulation 2009; 119: 2545-2552.
- Nasrallah MM, El-Shehaby AR, Salem MM et al. Fibroblast growth factor-17. 23 (FGF-23) is independently correlated to aortic calcification in haemodialysis patients. Nephrol Dial Transplant 2010; 25: 2679-2685.
- 18. Perwad F, Azam N, Zhang MY et al. Dietary and serum phosphorus regulate fibroblast growth factor 23 expression and 1,25-dihydroxyvitamin D metabolism in mice. Endocrinology 2005; 146: 5358-5364.
- Burnett SM, Gunawardene SC, Bringhurst FR et al. Regulation of 19. C-terminal and intact FGF-23 by dietary phosphate in men and women. J Bone Miner Res 2006; 21: 1187-1196.
- 20. Isakova T, Gutierrez OM, Wolf M. A blueprint for randomized trials targeting phosphorus metabolism in chronic kidney disease. Kidney Int 2009; 76: 705–716.
- 21. Shimada T, Urakawa I, Isakova T et al. Circulating fibroblast growth factor 23 in patients with end-stage renal disease treated by peritoneal dialysis is intact and biologically active. J Clin Endocrinol Metab 2010; 95: 578-585
- 22. Heijboer AC, Levitus M, Vervloet MG et al. Determination of fibroblast growth factor 23. Ann Clin Biochem 2009; 46: 338-340.
- 23 Gutiérrez OM, Mannstadt M, Isakova T et al. Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. N Engl J Med 2008; 359: 584-592.
- 24. Jean G, Terrat JC, Vanel T et al. High levels of serum fibroblast growth factor (FGF)-23 are associated with increased mortality in long haemodialysis patients. Nephrol Dial Transplant 2009; 24: 2792-2796.
- 25. Nagai S, Okazaki M, Segawa H et al. Acute down-regulation of sodiumdependent phosphate transporter NPT2a involves predominantly the cAMP/PKA pathway as revealed by signaling-selective parathyroid hormone analogs. J Biol Chem 2011; 286: 1618-1626.
- Cancela AL, Oliveira RB, Graciolli FG et al. Fibroblast growth factor 23 in 26. hemodialysis patients: effects of phosphate binder, calcitriol and calcium concentration in the dialysate. Nephron Clin Pract 2011; 117: c74-c82.

This work is licensed under the Creative Com-SOME RIGHTS RESERVED

mons Attribution-NonCommercial-Share Alike 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-sa/3.0/

60)