





## **Draft Genome Sequence of a Filamentous Anoxygenic** Phototrophic Bacterium, "Candidatus Roseilinea sp. Strain NK\_OTU-006," Recovered from Metagenomic Data of a Hot **Spring Microbial Mat**

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ABSTRACT We report here the metagenome-assembled draft genome of an uncultured filamentous anoxygenic phototroph of the phylum Chloroflexota (formerly Chloroflexi), "Candidatus Roseilinea sp. strain NK\_OTU-006," recovered from hot spring-associated microbial mats. The 3.6-Mb genome is estimated to be 94% complete and comprises 117 contigs encoding 3,203 predicted genes, including a full-length rRNA operon.

Il known culturable filamentous anoxygenic phototrophic (FAP) bacteria within the phylum Chloroflexota are affiliated with the class Chloroflexia. However, metagenomic studies of hot spring-associated microbial mats discovered a novel FAP outside this class (1, 2). This still-uncultured organism was first described as an Anaerolinea-like bacterium and was later tentatively named "Candidatus Roseilinea gracile" (3). A recent evolutionary study of a related uncultured representative, the metagenome-assembled genome (MAG) JP3\_7, proposed that both phototrophs from geographically different hot springs belong to the same novel "Candidatus" class, "Candidatus Thermofonsia," of which no validly described species have been reported so far (4). Here, we report the draft genome of a third hot spring-associated MAG representative of this novel class, "Candidatus Roseilinea sp. strain NK\_OTU-006."

Phototrophic microbial mats were collected from Nakabusa hot springs in Nagano Prefecture, Japan (36°23'33.1"N, 137°44'53.0"E), at an overlying water temperature of  $\sim$ 56°C. Genomic DNA was isolated from the mats using a PowerBiofilm DNA extraction kit (Qiagen, Inc.). Sequencing libraries were prepared using a KAPA Hyperplus library prep kit (Kapa Biosystems, Inc.) and were sequenced via an Illumina MiSeg instrument (2  $\times$  300 bp) at FASMAC (Japan). Raw metagenome sequences (32,921,964 reads) were quality filtered using Sickle v.1.33 (5), trimmed using FASTX-Toolkit v.0.0.13.2 (6), and assembled using SPAdes v.3.7.1 (7) with default parameters. Tetranucleotide binning of assembled contigs was employed using Emergent Self-Organizing Map (ESOM) as described previously (8). Bin identity and completeness were assessed based on the presence of phylogenetic marker genes and taxonomic affiliations implemented in AmphoraNet (9) and CheckM (10).

The 3.6-Mb draft genome sequence consists of 63% G+C content and is estimated to be 94.1% complete, as determined by CheckM (10). Genome annotation by Prokka v.1.2 (11) comprises 117 contigs, with 3,203 predicted genes, 3,061 protein-coding sequences, 44 tRNAs, and 3 rRNAs (5S, 16S, 23S). The full-length 16S rRNA gene Citation Martinez JN, Kawai S, Saini MK, Tank M, Hanada S, Thiel V. 2020. Draft genome sequence of a filamentous anoxygenic phototrophic bacterium, "Candidatus Roseilinea sp. strain NK\_OTU-006," recovered from metagenomic data of a hot spring microbial mat. Microbiol Resour Announc 9:e01104-20. https://doi.org/10.1128/MRA .01104-20.

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Martinez et al.

sequence from the NK\_OTU-006 bin shows the highest identity (95%) with "Ca. Roseilinea gracile" (3). However, the average nucleotide identity (ANI) values show 78% and 99% similarities to "Ca. Roseilinea gracile" and JP3\_7 MAGs, respectively. NK\_OTU-006 also shows low ANI ( $\leq$ 78%) and 16S rRNA gene sequence identity ( $\leq$ 87%) to any described isolates from the classes Anaerolinea (12, 13) and Chloroflexia (14, 15).

MAG NK\_OTU-006 contains genes encoding a type 2 photosynthetic reaction center (pufLMC) and light-harvesting complex 1 (pufAB), which is consistent with genes found in "Ca. Roseilinea gracile" (3) and JP3\_7 MAGs (4). Similar to the red filamentous anoxygenic phototroph Roseiflexus spp. (14, 16), no genes encoding chlorosomes were recovered, while genes required for bacteriochlorophyll a and carotenoid biosynthesis were detected. Homologous genes encoding the major electron transport complexes for aerobic respiration and ACIII-like genes needed for phototrophy were also found. This finding is consistent with other those for phototrophic Chloroflexia (16-18).

Unlike many phototrophic Chloroflexia members, no genes for any known autotrophic carbon fixation pathways were detected, indicating a heterotrophic lifestyle (14-16). Genes for sulfide oxidation and nitrogen fixation were not recovered from the binned genome. However, uptake and bidirectional hydrogenase genes were found in the MAG, suggesting the ability to metabolize H<sub>2</sub>. The genomic information of "Ca. Roseilinea sp. NK OTU-006" facilitates understanding of the lifestyle of the underrepresented lineage of phototrophic Chloroflexota. A metabolic model is accessible at https://kbase.us/n/59604/258/.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number NZ\_JAAFGM000000000. The raw sequence reads are available in the Sequence Read Archive (SRA) under database accession number SRR12784215 and BioProject accession number PRJNA603154.

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## **REFERENCES**

- 1. Klatt CG, Wood JM, Rusch DB, Bateson MM, Hamamura N, Heidelberg JF, Grossman AR, Bhaya D, Cohan FM, Kühl M, Bryant DA, Ward DM. 2011. Community ecology of hot spring cyanobacterial mats: predominant populations and their functional potential. ISME J 5:1262-1278. https:// doi.org/10.1038/ismej.2011.73.
- 2. Klatt CG, Inskeep WP, Herrgard MJ, Jay ZJ, Rusch DB, Tringe SG, Niki Parenteau M, Ward DM, Boomer SM, Bryant DA, Miller SR. 2013. Community structure and function of high-temperature chlorophototrophic microbial mats inhabiting diverse geothermal environments. Front Microbiol 4:106. https://doi.org/10.3389/fmicb.2013.00106.
- 3. Tank M, Thiel V, Ward DM, Bryant DA. 2017. A panoply of phototrophs: an overview of the thermophilic chlorophototrophs of the microbial mats of alkaline siliceous hot springs in Yellowstone National Park, WY, USA, p 87–137. In Hallenbeck P (ed), Modern topics in the phototrophic prokaryotes. Springer International Publishing, Cham, Switzerland.
- 4. Ward LM, Hemp J, Shih PM, McGlynn SE, Fischer WW. 2018. Evolution of phototrophy in the Chloroflexi phylum driven by horizontal gene transfer. Front Microbiol 9:260. https://doi.org/10.3389/fmicb.2018.00260.
- 5. Joshi N, Fass J. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files (Version 1.33) [Software]. https://github .com/najoshi/sickle.
- 6. Gordon A, Hannon GJ. 2010. FASTX-Toolkit. http://hannonlab.cshl.edu/ fastx\_toolkit/.
- 7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N,

- Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455-477. https://doi.org/10.1089/cmb.2012.0021.
- 8. Dick GJ, Andersson AF, Baker BJ, Simmons SL, Thomas BC, Yelton AP, Banfield JF. 2009. Community-wide analysis of microbial genome sequence signatures. Genome Biol 10:R85. https://doi.org/10.1186/gb-2009-10-8-r85.
- 9. Kerepesi C, Bánky D, Grolmusz V. 2014. AmphoraNet: the webserver implementation of the AMPHORA2 metagenomic workflow suite. Gene 533:538-540. https://doi.org/10.1016/j.gene.2013.10.015.
- 10. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043-1055. https:// doi.org/10.1101/gr.186072.114.
- 11. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- 12. Grégoire P, Fardeau ML, Joseph M, Guasco S, Hamaide F, Biasutti S, Michotey V, Bonin P, Ollivier B. 2011. Isolation and characterization of Thermanaerothrix daxensis gen. nov., sp. nov., a thermophilic anaerobic bacterium pertaining to the phylum "Chloroflexi", isolated from a deep hot aquifer in the Aquitaine Basin. Syst Appl Microbiol 34:494-497. https:// doi.org/10.1016/j.syapm.2011.02.004.
- 13. Sekiguchi Y, Yamada T, Hanada S, Ohashi A, Harada H, Kamagata Y. 2003. Anaerolinea thermophila gen. nov., sp. nov. and Caldilinea aerophila gen. nov., sp. nov., novel filamentous thermophiles that represent a previously



- uncultured lineage of the domain Bacteria at the subphylum level. Int J Syst Evol Microbiol 53:1843–1851. https://doi.org/10.1099/ijs.0.02699-0.
- 14. Hanada S, Takaichi S, Matsuura K, Nakamura K. 2002. Roseiflexus castenholzii gen. nov., sp. nov., a thermophilic, filamentous, photosynthetic bacterium that lacks chlorosomes. Int J Syst Evol Microbiol 52:187-193. https://doi.org/10.1099/00207713-52-1-187.
- 15. Hanada S, Hiraishi A, Shimada K, Matsuura K. 1995. *Chloroflexus aggregans* sp. nov., a filamentous phototrophic bacterium which forms dense cell aggregates by active gliding movement. Int J Syst Bacteriol 45:676-681. https://doi.org/10.1099/00207713-45-4-676.
- 16. Klatt CG, Liu Z, Ludwig M, Kühl M, Jensen SI, Bryant DA, Ward DM. 2013. Temporal metatranscriptomic patterning in phototrophic Chloroflexi
- inhabiting a microbial mat in a geothermal spring. ISME J 7:1775–1789. https://doi.org/10.1038/ismej.2013.52.
- 17. Tang K-H, Barry K, Chertkov O, Dalin E, Han CS, Hauser LJ, Honchak BM, Karbach LE, Land ML, Lapidus A, Larimer FW, Mikhailova N, Pitluck S, Pierson BK, Blankenship RE. 2011. Complete genome sequence of the filamentous anoxygenic phototrophic bacterium Chloroflexus aurantiacus. BMC Genomics 12:334. https://doi.org/10.1186/1471-2164-12-334.
- 18. Refojo PN, Ribeiro MA, Calisto F, Teixeira M, Pereira MM. 2013. Structural composition of alternative complex III: variations on the same theme. Biochim Biophys Acta 1827:1378–1382. https://doi.org/10.1016/j.bbabio.2013 .01.001.