



Reactive oxygen species in the world ocean and their impacts on marine ecosystems

J. Jeffrey Morris^{a,*}, Andrew L. Rose^b, Zhiying Lu^a

^a Department of Biology, University of Alabama at Birmingham, Birmingham, AL, USA

^b Faculty of Science and Engineering, Southern Cross University, New South Wales, Australia

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ABSTRACT

Reactive oxygen species (ROS) are omnipresent in the ocean, originating from both biological (e.g., unbalanced metabolism or stress) and non-biological processes (e.g. photooxidation of colored dissolved organic matter). ROS can directly affect the growth of marine organisms, and can also influence marine biogeochemistry, thus indirectly impacting the availability of nutrients and food sources. Microbial communities and evolution are shaped by marine ROS, and in turn microorganisms influence steady-state ROS concentrations by acting as the predominant sink for marine ROS. Through their interactions with trace metals and organic matter, ROS can enhance microbial growth, but ROS can also attack biological macromolecules, causing extensive modifications with deleterious results. Several biogeochemically important taxa are vulnerable to very low ROS concentrations within the ranges measured in situ, including the globally distributed marine cyanobacterium *Prochlorococcus* and ammonia-oxidizing archaea of the phylum *Thaumarchaeota*. Finally, climate change may increase the amount of ROS in the ocean, especially in the most productive surface layers. In this review, we explore the sources of ROS and their roles in the oceans, how the dynamics of ROS might change in the future, and how this change might impact the ecology and chemistry of the future ocean.

1. Introduction

The distribution, abundance, and productivity of organisms in the World Ocean is intimately tied to biogeochemical cycles. Most marine ecosystems are dependent on primary production by aquatic photoautotrophs such as diatoms, cyanobacteria, seaweeds and seagrasses, and the growth of these organisms is in turn limited by the availability of nitrogen and/or phosphorus, or in some cases trace nutrients such as iron or vitamins. These molecules exist in a variety of chemical forms in the ocean, and because of their foundational importance in marine ecology, the enzymatic and abiotic reactions that shuttle them between their various states have been studied extensively.

Despite its ubiquity in both the ocean environment and in living biomass, the cycling of oxygen has received comparatively little attention from marine ecologists. The oxygen cycle is routinely imagined as a simple back-and-forth between the release of O₂ as a by-product of photosynthetic water-splitting and the reduction of O₂ to water by aerobic heterotrophic metabolism. But this formulation underestimates the complexity of environmental reactions involving oxygen, which in fact cycles continually through a variety of oxygen-containing

intermediates known as reactive oxygen species (ROS) (see Ref. [1] for an overview). Sometimes these ROS act as potent toxins; sometimes they are created by organisms as part of their natural growth processes; and in many cases they interact with other biogeochemical cycles in critical ways [1–3].

In this review, we will explore the roles played by ROS in the oceans. We will begin with a description of the chemistry of ROS and a survey of the biotic and abiotic mechanisms that generate them, follow up with an exploration of how ROS interact with marine biogeochemical cycles, and then consider some specific positive and negative interactions ROS have with marine organisms. Finally, we will close with a consideration of how the dynamics of ROS might change in the future due to human activity, and how this might impact the ecology and chemistry of the future ocean.

2. Aquatic chemistry of ROS

ROS are oxygen-containing molecules in which the redox state of oxygen is intermediate between that of O₂ and H₂O, which form the dominant redox couple in most oxic natural waters. The co-existence of

* Corresponding author.

E-mail addresses: evolve@uab.edu (J.J. Morris), luzy05111036@yahoo.com (Z. Lu).

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O₂ and H₂O, despite their very different redox potentials, is a thermodynamically unfavorable situation that is enabled by two primary factors [4]:

1. O₂ is relatively unreactive because its most energetically favorable state (the ground state) is a triplet state with two unpaired electrons; and
2. The continuous input of free energy into the global aquatic system, primarily due to sunlight-driven photo(bio)chemical reactions, prevents the system from reaching a thermodynamic endpoint in which O₂ is fully consumed.

The triplet ground state of O₂ (denoted ³O₂) means that it is typically only able to react with other ground state molecules containing unpaired electrons, which forces the reduction of O₂ to occur via a series of one-electron transfer steps [4]. Intracellularly, these one-electron transfer steps are often mediated by enzymes such that they occur in quick succession and the intermediate reaction products are not released [5]. In contrast, in the extracellular milieu of natural aquatic systems the sequence of one-electron transfer steps during the reduction of O₂ can give rise to a range of ROS in free solution.

2.1. Thermodynamics

In oxic waters the overall thermodynamic drive is primarily for reduction of O₂ (and ROS) to H₂O, coupled to oxidation of other chemical species [3]. In other words, ROS are predominantly oxidants. Nonetheless, there are circumstances in which some ROS can also function as reductants and drive redox cycling of various biogeochemically important elements [3]. While ROS may often be found in free solution in the aquatic milieu as noted above, there are circumstances in which they are bound to other species, either because they are not released as free species during formation, or due to additional reactions with other suitable substrates [4]. In this case, the resulting bound species may also function as oxidants or facilitate redox cycling.

This stepwise reduction of O₂ to H₂O occurs via the following series of reactions [4]:



The thermodynamics of this sequence of redox reactions is illustrated in Fig. 1 under standard reaction conditions (unit activity and a temperature of 298 K) at pH 7.

As illustrated in Fig. 1, O₂^{•−} is mildly reducing under standard reaction conditions at pH 7. In contrast, H₂O₂ is a relatively powerful two-electron oxidant from a thermodynamic perspective, and is not constrained to one-electron reactions in the same way as ³O₂, but typically must overcome a large activation energy barrier [6]. Consequently, H₂O₂ reacts mostly as a one electron oxidant. One of the most well-known such reactions is the so-called Fenton reaction with Fe(II):



where Mⁿ⁺ is Fe(II) in the case of the Fenton reaction. This reaction, and similar “Fenton-like” reactions with other trace metals and certain non-metals, produce •OH or high-valent metal ions [6], both of which are very strong and highly reactive oxidants.

In addition to these species formed during reduction of O₂, the electronically excited ¹Δ_g singlet state of dioxygen is typically also considered as one of the ROS. While two electronically excited states of dioxygen are possible, the highest energy ¹Σ_g⁺ singlet state relaxes back

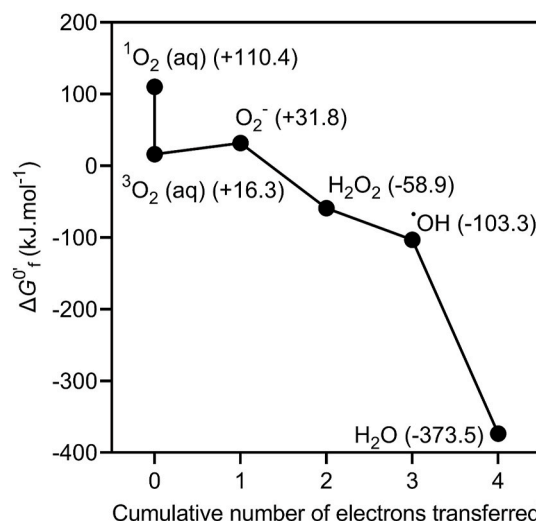


Fig. 1. Gibbs free energy of formation of ROS under standard conditions (1 M activity) in aqueous solution at pH 7. ¹O₂ refers to the ¹Δ_g excited state of dioxygen. Values were calculated from thermodynamic data reported in Sawyer (1991) [4].

to the ¹Δ_g state faster than it can undergo bimolecular reaction; hence the common notation ¹O₂ is used in this and other work in reference to the latter. While ¹O₂ is a relatively powerful oxidant, and not subject to the one-electron transfer constraints of ³O₂, in practice it is highly selective as an oxidant because it readily relaxes back to the ³O₂ ground state through either energy transfer (particularly to water in aqueous environments), or by partial charge transfer in which formation of an intermediate charge-transfer complex results in relaxation of ¹O₂ back to ³O₂ without completion of electron transfer [7].

When electron transfer is required to occur in single electron transfer steps, additional thermodynamic constraints are imposed on the system: not only must the overall reaction be thermodynamically favorable, but so too must each one-electron transfer step, else the reaction will be unable to proceed beyond the thermodynamic barrier imposed by a particular electron transfer step. Consequently, there are a limited set of redox reactions that are thermodynamically possible in oxic marine waters [3], as illustrated in Fig. 2. From a thermodynamic perspective, this explains why some intermediates in the redox reactions of various biogeochemically important species are only ever transient, and present at exceedingly low concentrations. While Fig. 2 does not provide an exhaustive examination of all possible redox couples that may be influenced by ROS, it does give an indication of some of the most important.

2.2. Reaction kinetics

Although thermodynamics is a critical control on the energetic favorability of potential reactions involving ROS, marine systems are dynamic and in many cases equilibrium is never reached. As such, the biogeochemistry of ROS is strongly influenced by reaction kinetics, which control their lifetimes and hence steady-state concentrations. The reactivity of the different ROS varies considerably. At the pH of seawater, O₂^{•−} exists predominantly in the deprotonated form, which possesses a resonance-stabilized form with a “three-electron bond”, and exhibits very little free radical character [5]. As such, it is relatively stable in marine waters and only reacts with other species possessing unpaired electrons in the valence shell, resulting in half-lives on the order of a few seconds to minutes in marine waters [1]. The reactivity of H₂O₂ is usually kinetically limited due to the high activation energy barrier noted previously, with the overall reaction kinetics often controlled by rate of the formation of reaction intermediates that lower

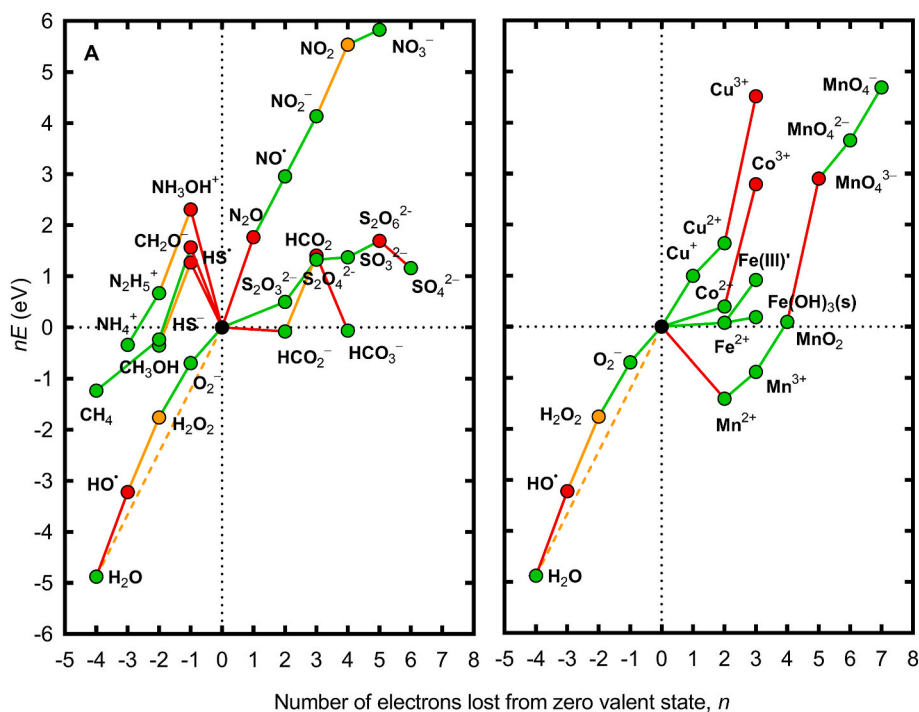


Fig. 2. Frost diagrams for some common redox active elements in natural waters at pH 8.1 accounting for typical activities of oxygen redox species in natural waters. (A) Representative species for the major elements C, N and S. (B) Trace metals Co, Cu, Fe and Mn. Red lines indicate redox couples that are absolutely unstable with respect to one-electron transfer processes involving the $\text{H}_2\text{O}/\cdot\text{OH}$ couple (lines with positive slope) or the H_2/H^+ couple (lines with negative slope); orange lines indicate redox couples that are conditionally unstable with respect to multi-electron transfer processes involving the $\text{H}_2\text{O}/\text{O}_2$ couple; and green lines indicate redox couples that are thermodynamically permissible in aqueous solution at pH 8.1. The dashed orange line represents the redox potential associated with the four-electron transfer for complete reduction of O_2 to H_2O . Red symbols represent species that are thermodynamically unable to exist due to absolute instability with respect to one-electron transfer processes involving the $\text{H}_2\text{O}/\cdot\text{OH}$ couple (lines with positive slope) or the H_2/H^+ couple; orange symbols represent species that are thermodynamically able to exist in the absence of processes to catalyze multi-electron transfer processes involving the $\text{H}_2\text{O}/\text{O}_2$ couple; and green symbols represent species that are thermodynamically able to coexist with both the H_2/H^+ and $\text{H}_2\text{O}/\text{O}_2$ couples. See Ref. [3] for details of concentrations and thermodynamic data used. Reproduced from Ref. [3] with permission.

the activation energy barrier [6]. For example, the Fenton reaction is thought to involve formation of an inner sphere complex between Fe(II) and H_2O_2 as the initial step, resulting in an overall reaction rate constant many orders of magnitude less than diffusion-controlled [6]. Consequently, typical half-lives of H_2O_2 in marine waters are considerably longer, on the order of hours to days [1]. In contrast, $\cdot\text{OH}$ reacts at diffusion-controlled rates with many organic and inorganic compounds, and as such typically has a very short lifetime in natural waters (half-life ~ 250 ns, computed using data from Ref. [8]). Similarly, $^1\text{O}_2$ is highly reactive in aqueous environments with a half-life of around $4 \mu\text{s}$ due to physical relaxation by solvent water [9].

Short half-lives and localized sources (as discussed further in Section III) may also result in spatially heterogeneous distributions of ROS. This is particularly the case for $\cdot\text{OH}$ and $^1\text{O}_2$, but also to some extent for $\text{O}_2^{\cdot-}$, which may be formed through highly localized processes (e.g. in cells or at particle surfaces) and unable to diffuse far from the site of production due to their short half-life. This can result in significant spatial variability in steady-state concentrations, with the potential for concentrations near the site of ROS production that are many orders of magnitude greater than in the bulk solution, and associated differences in reaction kinetics.

3. Sources and sinks of ROS in the ocean

Naturally occurring H_2O_2 in the ocean was first reported in the 1960's [10], and even earlier than that, researchers were aware that H_2O_2 added to seawater was not stable, but disappeared with enzyme-like kinetics [11]. Nevertheless, the ability to study the distribution, sources, and sinks of ROS in the ocean awaited the development of sufficiently sensitive detection methods which would not occur until the 1980's. For instance, classical titration-based methods for quantifying H_2O_2 had limits of detection in the millimolar range, but the development of fluorescence methods in the 1980s [12] and chemiluminescence methods in the 2000s [13] made it possible to accurately measure the nanomolar H_2O_2 commonly found in natural waters (Fig. 4). In this section we first give an overview of the major ROS production and destruction pathways along with their impact on

steady-state ROS concentrations, and finally we describe some of the most common methods used for assessing seawater ROS concentrations in the field.

3.1. Sources of marine ROS

3.1.1. Photochemical production

Many studies in a wide variety of waters have supported the hypothesis that most ROS in bulk surface waters arise by photochemistry, especially via the photooxidation of dissolved organic matter (DOM) [14–17]. Colored DOM can absorb photons and enter an excited triplet state capable of reducing O_2 to $\text{O}_2^{\cdot-}$, which can then undergo spontaneous dismutation to H_2O_2 and O_2 . Because of its relatively long residence time, H_2O_2 is the most easily measured ROS in the field and has been used as a proxy for the relative abundance of other ROS such as $\text{O}_2^{\cdot-}$ and $\cdot\text{OH}$. The rate of H_2O_2 accumulation is generally correlated with DOM concentration [18], and therefore steady-state H_2O_2 concentrations are generally greater in terrestrially-impacted waters like coasts and estuaries, as opposed to the oligotrophic ocean gyres (Table 1 and references therein). ROS fluxes in bulk water and H_2O_2 steady-state concentrations also decrease rapidly with depth and light level, dropping to near or below the limit of detection below the euphotic zone (Fig. 3a, Table 1). Because of the influence of sunlight, steady state H_2O_2 concentrations in surface waters tend to exhibit a diel cycle with afternoon maxima and early morning minima (Fig. 3c) [19–21], and also a seasonal cycle peaking in the summer (Fig. 3b) [22,23]. Such trends have not been widely observed for other ROS, however.

H_2O_2 production is stimulated by light in the visual range but is much faster under UV irradiation. While representing only a small fraction of the irradiance impacting natural seawater, photons in the UV-B range cause the majority of H_2O_2 accumulation [24]. H_2O_2 can also form in the atmosphere by the photolysis of water into hydrogen atoms and $\cdot\text{OH}$ [25]. These radicals can then join to form H_2O_2 in the gas phase, which can subsequently be recruited into rain droplets, resulting in H_2O_2 concentrations that are typically two orders of magnitude greater than that found in surface seawater [26]. Rain events have been shown to temporarily increase the in situ H_2O_2 of seawater [26–32].

Table 1
H₂O₂ concentrations in various waters.

Type	Location	[H ₂ O ₂]	Reference
Freshwater	Jacks Lake, Ontario, surface	10–800 nM	[19]
	Swedish Highland Lakes, surface	132–984 nM	[247]
	Lake Erie, surface	50–175 nM	[248]
Brackish	Patuxent Estuary, surface	12–350 nM	[22]
	Chesapeake Bay, surface	3–1700 nM	[15]
	Chesapeake Bay, surface	40–80 nM	[248]
	Coastal Germany, Intertidal salt flat	0.1–4.5 μM	[23]
Coastal	Southern Ocean	1.5 μM	[249]
	Antarctica	9–25 nM	[250]
	Peru	80–500 nM	[251]
	Southern California	44–370 nM	[252]
	Baja California	50–125 nM	[248]
	Texas	14–170 nM	[10]
	Florida	80–210 nM	[15]
	Gulf of Mexico	100–240 nM	[253]
	Bahamas	50–190 nM	[15]
	Amazon River plume	25–165 nM	[21]
Open ocean surface	Tropical Pacific	50–150 nM	[200]
	Tropical Pacific, during rain	100–300 nM	[28]
	North Pacific	25–120 nM	[254]
	South and Central Atlantic	16–68 nM	[21]
	Subtropical Atlantic	50–220 nM	[18]
	Equatorial Atlantic ITCZ	31–236 nM	[255]
	Caribbean Sea	50–100 nM	[256]
	Gulf of Mexico	90–140 nM	[253]
	Sargasso Sea	95–175 nM	[257]
	Western Atlantic	40–175 nM	[248]
	Eastern Atlantic	30–80 nM	[258]
	North Atlantic	135–483 nM	[32]
	Bermuda Atlantic Time Series	20–200 nM	[31]
	Mediterranean Sea	100–140 nM	[259]
Southern Ocean Indian Sector	5–20 nM	[260]	
Open ocean depths	North Pacific, >1000 m	<6 nM	[261]
	Mediterranean, South Atlantic and Pacific, >1000 m	<3 nM	[262]
	South and Central Atlantic, below euphotic zone	<1 nM	[21]
	Bermuda Atlantic Time Series, 150 m	<2 nM	[31]
Precipitation	South and Central Atlantic, rain	3.5–71 μM	[27]
	Equatorial Atlantic ITCZ	1.5–22.8 μM	[255]

Table 1 (continued)

Type	Location	[H ₂ O ₂]	Reference
	Gulf of Mexico, rain	40.2 μM	[263]
	Florida Keys, rain	28.4 μM	[263]
	Miami/Bahamas, rain	30.7 μM	[264]
	West Atlantic, rain	12.7 μM	[263]
	Bermuda Atlantic Time Series, rain	11–40 μM	[31]
	Bermuda Atlantic Time Series, rain	5–85 μM	[26]
	New Zealand, rain	10–30 μM	[30]
	Equatorial Pacific, rain	8.5 μM	[28]
	North Pacific ITCZ, rain	10.4 μM	[28]
	Jacks Lake, Ontario, rain	1.3–34 μM	[19]
	Southern Ocean, snow	10–14 μM	[249]

3.1.2. Stress production by organisms

In addition to direct photochemical ROS production, marine organisms can release H₂O₂ into the environment, often due to stress. Many enzymes capable of single-electron transfer to a substrate are also capable of reducing O₂, and the ubiquity of O₂ in oxic environments ensures that some H₂O₂ will escape. Common metabolic pathways that produce H₂O₂ as a by-product even under ideal conditions include the Mehler reaction (aka the water-water cycle) and photorespiration in photosynthetic organisms [33] as well as the electron transfer reactions of aerobic oxidative phosphorylation [34]. O₂ reduction is enhanced when a primary electron transport carrier is not available in sufficient quantities due either to excess electron flow, nutrient deficiency, or other problems. For instance, in Fe-starved cyanobacteria, ROS buildup occurs due to a lack of ferredoxin, the Fe-containing primary cytoplasmic electron acceptor for photosystem I [35]. When more electrons flow through the photosynthetic electron transport chain than can be accepted by ferredoxin, O₂ reduction becomes practically inevitable [33], and therefore excess ROS production and leakage of H₂O₂ into the environment is likely to be common in low-Fe marine habitats, particularly near the ocean surface under conditions of high light [36]. Release of H₂O₂ has also been observed in Antarctic diatoms when suddenly exposed to light following the long periods of uninterrupted darkness characteristic of the polar winter [37]. Importantly, H₂O₂ release by light-stressed photoautotrophs may contribute to the diel cycle in H₂O₂ observed at the ocean's surface, leading to overestimates of the role of DOM photooxidation in ROS production. ROS buildup from unbalanced metabolism could also explain the toxicity of high-nutrient culture media to marine microorganisms adapted to oligotrophic lifestyles [38].

Low levels of ROS are continuously produced by many organisms for signaling purposes and as normal, short-lived metabolic intermediates. However, intracellular ROS accumulation – a condition generally referred to as oxidative stress – can occur as a general consequence of diverse stresses such as temperature, osmotic, and dehydration stress [39–44]. Exposure to anthropogenic pollutants can also induce oxidative stress in marine organisms. For example, silver nanoparticles and microplastics like polyacrylonitrile induced lethal oxidative stress in the photosynthetic microorganisms *Prochlorococcus* and *Chlorella pyrenoidosa*, respectively [45,46]. Exposure to heavy metals, elevated temperatures, acidification, organic pollutants, allelochemicals, and other stressors lead to ROS accumulation and other biomarkers of oxidative stress (e.g. DNA strand breaks) in a wide variety of marine animals, plants, and microbes [45–57]. However, it is not clear how much of this intracellular ROS is able to escape the organism and contribute to bulk ROS concentrations in the environment. O₂^{•−}, for instance, is incapable of crossing cellular membranes due to its charge, while highly reactive ROS such as [•]OH have such short lifespans that they are unlikely to move far from their point of origin.

3.1.3. Extracellular ROS production by marine organisms

ROS are also released into the environment by healthy organisms as a normal part of their metabolism. For example, animals use H₂O₂ at low

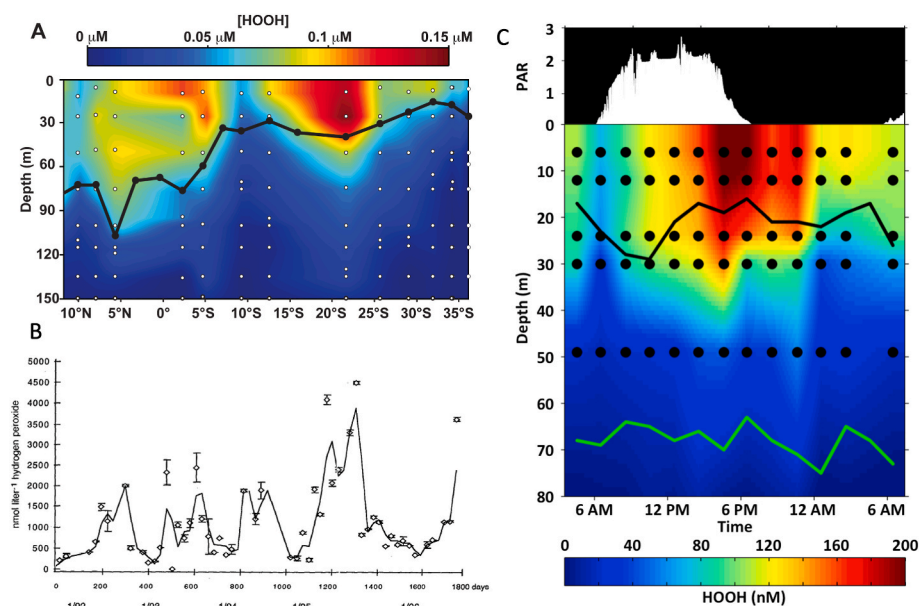


Fig. 3. H_2O_2 profiles across time and space. A) H_2O_2 depth profiles from samples across a Pacific transect in Jan–Feb 2007. B) Monthly H_2O_2 measurements of a tide pool in Germany from 1991 to 1996. C) H_2O_2 depth profiles and light intensity from a station in the South Pacific (Feb 2007) across a 24-h period; PAR, photosynthetically active radiation, with white bars indicating light intensity. In A and C dots represent sampled depths/times, with H_2O_2 values interpolated between the points; black lines represent the depth of the surface mixed layer; and green line in C represents the deep chlorophyll maximum. Reprinted with permission from Ref. [200] (A) [273], (B), and [20] (C).

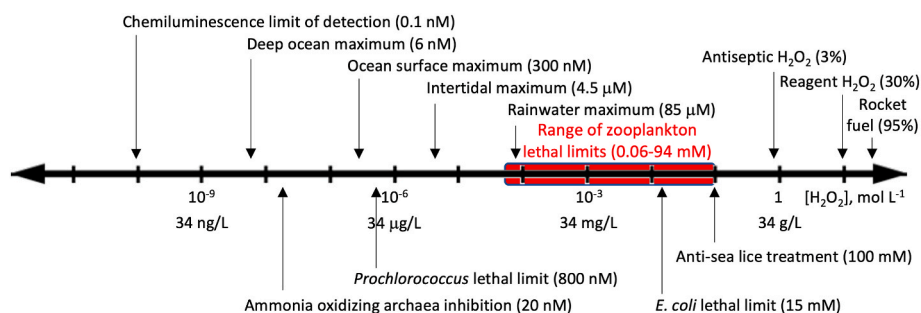


Fig. 4. H_2O_2 concentrations in different environments and their biological impacts on various organisms.

concentrations as a signaling compound [58] and in high concentrations as an antimicrobial defense [59,60]. The rapid production of H_2O_2 bursts, often facilitated by NADPH-dependent oxidases, is an important component of the innate immune system of land plants and is also observed in aquatic phototrophs, where oxidative bursts have been observed after wounding or exposure to grazers or pathogens in a wide variety of macroalgae [61–66].

H_2O_2 may also be produced as a weapon against prey or parasites. For instance, the coral *Stylophora pistillata* responded to physical stimulation (e.g. by pushing against its tissue) by releasing H_2O_2 from the site of stimulation, and the amount of H_2O_2 released was proportional to the intensity of the stimulus [67]. Contact with prey organisms also stimulated H_2O_2 release. The production of H_2O_2 was unaffected by the presence or absence of zooxanthellae, indicating it was a native function of the cnidarian organism, possibly serving as a protection against pathogens or a way to stun and capture prey more effectively. Similarly, H_2O_2 released as a response to wounding in the Antarctic seaweeds *Ascoseira mirabilis* and *Saccharina latissima* was able to inhibit grazing by amphipods, the primary herbivore feeding on these organisms in their native habitat [68,69].

Over the past decade, a variety of studies have also shown that extracellular production of $\text{O}_2^{\bullet-}$ by marine microbes is nearly ubiquitous (e.g. Ref. [70]). This body of work has been summarized in detail in several excellent recent reviews [1,2,71] and will not be repeated here. However, it is worth emphasizing that many of these studies have suggested that this process may be “intentional”, in the sense that the process may be exploited and even tightly regulated [72] to create or

maintain locally favorable biogeochemical conditions. For example, one of the most significant and potentially widespread uses of ROS in the marine environment is the facilitation of iron acquisition by extracellular $\text{O}_2^{\bullet-}$ —produced by microorganisms. The effectiveness of this process depends strongly on the concentration and speciation of iron but appears to be exploited by marine microbes under some conditions at least, as discussed in detail in Rose (2012) [73].

Algal blooms represent conspicuous hot spots for gross ROS production. Blooms are common in freshwater systems, but also occur in marine environments where they are often associated with the production of harmful toxins. H_2O_2 accumulation has been documented in the extracellular environment for a number of toxic species, albeit with substantial variation in production rates even between different strains of the same species [74–76]. It remains an open question how much of the ROS generation associated with blooms represents passive release or active production of ROS by the algae, and how much is passive photochemical generation caused by the high DOM conditions created by the bloom. For instance, production of $\text{O}_2^{\bullet-}$ and H_2O_2 was observed in filtered supernatants of cultures of several toxic algae [74], consistent with the same photochemical or other abiotic processes acting on DOM. On the other hand, the contribution of factors such as colony morphology [77], nutrient deficiency [78], CO_2 depletion [79], and lectin induction [76] to $\text{O}_2^{\bullet-}$ production all suggest active processes by the algae involved. Importantly however, some of these experiments were performed with axenic cultures whereas others also had undefined populations of associated bacteria; therefore it is not clear how much of the observed ROS production was caused by the algae and how much by

the bacterial component, or how the presence of different groups of bacteria may have altered the metabolism or ROS dynamics of the cultures during the experiments.

Regardless of the mechanisms underlying ROS production by algal blooms, they are capable of dramatically increasing ROS fluxes and steady-state H_2O_2 concentrations compared to normal conditions. For example, the bloom-forming seaweed *Ulva* secretes sufficient H_2O_2 to more than double the ambient concentration of H_2O_2 in coastal ecosystems where it dominates [80]. High concentrations of superoxide (~ 10 nM) were detected on the surface of coral *Porites astreoides* in the dark, independent of photosynthesis, and its symbionts also contributed extracellular superoxide in the dark and at low light levels [81]. ROS concentrations in a number of harmful algal blooms are so high that they have been considered a possible source of ichthyotoxicity [82–86] (although other studies such as [87] have argued against this conclusion). One possible factor that may exacerbate the impact of blooms on the prevailing H_2O_2 concentration is the fact that blooms have mechanisms for remaining at the sea surface, forming thin, very dense films at the air-water interface. One experiment using a recently developed microelectrode method for H_2O_2 determination found H_2O_2 concentrations 3–5 times greater in the upper few millimeters of the water column as compared to water a few centimeters deep during a freshwater *Microcystis* bloom that would have eluded older measurement protocols [88]; it is possible that such fine-scale measurements may reveal more high-ROS microenvironments in marine systems as well.

3.1.4. ROS production by non-photochemical abiotic processes

The final major source of ROS is via non-photochemical, abiotic redox processes, predominantly through the oxidation of reactive reduced species by O_2 . Such processes can potentially result in significant fluxes of ROS whenever there is a continuous supply of these reduced species to oxygenated waters, such as at the interface of reducing sediments with oxygenated waters. Murphy et al. demonstrated that, in the presence of hydrous ferric oxides, S^{2-} undergoes net oxidation by O_2 to produce $\text{O}_2^{\bullet-}$ and H_2O_2 [89]. More recently, Shaw et al. demonstrated a similar process for ROS production in hydrothermal vents [90] and calculated that the resulting flux of H_2O_2 to the global ocean via this pathway is of a similar magnitude to that produced by photochemical processes in the surface ocean. Non-photochemical, abiotic ROS production could also occur at the interface of reducing microenvironments within the oxic water column, such as at the surfaces of abiotic particles (e.g. minerals). However, studies examining dark, particle-associated ROS production in the marine water column have primarily found this source to be biological (e.g. Refs. [81,91,92]).

3.2. Microorganisms: the predominant sink for marine H_2O_2

As has been previously mentioned, most ROS have very short residence times, and are rapidly eliminated by reaction with other molecules near their point of generation. Those reactions involving DOM or metals, or involved in toxic attacks on biomolecules, will be considered in later sections. Here we exclusively consider the role of microorganisms in the removal of H_2O_2 from their environment. Microbes are generally considered the primary agents of active H_2O_2 quenching in the ocean [1] and indeed in freshwater systems as well. Cooper and Zepp [93] found that H_2O_2 decomposition in freshwater was arrested in chemically sterilized samples. Similarly, Petasne and Zika [94] found that 0.2 μm filtration of samples greatly reduced H_2O_2 decay rates, and that most of the H_2O_2 degradation ability was contained within the 0.2–1 μm size fraction, implicating bacteria as the predominant players in H_2O_2 removal. Both these studies found that re-seeding sterilized samples with cultured bacteria restored H_2O_2 degradation, confirming the important role played by microbes as the predominant sink for H_2O_2 in natural waters.

Microorganisms that inhabit oxic waters have evolved many strategies for protecting themselves from ubiquitous H_2O_2 . The most

conspicuous of these defenses are ROS-scavenging enzymes, including both catalases and peroxidases. These enzymes are widespread in both microbes and macroscopic life and have traditionally been thought to be essential for survival in the presence of atmospheric concentrations of O_2 , although recent evidence suggests that they are actually present in only a fraction of marine microbes [20,95]. Both catalase and peroxidase are phylogenetically diverse – there are multiple, independently evolved enzyme groups that catalyze these reactions – and many use redox-active metal cofactors containing Fe or Mn to facilitate the removal of H_2O_2 [96]. Canonical catalase disproportionates H_2O_2 directly into water and O_2 , but other catalases as well as metal-cofactor peroxidases are also capable of reducing H_2O_2 using electrons from other metabolites such as ascorbate. Peroxiredoxins, another group of peroxidases common in marine bacteria, do not have metal cofactors, but rather use redox-competent cysteine residues to reduce peroxides (e.g., H_2O_2 as well as lipid and other organic peroxides) using electrons from small molecules (e.g., glutathione and thioredoxin) that are common intermediaries in cellular redox processes [97]. While some peroxiredoxins are clearly front-line defenses against H_2O_2 (e.g., AhpC in *Escherichia coli* [98] and the recently-discovered 2-Cys peroxiredoxin from *Chattonella marina* [99–101]), the function of others appears to be directed more toward scavenging of lipid or other organic peroxides [102].

In the ocean, removal of H_2O_2 appears to occur primarily through the action of microbial catalases and peroxidases, although the relative importance of the two enzymes as well as the identities of the organisms expressing them remain incompletely understood. In a study of H_2O_2 decomposition using ^{18}O -labelled H_2O_2 , Moffett and Zafiriou [103] determined that the majority of H_2O_2 was converted to O_2 , indicating that most of the H_2O_2 was broken down using catalase. These results were recently revisited with improved methods, revealing that H_2O_2 removal processes proceed from catalase-like to peroxidase-like redox profiles with increasing depth [104]. Interestingly this same study also revealed that some intermediate depth samples exhibited unexpectedly high oxidation:reduction ratios, suggesting the existence of unknown powerful oxidants capable of scavenging H_2O_2 from the water column.

While catalase appears to be the predominant sink for H_2O_2 , at least at the ocean's surface, only a minority of marine microorganisms produce it. In a re-examination of several diel metatranscriptomic studies from the surface ocean, the bacterial enzyme catalase-peroxidase (KatG) was much more abundant than the monofunctional catalase found in both bacteria and eukaryotes in all samples [20]. Algal ascorbate peroxidases were also widespread, and the expression of both enzymes was found to oscillate over the diel cycle, with higher expression during the day than at night, corresponding with increased rates of H_2O_2 removal during the day than at night. The taxonomic composition of the two enzymes varied between stations, but KatG sequences like those from heterotrophic bacterial clades Alteromonadales, SAR11, and SAR116 and green algal ascorbate peroxidase sequences were always abundant and over-represented relative to their share of the overall microbial population. Thus, a small subset of total microbial diversity in all these samples appeared to be responsible for the majority of the effort required to remove H_2O_2 from the environment.

If only a fraction of the total microbial community is engaged in ROS removal and most of the removal is performed by intracellular enzymes, how do the other organisms survive? First, it is possible that some ROS scavenging enzymes are actually secreted into the environment; extracellular SODs are common in many bacteria and fungi [105], and a few organisms also express extracellular catalases [106–108]. Cellular contents may also be released into the environment as a result of sloppy feeding or viral lysis [109]. Residual H_2O_2 degradation has been detected in sterile-filtered seawater in a variety of studies as well [94, 110]. More likely, however, is that intracellular H_2O_2 degradation unavoidably also reduces the extracellular H_2O_2 concentration due to the fact that H_2O_2 is freely membrane permeable [111]. Thus, it is possible that the entire microbial community can be protected from

H₂O₂ by a minority of H₂O₂-scavenging organisms. This phenomenon creates a feedback mechanism that may be responsible for the evolution of complex, market-like interdependencies in marine microbial communities wherein H₂O₂-removing “helpers” protect their neighbors who in some cases “pay” them with other products, such as photosynthate or vitamins, a process described by the Black Queen Hypothesis [95,112].

4. The impact of ROS on marine biogeochemistry

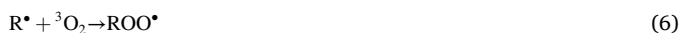
As summarized previously, each of the various ROS exhibit quite different characteristics in terms of their reactivity. While all can act as oxidants, some are also able to act as reductants under different circumstances. Thus, ROS impact the biogeochemical cycling of a wide range of elements. Some of the major mechanisms by which ROS react with various elements, and the impact on their biogeochemical cycles, are discussed in further detail below.

4.1. ROS and organic carbon

Organic carbon (OC) molecules in their ground state are relatively stable against direct oxidation by ³O₂, despite the thermodynamic favorability of the overall reaction with respect to the O₂/H₂O couple, due to the barrier imposed by the one-electron transfer requirement. This constraint is overcome when OC molecules are in an excited (typically singlet) state [4], as can occur by photochemical excitation of CDOM (although the majority of excited CDOM molecules transfer energy rather than electrons to ³O₂, yielding ¹O₂ [9]). While ¹O₂ may itself oxidize certain OC compounds, the effectiveness of this reaction is limited by its very short lifetime due to relaxation by solvent water [9]. Consequently, ¹O₂ is unable to diffuse far from the site of its formation, and the highest concentrations are typically found in hydrophobic microenvironments within large CDOM molecules (e.g. humic acids) [113]. In these localized environments, oxidation by ¹O₂ may represent a significant pathway for degradation of certain OC compounds, including organic pollutants [9].

OC is also readily oxidized by •OH. Due to its high reactivity, •OH is typically short-lived and thus this process is only significant where there is a continuous flux of •OH production, usually from photochemical processes in marine systems. •OH can also be formed through the reduction of H₂O₂ via Fenton or Fenton-like reactions, but these are unlikely to occur at biogeochemically significant rates in most marine systems unless there is a continuous source of reduced trace metals. This can occur in the photic zone through photochemical reduction of some metals, such as Fe, giving rise to the so-called photo-Fenton reaction. The photo-Fenton reaction may be a major pathway for OC degradation under conditions like those found in marine waters [114], although evidence suggests that the primary oxidant may not necessarily be •OH. This is consistent with the fact that the Fenton reaction does not necessarily produce free •OH, but may instead yield other oxidants such as Fe(IV) (ferryl iron), depending on the complex speciation of iron [115]. While the reactivity of Fe(IV) towards OC may differ from that of free •OH, it is still a powerful oxidant [116]. This may also be true for the Fenton-like system involving Cu(I) and H₂O₂ [117].

The reaction between •OH and OC may be a significant sink for OC in marine waters [118]. This reaction initially produces carbon-centered radicals, which (unlike bulk OC) may subsequently react with ³O₂. This process typically involves addition of O₂ to the carbon-centered radical to yield a peroxy radical:



where R represents an organic group and ROO• the peroxy radical. Peroxy radicals are typically far more stable than carbon-centered radicals under oxic conditions, since the latter typically react with ³O₂ at near diffusion-controlled rates [119], but may undergo further reaction with O₂ or other peroxy radicals to yield organic peroxides:



where ROOH and ROOR' are organic peroxides. The fate of these organic peroxides is discussed further below.

In contrast to both ¹O₂ and •OH, O₂^{•-} is highly selective in its reactions due to its unusual electronic structure, such that it readily reacts only with molecules possessing unpaired valence shell electrons, including a range of redox-active trace metals and organic radicals. O₂^{•-} is also unique among the ROS in that it is thermodynamically more favorable as a reductant than as an oxidant under typical conditions found in oxic marine waters (Fig. 1). As such, O₂^{•-} does not readily react with bulk OC, but may facilitate redox cycling of stable radical groups, such as semiquinones [120] and phenoxy radicals [121,122], which are thought to be present in natural organic matter [122,123]. Goldstone and Voelker [124] first reported a non-metallic sink for O₂^{•-} in natural waters associated with the presence of humic and fulvic substances. Heller and Croot [125] showed that reactions with CDOM represented a significant sink of O₂^{•-} in tropical Atlantic surface waters, but that such a pathway was not a major sink for O₂^{•-} in the Southern Ocean [126]. King et al. [127] also showed that unidentified antioxidant compounds able to consume O₂^{•-} were present at concentrations of 100–400 pM in seawater samples from the South Atlantic Ocean, but only during daylight hours.

While H₂O₂ is a relatively powerful oxidant from a thermodynamic perspective [4], it does not readily oxidize OC under ambient conditions due to a high activation energy barrier for the reaction [6]. The oxidation of semiquinone-type radicals via a “metal-independent Fenton reaction” has been reported in aqueous solutions [128] and the formation of •OH by such a pathway has been reported in solutions of irradiated humic acids [129]. However, this process has not been widely examined in marine waters.

Organic peroxides may reach concentrations comparable to H₂O₂ of up to several hundred nM in surface nearshore marine waters [130], suggesting that they are similarly relatively unreactive. Nonetheless, relatively little is known about the fate of organic peroxides in the ocean, or their influence on biogeochemical cycles.

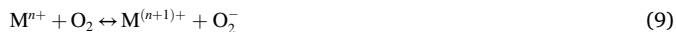
4.2. ROS and trace metals

ROS can potentially react with, and thus influence the redox speciation, of a range of redox-active trace metals in seawater. Among the biogeochemically relevant trace metals found in marine waters, Fe, Cu, Mn, Mo and V are particularly notable in terms of their ability to cycle between metastable redox states, at least in part because of the thermodynamic constraints imposed by the one electron transfer requirement in systems dominated by the O₂/H₂O redox couple [3]. This section will thus focus on these five trace metals as examples of trace metals with particular biological significance whose redox cycling in marine waters is well known to be influenced by ROS. This list is not intended to be exhaustive, however; there are a range of other redox active trace metals that are known to react with ROS under aqueous conditions (see, for example, compilations of reactions of O₂^{•-} [131] and •OH [132] with various trace metals and other compounds), including in seawater (for example oxidation of Cr(III) by H₂O₂ [133] or the suggested involvement of ROS in Hg cycling [134]).

As stated previously, H₂O₂ and •OH are predominantly oxidizing in nature at the pH of typical marine waters, with the notable exception of the reduction of Cu(II) by H₂O₂ (discussed further below). In contrast, O₂^{•-} is capable of functioning as both an oxidant and reductant and, as such, plays a somewhat special role in relation to the redox cycling of trace metals. The role of ¹O₂ in redox cycling of trace metals is likely to be limited due to its low concentrations and localization near the site of its predominantly photochemical generation, but due to their unpaired

valence shell electrons the trace metals discussed above may all react directly with $^3\text{O}_2$, albeit at varying rates.

The general sequence of reactions between ROS and the trace metals may be described as follows:



where M represents the metal and M^{n+} and $\text{M}^{(n+1)+}$ are reduced and oxidized redox states, respectively. (The first reaction is shown as a two-way reaction due to the commonly reducing behavior of O_2^- , in contrast to the other reactions). A critical control on the nature of ROS-induced redox cycling of trace metals is the relative rates of each of these reactions, leading to three general scenarios:

Scenario 1 – The forward reactions in eqs. (9)–(11) and/or 12 are fast relative to the backward reaction in eq. (9): the trace metal will exist primarily in its oxidized state and does not readily undergo ROS-induced redox cycling. The relative importance of $^3\text{O}_2$ compared to other ROS as oxidants of the trace metals will depend on the relative rates of the forward reactions in eqs. (9)–(12).

Scenario 2 – The forward reactions in eqs. (9)–(12) are all slow compared to backward reaction in eq. (9): the trace metal will exist primarily in its reduced state and does not readily undergo ROS-induced redox cycling.

Scenario 3 – The forward and backward reactions in eq. (9) are both fast relative to the reactions in eqs. (10)–(12): the trace metal will be rapidly cycled between the reduced and oxidized states and reaches a steady-state redox equilibrium with the O_2/O_2^- redox couple.

It is important to note that the relative reaction rates depend on both the rate constant, which is an intrinsic property of the reacting species, and the concentrations of $^3\text{O}_2$ and the various other ROS. Thus, the applicability of each of these scenarios will depend on both the chemistry of the trace metal involved and local conditions and concentrations.

4.2.1. Iron

The reactions of ROS with Fe are perhaps the most well-studied among these trace metals. In typical oxic marine waters, Fe falls into scenario 1 above. While Fe(III) is the thermodynamically stable redox state of free Fe in oxic marine waters, Fe(II) has been measured in a range of surface and deep ocean environments even under fully oxygenated conditions (e.g. Refs. [135,136]). The existence of measurable Fe(II) in these environments can result from photochemical and biological processes, in addition to physical transport from more reducing waters (e.g. in oxygen deficient zones [137], or potentially in reducing microenvironments in sinking particles [138]). The biogeochemical cycling of Fe in these oxic waters is tightly coupled to the cycling of O_2 and O_2^- via the two-way reaction:



The tight coupling between these redox pairs means that it is often difficult to resolve whether the redox cycling is initiated by the formation of Fe(II) or O_2^- [73], although arguably this may be unimportant in terms of the overall biogeochemistry. The dynamics of this system additionally depends on the chemical speciation of Fe, which is strongly influenced by the complexation of both Fe(II) and Fe(III) by organic matter [e.g. Refs. [139–141]]. This influence of iron speciation on reactivity with the O_2/O_2^- couple has been examined in detail previously [73], and as such will not be explored further here.

Fe(II) may also be oxidized by H_2O_2 and OH^\bullet according to the series of reactions shown in eqs. (10)–(12). Both the rates and mechanisms of these reactions depend strongly on the speciation of Fe(II), in addition to

the relative concentrations of the various ROS. For example [142], showed that humic-type organic matter can inhibit oxidation of Fe(II) by H_2O_2 [115]. Additionally showed that the oxidation of Fe(II) by H_2O_2 yields OH^\bullet when the Fe(II) is complexed by certain organic ligands, but in the absence of organic complexation at circumneutral pH appears to yield a different (but still highly oxidizing) species. In lake waters, the reduction of organically complexed Fe(III) by O_2^- was observed to generate an “autocatalytic Fenton reaction” [143], resulting in significant OH^\bullet production due to redox cycling of Fe(II), and subsequent oxidation of OC by the generated OH^\bullet . Such processes are also likely to occur in marine waters.

4.2.2. Copper

Many of the reactions of Cu with ROS are analogous to those of Fe with ROS, although with some important differences. Like Fe, Cu may exist in both reduced and oxidized states in marine waters, with the oxidized Cu(II) state the most thermodynamically stable. Unlike Fe(II), the reduced Cu(I) state reacts at near diffusion-limited rates with $^3\text{O}_2$ except when complexed to certain stabilizing ligands, notably including Cl^- , which is an important factor in why Cu(I) is able to persist at measurable concentrations in marine waters in some circumstances [144,145]. However, Cu(I) and Cu(II), including their organic complexes, react at near-diffusion controlled rates with O_2^- [146]. As such Cu also tends to fall into scenario 1 above, in which both Cu(I) and Cu(II) may coexist at biogeochemically significant concentrations particularly in surface waters [147,148]. Also similarly to Fe, both Cu(I) and Cu(II) may be complexed by organic matter in marine waters (e.g. Refs. [149, 150]), which influences the kinetics of the various reactions shown in eqs (1)–(4) and hence its overall redox cycling dynamics [151]. However, unlike Fe(III), Cu(II) can be reduced by H_2O_2 at biogeochemically significant rates according to the reaction [152]:



This reaction introduces an additional mechanism by which ROS can induce redox cycling of Cu between the Cu(I) and Cu(II) states. Cu can also participate in Fenton-like processes under marine conditions, but as with Fe the formation of OH^\bullet through this process depends on factors such as pH and complex speciation of Cu [117,153].

4.2.3. Manganese

Manganese has been measured to exist in three redox states in marine systems: Mn(II), Mn(III) and Mn(IV). The most oxidized Mn(IV) state is thermodynamically favored under oxic conditions with respect to the overall $\text{O}_2/\text{H}_2\text{O}$ redox couple, but the first electron transfer step in the oxidation of Mn(II) by $^3\text{O}_2$ is thermodynamically unfavorable (i.e. with respect to the O_2/O_2^- redox couple) [154]. Hence, the oxidation of free, dissolved Mn(II) by $^3\text{O}_2$ under conditions typical of marine waters is negligibly slow [155], but can be accelerated by microbes, organic complexation or adsorption to mineral surfaces (see Refs. [156, 157] for detailed reviews). Mn(II) is nonetheless often the most abundant redox state measured in oxic marine waters [157].

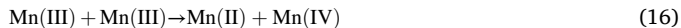
In contrast, the oxidation of Mn(II) to Mn(III) by O_2^- is thermodynamically favorable under conditions found in marine waters [154], and has been shown to occur experimentally under similar conditions via either direct addition of O_2^- [158,159] or by microbial O_2^- production [160]. However, oxidation of Mn(II) via this pathway appears to be readily reversible, such that net oxidation of Mn(II) to Mn(IV) may not necessarily occur, due to the reverse reaction in which intermediate Mn(III) is reduced back to Mn(II) by H_2O_2 [157,158,161]:



As such, for net oxidation of Mn(II) to Mn(III) to occur via this pathway, H_2O_2 produced by the reaction must be rapidly consumed via other processes.

Mn(III) may also be able to persist in marine waters when complexed

[157], and has been shown to represent a substantial proportion of the total dissolved Mn even under oxic conditions [162]; however there is currently only limited evidence for this. The major fate of Mn(III) is thought to be disproportionation:



Mn(IV) is highly insoluble in marine waters, and predominantly exists as Mn(IV), or in mixed valence Mn(III)/Mn(IV), oxides. Reductive dissolution of these Mn oxides has been shown to be induced by reactions with both O_2^- and H_2O_2 [163]. The ranges of potential reactions of Mn(II), Mn(III) and Mn(IV) with O_2^- and H_2O_2 are consistent with observations of dynamic redox cycling of Mn in marine systems (e.g. Ref. [164]), and the suggestion that redox cycling of Mn may be a potential mechanism for the observed decay of H_2O_2 in marine waters [104].

4.2.4. Vanadium

Vanadium can exist in the V(III), V(IV) or V(V) redox states in aqueous systems; however, the V(III) state is only thermodynamically favored under strongly reducing conditions [165]. In oxic marine systems, V(V) is the most thermodynamically stable state, but both the V(IV) or V(V) states have been measured at abundances of >10% of total dissolved V in oxic marine waters (e.g. Refs. [166,167]).

The rate constant for oxidation of VO(OH)^+ , the dominant V(IV) species at circumneutral pH, by $^3\text{O}_2$ is around $1.1 \text{ M}^{-1}\text{s}^{-1}$ [168], which is only slightly lower than that for inorganic Fe(II) under similar conditions. However, as with many other trace metals discussed here, both V(IV) and V(V) form complexes with organic matter ([169] and references therein), which likely influences V redox dynamics. Given that oxidation by $^3\text{O}_2$ proceeds via one-electron transfer steps, this reaction would be expected to yield O_2^- , but this has not been experimentally confirmed. V(IV) has been reported to react with HO_2 (the conjugate acid of O_2^-) under highly acidic conditions in aqueous solution [170]. It is also oxidized by H_2O_2 under circumneutral conditions [171]. However, there remains a considerable knowledge gap around the dynamics of ROS-induced vanadium redox cycling in marine waters, including its reactions with O_2^- and H_2O_2 , which have not been well studied under these conditions.

4.2.5. Molybdenum

Like vanadium, molybdenum can exist in three redox states in aqueous systems: Mo(IV), Mo(V) and Mo(VI) [172]. Of these, Mo(IV) is known to occur only under highly reducing conditions, with Mo(VI) being the most thermodynamically stable and dominant form under oxic conditions, and Mo(V) typically found in mildly reducing microenvironments [172]. Relatively few studies have examined Mo redox speciation in oxic waters. Wang et al. observed the coexistence of both Mo(V) and Mo(VI) in both fully oxic and low oxygen (<10 μM dissolved O_2) conditions in the Peconic River Estuary, with the reduced Mo(V) present as a greater proportion (up to 15%) of total dissolved Mo under the lower oxygen conditions but nonetheless present at measurable concentrations in some fully oxygenated surface waters. However, some doubts have been raised about the existence of Mo(V) in the oxygenated water column [173]. In any case, the diffusion of Mo(V) from reducing environments (e.g. sediments) into oxygenated waters would likely result in oxidation of Mo(V) to Mo(VI) by $^3\text{O}_2$, given that reaction is thermodynamically favorable to proceed via one-electron steps [3]. This would suggest the involvement of ROS in redox cycling between Mo(V) and Mo(VI); however we are unaware of any experimental evidence for such processes in the literature at this time.

4.3. ROS and other inorganic species

ROS have been implicated in the biogeochemical cycling of a range of other elements in marine systems. In the case of iodine, I^- reacts

exceedingly slowly with $^3\text{O}_2$, but reacts readily with O_2^- [174], H_2O_2 [175] and organic peroxides [176]. Consistent with these observations, Luther [177] demonstrated that the oxidation of halides under conditions typical of seawater is thermodynamically feasible by a range of ROS including $^1\text{O}_2$, O_2^- , H_2O_2 and $^{\bullet}\text{OH}$, with the effectiveness of each depending on the exact pathway.

ROS are additionally known to react with transient inorganic species that are found in seawater, including nitrogen radicals such as NO^{\bullet} and sulfur radicals such as thiols [178]. While the reaction of O_2^- with NO^{\bullet} to yield peroxyxynitrite in alkaline solutions has been known for decades [179], recent work has shown this reaction is a major sink for photochemically generated NO^{\bullet} in marine waters [180]. Where these radicals exist in oxygenated waters, often as transient species, reactions with ROS are likely through so-called 'cryptic cycles' in which the rapid turnover and low concentrations of the species involved renders them largely undetectable [181].

5. ROS impacts in marine ecosystems

In addition to the roles that ROS play in biogeochemical cycling, they also can create profound biological effects. ROS have predominantly been considered to have negative impacts on living organisms due to their ability to react, often indiscriminately, with biological macromolecules. However, increasing evidence indicates that ROS can also play beneficial roles for individual organisms and at the ecosystem level, including through potentially favorable moderation of the local redox environment (e.g. by increasing the bioavailability of micronutrients like iron [73]), but also through biological processes such as regulating growth and resisting pathogens. The beneficial effects of extracellular ROS have recently been reviewed by Hansel and Diaz [2], so will not be considered in further detail here. This section will therefore focus primarily on the potential negative impacts of ROS in marine ecosystems.

Steady-state concentrations and fluxes of ROS in the ocean are generally well below levels that cause toxicity in laboratory studies. For instance, the bacterium *Escherichia coli* was able to tolerate acute exposure to 15 mM H_2O_2 with only a moderate loss of viable counts and a short lag period [182]; this value is almost five orders of magnitude higher than any in situ concentration measured in the surface ocean. Animals easily tolerate exposure to even higher concentrations of H_2O_2 , with 3% (~100 mM) H_2O_2 routinely used as a topical first-aid antiseptic in humans, for example. Environmental concentrations of free and $^{\bullet}\text{OH}$ are even lower, and orders of magnitude less than concentrations measured in healthy animal cells. Despite this, research over the past decade has revealed that some important marine microbial groups are vulnerable to even these low ROS concentrations. Moreover, some marine microenvironments, such as saltmarsh pools and certain kinds of algal blooms, may accumulate high concentrations of ROS capable of damaging even relatively well-protected organisms. In this section, we review the cellular targets of ROS attack and consider several key organisms that are particularly at risk.

5.1. Targets of ROS attack in marine organisms

ROS, especially $^{\bullet}\text{OH}$, can attack all classes of biological macromolecules. $^{\bullet}\text{OH}$ may attack the backbone deoxyribose sugars in DNA, fragmenting them in a variety of ways leading to single- or double-strand breaks [105,183,184]. Also, the nucleotide bases themselves may be modified, either before or after incorporation into a DNA molecule. 8-Hydroxyguanine, one result of $^{\bullet}\text{OH}$ attack on guanine, base pairs with thymine rather than cytosine, and if not repaired may lead to transversion mutations. RNA, including relatively long-lived rRNA and tRNA molecules, is even more vulnerable to oxidation than DNA [185], and oxidative mRNA damage predictably correlates with translational errors [186].

$^{\bullet}\text{OH}$ is also capable of many non-specific reactions on proteins,

oxidizing both side chains and polypeptide backbones. The typical result of such oxidation is the formation of carbonyl groups, and the detection of this “carbonylation” using antibodies is a common method for measuring oxidative protein damage in vitro [105]. Compared to $\bullet\text{OH}$, other ROS have more well-defined protein targets. O_2^- degrades proteins that bind iron, such as ferritin, catalase, and Fe–S cluster enzymes [105,187,188], causing the release of free Fe into the cytoplasm, allowing destructive Fenton chemistry. H_2O_2 , while more stable than either O_2^- or $\bullet\text{OH}$, attacks certain proteins even in the absence of free Fe, including important, constitutively expressed proteins such as the β -subunit of ATPase and prokaryotic translation elongation factor G [189,190].

Lipids are also prone to attack by $\bullet\text{OH}$, and oxidized lipids are capable of dramatically magnifying the effects of ROS attack via radical propagation. In a typical scenario, $\bullet\text{OH}$ attacks a double bond in an unsaturated lipid and abstracts a hydrogen atom, leaving a carbon radical and water. This new lipid radical may react with any other radical with no energy barrier. One possible reaction is with a second lipid radical, quenching the radical character of the system and forming a stable, cross-linked species that terminates radical propagation. This reaction is rare in oxic systems, however, compared to reaction with $^3\text{O}_2$ to form a lipid hydroperoxide and a new lipid radical. This propagation step may be carried out many times before the original lipid radical is quenched by a termination reaction. Lipid hydroperoxides are capable of attacking membrane proteins as well as facilitating the transition of $^3\text{O}_2$ to $^1\text{O}_2$ in the cytoplasm [191].

5.2. Major ROS-vulnerable marine populations

There are two ways that an organism living in the ocean may experience oxidative stress. First, the stress may arise intracellularly because of unbalanced metabolism, increased temperature or pH stress, or exposure to toxic substances in the environment such as xenobiotics and heavy metals. Some of these stressors were described above in section II. Second, the stress may arise through direct exposure to ROS in the environment. As described above, extracellular ROS fluxes in marine systems tend to be substantially lower than doses shown to negatively impact organisms in the laboratory. However, these observations may underestimate the direct risk organisms face from ROS in their native habitats. For instance, an *E. coli* population in natural river water exposed to light experienced $\sim 99\%$ mortality after 3 days unless catalase or sodium pyruvate (a $\bullet\text{OH}$ trap) was added to the water prior to illumination [192]. Based on other studies, the likely H_2O_2 concentration of the river water was no greater than $10\ \mu\text{M}$ (Table 1), more than 1000-fold less than the concentration necessary to kill laboratory populations of *E. coli* [182]. Laboratory culture media often contain hidden sources of ROS [193–196] which may select for disproportionately high levels of resistance over time, obscuring the sensitivity of native populations. More broadly, Xenopoulos and Bird [197] reported $>50\%$ reduction of bacterial productivity in a lake following exposure to $100\ \text{nM}$ H_2O_2 as compared to a catalase-treated control. Similar to these freshwater observations, a study in a coastal macrophyte-dominated ecosystem found that additions of as little as $20\ \text{nM}$ H_2O_2 significantly reduced bacterial productivity, with $1\ \mu\text{M}$ completely eliminating productivity over a 1 h timescale [80]; importantly however, other studies have shown no effect on bacterial production with H_2O_2 addition [198].

Among the first marine organisms to be conclusively demonstrated to have high sensitivity to H_2O_2 inhibition was *Prochlorococcus*, the numerically dominant phytoplankton genus throughout the temperate and tropical oligotrophic oceans. When grown in pure culture, *Prochlorococcus* is completely inhibited by $800\ \text{nM}$ H_2O_2 , but when grown in co-culture with a wide variety of heterotrophic bacteria, it can tolerate much higher exposures [199,200]. In the absence of bacteria, open ocean seawater from areas that are rich in *Prochlorococcus* rapidly accumulates H_2O_2 concentrations at least this high when exposed to sunlight, suggesting that *Prochlorococcus* is obligately dependent on other

members of its community to survive. This sensitivity likely exists because *Prochlorococcus* has an extensively streamlined genome, which contains no genes for catalase or any heme peroxidases, essentially eliminating its capacity to defend itself from exogenous H_2O_2 [201]. This lack of intrinsic H_2O_2 resistance is reflected in conspicuous vulnerability of pure *Prochlorococcus* cultures to a wide variety of stresses, including dark exposure [202,203] and suboptimal temperatures [204]. While *Prochlorococcus* is more vulnerable than most, cyanobacteria in general appear to be vulnerable to lower doses of H_2O_2 than most other studied organisms. Both unicellular cyanobacteria and filamentous forms experienced high mortality when exposed to $\sim 10\ \mu\text{M}$ H_2O_2 , a value that was consistent across numerous experiments in different laboratories [205–208]. On the other hand, eukaryotic microalgae (green algae, diatoms) survived doses as high as $1\ \text{mM}$ [206, 207].

H_2O_2 can also inhibit phytoplankton growth by altering the availability of limiting nutrients. The primary source of Fe in surface waters of the Atlantic ocean is atmospheric deposition, e.g. via dust from the Sahara Desert. Much of this Fe enters the ocean in the form of rainwater, which can also carry H_2O_2 at concentrations hundreds of times higher than surface seawater (Table 1). Experiments in March 2000 at the Bermuda Atlantic Time Series stations showed that additions of either FeCl_2 or FeCl_3 to seawater increased chlorophyll-*a* concentrations over a three-day incubation, indicating Fe limitation of primary production in the absence of seasonal dust deposition [29]. However, when the Fe was added as a synthetic rainwater mixture also containing H_2O_2 , the increase in phytoplankton growth compared to unamended controls was completely eliminated. The presence of H_2O_2 effectively made the Fe unavailable, presumably through Fenton chemistry and precipitation of insoluble Fe(III) complexes. It is also possible that the increase in H_2O_2 (to a final concentration of approximately $400\ \text{nM}$) may have directly damaged or killed phytoplankton in this experiment.

Another ecologically important group of organisms, the ammonia-oxidizing archaea (AOA) of the phylum *Thaumarchaeota*, are also catalase and peroxidase deficient and conspicuously vulnerable to H_2O_2 [209]. AOA are important regulators of the speciation of bioavailable nitrogen in the ocean, representing up to 40% of the total bacterioplankton in some parts of the World Ocean and directly or indirectly contributing significant fractions of the greenhouse gases N_2O and CH_4 to the atmosphere [210]. Some AOA strains are also among the most H_2O_2 -vulnerable aerobic organisms known. For instance, the growth of one Southern Ocean strain was inhibited by $10\ \text{nM}$ H_2O_2 , nearly two orders of magnitude lower concentration than the lethal dose for *Prochlorococcus*, and the addition of only $6\ \text{nM}$ H_2O_2 to a field population completely eliminated nitrification while not affecting overall bacterial production at all [211]. Inhibition of AOA by nanomolar H_2O_2 was also observed in isolates from coastal temperate seas and the deep ocean [212,213], and enhancement of growth was frequently seen after addition of H_2O_2 -scavenging pyruvate or catalase, or by co-culture with helper bacteria [213–215]. AOA isolates have been observed to secrete H_2O_2 into their growth media in proportion to the amount of ammonia oxidized, suggesting that their vulnerability may arise as a side effect of their primary metabolism [215]. If true, this may represent a difficult-to-overcome constraint on AOA in the ocean, and could account for the fact that they are primarily active in deep water, only thriving at the surface during polar winters when photochemically generated H_2O_2 is at a minimum [211].

In general, macroscopic organisms, and even multicellular microscopic organisms such as fish and invertebrate larvae, are much less vulnerable to H_2O_2 in the nanomolar-micromolar range than these bacteria, although a number of them are inhibited by doses low enough to be of concern in the presence of some human activities (see section VI below). However, there are exceptions. For instance, one study demonstrated substantial variability in H_2O_2 tolerance in different clonal populations of the starlet sea anemone *Nematostella vectensis*, which inhabits coastal salt marshes where natural H_2O_2 concentrations

can reach the 100 μM range. When anemones were sampled from South Carolina, New Jersey, and Nova Scotia, a clear north-south gradient in tolerance for acute H_2O_2 exposure was observed, supporting the hypothesis that elevated temperatures correlate with greater exposure to H_2O_2 [216]. Interestingly, when the same researchers sampled anemones from a variety of saltmarsh ponds in Massachusetts, they found substantial variation in H_2O_2 tolerance between populations. Notably, these organisms were severely impacted by H_2O_2 concentrations within the range observed in the estuarine waters they inhabit (Fig. 4), so this high variation over a short spatial scale suggests that H_2O_2 in these ranges represents a sufficiently potent stress to affect local adaptation, at least in this species.

5.3. Impacts of ROS on marine biology experiments

Along with the discovery that some marine organisms are negatively affected by naturally occurring concentrations of ROS comes the possibility that cryptic ROS production caused by experimental procedures may impact results. For example, in a series of in situ mesocosm experiments in the Canary Islands in 2016 designed to test the effects of different degrees of ocean acidification on the structure and function of natural plankton assemblages, H_2O_2 concentrations within the mesocosm enclosures were observed to increase significantly (>200 nM) in comparison to ambient conditions (~ 50 nM) after several days [198, 217]. The authors concluded that these increases were caused by the mesocosm enclosures inhibiting vertical mixture of surface and deep waters, causing H_2O_2 formed near the surface to remain concentrated relative to conditions outside the enclosures. Elevated H_2O_2 correlated with decreased bacterial and phytoplankton abundances in these experiments, possibly indicating H_2O_2 toxicity that may have killed or otherwise inhibited these cells, but also confirming that reduced bacterial concentrations exerted a positive feedback on H_2O_2 concentration by removing the primary H_2O_2 sink in the mesocosms. Further, the elevated H_2O_2 concentrations likely impacted the lability of DOM and trace metals and, as a consequence, the availability of important growth substrates.

Unexpected ROS production can also influence laboratory experiments with marine microorganisms. For example, HEPES and other zwitterionic pH buffers commonly used in culture media for algae and bacteria generate H_2O_2 continuously when exposed to light, and can produce lethal concentrations for some of the more sensitive species (e.g. *Prochlorococcus*) in less than 24 h [194]. Collecting *Thalassiosira weissflogii* diatoms by syringe instead of peristaltic pump increased their release of H_2O_2 , which could influence transcriptomic studies of these organisms by artifactually increasing oxidative stress gene expression, for example [75]. Based on these observations, researchers working in these systems need to consider H_2O_2 concentrations, and the effect of experimental procedures on them, in the same manner they measure light, temperature, and other physicochemical properties that may be important confounding variables, and should also consider the possibility that laboratory strains are substantially more H_2O_2 resistant than their relatives in the ocean.

6. Impacts of human activity on marine ROS

There are several reasons to expect that the threat of oxidative stress will increase for marine organisms in the future as secondary effects of anthropogenic changes, including both the introduction of pollutants into marine systems as well as the broader impacts of CO_2 emissions on the physicochemical environment of the ocean. In this section we consider two vectors for increased oxidative stress, specifically the natural increase in ROS due to anthropogenic CO_2 increase, and secondarily human activities that may introduce or concentrate novel ROS sources in the environment.

6.1. Global CO_2 increase and ROS

The CO_2 concentration of Earth's atmosphere is increasing at an unprecedented pace due to human consumption of fossil fuels. The two most profound sequelae of this change are i) an increase in global average temperature, both in the atmosphere and the ocean, and ii) an imbalance in the carbonate buffering system of aquatic ecosystems, leading to acidification. Both conditions could lead directly to increased ROS exposure for marine organisms, as well as creating secondary changes to marine environments that may exacerbate oxidative stresses.

First, higher temperatures are likely to lead to greater rates of biological H_2O_2 production due to temperature stress on marine organisms, at least in the short term before local populations either adapt or are outcompeted by more temperature-tolerant competitors. When West Antarctic Peninsula seaweeds were exposed to 2-8 $^\circ\text{C}$ of warming they increased their rate of H_2O_2 release into the surrounding seawater as well as exhibiting metabolic signs of oxidative stress [49]. Similarly, the microscopic gametophytes of three Arctic kelp species released greater amounts of $\text{O}_2^{\bullet -}$ into the surrounding medium when exposed to elevated temperatures [218]. In both cases, the seaweeds themselves appeared to be unharmed by the conditions, but the possibility of enhanced ROS release negatively impacting the surrounding community was not explored.

Increased H_2O_2 release is also observed in sessile animals. For example, 16 different *Symbiodinium* coral symbionts exposed to heat stress showed substantial variability in signs of oxidative stress. Some strains exhibited severe electron transport chain damage resulting in production of large amounts of H_2O_2 , whereas others avoided H_2O_2 accumulation by downregulating PSII [219]. Like the anemones described earlier [216], this observation suggests a substantial amount of intraspecific variation in ROS tolerance that may increase the rate of evolution in a changing ROS environment, but perhaps at the cost of reduced photosynthesis or other unforeseen physiological trade-offs.

Increased temperature may also lead to higher ROS fluxes in future waters. Studies across time and space under present conditions reveal that shallower, hotter waters accumulate greater H_2O_2 concentrations (Table 1, Fig. 3). One predicted effect of global warming on the ocean is the shoaling of the mixed layer, where higher temperatures strengthen thermal stratification of the ocean and yield shallower thermoclines [220]. Fig. 3 shows clearly that H_2O_2 concentrations drop off rapidly below the thermocline, suggesting that most production occurs near the surface and only accumulates near the surface because H_2O_2 diffuses slowly across this barrier. Thus, shallower thermoclines may yield greater steady-state H_2O_2 concentrations at the ocean's surface, which may negatively impact ROS-sensitive organisms such as *Prochlorococcus*.

Ocean acidification may additionally enhance the susceptibility of marine organisms to environmental ROS. Most acidification research has focused on its interference with calcification in corals and other shelled organisms, but acidification also creates oxidative stress in a wide variety of both calcified and non-calcified species by imbalancing intracellular redox states, resulting in oxidative stress (e.g. Refs. [52,54, 221]) and enhanced vulnerability of some keystone species to a wide variety of both biotic and abiotic challenges [48,50,56]. In some cases, acidification may disrupt important symbiotic interactions as well. For instance, the synergistic "helper" interaction between *Prochlorococcus* and the bacterium *Alteromonas* appeared to shift from a commensal or mutualistic state under present-day conditions to an exploitative state under elevated CO_2 , with *Alteromonas* significantly downregulating its catalase genes and causing a dramatic increase in oxidative stress and cell death for *Prochlorococcus* [222]. Moreover, rates of hydroxyl radical formation by the Fenton reaction increase with H^+ concentration [223] as well as with temperature, suggesting that not just the concentrations, but also the lethality of ROS will be enhanced in future oceans, and that oxidative stress enhancement may underlie some of the synergistic effects of "multiple stressors" on marine life.

We must note, however, that much remains unknown about the

impact of CO₂ increase on the World Ocean in general, and other effects of acidification or temperature increase may reduce ROS stress on marine organisms. For instance, higher temperatures will increase the rates of all chemical, and most enzymatic, reactions, which will simultaneously increase both ROS production and removal processes and may also increase organismal maximum growth rates. All biogeochemical cycles, including both the biotic and abiotic cycling of DOM and other C compounds, will speed up, potentially increasing the bioavailability of many nutrients, which could reduce nutrient limitation stresses and/or improve the ability of organisms to protect themselves from ROS. Much more research into these processes will be necessary before we can draw any confident conclusions about ROS exposure will affect life in future oceans.

6.2. Pollution and marine ROS

In addition to the global impacts of CO₂ release, human activity is increasing the exposure of marine life to a wide variety of compounds that may increase oxidative stress. We have already discussed the effects of allelochemicals such as microplastics and heavy metals on oxidative stress in invertebrates, but in this section we will focus specifically on pollutants that may lead directly to increased ROS fluxes, particularly in coastal ecosystems.

6.2.1. Use of H₂O₂ in aquaculture

Since 1993, H₂O₂ has been used to control the growth of parasitic “sea lice” copepods and other ectoparasites in salmon aquaculture operations in Norway and elsewhere, with usage peaking in 2015 and remaining common today [224]. H₂O₂ has been popular because of its relatively low toxicity profile compared to other effective pesticides and the fact that it is removed from water bodies after a few days by microbes as described above [225]. Relatively high H₂O₂ dosages are necessary for complete lice control, however, with approximately 100 mM necessary to achieve 98% reduction in parasite load (Fig. 4). Because water from salmon farms is released directly into coastal seas, there is concern that these concentrations, which are orders of magnitude higher than naturally occurring aquatic H₂O₂ concentrations, may have toxic effects on off-target organisms. Indeed, studies have shown that several marine invertebrates and fish can be killed by lesser exposures, particularly in their immature forms (Table 2). For example, the LC50 for a 1-h H₂O₂ exposure for European lobster *Homarus gammarus* larvae increased from 5 mM to 22 mM as the larvae progressed through stage I-IV molts [226], although one study showed that adult *Calanus* copepods were killed by significantly lower H₂O₂ doses than adults [227]. Notably, assays that utilized longer exposures typically reported lower LC50 values, indicating that chronic exposure to lower doses of H₂O₂ can cumulatively increase damage even to complex multicellular organisms. For many of these species, lethal H₂O₂ doses were substantially lower than the recommended dosing for delousing of salmon enclosures, but it is unclear whether concentrations of H₂O₂ released from these ponds into the ocean is sufficiently high to kill significant numbers of animals, or if instead dilution by the bulk seawater is sufficient to eliminate their negative impacts. Importantly, however, much lower sublethal doses of H₂O₂ contribute to other syndromes for these organisms, including impairing their ability to find refuge or otherwise avoid predation [226,227], reducing their feeding rate ([228], Japanese language paper cited in Ref. [216]), or slowing healing processes [216]. These impediments may indirectly contribute to increased mortality for species living in the vicinity of aquaculture operations or other environments where humans use H₂O₂. Combined with the facts that repeated H₂O₂ treatment has negative impacts on salmon health [229] and appears to select for H₂O₂-resistant sea louse varieties [230], H₂O₂ has lost popularity in recent years in favor of mechanical and biocontrol delousing methods [225], but the practice remains widespread and worthy of further attention regarding its impacts on the surrounding natural environment.

Table 2
Lethal limits for H₂O₂ exposure in various organisms.

	Species	Type of assay	Effective dosage	Reference
Cnidarians	<i>Nematostella</i> sp.	14-d 100% mortality	163 μM	[216]
Crustaceans	<i>Mysis</i> sp.	1-h LC50	29 mM	[265]
	<i>Calanus finmarchicus</i>	1-h LC50	180 μM	[266]
	<i>Calanus</i> spp. Stage V larvae	1-h LC50	6.3 mM	[227]
	<i>Calanus</i> spp. Stage V larvae	25-h LC50	2.3 mM	[227]
	<i>Calanus</i> spp. Adult	1-h LC50	1.4 mM	[227]
	<i>Calanus</i> spp. Adult	25-h LC50	900 μM	[227]
	<i>Crangon septemspinosa</i>	1-h LC50	94 mM	[265]
	<i>Atemia salina</i>	24-h LC50	24 mM	[267]
	<i>Crophium volutator</i>	96-h LC50	1.4 mM	[268]
	<i>Homarus americanus</i> , stage I larva	1-h LC50	48 mM	[265]
Molluscs	<i>Homarus gammarus</i> , larvae stages I-IV	1-h LC50	5.2–22 mM	[226]
	<i>Dreissena polymorpha</i>	72-h LC50	890 μM	[55]
Fish	<i>Siganus fuscescens</i>	24-h LC50	6.6 mM	[269]
	<i>Trachurus japonicus</i>	24-h LC50	2.6 mM	[269]
	<i>Oncorhynchus mykiss</i> juveniles	3-h 90% mortality	~9 μM ^a	[270]
	<i>Microcystis aeruginosa</i>	3-h LC50	7.9–195 μM	[207]
Phytoplankton	<i>Selenastrum capricornutum</i>	3-h LC50	119–625.3 μM	[207]
	<i>Raphidocelis subcapitata</i>	3-h LC50	359–2088 μM	[207]
	<i>Prochlorococcus</i>	24-h 100% mortality	800 nM	[200]
	<i>Daphnia</i> sp.	48-h LC50	164 μM	[271]
Zooplankton	<i>Moina</i> sp.	48-h LC50	58.8 μM	[271]
	<i>Escherichia coli</i>	3 h-LC50	7.8 mM	[272]
	<i>Escherichia coli</i>	6 h 100% mortality	15 mM	[182]
Bacteria	<i>Vibrio coralliilyticus</i>	0.5 h-LC50	~15 μM	[67]

^a Measured in the presence of harmful alga *Heterosigma carterae*; algal toxins likely enhanced ROS lethality.

6.2.2. Use of H₂O₂ to combat harmful algal blooms

Harmful cyanobacterial blooms are increasing globally, and this trend may continue in the next decades [231]. One study showed that in 100 lakes investigated in North America and Europe, 60% have observed a substantial increase in cyanobacteria since the industrial revolution [232], and recently, the model of Chapra et al. [233] predicted that the average days per year with cyanobacterial blooms in USA will increase from ~7 d to 18–39 d by 2090. Rising temperatures and atmospheric CO₂ as well as the increasing frequency of extreme hydrologic events increase the growth rates of algae and alter critical nutrient thresholds for the development of algal blooms [234,235]. The expansion of agriculture in developing countries also accelerates the input of nitrogen and phosphorus into coast area, which also promotes the occurrence of algal blooms.

A large body of work from Russia and the former Soviet Union, summarized in English by Skurlatov and Ernestova [236], suggested that environmental H₂O₂ has the positive effect of reducing the frequency and severity of toxic algal blooms. Consequently, toxin-producing freshwater cyanobacteria were rare in waters where the concentration of H₂O₂ exceeded 0.3 μM. When waters became loaded with ROS “traps” from wastewater effluents, however, toxic blooms frequently formed. As H₂O₂ is generally toxic at lower concentrations to microorganisms than macroorganisms, it has been proposed as a very promising algicide in the control of cyanobacterial blooms [231,234,235]. Researchers in 1986 discovered that as little as 50 μM H₂O₂ strongly suppressed

photosynthesis and growth in the cyanobacterium *Oscillatoria rubescens* [237]. Afterwards, laboratory and field tests [206,238,239] proved the effectiveness of H₂O₂ in selectively eliminating cyanobacterial blooms and preventing bloom-forming species from surviving during winter [214]. Both high light and blue light were found to enhance the inhibition of H₂O₂ to *Microcystis aeruginosa* PCC 7806 [240]. A range of authors observed a dose-effect response between H₂O₂ and their effectiveness in the inhibition of different cyanobacteria. Considering the cost and ecological risk, a maximum dose of 5 mg L⁻¹ (~150 μM) was recommended for use in freshwater systems [241].

One major concern of using H₂O₂ to control *Microcystis* blooms was the release of the toxin microcystin from the lysing cells. However, a field experiment showed that the particulate and water soluble microcystin degraded 90% in less than 3 days. The decomposition of microcystin may be related to microbial degradation [242] but also to •OH catalyzation due to Fenton chemistry involving the added H₂O₂ [243]. Though H₂O₂ could not completely eliminate the harmful cyanobacterial bloom, it created a possibility for the rapid succession of phytoplankton populations from HAB species to non-harmful species, which could be readily consumed in classic microbial food chains [244].

Most applications of H₂O₂ to control algal blooms have been in freshwater systems, but recent efforts have also shown promise in marine systems. In the Norfolk Broads, a series of shallow saltwater lakes in Great Britain, the eukaryotic chrysophyte *Prymnesium parvum* is a toxic eukaryotic alga that has been responsible for massive fish kills. Administration of approximately 1.5 mM H₂O₂ to the Broads during a period between blooms reduced the ambient *P. parvum* levels by >90%, with effects starting only 2 h after exposure [245]. The H₂O₂ treatment also led to sizable changes in the bacterial community, with especially conspicuous losses of unicellular cyanobacterial taxa. Interestingly, the community rebounded within 4 d after treatment, suggesting that H₂O₂ did not cause a permanent shift in community structure although it could possibly help to reduce the magnitude of toxic taxa during a bloom. Similar concentrations were applied to a bloom of the toxic dinoflagellate *Alexandrium* in a brackish creek in the Netherlands, resulting in 99.8% reduction of algal biomass as well as elimination of the algal toxin from the water column [246]. The dosage necessary to destroy *Alexandrium* also caused severe reduction in zooplankton as well as altering the macroinvertebrate and even fish communities, however, reflecting the concerns raised in the previous section on the trade-offs in using H₂O₂ as a biocontrol agent.

7. Conclusions

The oxygen cycle in the ocean is considerably more complex than the simple shuttling of O between water and organic matter, with reactive intermediates engaged in a wide variety of reactions that have critical importance for the biogeochemistry of the sea. Improvements in our technological capability to measure ROS fluxes have led to an improved understanding of the cycling of DOM and trace metals in the ocean and have revealed previously unsuspected mortality pathways for microorganisms as well as potential confounding factors for oceanographic experiments. However, we still know comparably little about the mechanistic reasons for why some organisms are so much more susceptible to ROS damage than others, or how changing environmental conditions impact ROS fluxes and their effect on marine life. Critically, we also do not understand fully how human activity will affect ROS fluxes, nor how our use of ROS such as H₂O₂ for biocontrol may affect ecosystems. Because many stress pathways involve ROS, future efforts to mitigate harm to the environment from human activity must involve a deeper understanding of the pathways of formation and degradation of these enigmatic molecules.

Declaration of interests

The authors declare that they have no known competing financial

interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- [1] E.R. Zinser, The microbial contribution to reactive oxygen species dynamics in marine ecosystems, *Environmental Microbiology Reports* 10 (4) (2018) 412–427.
- [2] C.M. Hansel, J.M. Diaz, Production of extracellular reactive oxygen species by marine biota, *Ann. Rev. Mar. Sci.* 13 (2021) 177–200.
- [3] A.L. Rose, The influence of reactive oxygen species on local redox conditions in oxygenated natural waters, *Front. Earth Sci.* 4 (2016).
- [4] D.T. Sawyer, *Oxygen Chemistry*, Oxford University Press, Oxford, 1991, p. 223.
- [5] I. Fridovich, Oxygen toxicity: a radical explanation, *J. Exp. Biol.* 201 (8) (1998) 1203–1209.
- [6] C.C. Winterbourn, The biological chemistry of hydrogen peroxide, *Methods Enzymol.* 528 (2013) 3–25.
- [7] C.S. Foote, E.L. Cleannan, Properties and reactions of singlet dioxygen, in: *Active Oxygen in Chemistry*, Springer, 1995, pp. 105–140.
- [8] K. Mopper, X. Zhou, Hydroxyl radical photoproduction in the sea and its potential impact on marine processes, *Science* 250 (1990) 661–664.
- [9] D. Vione, et al., Indirect photochemistry in sunlit surface waters: photoinduced production of reactive transient species, *Chem. Eur. J.* 20 (34) (2014) 10590–10606.
- [10] C. Van Baalen, J.E. Marler, Occurrence of hydrogen peroxide in sea water, *Nature* 211 (5052) (1966) 951.
- [11] H.W. Harvey, Oxidation in sea water, *J. Mar. Biol. Assoc. U. K.* 13 (4) (1925) 953–969.
- [12] W.L. Miller, D.R. Kester, Hydrogen peroxide measurement in seawater by (p-hydroxyphenyl)acetic acid dimerization, *Anal. Chem.* 60 (1988) 2711–2715.
- [13] D.W. King, et al., Flow injection analysis of H₂O₂ in natural waters using acridinium ester chemiluminescence: method development and optimization using a kinetic model, *Anal. Chem.* 79 (2007) 4169–4176.
- [14] W.M. Draper, D.G. Crosby, The photochemical generation of hydrogen peroxide in natural waters, *Arch. Environ. Contam. Toxicol.* 12 (1983) 121–126.
- [15] W.J. Cooper, et al., Photochemical formation of H₂O₂ in natural waters exposed to sunlight, *Environ. Sci. Technol.* 22 (1988) 1156–1160.
- [16] W.J. Cooper, R.G. Zika, Photochemical formation of hydrogen peroxide in surface and ground waters exposed to sunlight, *Science* 220 (1983) 711–712.
- [17] C.L. Wilson, N.W. Hinman, R.P. Sheridan, Hydrogen peroxide formation and decay in iron-rich geothermal waters: the relative roles of abiotic and biotic mechanisms, *Photochem. Photobiol.* 71 (6) (2000) 691–699.
- [18] I. Obernosterer, P. Ruardij, G.J. Herndl, Spatial and diurnal dynamics of dissolved organic matter (DOM) fluorescence and H₂O₂ and the photochemical oxygen demand of surface water DOM across the subtropical Atlantic Ocean, *Limnol. Oceanogr.* 46 (3) (2001) 632–643.
- [19] W.J. Cooper, D.R.S. Lean, Hydrogen peroxide concentration in a northern lake: photochemical formation and diel variability, *Environ. Sci. Technol.* 23 (1989) 1425–1428.
- [20] J.J. Morris, et al., Diel regulation of hydrogen peroxide defenses by open ocean microbial communities, *J. Plankton Res.* 38 (4) (2016) 1103–1114.
- [21] J. Yuan, A.M. Shiller, The distribution of hydrogen peroxide in the southern and central Atlantic ocean, *Deep-Sea Res.* 48 (2001) 2947–2970.
- [22] R.J. Kieber, G.R. Helz, Temporal and seasonal variations of hydrogen peroxide levels in estuarine waters, *Estuar. Coast Shelf Sci.* 40 (1995) 495–503.
- [23] D. Abele-Oeschger, H. Tug, R. Rottgers, Dynamics of UV-driven hydrogen peroxide formation on an intertidal sandflat, *Limnol. Oceanogr.* 42 (6) (1997) 1406–1415.
- [24] L.J.A. Gerringa, et al., The influence of solar ultraviolet radiation on the photochemical production of H₂O₂ in the equatorial Atlantic Ocean, *J. Sea Res.* 51 (2004) 3–10.
- [25] J.F. Kasting, H.D. Holland, J.P. Pinto, Oxidant abundances in rainwater and the evolution of atmospheric oxygen, *J. Geophys. Res. Atmos.* 90 (ND6) (1985) 10497–10510.
- [26] R.J. Kieber, et al., Hydrogen peroxide at the Bermuda Atlantic Time Series Station. Part 1: temporal variability of atmospheric hydrogen peroxide and its influence on seawater concentrations, *J. Atmos. Chem.* 39 (2001) 1–13.
- [27] J. Yuan, A.M. Shiller, The variation of hydrogen peroxide in rainwater over the South and Central Atlantic Ocean, *Atmos. Environ.* 34 (2000) 3973–3980.
- [28] A.K. Hanson, N.W. Tindale, M.A.R. Abdel-Moati, An equatorial Pacific rain event: influence on the distribution of iron and hydrogen peroxide in surface waters, *Mar. Chem.* 75 (2001) 69–88.
- [29] J.D. Willey, R.J. Kieber, G.B. Avery Jr., Effects of rainwater iron and hydrogen peroxide on iron speciation and phytoplankton growth in seawater near Bermuda, *J. Atmos. Chem.* 47 (2004) 209–222.

- [30] R.J. Kieber, et al., Iron speciation and hydrogen peroxide concentrations in New Zealand rainwater, *Atmos. Environ.* 35 (2001) 6041–6048.
- [31] G.B.J. Avery, et al., Hydrogen peroxide at the Bermuda Atlantic Time Series Station: temporal variability of seawater hydrogen peroxide, *Mar. Chem.* 97 (2005) 236–244.
- [32] J.D. Willey, H.W. Paerl, M. Go, Impact of rainwater hydrogen peroxide on chlorophyll a content of surface Gulf Stream seawater off North Carolina, USA, *Mar. Ecol. Prog. Ser.* 178 (1999) 145–150.
- [33] K. Asada, Production and scavenging of reactive oxygen species in chloroplasts and their functions, *Plant Physiol.* 141 (2) (2006) 391–396.
- [34] J.A. Imlay, The molecular mechanisms and physiological consequences of oxidative stress: lessons from a model bacterium, *Nat. Rev. Microbiol.* 11 (7) (2013) 443–454.
- [35] A. Latifi, et al., Iron starvation leads to oxidative stress in *Anabaena* sp. strain PCC 7120, *J. Bacteriol.* 187 (18) (2005) 6596–6598.
- [36] H. Schoffman, et al., Iron-nutrient interactions within phytoplankton, *Front. Plant Sci.* 7 (2016).
- [37] F. Kennedy, A. Martin, A. McMin, Rapid changes in spectral composition after darkness influences nitric oxide, glucose and hydrogen peroxide production in the Antarctic diatom *Fragilariopsis cylindrus*, *Polar Biol.* 44 (7) (2021) 1289–1303.
- [38] D.L. Kirchman (Ed.), *Microbial Ecology of the Oceans*, second ed., Wiley, Hoboken, NJ, 2008.
- [39] C. Ross, K.L.V. Alstyne, Intraspecific variation in stress-induced hydrogen peroxide scavenging by the ulvoid macroalga *Ulva lactuca*, *J. Phycol.* 43 (3) (2007) 466–474.
- [40] T.K. Prasad, et al., Evidence for chilling-induced oxidative stress in maize seedlings and a regulatory role for hydrogen peroxide, *Plant Cell* 6 (1994) 65–74.
- [41] I.S. Kong, et al., Role of catalase and oxyR in the viable but nonculturable state of *Vibrio vulnificus*, *FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Ecol.* 50 (3) (2004) 133–142.
- [42] T. Higuchi, et al., The synergistic effects of hydrogen peroxide and elevated seawater temperature on the metabolic activity of the coral *Galaxea fascicularis*, *Mar. Biol.* 156 (4) (2009) 589–596.
- [43] D.J. Suggett, et al., Photosynthesis and production of hydrogen peroxide by Symbiodinium (Pyrrophyta) phylotypes with different thermal tolerances, *J. Phycol.* 44 (4) (2008) 948–956.
- [44] M.B. Franca, A.D. Panek, E.C.A. Eleutherio, Oxidative stress and its effects during dehydration, *Comparative Biochemistry and Physiology A - Molecular & Integrative Physiology* 146 (4) (2007) 621–631.
- [45] C.J. Dedman, et al., Mechanisms of silver nanoparticle toxicity on the marine cyanobacterium *Prochlorococcus* under environmentally-relevant conditions, *Sci. Total Environ.* 747 (2020), 141229.
- [46] W. Lin, et al., Effect of microplastics PAN polymer and/or Cu²⁺ pollution on the growth of *Chlorella pyrenoidosa*, *Environ. Pollut.* 265 (Pt A) (2020), 114985.
- [47] L.G.A. Barboza, et al., Microplastics in wild fish from North East Atlantic Ocean and its potential for causing neurotoxic effects, lipid oxidative damage, and human health risks associated with ingestion exposure, *Sci. Total Environ.* 717 (2020), 134625.
- [48] R. Cao, et al., Seawater acidification increases copper toxicity: a multi-biomarker approach with a key marine invertebrate, the Pacific Oyster *Crassostrea gigas*, *Aquat. Toxicol.* 210 (2019) 167–178.
- [49] P.S.M. Celis-Plá, et al., Antarctic intertidal macroalgae under predicted increased temperatures mediated by global climate change: would they cope? *Sci. Total Environ.* 740 (2020), 140379.
- [50] J. Engström-Öst, et al., Eco-physiological responses of copepods and pteropods to ocean warming and acidification, *Sci. Rep.* 9 (1) (2019) 4748.
- [51] D. Fu, et al., Aged microplastics polyvinyl chloride interact with copper and cause oxidative stress towards microalgae *Chlorella vulgaris*, *Aquat. Toxicol.* 216 (2019), 105319.
- [52] Y.H. Lee, et al., Effects of ocean acidification on life parameters and antioxidant system in the marine copepod *Tigriopus japonicus*, *Aquat. Toxicol.* 212 (2019) 186–193.
- [53] Q. Liu, et al., ROS changes are responsible for tributyl phosphate (TBP)-induced toxicity in the alga *Phaeodactylum tricoratum*, *Aquat. Toxicol.* 208 (2019) 168–178.
- [54] D.C. Luz, et al., Oxidative stress in the hydrocoral *Millepora alcornis* exposed to CO₂-driven seawater acidification, *Coral Reefs* 37 (2) (2018) 571–579.
- [55] I.D. Martin, G.L. Mackie, M.A. Baker, Acute toxicity tests and pulsed-dose delayed mortality at 12 °C and 22 °C in the zebra mussel (*Dreissena polymorpha*), *Arch. Environ. Contam. Toxicol.* 24 (3) (1993) 389–398.
- [56] A. Nardi, et al., Oxidative and interactive challenge of cadmium and ocean acidification on the smooth scallop *Flexopecten glaber*, *Aquat. Toxicol.* 196 (2018) 53–60.
- [57] Y.M. Zhang, et al., Nitrogen starvation induced oxidative stress in an oil-producing green alga *Chlorella sorokiniana* C3, *PLoS One* 8 (7) (2013).
- [58] S.G. Rhee, H₂O₂, a necessary evil for cell signaling, *Science* 312 (5782) (2006) 1882–1883.
- [59] K.L. Visick, E.G. Ruby, The periplasmic, group III catalase of *Vibrio fischeri* is required for normal symbiotic competence and is induced both by oxidative stress and by approach to stationary phase, *J. Bacteriol.* 180 (8) (1998) 2087–2092.
- [60] B.M. Babior, Phagocytes and oxidative stress, *Am. J. Med.* 109 (1) (2000) 33–44.
- [61] R.E. McDowell, et al., Reactive oxygen species and the antarctic macroalgal wound response, *J. Phycol.* 50 (1) (2014) 71–80.
- [62] J. Collen, M. Pedersen, A stress-induced oxidative burst in *Euchemum platycladum* (Rhodophyta), *Physiol. Plantarum* 92 (3) (1994) 417–422.
- [63] F.C. Kupper, et al., Oligoalginate recognition and oxidative burst play a key role in natural and induced resistance of sporophytes of Laminariales, *J. Chem. Ecol.* 28 (10) (2002) 2057–2081.
- [64] K. Bouarab, et al., Sulfated oligosaccharides mediate the interaction between a marine red alga and its green algal pathogenic endophyte, *Plant Cell* 11 (9) (1999) 1635–1650.
- [65] F. Weinberger, M. Friedlander, H.G. Hoppe, Oligoagars elicit a physiological response in *Gracilaria conferta* (Rhodophyta), *J. Phycol.* 35 (4) (1999) 747–755.
- [66] C. Ross, et al., Evidence of a latent oxidative burst in relation to wound repair in the giant unicellular chlorophyte *Dasycladus vermicularis*, *J. Phycol.* 41 (3) (2005) 531–541.
- [67] R. Armoza-Zvuloni, et al., Rapid hydrogen peroxide release from the coral *Stylophora pistillata* during feeding and in response to chemical and physical stimuli, *Sci. Rep.* 6 (2016).
- [68] R.E. McDowell, et al., Reactive oxygen species as a marine grazing defense: H₂O₂ and wounded *Ascoseira mirabilis* both inhibit feeding by an amphipod grazer, *J. Exp. Mar. Biol. Ecol.* 458 (2014) 34–38.
- [69] R.E. McDowell, et al., Control of grazing by light availability via light-dependent, wound-induced metabolites: the role of reactive oxygen species, *J. Exp. Mar. Biol. Ecol.* 477 (2016) 86–91.
- [70] K.M. Sutherland, et al., Extracellular superoxide production by key microbes in the global ocean, *Limnol. Oceanogr.* 64 (6) (2019) 2679–2693.
- [71] J.M. Diaz, S. Plummer, Production of extracellular reactive oxygen species by phytoplankton: past and future directions, *J. Plankton Res.* 40 (6) (2018) 655–666.
- [72] C.M. Hansel, J.M. Diaz, S. Plummer, Tight regulation of extracellular superoxide points to its vital role in the physiology of the globally relevant Roseobacter clade, *mBio* 10 (2) (2019).
- [73] A.L. Rose, The influence of extracellular superoxide on iron redox chemistry and bioavailability to aquatic microorganisms, *Front. Microbiol.* 3 (2012).
- [74] J.M. Diaz, et al., Production of extracellular superoxide and hydrogen peroxide by five marine species of harmful bloom-forming algae, *J. Plankton Res.* 40 (6) (2018) 667–677.
- [75] R.J. Schneider, et al., Species-level variability in extracellular production rates of reactive oxygen species by diatoms, *Front. Chem.* 4 (2016).
- [76] D. Kim, et al., Extremely high level of reactive oxygen species (ROS) production in a newly isolated strain of the dinoflagellate *Karenia mikimotoi*, *Eur. J. Phycol.* 54 (4) (2019) 632–640.
- [77] K. Cho, et al., Haemolytic activity and reactive oxygen species production of four harmful algal bloom species, *Eur. J. Phycol.* 52 (3) (2017) 311–319.
- [78] K. Yuasa, et al., Nutrient deficiency stimulates the production of superoxide in the noxious red-tide-forming raphidophyte *Chattonella antiqua*, *Harmful Algae* 99 (2020).
- [79] A. Vardi, et al., Programmed cell death of the dinoflagellate *Peridinium gatunense* is mediated by CO₂ limitation and oxidative stress, *Curr. Biol.* 9 (18) (1999) 1061–1064.
- [80] I.M. Twigg, et al., Revealing hydrogen peroxide as an external stressor in macrophyte-dominated coastal ecosystems, *Oecologia* 193 (3) (2020) 583–591.
- [81] T. Zhang, et al., Extensive dark biological production of reactive oxygen species in brackish and freshwater ponds, *Environ. Sci. Technol.* 50 (6) (2016) 2983–2993.
- [82] M.J. Twiner, C.G. Trick, Possible physiological mechanisms for production of hydrogen peroxide by the ichthyotoxic flagellate *Heterosigma akashiwo*, *J. Plankton Res.* 22 (10) (2000) 1961–1975.
- [83] T. Oda, et al., Hydroxyl radical generation by red tide algae, *Arch. Biochem. Biophys.* 294 (1) (1992) 38–43.
- [84] T. Oda, et al., Generation of reactive oxygen species by raphidophcean phytoplankton, *Biosci. Biotech. Biochem.* 61 (10) (1997) 1658–1662.
- [85] C. Yang, L. Albright, A. Yousif, Oxygen-radical-mediated effects of the toxic phytoplankter *Heterosigma carterae* on juvenile rainbow trout *Oncorhynchus mykiss*, *Dis. Aquat. Org.* 23 (2) (1995) 101–108.
- [86] K. Cho, et al., Generation of reactive oxygen species (ROS) by harmful algal bloom (HAB)-forming phytoplankton and their potential impact on surrounding living organisms, *Antioxidants* 11 (2) (2022) 206.
- [87] J.J. Dorantes-Aranda, et al., Progress in understanding algal bloom-mediated fish kills: the role of superoxide radicals, phycotoxins and fatty acids, *PLoS One* 10 (7) (2015), e0133549.
- [88] H. Urakawa, et al., Subtropical freshwater cyanobacterial blooms as hydrogen peroxide hot spots, *Environ. Sci. Technol. Lett.* 8 (10) (2021) 911–917.
- [89] S.A. Murphy, et al., Hydrous ferric oxides in sediment catalyze formation of reactive oxygen species during sulfide oxidation, *Front. Mar. Sci.* 3 (2016).
- [90] T.J. Shaw, et al., Fe-catalyzed sulfide oxidation in hydrothermal plumes is a source of reactive oxygen species to the ocean, *Proc. Nat. Acad. Sci. U.S.A.* vol. 118 (40) (2021).
- [91] A.L. Rose, et al., Dynamics of nonphotochemical superoxide production and decay in the Great Barrier Reef lagoon, *Limnol. Oceanogr.* 55 (4) (2010) 1521–1536.
- [92] K.M. Sutherland, et al., Spatial heterogeneity in particle-associated, light-independent superoxide production within productive coastal waters, *Journal of Geophysical Research-Oceans* 125 (10) (2020).
- [93] W.J. Cooper, R.G. Zepp, Hydrogen peroxide decay in waters with suspended soils: evidence for biologically mediated processes, *Can. J. Fish. Aquat. Sci.* 47 (1990) 888–893.
- [94] R.G. Petasne, R.G. Zika, Hydrogen peroxide lifetimes in south Florida coastal and offshore waters, *Mar. Chem.* 56 (1997) 215–225.
- [95] J.J. Morris, R.E. Lenski, E.R. Zinser, The Black Queen Hypothesis: evolution of dependencies through adaptive gene loss, *mBio* 3 (2) (2012) e00036-12.

- [196] G. Regelsberger, et al., Occurrence and biochemistry of hydroperoxidases in oxygenic phototrophic prokaryotes (cyanobacteria), *Plant Physiol. Biochem.* 40 (2002) 479–490.
- [197] B. Hofmann, H.J. Hecht, L. Flohe, Peroxiredoxins, *Biol. Chem.* 383 (3–4) (2002) 347–364.
- [198] L.C. Seaver, J.A. Imlay, Alkyl hydroperoxide reductase is the primary scavenger of endogenous hydrogen peroxide in *Escherichia coli*, *J. Bacteriol.* 183 (24) (2001) 7173–7181.
- [199] K. Mukai, et al., Effects of light and hydrogen peroxide on gene expression of newly identified antioxidant enzymes in the harmful algal bloom species *Chattonella marina*, *Eur. J. Phycol.* 54 (3) (2019) 393–403.
- [200] K. Mukai, et al., Gene structure and cDNA sequence of 2-Cys peroxiredoxin in the harmful algal bloom species *Chattonella marina* and its gene transcription under different light intensities, *Eur. J. Phycol.* 53 (1) (2017) 29–38.
- [201] X. Qiu, et al., Time-series responses in photosynthetic activity, 2-cysteine peroxiredoxin gene expression, and proteomics of *Chattonella marina* var. *antiqua* under different oxidative stress, *Harmful Algae* 94 (2020), 101808.
- [202] M.E. Perez-Perez, et al., A comprehensive analysis of the peroxiredoxin reduction system in the cyanobacterium *Synechocystis* sp. strain PCC 6803 reveals that all five peroxiredoxins are thioredoxin dependent, *J. Bacteriol.* 191 (24) (2009) 7477–7489.
- [203] J.W. Moffett, O.C. Zafriou, An investigation of hydrogen peroxide chemistry in surface waters of Vineyard Sound with $H_2^{18}O_2$, *Limnol. Oceanogr.* 35 (6) (1990) 1221–1229.
- [204] K.M. Sutherland, et al., The redox fate of hydrogen peroxide in the marine water column, *Limnol. Oceanogr.* 66 (10) (2021), 3828–2841.
- [205] B. Halliwell, J.M.C. Gutteridge, *Free Radicals in Biology and Medicine*, fourth ed., Oxford University Press, New York, 2007, p. 851.
- [206] G. Naclerio, et al., *Bacillus subtilis* vegetative catalase is an extracellular enzyme, *Appl. Environ. Microbiol.* 61 (12) (1995) 4471–4473.
- [207] D.J. Hassett, et al., A protease-resistant catalase, KatA, released upon cell lysis during stationary phase is essential for aerobic survival of a *Pseudomonas aeruginosa* *oxyR* mutant at low cell densities, *J. Bacteriol.* 182 (16) (2000) 4557–4563.
- [208] A.N. Eremin, R.V. Mikhailova, D.I. Metelitsa, Comparative kinetic characterization of catalases of the genus *Penicillium*, *Appl. Biochem. Microbiol.* 36 (3) (2000) 221–226.
- [209] J.R. Brum, et al., Mortality in the oceans: causes and consequences, in: P.F. Kemp (Ed.), *Eco-DAS IX Symposium Proceedings*, ASLO, 2014, pp. 16–48.
- [210] D.L. Angel, et al., Catalase activity in macro- and microorganisms as an indicator of biotic stress in coastal waters of the eastern Mediterranean Sea, *Helgol. Mar. Res.* 53 (1999) 209–218.
- [211] L.C. Seaver, J.A. Imlay, Hydrogen peroxide fluxes and compartmentalization inside growing *Escherichia coli*, *J. Bacteriol.* 183 (24) (2001) 7182–7189.
- [212] J.J. Morris, Black Queen evolution: the role of leakiness in structuring microbial communities, *Trends Genet.* 31 (8) (2015) 475–482.
- [213] D.E. Latch, K. McNeill, Microheterogeneity of singlet oxygen distributions in irradiated humic acid solutions, *Science* 311 (5768) (2006) 1743–1747.
- [214] A.W. Vermilyea, T.C. Dixon, B.M. Voelker, Use of $H_2O_2-^{18}O$ to measure absolute rates of dark H_2O_2 production in freshwater systems, *Environ. Sci. Technol.* 44 (8) (2010) 3066–3072.
- [215] C.J. Miller, A.L. Rose, T.D. Waite, Importance of iron complexation for fenton-mediated hydroxyl radical production at circumneutral pH, *Front. Mar. Sci.* 3 (134) (2016).
- [216] W.H. Koppenol, J.F. Liebman, The oxidizing nature of the hydroxyl radical. A comparison with the ferryl ion (FeO^{2+}), *J. Phys. Chem.* 88 (1) (1984) 99–101.
- [217] A.N. Pham, et al., Fenton-like copper redox chemistry revisited: hydrogen peroxide and superoxide mediation of copper-catalyzed oxidant production, *J. Catal.* 301 (2013) 54–64, 0.
- [218] S.E. Page, et al., Evidence for dissolved organic matter as the primary source and sink of photochemically produced hydroxyl radical in arctic surface waters, *Environ. Sci.: Processes & Impacts* 16 (4) (2014) 807–822.
- [219] C. von Sonntag, et al., The fate of peroxy radicals in aqueous solution, *Water Sci. Technol.* 35 (4) (1997) 9–15.
- [220] S. Garg, A.L. Rose, T.D. Waite, Production of reactive oxygen species on photolysis of dilute aqueous quinone solutions, *Photochem. Photobiol.* 83 (4) (2007) 904–913.
- [221] N. d'Alessandro, et al., Reaction of superoxide with phenoxyl-type radicals, *J. Chem. Soc. Perkin Transac.* 2 (9) (2000) 1862–1867.
- [222] D.M. Le Roux, L.C. Powers, N.V. Blough, Photoproduction rates of one-electron reductants by chromophoric dissolved organic matter via fluorescence spectroscopy: comparison with superoxide and hydrogen peroxide rates, *Environ. Sci. Technol.* 55 (17) (2021) 12095–12105.
- [223] R.M. Cory, D.M. McKnight, Fluorescence spectroscopy reveals ubiquitous presence of oxidized and reduced quinones in dissolved organic matter, *Environ. Sci. Technol.* 39 (21) (2005) 8142–8149.
- [224] J.V. Goldstone, B.M. Voelker, Chemistry of superoxide radical in seawater: CDOM associated sink of superoxide in coastal waters, *Environ. Sci. Technol.* 34 (2000) 1043–1048.
- [225] M.I. Heller, P.L. Croot, Kinetics of superoxide reactions with dissolved organic matter in tropical Atlantic surface waters near Cape Verde (TENATSO), *J. Geophys. Res.* 115 (C12) (2010), C12038.
- [226] M.I. Heller, P.L. Croot, Superoxide decay kinetics in the Southern Ocean, *Environ. Sci. Technol.* 44 (1) (2010) 191–196.
- [227] D.W. King, et al., Measurement of antioxidant activity toward superoxide in natural waters, *Front. Mar. Sci.* 3 (2016).
- [228] P. Sanchez-Cruz, et al., Metal-independent reduction of hydrogen peroxide by semiquinones, *Chem. Res. Toxicol.* 27 (8) (2014) 1380–1386.
- [229] S.E. Page, et al., Hydroxyl radical formation upon oxidation of reduced humic acids by oxygen in the dark, *Environ. Sci. Technol.* 46 (3) (2012) 1590–1597.
- [230] K.M.G. Mostofa, H. Sakugawa, Spatial and temporal variations and factors controlling the concentrations of hydrogen peroxide and organic peroxides in rivers, *Environ. Chem.* 6 (6) (2009) 524–534.
- [231] B.H.J. Bielski, et al., Reactivity of HO_2/O_2^- radicals in aqueous solution, *J. Phys. Chem. Ref. Data* 14 (4) (1985) 1041–1100.
- [232] G.V. Buxton, et al., Critical review of rate constants for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals ($-OH/O^-$) in aqueous solution, *J. Phys. Chem. Ref. Data* 17 (2) (1988) 513–886.
- [233] M. Pettine, Redox processes of chromium in sea water, in: A. Gaianguzza, E. Pelizzetti, S. Sammartano (Eds.), *Chemical Processes in Marine Environments*, Springer, Berlin, 2000, pp. 281–296.
- [234] C.H. Lamborg, et al., Dark reduction drives evasion of mercury from the ocean, *Front. Environ. Chem.* 2 (2021).
- [235] S.P. Hansard, et al., Dissolved iron(II) in the Pacific Ocean: Measurements from the PO2 and P16N CLIVAR/CO2 repeat hydrography expeditions, *Deep Sea Res. Oceanogr. Res. Pap.* 56 (7) (2009) 1117–1129.
- [236] E.G. Roy, M.L. Wells, D.W. King, Persistence of iron(II) in surface waters of the western subarctic Pacific, *Limnol. Oceanogr.* 53 (1) (2008) 89–98.
- [237] J.W. Moffett, Iron(II) in the world's oxygen deficient zones, *Chem. Geol.* 580 (2021), 120314.
- [238] D. Bianchi, et al., Global niche of marine anaerobic metabolisms expanded by particle microenvironments, *Nat. Geosci.* 11 (4) (2018) 263–+.
- [239] K.A. Hunter, P.W. Boyd, Iron-binding ligands and their role in the ocean biogeochemistry of iron, *Environ. Chem.* 4 (4) (2007) 221–232.
- [240] P.W. Boyd, M.J. Ellwood, The biogeochemical cycle of iron in the ocean, *Nat. Geosci.* 3 (10) (2010) 675–682.
- [241] M.J. Hopwood, et al., Dissolved iron(II) ligands in river and estuarine water, *Mar. Chem.* 173 (2015) 173–182.
- [242] C.J. Miller, A.L. Rose, T.D. Waite, Hydroxyl radical production by H_2O_2 -mediated oxidation of Fe(II) complexed by Suwannee River fulvic acid under circumneutral freshwater conditions, *Environ. Sci. Technol.* 47 (2) (2013) 829–835.
- [243] Y. Xiao, et al., Superoxide-driven autocatalytic dark production of hydroxyl radicals in the presence of complexes of natural dissolved organic matter and iron, *Water Res.* 177 (2020), 115782.
- [244] J.W. Moffett, R.G. Zika, Oxidation kinetics of Cu(I) in seawater: implications for its existence in the marine environment, *Mar. Chem.* 13 (3) (1983) 239–251.
- [245] M. González-Dávila, et al., Oxidation of copper(I) in seawater at nanomolar levels, *Mar. Chem.* 115 (1–2) (2009) 118–124.
- [246] B.M. Voelker, D.L. Sedlak, O.C. Zafriou, Chemistry of superoxide radical in seawater: reactions with organic Cu complexes, *Environ. Sci. Technol.* 34 (6) (2000) 1036–1042.
- [247] J.W. Moffett, R.G. Zika, Measurement of copper(I) in surface waters of the subtropical Atlantic and Gulf of Mexico, *Geochem. Cosmochim. Acta* 52 (7) (1988) 1849–1857.
- [248] D. Buerge-Weirich, B. Sulzberger, formation of Cu(I) in estuarine and marine waters: application of a new solid-phase extraction method to measure Cu(I), *Environ. Sci. Technol.* 38 (6) (2004) 1843–1848.
- [249] M.I. Heller, P.L. Croot, Copper speciation and distribution in the Atlantic sector of the Southern Ocean, *Mar. Chem.* 173 (2015) 253–268.
- [250] Wiwit, et al., Wide-range detection of Cu-binding organic ligands in seawater using reverse titration, *Mar. Chem.* 230 (2021), 103927.
- [251] G. Xing, et al., Effect of chloride and suwannee river fulvic acid on Cu speciation: implications to Cu redox transformations in simulated natural waters, *Environ. Sci. Technol.* 54 (4) (2020) 2334–2343.
- [252] F.J. Millero, V.K. Sharma, B. Karn, The rate of reduction of copper(II) with hydrogen peroxide in seawater, *Mar. Chem.* 36 (1–4) (1991) 71–83.
- [253] G. Xing, et al., pH-dependence of production of oxidants (Cu(III) and/or HO•) by copper-catalyzed decomposition of hydrogen peroxide under conditions typical of natural saline waters, *Geochem. Cosmochim. Acta* 232 (2018) 30–47.
- [254] G.W. Luther III, The role of one- and two-electron transfer reactions in forming thermodynamically unstable intermediates as barriers in multi-electron redox reactions, *Aquat. Geochem.* 16 (3) (2010) 395–420.
- [255] J.J. Morgan, Kinetics of reaction between O_2 and Mn(II) species in aqueous solutions, *Geochem. Cosmochim. Acta* 69 (1) (2005) 35–48.
- [256] B.M. Tebo, et al., Geomicrobiology of manganese(II) oxidation, *Trends Microbiol.* 13 (9) (2005) 421–428.
- [257] C.M. Hansel, Chapter two - manganese in marine microbiology, in: R.K. Poole (Ed.), *Advances in Microbial Physiology*, Academic Press, 2017, pp. 37–83.
- [258] D. Learman, et al., Constraints on superoxide mediated formation of manganese oxides, *Front. Microbiol.* 4 (262) (2013).
- [259] S.P. Hansard, H.D. Easter, B.M. Voelker, Rapid reaction of nanomolar Mn(II) with superoxide radical in seawater and simulated freshwater, *Environ. Sci. Technol.* 45 (7) (2011) 2811–2817.
- [260] D.R. Learman, et al., Formation of manganese oxides by bacterially generated superoxide, *Nat. Geosci.* 4 (2) (2011) 95–98.
- [261] K. Wuttig, M.I. Heller, P.L. Croot, Reactivity of inorganic Mn and Mn desferrioxamine B with O_2 , O_2^- , and H_2O_2 in seawater, *Environ. Sci. Technol.* 47 (18) (2013) 10257–10265.
- [262] V.E. Oldham, et al., Soluble Mn(III)–L complexes are abundant in oxygenated waters and stabilized by humic ligands, *Geochem. Cosmochim. Acta* 199 (2017) 238–246.

- [163] W.G. Sunda, S.A. Huntsman, Photoreduction of manganese oxides in seawater, *Mar. Chem.* 46 (1–2) (1994) 133–152.
- [164] V.E. Oldham, et al., Oxidative and reductive processes contributing to manganese cycling at oxic-anoxic interfaces, *Mar. Chem.* 195 (2017) 122–128.
- [165] J.P. Gustafsson, Vanadium geochemistry in the biogeosphere –speciation, solid-solution interactions, and ecotoxicity, *Appl. Geochem.* 102 (2019) 1–25.
- [166] D. Wang, S.A. Sañudo Wilhelmy, Vanadium speciation and cycling in coastal waters, *Mar. Chem.* 117 (1–4) (2009) 52–58.
- [167] L. Knežević, et al., Redox speciation of vanadium in estuarine waters using improved methodology based on anion exchange chromatography coupled to HR ICP-MS system, *Molecules* 26 (9) (2021).
- [168] B. Wehrli, W. Stumm, Oxygenation of vanadyl(IV). Effect of coordinated surface hydroxyl groups and hydroxide ion, *Langmuir* 4 (3) (1988) 753–758.
- [169] J.-H. Huang, et al., Vanadium: global (bio)geochemistry, *Chem. Geol.* 417 (2015) 68–89.
- [170] B.H.J. Bielski, et al., Reactivity of HO₂/O₂⁻ radicals in aqueous solution, *J. Phys. Chem. Ref. Data* 14 (4) (1985) 1041–1100.
- [171] H.N.R. Shankar, T. Ramasarma, Multiple reactions in vanadyl-V(IV) oxidation by H₂O₂, *Mol. Cell. Biochem.* 129 (1) (1993) 9–29.
- [172] D. Wang, Redox chemistry of molybdenum in natural waters and its involvement in biological evolution, *Front. Microbiol.* 3 (2012) 427.
- [173] P.L. Smedley, D.G. Kinniburgh, Molybdenum in natural waters: a review of occurrence, distributions and controls, *Appl. Geochem.* 84 (2017) 387–432.
- [174] H.-P. Li, et al., Superoxide production by a manganese-oxidizing bacterium facilitates iodide oxidation, *Appl. Environ. Microbiol.* 80 (9) (2014) 2693–2699.
- [175] G.T.F. Wong, L.-S. Zhang, The kinetics of the reactions between iodide and hydrogen peroxide in seawater, *Mar. Chem.* 111 (1) (2008) 22–29.
- [176] H.-P. Li, et al., Bacterial production of organic acids enhances H₂O₂-dependent iodide oxidation, *Environ. Sci. Technol.* 46 (9) (2012) 4837–4844.
- [177] G.W. Luther, Thermodynamic redox calculations for one and two electron transfer steps: implications for halide oxidation and halogen environmental cycling, in: *Aquatic Redox Chemistry*, American Chemical Society, 2011, pp. 15–35.
- [178] C.C. Winterbourn, D. Metodieva, Reactivity of biologically important thiol compounds with superoxide and hydrogen peroxide, *Free Radic. Biol. Med.* 27 (3–4) (1999) 322–328.
- [179] N.V. Blough, O.C. Zafriou, Reaction of superoxide with nitric oxide to form peroxynitrite in alkaline aqueous solution, *Inorg. Chem.* 24 (22) (1985) 3502–3504.
- [180] A.O. Adesina, H. Sakugawa, Photochemically generated nitric oxide in seawater: the peroxynitrite sink and its implications for daytime air quality, *Sci. Total Environ.* 781 (2021), 146683.
- [181] C.M. Hansel, T.G. Ferdelman, B.M. Tebo, Cryptic cross-linkages among biogeochemical cycles: novel insights from reactive intermediates, *Elements* 11 (6) (2015) 409–414.
- [182] Y. Yoshpeper, Y. Henis, Factors affecting catalase level and sensitivity to hydrogen peroxide in *Escherichia coli*, *Appl. Environ. Microbiol.* 32 (4) (1976) 465–469.
- [183] J.M.C. Gutteridge, Reactivity of hydroxyl and hydroxyl-like radicals discriminated by release of thiobarbituric acid-reactive material from deoxy sugars, nucleosides and benzoate, *Biochem. J.* 224 (1984) 761–767.
- [184] Z. Lu, et al., Effects of pyrogallol acid on *Microcystis aeruginosa*: oxidative stress related toxicity, *Ecotoxicol. Environ. Saf.* 132 (2016) 413–419.
- [185] T. Hofer, et al., Hydrogen peroxide causes greater oxidation in cellular RNA than in DNA, *Biol. Chem.* 386 (4) (2005) 333–337.
- [186] M. Tanaka, P.B. Chock, E.R. Stadtman, Oxidized messenger RNA induces translation errors, *Proc. Natl. Acad. Sci. U.S.A.* vol. 104 (1) (2007) 66–71.
- [187] K. Keyer, J.A. Imlay, Superoxide accelerates DNA damage by elevating free iron levels, *Proc. Natl. Acad. Sci. U.S.A.* 93 (1996) 13635–13640.
- [188] B.J. Bolann, R.J. Ulvik, Release of iron from ferritin by xanthine oxidase, *Biochem. J.* 243 (1987) 55–59.
- [189] K. Kojima, et al., Oxidation of elongation factor G inhibits the synthesis of the D1 protein of photosystem II, *Mol. Microbiol.* 65 (4) (2007) 936–947.
- [190] J. Tamarit, E. Cabiscol, J. Ros, Identification of the major oxidatively damaged proteins in *Escherichia coli* cells exposed to oxidative stress, *J. Biol. Chem.* 273 (5) (1998) 3027–3032.
- [191] J.M.C. Gutteridge, B. Halliwell, The measurement and mechanism of lipid peroxidation in biological systems, *Trends Biochem. Sci.* 15 (4) (1990) 129–135.
- [192] I. Arana, et al., Role of hydrogen peroxide in loss of culturability mediated by visible light in *Escherichia coli* in a fresh water ecosystem, *Appl. Environ. Microbiol.* 58 (12) (1992) 3903–3907.
- [193] T. Tanaka, et al., A hidden pitfall in the preparation of agar media undermines microorganism cultivability, *Appl. Environ. Microbiol.* 80 (24) (2014) 7659–7666.
- [194] J.J. Morris, E.R. Zinser, Continuous hydrogen peroxide production by organic buffers in phytoplankton culture media, *J. Phycol.* 49 (6) (2013) 1223–1228.
- [195] V.C. Barry, et al., Peroxide formation in bacteriological media, *Nature* 178 (4533) (1956) 596–597.
- [196] L.C. McDonald, C.R. Hackney, B. Ray, Enhanced recovery of injured *Escherichia coli* by compounds that degrade hydrogen peroxide or block its formation, *Appl. Environ. Microbiol.* 45 (2) (1983) 360–365.
- [197] M.A. Xenopoulos, D.F. Bird, Effect of acute exposure to hydrogen peroxide on the production of phytoplankton and bacterioplankton in a mesohumic lake, *Photochem. Photobiol.* 66 (4) (1997) 471–478.
- [198] M.J. Hopwood, et al., Experiment design and bacterial abundance control extracellular H₂O₂ concentrations during four series of mesocosm experiments, *Biogeosciences* 17 (5) (2020) 1309–1326.
- [199] J.J. Morris, et al., Facilitation of robust growth of *Prochlorococcus* colonies and dilute liquid cultures by “helper” heterotrophic bacteria, *Appl. Environ. Microbiol.* 74 (14) (2008) 4530–4534.
- [200] J.J. Morris, et al., Dependence of the cyanobacterium *Prochlorococcus* on hydrogen peroxide scavenging microbes for growth at the ocean’s surface, *PLoS One* 6 (2) (2011), e16805.
- [201] D.J. Scanlan, et al., Ecological genomics of marine picocyanobacteria, *Microbiol. Mol. Biol. Rev.* 73 (2) (2009) 249–299.
- [202] A. Coe, et al., Survival of *Prochlorococcus* in extended darkness, *Limnol. Oceanogr.* 61 (4) (2016) 1375–1388.
- [203] S.J. Biller, et al., Heterotroph interactions alter *Prochlorococcus* transcriptome dynamics during extended periods of darkness, *mSystems* 3 (3) (2018).
- [204] L. Ma, et al., Degradation of hydrogen peroxide at the ocean’s surface: the influence of the microbial community on the realized thermal niche of *Prochlorococcus*, *ISME J.* 12 (2018) 473–484.
- [205] V.D. Samuilov, et al., Hydrogen peroxide inhibits the growth of cyanobacteria, *Biochemistry-Moscow* 64 (1) (1999) 47–53.
- [206] M. Drabkova, W. Admiraal, B. Marsalek, Combined exposure to hydrogen peroxide and light – selective effects on cyanobacteria, green algae, and diatoms, *Environ. Sci. Technol.* 41 (2007) 309–314.
- [207] M. Drabkova, et al., Selective effects of H₂O₂ on cyanobacterial photosynthesis, *Photosynthetica* 45 (3) (2007) 363–369.
- [208] M. Tichy, W. Vermaas, In vivo role of catalase-peroxidase in *Synechocystis* sp. Strain PCC 6803, *J. Bacteriol.* 181 (6) (1999) 1875–1882.
- [209] J.T. Hollibaugh, Oxygen and the activity and distribution of marine Thaumarchaeota, *Environmental Microbiology Reports* 9 (3) (2017) 186–188.
- [210] D.A. Stahl, J.R.d.l. Torre, Physiology and diversity of ammonia-oxidizing archaea, *Annu. Rev. Microbiol.* 66 (1) (2012) 83–101.
- [211] B.B. Tolar, et al., Ammonia oxidation in the ocean can be inhibited by nanomolar concentrations of hydrogen peroxide, *Front. Mar. Sci.* 3 (237) (2016).
- [212] W. Qin, et al., Influence of oxygen availability on the activities of ammonia-oxidizing archaea, *Environ. Microbiol. Rep.* 9 (3) (2017) 250–256.
- [213] B. Bayer, et al., Proteomic response of three marine ammonia-oxidizing archaea to hydrogen peroxide and their metabolic interactions with a heterotrophic alphaproteobacterium, *mSystems* 4 (4) (2019), e00181-19.
- [214] C. Chen, et al., Growth, physicochemical and antioxidant responses of overwintering benthic cyanobacteria to hydrogen peroxide, *Environ. Pollut.* 219 (2016) 649–655.
- [215] J.-G. Kim, et al., Hydrogen peroxide detoxification is a key mechanism for growth of ammonia-oxidizing archaea, *Proc. Natl. Acad. Sci. Unit. States Am.* 113 (28) (2016) 7888–7893.
- [216] L.E. Friedman, T.D. Gilmore, J.R. Finnerty, Intraspecific variation in oxidative stress tolerance in a model cnidarian: differences in peroxide sensitivity between and within populations of *Nematostella vectensis*, *PLoS One* 13 (1) (2018).
- [217] M.J. Hopwood, et al., Photochemical vs. Bacterial control of H₂O₂ concentration across a pCO₂ gradient mesocosm experiment in the subtropical north Atlantic, *Front. Mar. Sci.* 5 (105) (2018).
- [218] R. Müller, et al., UV-radiation and elevated temperatures induce formation of reactive oxygen species in gametophytes of cold-temperate/Arctic kelps (Laminariales, Phaeophyceae), *Phycol. Res.* 60 (1) (2012) 27–36.
- [219] S. Goyen, et al., A molecular physiology basis for functional diversity of hydrogen peroxide production amongst *Symbiodinium* spp. (Dinophyceae), *Mar. Biol.* 164 (3) (2017).
- [220] C.J. Jang, et al., Response of the ocean mixed layer depth to global warming and its impact on primary production: a case for the North Pacific Ocean, *ICES (Int. Counc. Explor. Sea) J. Mar. Sci.* 68 (6) (2011) 996–1007.
- [221] A.R. Lopes, et al., Transgenerational exposure to ocean acidification induces biochemical distress in a keystone amphipod species (*Gammarus locusta*), *Environ. Res.* 170 (2019) 168–177.
- [222] G.M.M. Hennon, et al., The impact of elevated CO₂ on *Prochlorococcus* and microbial interactions with ‘helper’ bacterium *Alteromonas*, *ISME J.* 12 (2018) 520–531.
- [223] R.G. Zepp, B.C. Faust, J. Hoigne, Hydroxyl radical formation in aqueous reactions (ph 3–8) of iron(II) with hydrogen-peroxide - the photo-fenton reaction, *Environ. Sci. Technol.* 26 (2) (1992) 313–319.
- [224] E. Myhre Jensen, et al., Trends in de-lousing of Norwegian farmed salmon from 2000–2019—consumption of medicines, salmon louse resistance and non-medical control methods, *PLoS One* 15 (10) (2020), e0240894.
- [225] L. Duchene, In Sea Lice Fight, Salmon Farmers Phasing Out Hydrogen Peroxide, *Global Aquaculture Advocate*, 2017.
- [226] R.H. Escobar-Lux, et al., Short-term exposure to hydrogen peroxide induces mortality and alters exploratory behaviour of European lobster (*Homarus gammarus*), *Ecotoxicology and Environmental Safety*, 2020, p. 204.
- [227] R.H. Escobar-Lux, et al., The Effects of Hydrogen Peroxide on Mortality, Escape Response, and Oxygen Consumption of *Calanus* spp.F. *Facets*, vol. 4, 2019, pp. 626–637.
- [228] D. Abele-Oeschger, T. Buchner, Effect of environmental hydrogen peroxide accumulation on filtration rates of the intertidal bivalve clam *Cerastoma edule*, *Verh. Dtsch. Zool. Ges.* 88 (1995) 93.
- [229] L.M. Vera, H. Migaud, Hydrogen peroxide treatment in Atlantic salmon induces stress and detoxification response in a daily manner, *Chronobiol. Int.* 33 (5) (2016) 530–542.
- [230] K.O. Helgesen, et al., First report of reduced sensitivity towards hydrogen peroxide found in the salmon louse *Lepeophtheirus salmonis* in Norway, *Aquaculture Reports* 1 (2015) 37–42.

- [231] J. Huisman, et al., Cyanobacterial blooms, *Nat. Rev. Microbiol.* 16 (8) (2018) 471–483.
- [232] Z.E. Taranu, et al., Acceleration of cyanobacterial dominance in north temperate-subarctic lakes during the Anthropocene, *Ecol. Lett.* 18 (4) (2015) 375–384.
- [233] S.C. Chapra, et al., Climate change impacts on harmful algal blooms in US freshwaters: a screening-level assessment, *Environ. Sci. Technol.* 51 (16) (2017) 8933–8943.
- [234] H.W. Paerl, T.G. Otten, R. Kudela, Mitigating the expansion of harmful algal blooms across the freshwater-to-marine continuum, *Environ. Sci. Technol.* 52 (10) (2018) 5519–5529.
- [235] H.W. Paerl, Mitigating harmful cyanobacterial blooms in a human- and climatically-impacted world, *Life* 4 (4) (2014) 988–1012.
- [236] Y.I. Skurlatov, L.S. Ernestova, The impact of human activities on freshwater aquatic systems, *Acta Hydrochim. Hydrobiol.* 26 (1) (1998) 5–12.
- [237] G. Barroin, M. Feuillade, Hydrogen peroxide as a potential algicide for *Oscillatoria rubescens* D.C., *Wat. Res.* 20 (5) (1986) 619–623.
- [238] E.F. Weenink, et al., Combatting cyanobacteria with hydrogen peroxide: a laboratory study on the consequences for phytoplankton community and diversity, *Front. Microbiol.* 6 (2015) 714.
- [239] H.C. Matthijs, et al., Selective suppression of harmful cyanobacteria in an entire lake with hydrogen peroxide, *Water Res.* 46 (5) (2012) 1460–1472.
- [240] T. Piel, et al., Suppressing cyanobacteria with hydrogen peroxide is more effective at high light intensities, *Toxins* 12 (1) (2019).
- [241] H.C.P. Matthijs, et al., Existing and emerging cyanocidal compounds: new perspectives for cyanobacterial bloom mitigation, *Aquat. Ecol.* 50 (3) (2016) 443–460.
- [242] D. Dziga, et al., Microbial degradation of microcystins, *Chem. Res. Toxicol.* 26 (6) (2013) 841–852.
- [243] X. Huo, et al., Exposure of *Microcystis aeruginosa* to hydrogen peroxide under light: kinetic modeling of cell rupture and simultaneous microcystin degradation, *Environ. Sci. Technol.* 49 (9) (2015) 5502–5510.
- [244] L.K. Ndungu, et al., Hydrogen peroxide measurements in subtropical aquatic systems and their implications for cyanobacterial blooms, *Ecol. Eng.* 138 (2019) 444–453.
- [245] B.A. Wagstaff, et al., Dissecting the toxicity and mitigating the impact of harmful *Prymnesium* blooms in the UK waters of the Norfolk Broads, *bioRxiv*, 2020, 2020.03.26.010066.
- [246] A. Burson, et al., Termination of a toxic *Alexandrium* bloom with hydrogen peroxide, *Harmful Algae* 31 (2014) 125–135.
- [247] P.J. Häkkinen, A.M. Anesio, W. Granéli, Hydrogen peroxide distribution, production, and decay in boreal lakes, *Can. J. Fish. Aquat. Sci.* 61 (2004) 1520–1527.
- [248] J.J. Morris, The ‘helper’ phenotype: a symbiotic interaction between *Prochlorococcus* and hydrogen peroxide scavenging microorganisms, in: *Microbiology*, University of Tennessee, Knoxville, 2011, p. 187.
- [249] D. Abele, G.A. Ferreyra, I. Schloss, H₂O₂ accumulation from photochemical production and atmospheric wet deposition in Antarctic coastal and off-shore waters of Potter Cove, King George Island, South Shetland Islands, *Antarct. Sci.* 11 (2) (1999) 131–139.
- [250] J. Resing, et al., L.T.E.R. Palmer, Hydrogen peroxide in the Palmer LTER region: II. Water column distribution, *Antarct. J. U. S.* 28 (1993) 227–228.
- [251] R.G. Zika, E.S. Saltzman, W.J. Cooper, Hydrogen peroxide concentrations in the Peru upwelling area, *Mar. Chem.* 17 (1985) 265–275.
- [252] C.D. Clark, et al., Hydrogen peroxide measurements in recreational marine bathing waters in Southern California, USA, *Water Res.* 44 (7) (2010) 2203–2210.
- [253] R.G. Zika, et al., Spatial and temporal variations of hydrogen peroxide in Gulf of Mexico waters, *Geochem. Cosmochim. Acta* 49 (1985) 1173–1184.
- [254] J. Yuan, A.M. Shiller, Distribution of hydrogen peroxide in the northwest Pacific Ocean, *G-cubed* 6 (9) (2005) 1–3.
- [255] P.L. Croot, et al., Influence of the ITCZ on H₂O₂ in near surface waters in the equatorial Atlantic Ocean, *Geophys. Res. Lett.* 31 (23) (2004).
- [256] C.A. Moore, C.T. Farmer, R.G. Zika, Influence of the Orinoco River on hydrogen peroxide distribution and production in the eastern Caribbean, *J. Geophys. Res.: Oceans* 98 (C2) (1993) 2289–2298.
- [257] W.L. Miller, D.R. Kester, Peroxide variations in the sargasso sea, *Mar. Chem.* 48 (1994) 17–29.
- [258] S. Steigenberger, P.L. Croot, Identifying the processes controlling the distribution of H₂O₂ in surface waters along a meridional transect in the eastern Atlantic, *Geophys. Res. Lett.* 35 (3) (2008).
- [259] K.S. Johnson, et al., Hydrogen peroxide in the western Mediterranean Sea: a tracer for vertical advection, *Deep-Sea Res.* 36 (2) (1989) 241–254.
- [260] G. Sarthou, et al., Fe and H₂O₂ distributions in the upper water column in the Indian sector of the Southern Ocean, *Earth Planet Sci. Lett.* 147 (1–4) (1997) 83–92.
- [261] J.C. Yuan, A.M. Shiller, Hydrogen peroxide in deep waters of the north pacific ocean, *Geophys. Res. Lett.* 31 (1) (2004).
- [262] M.J. Hopwood, et al., Hydrogen peroxide in deep waters from the mediterranean sea, South Atlantic and South Pacific oceans, *Sci. Rep.* 7 (1) (2017) 43436.
- [263] W.J. Cooper, E.S. Saltzman, R.G. Zika, The contribution of rainwater to variability in surface ocean hydrogen peroxide, *J. Geophys. Res.: Oceans* 92 (C3) (1987) 2970–2980.
- [264] R. Zika, et al., H₂O₂ levels in rainwater collected in south Florida and the Bahama Islands, *J. Geophys. Res.: Oceans* 87 (C7) (1982) 5015–5017.
- [265] L.E. Burridge, et al., *The Acute Lethality of Three Anti-sea Lice Formulations: AlphaMax®, Salmosan®, and Interlox®Paramove™50 to Lobster and Shrimp*. *Aquaculture*, vols. 420–421, 2014, pp. 180–186.
- [266] B.H. Hansen, et al., Acute hydrogen peroxide (H₂O₂) exposure does not cause oxidative stress in late-copepodite stage of *Calanus finmarchicus*, *J. Toxicol. Environ. Health* 80 (16–18) (2017) 820–829.
- [267] R.S. Matthews, *Artemia salina* as a test organism for measuring superoxide-mediated toxicity, *Free Radic. Biol. Med.* 18 (5) (1995) 919–922.
- [268] M.G.D. Smit, et al., Time and concentration dependency in the potentially affected fraction of species: the case of hydrogen peroxide treatment of ballast water, *Environ. Toxicol. Chem.* 27 (3) (2008) 746–753.
- [269] T. Kanda, H. Murata, A. Kuroki, Toxicity of removal agents of red tide plankton to the fishes with special reference to the toxicity of hydrogen peroxide, iron sulfate, and iron chloride, *Suisan zoshoku* 37 (3) (1989) 221–224.
- [270] C.Z. Yang, L.J. Albright, A.N. Yousif, Oxygen-radical-mediated effects of the toxic phytoplankter *Heterosigma carterae* on juvenile rainbow trout *Oncorhynchus mykiss*, *Dis. Aquat. Org.* 23 (2) (1995) 101–108.
- [271] E.S. Reichwaldt, et al., Acute toxicological response of *Daphnia* and *Moina* to hydrogen peroxide, *J. Environ. Eng.* 138 (5) (2012) 607–611.
- [272] G.V. Peixoto, et al., Safe disposal of *E. coli* DH5α strain from liquid broth and solid substrate, *J. Environ. Stud.* 23 (1) (2020) 1–12.
- [273] D. Abele-Oeschger, H. Tüg, R. Röttgers, Dynamics of UV-driven hydrogen peroxide formation on an intertidal sandflat, *Limnol. Oceanogr.* 42 (6) (1997) 1406–1415.