



Neurotrophic, Gene Regulation, and Cognitive Functions of Carboxypeptidase E-Neurotrophic Factor-α1 and Its Variants

Lan Xiao, Xuyu Yang and Y. Peng Loh*

Section on Cellular Neurobiology, Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, United States

Carboxypeptidase E, also known as neurotrophic factor- α 1 (CPE-NF α 1), was first discovered as an exopeptidase and is known to work by cleaving C-terminal basic amino acids from prohormone intermediates to produce mature peptide hormones and neuropeptides in the endocrine and central nervous systems, respectively. CPE-NF α 1 also plays a critical role in prohormone sorting and secretory vesicle transportation. Recently, emerging studies have indicated that CPE-NF α 1 exerts multiple non-enzymatic physiological roles in maintaining normal central nervous system function and in neurodevelopment. This includes potent neuroprotective and anti-depressant activities, as well as stem cell differentiation functions. In addition, N-terminal truncated variants of CPE-NF α 1 have been identified to regulate expression of important neurodevelopmental genes. This mini-review summarizes recent advances in understanding the mechanisms underlying CPE-NF α 1's function in neuroprotection during stress and aspects of neurodevelopment.

OPEN ACCESS

Edited by:

Limei Zhang, National Autonomous University of Mexico, Mexico

Reviewed by:

Illana Gozes, Tel Aviv University, Israel Maite Montero-Hadjadje, Université de Rouen, France

> *Correspondence: Y. Peng Loh

lohp@mail.nih.gov

Specialty section:

This article was submitted to Neuroendocrine Science, a section of the journal Frontiers in Neuroscience

Received: 25 January 2019 Accepted: 01 March 2019 Published: 19 March 2019

Citation:

Xiao L, Yang X and Loh YP (2019) Neurotrophic, Gene Regulation, and Cognitive Functions of Carboxypeptidase E-Neurotrophic Factor-a1 and Its Variants. Front. Neurosci. 13:243. doi: 10.3389/fnins.2019.00243 Keywords: neuroprotection, neurotrophic factor, hippocampus, neurodevelopment, FGF2, BCL2, WNT/betacatenin, stress

INTRODUCTION

CPE-NF α 1, a member of the M14 metallocarboxypeptidase family was discovered in 1982 in bovine adrenal medulla and named as enkephalin convertase due to its enzymatic activity in processing enkephalin precursor into its mature form (Fricker and Snyder, 1983). Since then, CPE-NF α 1 has been shown to cleave C-terminal basic amino acids from the intermediates generated by proprotein convertases' action on prohormones, thereby producing bioactive hormones and neuropeptides (Hook et al., 1982; Fricker and Snyder, 1983; Fricker, 1988). In the central nervous system, CPE-NF α 1 also functions as a regulated secretory pathway sorting receptor, secretory vesicle transport regulator and mediates synaptic vesicle localization to the active zone for release (Cawley et al., 2012; Ji et al., 2017). Recent studies have indicated that CPE-NF α 1 is a new neurotrophic factor functioning extracellularly, independent of its enzymatic activity, in the adult and embryonic central nervous system (Ji et al., 2017). Human mutations of CPE-NF α 1 have been associated with obesity, diabetes, infertility, learning disabilities, and Alzheimer's disease (AD) (Alsters et al., 2015; Cheng et al., 2016a). This review presents recent advances concerning the molecular structure, distribution, and multifunctional roles of CPE-NF α 1 and its variants in the central nervous system in health and disease.

MOLECULAR STRUCTURE AND BIOSYNTHESIS OF CPE-NFα1 AND ITS VARIANTS

In mammals, WT-CPE-NFa1 gene consists of nine exons and encodes a 476 amino acid polypeptide (Jung et al., 1991; Cawley et al., 2012). Two CPE-NFa1 mRNA transcripts, 2.4 and 1.7 kb in size, respectively, have been identified by Northern Blot analysis and DNA sequencing from human cancer cells. The 2.4 kb CPE-NFa1 transcript encodes a 53 kD WT-CPE/NFa1 and the 1.7 kb transcript encodes an N-terminal truncated 40 kD CPE-NFa1 due to the intra-exonic splicing in exon1 (Yang et al., 2017). Both transcripts have also been detected in Northern blot of human hippocampal mRNA extract (Yang, our unpublished data). Three CPE-NFa1 transcripts, a 2.3 kb WT transcript and two CPE-NFα1 mRNA variants, 1.9 and 1.73 kb in size (Figure 1) have been identified in mouse embryonic brain. Unlike in human cancer cells, the two mouse CPE-NFa1 mRNA variants are generated by using alternative transcription start sites, each of them encodes an N-terminal truncated CPE-NFa1 with a molecular weight of 47 and 40 kD, respectively (Yang et al., 2017; Xiao et al., 2018).

The CPE-NFα1 protein consists of a signal peptide, a catalytic domain, and a C-terminal domain. A 3-D structure model of CPE-NFa1 shows a zinc binding site in the enzymatic domain, a prohormone sorting signal binding site, an amphipathic α-helical transmembrane domain, and a cytoplasmic tail that interacts with microtubule proteins for vesicle transport (Cawley et al., 2012). Mature CPE-NF α1 has 476-amino acids. It is synthesized as preproCPE-NFa1 that has a 25-amino acid signal peptide located at the N-terminal that directs proCPE-NFa1 to the cisternae of the rough endoplasmic reticulum (ER) where it is removed (Figure 2). The proCPE-NFa1 is then transported to the granules of the regulated secretory pathway through the Golgi complex where 17-amino acids comprising the pro-region are removed (Song and Fricker, 1995). Further processing at Arg455-Lysine 456 (Fricker and Devi, 1993) in the secretory granules yields a soluble form of CPE-NFa1 (50 kDa), while the 53 kD unprocessed form is associated with the granule membrane, with some molecules assuming a transmembrane orientation (Cawley et al., 2012).

DISTRIBUTION OF CPE-NF α 1 AND CPE-NF α 1- Δ N

In mammals, CPE-NF α 1 is generally distributed in the endocrine and nervous system, with the highest concentration in the brain (Strittmatter et al., 1984). Immunohistochemical studies showed CPE-NF α 1 is located in neuropeptide-rich regions of the brain and endocrine system, for example, in the hypothalamus, pituitary, adrenal medulla, paraventricular nucleus, hippocampus, and amygdala (Hook et al., 1985; Lynch et al., 1990). Consistently, CPE-NF α 1 mRNA is also found

to be highly expressed in these brain regions (Birch et al., 1990; MacCumber et al., 1990; Zheng et al., 1994). The retina and olfactory bulb are also highly enriched in CPE-NFa1 (Zhu et al., 2005). In the retina, CPE-NFa1 is localized to the photoreceptors and is involved in synaptic transmission to the inner retina (Zhu et al., 2005). At the cellular level, CPE-NFa1 is mainly distributed in the trans-Golgi network and dense core secretory granules of endocrine cells and peptidergic neurons (Cawley et al., 2012). CPE-NFa1 exists as two active forms, the soluble and membrane form (Figure 2). The soluble form of CPE-NFa1 functions as an enzyme that cleaves C-terminal basic residues such as arginine and lysine from intermediates derived from neuropeptide precursors or prohormones to produce biologically active peptides or hormones. The membrane form of CPE-NFa1 acts as a sorting receptor at the trans-Golgi network to direct prohormones to the regulated secretory pathway granules, and mediates secretory granule movement (Cawley et al., 2012). In contrast, the N-terminal truncated (CPE-NF α 1- Δ N) forms which lack a signal peptide do not enter the secretory pathway. The 40 kD form of CPE-NF α 1- Δ N has been shown to be localized in the cytoplasm and to be able to translocate into the nucleus where it exerts its effects on gene regulation (Qin et al., 2014) (see summary in Table 1).

MICE AND HUMANS WITH CPE-NF α 1 MUTATION EXHIBIT NEUROLOGICAL DEFICITS

CPE-NFa1 knockout (KO) mice, in addition to exhibiting endocrinological deficits such as obesity, diabetes, and infertility due to lack of enzyme activity, display a variety of behavioral abnormalities as evidenced by deficits in learning and memory in Morris water maze, object preference, and social transmission of food preference tests (Woronowicz et al., 2008). They also display depressive-like behavior (Cheng et al., 2015, 2016a). CPE^{fat/fat} mice which have a Ser202Pro mutation and lack CPE-NFa1 have a similar phenotype to the knockout mouse and exhibit anxiety-like and depression-like behaviors (Rodriguiz et al., 2013). Interestingly, a human with a null mutation also demonstrated similar symptoms, such as obesity, diabetes, hypogonadotropic hypogonadism, and impaired intellectual ability (Alsters et al., 2015). In another study, a human mutation of CPE-NFa1 with three adenosines insertions was identified, this mutation was named QQ-CPE-NFa1. Mice bearing this mutation showed neurodegeneration in the hippocampus and prefrontal cortex, deficits in neurogenesis at the dentate gyrus and hyperphosphorylation of tau (Cheng et al., 2016a). Another human CPE-NFa1 mutation, T980C, which introduces a W235R change in the catalytic domain of CPE-NFa1 caused a loss of enzyme activity, neurotoxic accumulation in the ER, resulting in ER stress and cell death when overexpressed in Neuro 2A cells. This novel single nucleotide polymorphism in the CPE-NFα1 gene found in 12.5% of the AGI_ASP population may confer neurological disorders in humans (Cong et al., 2017). These observations suggest that CPE-NFa1 plays an important

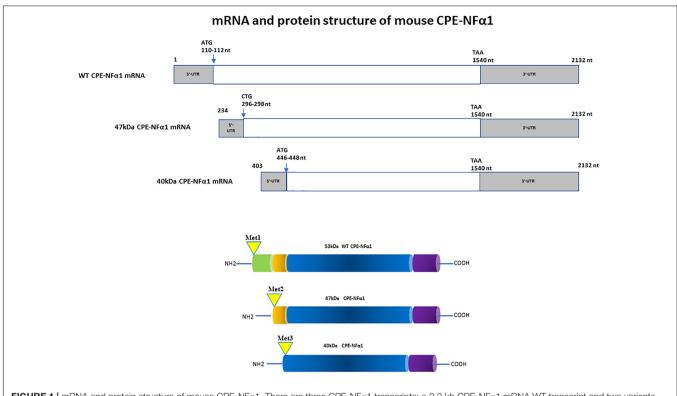


FIGURE 1 | mRNA and protein structure of mouse CPE-NF α 1. There are three CPE-NF α 1 transcripts: a 2.3 kb CPE-NF α 1 mRNA WT transcript and two variants, 1.9 and 1.73 kb transcripts generated by alternative transcription start sites in mouse embryonic brain. CPE-NF α 1 mRNA WT transcript produces a 53 kD protein, while the two mouse CPE-NF α 1 mRNA variants produce two N-terminal truncated CPE-NF α 1 proteins with 47 and 40 kD, respectively. Numbers in the 47 and 40 kD CPE mRNA refer to their relative position in the wt-CPE mRNA.

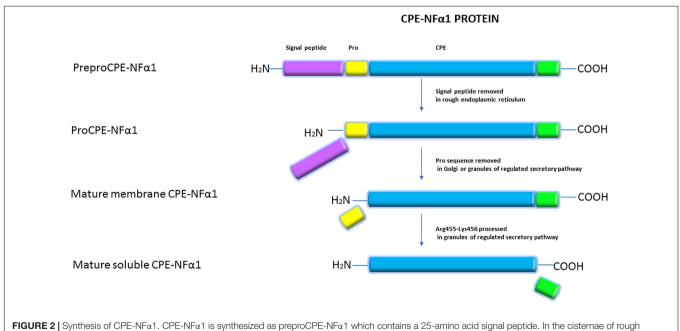


FIGURE 2 Synthesis of CPE-NF α 1 is synthesized as preproCPE-NF α 1 which contains a 25-amino acid signal peptide. In the cisternae of rough endoplasmic reticulum, the signal peptide is removed. Then, the 17-amino acid pro peptide is cleaved in the Golgi complex or granules of the regulated secretory pathway. Further processing at Arg455-Lysine 456 produces a soluble form of CPE-NF α 1, with the unprocessed form being membrane associated.

3

role in neurological function, and mutation of this gene can lead to cognitive and neurodegenerative disorders.

$\label{eq:cpe-nfall} \begin{array}{l} \text{CPE-Nf} \alpha 1 \text{ IN NEURODEGENERATIVE} \\ \text{DISORDERS AND STRESS} \end{array}$

In the cerebral cortex of AD patients, an abnormal accumulation of CPE-NF a1 was detected in dystrophic neurites surrounding amyloid plaques (Pla et al., 2013). In an AD animal model of APPswe/PS1dE9 mice, similar pattern of changes were also found. Amyloid plaques were surrounded by aberrant accumulation of CPE-NFa1 and Secretogranin III (SgIII) (Pla et al., 2013). Peptidase activity analysis of the postmortem brain from AD patients showed that soluble enzymatic activity of CPE-NFa1 was significantly increased in the Brodmann Area 21 compared with control (Weber et al., 1992). In a Cathepsins B and L double knockout mouse model that demonstrated early-onset neurodegeneration and reduction in brain size, the expression of CPE-NFa1 was upregulated by 10-fold (Stahl et al., 2007). In experimental autoimmune encephalomyelitis (EAE) animal model, CPE-NFa1 has been mapped as a EAE-linked trait loci which is associated to the severity of the disease and shown to be downregulated (Ibrahim et al., 2001; Mazon Pelaez et al., 2005). These observations suggest that CPE-NFa1 is associated to the pathophysiology of neurodegenerative diseases.

Studies have shown that CPE-NF α 1 expression is up-regulated after different types of stress. CPE-NF α 1 mRNA and protein were both increased in the rat hippocampal CA1, CA3, and cortex regions 15 min after transient global ischemia followed by 8 h reperfusion (Jin et al., 2001). An accumulation of CPE-NF α 1 precursor was found in mouse cortex after focal cerebral ischemia *in vivo* and in ischemic cortical neurons *in vitro*.

TABLE 1 Comparison of CPE-NF α 1 WT and CPE-NF α 1- Δ N.

| | CPE-NFα1 WT | CPE-NFα1-ΔN |
|---|--|--|
| Biosynthesis | First transcription start site | Alternative transcription start sites |
| Intracellular location | Regulated secretory vesicles | Cytoplasm and nucleus |
| Distribution | Endocrine and CNS. Embryonic and adult brain | Embryonic, not adult brain in mouse |
| Soluble and membrane forms | Soluble and membrane associated | Soluble |
| Extracellular secretion | Secreted from vesicles to extracellular space | Not secreted |
| Signal peptide | Yes | No |
| Structure | - | Lacking N-terminal |
| Enzymatic activity | Yes | Weak enzymatic activity |
| Intracellular sorting receptor to regulatory secreted pathway | Yes | No |
| Facilitates vesicle trafficking via cytoplasmic tail | Yes | No |
| | | |

Vice versa, mice lacking CPE-NFa1 were more vulnerable to focal cerebral ischemia (Zhou et al., 2004). Stress such as cat odor upregulated CPE-NFa1 gene expression in rat amygdala (Koks et al., 2004). Mild chronic restraint stress for 7 days upregulated CPE-NF a1 expression in mouse hippocampal CA3 region (Murthy et al., 2013). Reduced levels of CPE-NFa1 were found in the offspring of pregnant ewes that experienced aversive interaction with human handling, along with abnormal corticolimbic dendritic spine morphology (Coulon et al., 2013). Primary cultured hippocampal neurons derived from CPE-NFα1-KO mice tend to die more rapidly than that from wild type (WT) mice (Cheng et al., 2013). Low-potassium-induced apoptosis in cultured primary cerebellar granule neurons from CPE-NF α 1^{+/-} mice are much higher than that from CPE-NF α 1^{+/+} mice (Koshimizu et al., 2009). Overexpression of CPE-NFα1 in rat primary hippocampal neurons prevented hydrogen peroxide-induced neurotoxicity (Woronowicz et al., 2008). All these studies indicate that stress modulates CPE-NFa1 expression in the central nervous system (CNS).

CPE-NFα1 AS A NEUROTROPHIC FACTOR IN NEUROPROTECTION AND ANTI-DEPRESSION

CPE-NFa1 plays multiple intracellular roles in the CNS independent of its enzymatic activity. It functions as a regulated secretory pathway sorting receptor for proneuropeptides (Cool et al., 1997) and proBDNF (Lou et al., 2005), mediates anterograde transport of neuropeptide and BDNF vesicles to the plasma membrane via cytoplasmic tail interaction with dynactin/dynein (Park et al., 2008), and localizes synaptic vesicles to the actin-rich pre-active zone in hypothalamic neurons through actin binding protein y-adducin (Lou et al., 2010). While the mechanisms underlying these cell biological actions of CPE-NFa1 are well understood, those governing the neuroprotective effects of CPE-NFa1 during stress are just emerging. In 2013, a study showed that CPE-NFa1 secreted from hippocampal neurons, acts as a neuroprotective factor extracellularly, independent of its enzymatic activity (Cheng et al., 2013). Secreted rat WT CPE-NFa1 and CPE-NFa1-E300Q (mutant form with no enzymatic activity) (Qian et al., 1999) in medium from transduced cultured primary rat hippocampal neurons protected them from cell death induced by H2O2mediated oxidative stress and glutamate excitotoxicity. Results from experiments with recombinant WT CPE-NFa1 or CPE-NFa1-E342Q (mouse homolog of E300Q) demonstrated similar effects. In addition, hippocampal neurons from CPE-NFa1-KO mice at embryonic day 17 (E17) in culture exhibited higher death rate than WT E17 neurons, and this was reversed by treatment with recombinant WT CPE-NFa1 (Cheng et al., 2013). Studies on the mechanism underlying the neuroprotective action of CPE-NFα1 using rat primary hippocampal neurons in culture revealed that when these cells were stressed with H2O2, it activated the ERK and AKT signaling pathways, presumably by binding to a cognate receptor, which then up-regulated expression of Bcl2, a pro-survival mitochondrial protein, and down-regulated

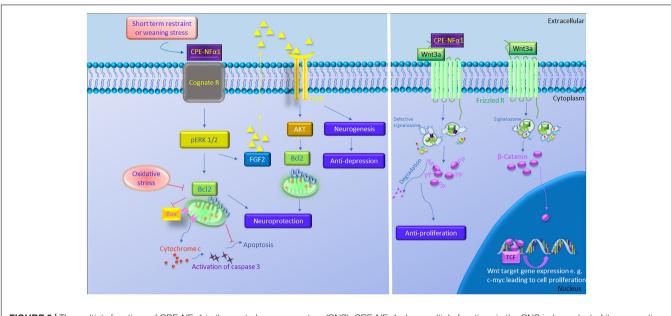
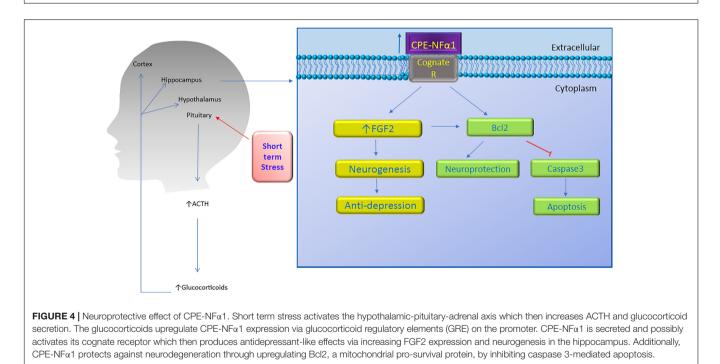


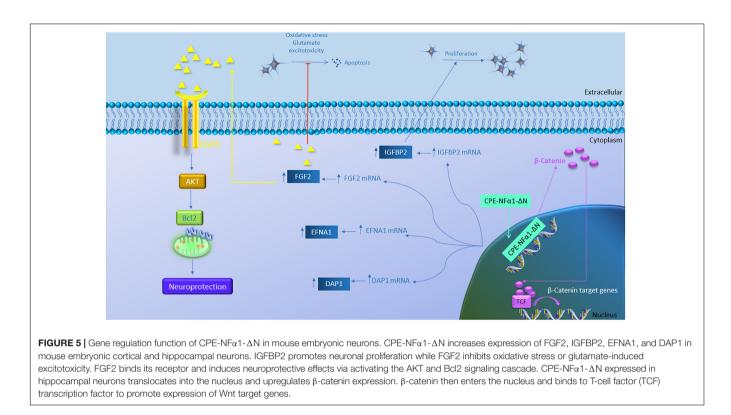
FIGURE 3 The multiple functions of CPE-NF α 1 in the central nervous system (CNS). CPE-NF α 1 plays multiple functions in the CNS independent of its enzymatic activity. Short term restraint or weaning stress increases CPE-NF α 1 which binds to a cognate receptor and upregulates ERK1/2 and Bcl2, and decreases Bax and cytochrome c-induced activation of caspase 3 and apoptosis. CPE-NF α 1 also increases FGF2 that activates AKT and Bcl2 signaling and increases hippocampal neurogenesis and produces antidepressive-like effect. CPE-NF α 1 can also bind to Frizzled receptor – Wnt3a complex, enters the endosome that contains Wnt signaling factors and subsequently impairs the formation of signalosome. This results in β -catenin being degraded, rather than translocation into the nucleus and binds to T-cell factor transcription factor (TCF) to activate expression of Wnt target genes. Signalosome Wnt Signalosome ACSK-3 β Axin.



expression of Caspase 3 to mediate neuroprotection (Cheng et al., 2013) (see Figure 3).

In an *in vivo* mouse model, complete degeneration of hippocampal CA3 region was observed in CPE-NF α 1-KO mice at 4-weeks of age after weaning stress which included maternal separation, ear tagging, and tail snipping at 3 weeks

of age (Woronowicz et al., 2008). CPE-NF α 1-KO mice that were not subjected to the weaning process at 3 weeks of age showed no degeneration of the CA3 region examined at week 4, indicating that this neurodegeneration was not due to a neurodevelopmental defect. Treatment with oral carbamazepine at 50 mg/kg daily for 2 weeks beginning at 2 weeks of age



with the weaning process at 3 weeks of age, revealed no degeneration of the CA3 region when examined at 4 weeks of age (Woronowicz et al., 2012). These observations suggested that the CA3 pyramidal neurons underwent apoptosis due to glutamate excitotoxicity during the social and physical stress following the weaning protocol. In contrast, WT-mice, did not show any degeneration of the CA3 region after weaning stress. In another mouse model, adult CPE^{fat/fat} mice lacking CPE-NFa1, also showed degeneration of the CA3 region (Zhou, personal communication). These findings together with the in vitro studies (Cheng et al., 2013) support the hypothesis that CPE-NFα1 acts as a neurotrophic factor to protect the pyramidal neurons in the CA3 region in the hippocampus from stressinduced degeneration. Since BDNF is expressed in CPE-NFa1-KO mice (Xiao et al., 2017), yet they showed neurodegeneration after weaning, indicates that BDNF could not protect the CA3 neurons in lieu of CPE-NFa1. Further demonstration that CPE-NFa1 is a neuroprotective factor in vivo during stress came from studies showing that chronic restraint stress of mice for 1 h/day for 7 days resulted in an increase in CPE-NFa1 mRNA and protein expression in the hippocampus, with no evidence of neurodegeneration despite increased circulating corticosterone levels under this stress paradigm (Murthy et al., 2013). This up-regulation in expression of CPE-NFα1 concurrs with in vitro evidence showing that the CPE-NFa1 promoter has a glucocorticoid binding domain and that dexamethasone up-regulated the expression of CPE-NFα1 (Murthy et al., 2013). These mice also showed an increase in phosphorylation of Akt and Bcl2 expression; however, Bax, a pro-apoptotic mitochondria protein was decreased in the hippocampus (Murthy et al., 2013).

In contrast, CPE-NFa1-KO mice subjected to the same stress paradigm showed no change in Akt phosphorylation, a decrease in expression of Bcl2 protein and an increase in Bax protein in the hippocampus compared to WT mice (Murthy et al., 2013). In vitro and in vivo evidence taken together indicate that during emotional and physical stress, secretion of glucocorticoid increases CPE-NFa1 expression at the transcriptional and translational level in the hippocampus which in turn leads to neuroprotection of the CA1-3 neurons by acting extracellularly as a trophic factor to activate Erk or Akt signaling and increase Bcl2 pro-survival protein expression (see Figures 3, 4). CPE-NFa1 also up-regulates the expression of FGF2 (Cheng et al., 2015), which has been shown to mediate protection against amyloid beta- or glutamate-induced neurotoxicity in hippocampal or cortical neurons via the Akt-Bcl2 signaling pathway (Qin et al., 2014; Cheng et al., 2016b; Figures 3, 5).

Behavioral analysis revealed that the CPE-NF α 1-KO and the Cpe^{fat/fat} mice exhibited depressive-like behavior as evidenced by increased immobility time in the forced-swimming test (Rodriguiz et al., 2013; Cheng et al., 2015). CPE-NF α 1-KO mice showed decreased levels of FGF2 (Cheng et al., 2015), a protein demonstrated to be decreased in post mortem brains of patients with major depressive disorder (Evans et al., 2004). In addition, after long-term chronic restraint stress, mice showed depressive-like behavior, and reduced CPE-NF α 1, FGF2 and doublecortin, a marker for neuroblasts in their hippocampus. Interestingly, injection of 5 ng/g recombinant FGF2 into CPE-NF α 1-KO mouse for 30 consecutive days completely reversed the decreased number of doublecortin positive neurons in the subgranular zone of hippocampus and depressive-like behaviors as evidenced by

decreased immobility time in the forced-swim test (Cheng et al., 2015). *In vitro* studies showed that incubation of rat cultured hippocampal neurons with CPE-NF α 1 enhanced FGF2 mRNA and protein expression which was inhibited by the Sp1 inhibitor mithramycin A, a transcription inhibitor actinomycin, and ERK inhibitor U0126, suggesting that CPE-NF α 1 can upregulate FGF2 via ERK-Sp1 signaling cascade (Cheng et al., 2015). Thus, CPE-NF α 1 may act as an anti-depressant through up-regulating FGF2 expression in the hippocampus.

ROLE OF CPE-NF α 1 AND CPE-NF α 1- Δ N VARIANTS IN NEURODEVELOPMENT

In mice and rats, CPE-NFa1 mRNA is expressed during embryogenesis as early as E5.5 mainly in the developing nervous system (Zheng et al., 1994; Selvaraj et al., 2017). In mice, the expression of CPE-NF α 1-WT and the two CPE-NF α 1- Δ N transcripts were detected in E8.5 embryonic brain and their expression peaked at embryonic day (E)10.5, then decreased from E12.5 to E16.5, and increased again at postnatal day 1 (P1). Interestingly, none of the CPE-NF α 1- Δ N mRNAs were found in adult mouse hippocampus or other organs such as liver, heart or lung, suggesting their critical role during embryonic neurodevelopment (Xiao et al., 2018). Studies of dendritic arborization in pyramidal layer V of cerebral cortex and hippocampal CA1 region in 14-week-old CPE-NFa1-KO mice revealed abnormal dendritic pruning and increased number of non-functional D-type spines (Woronowicz et al., 2010), suggesting a role of CPE-NFa1 in modeling the cytoarchitecture of the brain during development. Indeed, CPE-NFa1 has been shown to regulate NGF-induced neurite outgrowth via interacting with Wnt-3a and Wnt-5a in PC12 cells and mouse primary cortical neurons (Selvaraj et al., 2015). CPE-NFa1 is highly expressed in neural stem cells (Lee et al., 2012) and has been shown to inhibit embryonic neural stem cell proliferation by downregulating Wnt signaling pathway and β-catenin (Selvaraj et al., 2017). Studies have revealed that CPE-NFa1 forms a complex with Wnt3a ligand and Frizzled receptor to inhibit the Wnt3a signaling cascade (Figure 3; Skalka et al., 2013). Moreover, exogenous application of CPE-NFa1 and a nonenzymatic form of CPE-NF a1(E300Q) induced differentiation of E14.5 cortical neural stem cells into astrocytes via activating the ERK1/2-Sox9 signaling pathway to enhance glial fibrillary acidic protein (GFAP) expression (Selvaraj et al., 2017). This corroborates the 49% decrease in astrocyte numbers observed in the neocortex of mouse neonates in vivo at postnatal day 1 (P1). In another study, electroporation of CPE-NFa1 shRNA into E14.5 mouse embryos significantly decreased migration of neurons to cortex plate at E17.5, possibly due to the failure of transition from multipolar neurons into a bipolar morphology in the intermediate zone (Liang et al., 2018). Neuronal migration seems to depend on the function of the CPE-NFa1 cytoplasmic tail which can interact with dynactin, an adaptor protein that interacts with microtubules. Furthermore, this study also showed that CPE-NFα1 regulates dendrite morphology in vivo and in rat primary cultured hippocampal neurons (Liang et al., 2018).

While CPE-NFa1 acts extracellularly as an embryonic stem cell differentiation and anti-proliferation factor, studies have indicated that CPE-NF α 1- Δ N could serve as a master regulator of genes involved in neurodevelopment (Figure 5 and Table 2). CPE-NF α 1- Δ N (40 kD variant), but not CPE-NF α 1 (Xiao, our unpublished data) was found to significantly up-regulate expression of four genes, fibroblast growth factor 2 (FGF2) (Qin et al., 2014), insulin-like growth factor binding protein2 (IGFBP2), death associated protein (DAP1), and EphrinA1 in HT22, a hippocampal cell line and in primary mouse cortical neurons (Xiao et al., 2018). These three genes are known to mediate neuronal proliferation, cell death and neuronal migration, respectively (Murphy et al., 1990; Deiss et al., 1995; Feinstein et al., 1995; Levy-Strumpf and Kimchi, 1998; Martinez et al., 2004; Martinez and Soriano, 2005; Russo et al., 2005; Zechel et al., 2010). Overexpressing of CPE-NF α 1- Δ N in HT22 cells promoted proliferation, but was inhibited by IGFBP2 siRNA, suggesting CPE-NF α 1- Δ N regulates embryonic neuronal proliferation through increasing IGFBP2. In addition, CPE-NF α 1- Δ N-induced FGF2 expression has a neuroprotective role against glutamate and H₂O₂ neurotoxicity in rat embryonic cortical neurons (Qin et al., 2014). Since FGF2 is also known to play a role in neuronal proliferation and neurogenesis in the developing mouse cortex, CPE-NF α 1- Δ N may also regulate these events in the embryonic brain (Raballo et al., 2000). Moreover, CPE-NF α 1- Δ N has been shown to up-regulate the expression of β-catenin in the Wnt pathway in HEK-293 cells (Skalka et al., 2013) and osteosarcoma cell lines, which promoted migration of these cells (Fan et al., 2018). We have shown that neuroprotection by CPE-NF α 1- Δ N against glutamate excitotoxicity may also involve β-catenin and Wnt pathway. Glutamate treatment resulted in a decrease in β -catenin

TABLE 2 | Neuroprotective and neurodevelopmental functions of CPE-NF α 1 WT and CPE-NF α 1- Δ N.

| | Function | Reference |
|------------------------|--|----------------------------------|
| CPE-NFα1 WT | Regulates NGF-induced neurite outgrowth | Selvaraj et al., 2015 |
| | Inhibits neuronal stem cell proliferation | Selvaraj et al., 2017 |
| | Induces differentiation of neural stem cells into astrocytes | Selvaraj et al., 2017 |
| | Regulates cortical neuron migration and dendrite arborization | Liang et al., 2018 |
| | Protects primary cultured rat hippocampal neurons from H ₂ O ₂ -induced oxidative stress | Cheng et al., 2013 |
| | CA3 completely degenerated after weaning, ear tag and tail clipping at 3-week of age and prevented by carbamazepine | Woronowicz et al., 2008, 2012 |
| | Upregulates FGF2 and produces antidepressant-like effects | Cheng et al., 2015 |
| CPE-NFα1-ΔN (40 kD) | Protects primary rat embryonic cortical neurons from glutamate and H ₂ O ₂ -induced apoptosis via FGF2 | Qin et al., 2014 |
| | Increases proliferation via upregulating IGFBP2 | Xiao et al., 2018 |

level and poor cell viability compared to untreated control embryonic rat cortical neurons. However, in CPE-NF α 1- Δ Ntransduced neurons treated with glutamate, this decrease in β -catenin did not occur and the cell viability was similar to control. Treatment of cortical neurons with XAV939 which stimulates degradation of β -catenin (Huang et al., 2009) showed that in the presence of XAV939, the neuroprotective effect of CPE-NF α 1- Δ N against glutamate neurotoxicity was abolished. This suggests that the activation of the Wnt/ β -catenin pathway may also contribute to the neuroprotective mechanism of CPE-NF α 1- Δ N in embryonic neurons (Qin et al., our unpublished data). Additionally, β -catenin has been reported to mediate neuronal proliferation and differentiation of stem cells (Israsena et al., 2004). Thus, CPE-NF α 1- Δ N may also regulate these processes during development via the Wnt signaling pathway.

CONCLUSION

CPE-NFa1 is a multifunction protein (Table 2). In addition to being a prohormone and proneuropeptide processing enzyme, it is a sorting receptor and vesicle transport mediator. Recent studies have shown that it also plays a fundamental role in neurodevelopment, neurodegeneration, and cognitive functions. Null mutation of CPE-NFa1 in animal models and a human both produced similar deficits, including obesity, diabetes, and compromised cognitive function and memory. In addition, abnormal accumulation in the ER of a human mutant of CPE-NFα1 from an AD patient was found which caused neurotoxicity, neurodegeneration, and cognitive impairment in an animal model. In vitro studies using primary neurons in culture indicate that CPE-NFa1 acts extracellularly as a trophic factor, independent of enzymatic activity to mediate neuroprotection via activating several pro-survival signaling cascades such as upregulating Bcl2 and FGF2 signaling. Studies to demonstrate the neurotrophic effect in vivo will be necessary using transgenic

REFERENCES

- Alsters, S. I., Goldstone, A. P., Buxton, J. L., Zekavati, A., Sosinsky, A., Yiorkas, A. M., et al. (2015). Truncating Homozygous Mutation of Carboxypeptidase E (CPE) in a morbidly obese female with type 2 diabetes mellitus, intellectual disability and hypogonadotrophic hypogonadism. *PLoS One* 10:e0131417. doi: 10.1371/journal.pone.0131417
- Birch, N. P., Rodriguez, C., Dixon, J. E., and Mezey, E. (1990). Distribution of carboxypeptidase H messenger RNA in rat brain using in situ hybridization histochemistry: implications for neuropeptide biosynthesis. *Brain Res. Mol. Brain Res.* 7, 53–59. doi: 10.1016/0169-328X(90)90073-M
- Cawley, N. X., Wetsel, W. C., Murthy, S. R., Park, J. J., Pacak, K., and Loh, Y. P. (2012). New roles of carboxypeptidase E in endocrine and neural function and cancer. *Endocr. Rev.* 33, 216–253. doi: 10.1210/er.2011-1039
- Cheng, Y., Cawley, N. X., and Loh, Y. P. (2013). Carboxypeptidase E/NFalpha1: a new neurotrophic factor against oxidative stress-induced apoptotic cell death mediated by ERK and PI3-K/AKT pathways. *PLoS One* 8:e71578. doi: 10.1371/ journal.pone.0071578
- Cheng, Y., Cawley, N. X., Yanik, T., Murthy, S. R., Liu, C., Kasikci, F., et al. (2016a). A human carboxypeptidase E/NF-alpha1 gene mutation in an Alzheimer's disease patient leads to dementia and depression in mice. *Transl. Psychiatry* 6:e973. doi: 10.1038/tp.2016.237

mouse models to further support this hypothesis. Additionally, identifying a membrane receptor for CPE-NF α 1 will be required to substantiate a receptor-mediated mechanism for activating signaling pathways for neuroprotection and stem cell differentiation. Furthermore, CPE-NF α 1 and its derived peptides can be developed as potential therapeutic agents to treat neurodegenerative diseases. The 40 kD CPE-NF α 1- Δ N variant has been shown to activate genes involved in neurodevelopment (**Table 2**). Further studies will be required to determine the precise molecular mechanism underlying CPE-NF α 1- Δ N's role in activating such genes during embryonic development.

DATA AVAILABILITY

The datasets generated for this study are available on request to the corresponding author.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

This research was supported by the Intramural Research Program of the Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD), National Institutes of Health, United States.

ACKNOWLEDGMENTS

We thank Drs. Hong Lou, Vinay Sharma, and Sangeetha Hareendran, (NICHD) for helpful discussions of the manuscript.

- Cheng, Y., Li, Z., Kardami, E., and Loh, Y. P. (2016b). Neuroprotective effects of LMW and HMW FGF2 against amyloid beta toxicity in primary cultured hippocampal neurons. *Neurosci. Lett.* 632, 109–113. doi: 10.1016/j.neulet.2016. 08.031
- Cheng, Y., Rodriguiz, R. M., Murthy, S. R., Senatorov, V., Thouennon, E., Cawley, N. X., et al. (2015). Neurotrophic factor-alpha1 prevents stressinduced depression through enhancement of neurogenesis and is activated by rosiglitazone. *Mol. Psychiatry* 20, 744–754. doi: 10.1038/mp.2014.136
- Cong, L., Cheng, Y., Cawley, N. X., Murthy, S. R., and Loh, Y. P. (2017). A novel single nucleotide T980C polymorphism in the human carboxypeptidase E gene results in loss of neuroprotective function. *PLoS One* 12:e0170169. doi: 10.1371/ journal.pone.0170169
- Cool, D. R., Normant, E., Shen, F., Chen, H. C., Pannell, L., Zhang, Y., et al. (1997). Carboxypeptidase E is a regulated secretory pathway sorting receptor: genetic obliteration leads to endocrine disorders in Cpe(fat) mice. *Cell* 88, 73–83. doi: 10.1016/S0092-8674(00)81860-7
- Coulon, M., Wellman, C. L., Marjara, I. S., Janczak, A. M., and Zanella, A. J. (2013). Early adverse experience alters dendritic spine density and gene expression in prefrontal cortex and hippocampus in lambs. *Psychoneuroendocrinology* 38, 1112–1121. doi: 10.1016/j.psyneuen.2012.10.018
- Deiss, L. P., Feinstein, E., Berissi, H., Cohen, O., and Kimchi, A. (1995). Identification of a novel serine/threonine kinase and a novel 15-kD protein as

potential mediators of the gamma interferon-induced cell death. *Genes Dev.* 9, 15–30. doi: 10.1101/gad.9.1.15

- Evans, S. J., Choudary, P. V., Neal, C. R., Li, J. Z., Vawter, M. P., Tomita, H., et al. (2004). Dysregulation of the fibroblast growth factor system in major depression. *Proc. Natl. Acad. Sci. U.S.A.* 101, 15506–15511. doi: 10.1073/pnas. 0406788101
- Fan, S., Gao, X., Chen, P., and Li, X. (2018). Carboxypeptidase E-DeltaN promotes migration, invasion and epithelial-mesenchymal transition of human osteosarcoma cell lines through the Wnt/beta-catenin pathway. *Biochem. Cell Biol.* doi: 10.1139/bcb-2018-0236 [Epub ahead of print].
- Feinstein, E., Druck, T., Kastury, K., Berissi, H., Goodart, S. A., Overhauser, J., et al. (1995). Assignment of DAP1 and DAPK-genes that positively mediate programmed cell death triggered by IFN-gamma-to chromosome regions 5p12.2 and 9q34.1, respectively. *Genomics* 29, 305–307. doi: 10.1006/geno.1995. 1255
- Fricker, L. D. (1988). Carboxypeptidase E. Annu. Rev. Physiol. 50, 309–321. doi: 10.1146/annurev.ph.50.030188.001521
- Fricker, L. D., and Devi, L. (1993). Posttranslational processing of carboxypeptidase E, a neuropeptide-processing enzyme, in AtT-20 cells and bovine pituitary secretory granules. J. Neurochem. 61, 1404–1415. doi: 10.1111/j.1471-4159. 1993.tb13634.x
- Fricker, L. D., and Snyder, S. H. (1983). Purification and characterization of enkephalin convertase, an enkephalin-synthesizing carboxypeptidase. J. Biol. Chem. 258, 10950–10955.
- Hook, V. Y., Eiden, L. E., and Brownstein, M. J. (1982). A carboxypeptidase processing enzyme for enkephalin precursors. *Nature* 295, 341–342. doi: 10. 1038/295341a0
- Hook, V. Y., Mezey, E., Fricker, L. D., Pruss, R. M., Siegel, R. E., and Brownstein, M. J. (1985). Immunochemical characterization of carboxypeptidase B-like peptide-hormone-processing enzyme. *Proc. Natl. Acad. Sci. U.S.A.* 82, 4745–4749. doi: 10.1073/pnas.82.14.4745
- Huang, S. M., Mishina, Y. M., Liu, S., Cheung, A., Stegmeier, F., Michaud, G. A., et al. (2009). Tankyrase inhibition stabilizes axin and antagonizes Wnt signalling. *Nature* 461, 614–620. doi: 10.1038/nature08356
- Ibrahim, S. M., Mix, E., Bottcher, T., Koczan, D., Gold, R., Rolfs, A., et al. (2001). Gene expression profiling of the nervous system in murine experimental autoimmune encephalomyelitis. *Brain* 124, 1927–1938. doi: 10.1093/brain/124. 10.1927
- Israsena, N., Hu, M., Fu, W., Kan, L., and Kessler, J. A. (2004). The presence of FGF2 signaling determines whether beta-catenin exerts effects on proliferation or neuronal differentiation of neural stem cells. *Dev. Biol.* 268, 220–231. doi: 10.1016/j.ydbio.2003.12.024
- Ji, L., Wu, H. T., Qin, X. Y., and Lan, R. (2017). Dissecting carboxypeptidase E: properties, functions and pathophysiological roles in disease. *Endocr. Connect.* 6, R18–R38. doi: 10.1530/EC-17-0020
- Jin, K., Graham, S. H., Nagayama, T., Goldsmith, P. C., Greenberg, D. A., Zhou, A., et al. (2001). Altered expression of the neuropeptide-processing enzyme carboxypeptidase E in the rat brain after global ischemia. *J. Cereb. Blood Flow Metab.* 21, 1422–1429. doi: 10.1097/00004647-200112000-00006
- Jung, Y. K., Kunczt, C. J., Pearson, R. K., Dixon, J. E., and Fricker, L. D. (1991). Structural characterization of the rat carboxypeptidase-E gene. *Mol. Endocrinol.* 5, 1257–1268. doi: 10.1210/mend-5-9-1257
- Koks, S., Luuk, H., Nelovkov, A., Areda, T., and Vasar, E. (2004). A screen for genes induced in the amygdaloid area during cat odor exposure. *Genes Brain Behav.* 3, 80–89. doi: 10.1046/j.1601-183x.2003.00047.x
- Koshimizu, H., Senatorov, V., Loh, Y. P., and Gozes, I. (2009). Neuroprotective protein and carboxypeptidase E. J. Mol. Neurosci. 39, 1–8. doi: 10.1007/s12031-008-9164-5
- Lee, C., Hu, J., Ralls, S., Kitamura, T., Loh, Y. P., Yang, Y., et al. (2012). The molecular profiles of neural stem cell niche in the adult subventricular zone. *PLoS One* 7:e50501. doi: 10.1371/journal.pone.0050501
- Levy-Strumpf, N., and Kimchi, A. (1998). Death associated proteins (DAPs): from gene identification to the analysis of their apoptotic and tumor suppressive functions. *Oncogene* 17, 3331–3340. doi: 10.1038/sj.onc.1202588
- Liang, C., Carrel, D., Omelchenko, A., Kim, H., Patel, A., Fanget, I., et al. (2018). Cortical neuron migration and dendrite morphology are regulated by carboxypeptidase E. *Cereb. Cortex* doi: 10.1093/cercor/bhy155 [Epub ahead of print].

- Lou, H., Kim, S. K., Zaitsev, E., Snell, C. R., Lu, B., and Loh, Y. P. (2005). Sorting and activity-dependent secretion of BDNF require interaction of a specific motif with the sorting receptor Carboxypeptidase E. *Neuron* 45, 245–255. doi: 10.1016/j.neuron.2004.12.037
- Lou, H., Park, J. J., Cawley, N. X., Sarcon, A., Sun, L., Adams, T., et al. (2010). Carboxypeptidase E cytoplasmic tail mediates localization of synaptic vesicles to the pre-active zone in hypothalamic pre-synaptic terminals. *J. Neurochem.* 114, 886–896. doi: 10.1111/j.1471-4159.2010.06820.x
- Lynch, D. R., Braas, K. M., Hutton, J. C., and Snyder, S. H. (1990). Carboxypeptidase E (CPE): immunocytochemical localization in the rat central nervous system and pituitary gland. *J. Neurosci.* 10, 1592–1599. doi: 10.1523/ JNEUROSCI.10-05-01592.1990
- MacCumber, M. W., Snyder, S. H., and Ross, C. A. (1990). Carboxypeptidase E (enkephalin convertase): mRNA distribution in rat brain by in situ hybridization. J. Neurosci. 10, 2850–2860. doi: 10.1523/JNEUROSCI.10-08-02850.1990
- Martinez, A., Otal, R., and Soriano Garcia, E. (2004). [Ephrins, neuronal development and plasticity]. *Rev. Neurol.* 38, 647–655. doi: 10.33588/rn.3807. 2004068
- Martinez, A., and Soriano, E. (2005). Functions of ephrin/Eph interactions in the development of the nervous system: emphasis on the hippocampal system. *Brain Res. Brain Res. Rev.* 49, 211–226. doi: 10.1016/j.brainresrev.2005.02.001
- Mazon Pelaez, I., Vogler, S., Strauss, U., Wernhoff, P., Pahnke, J., Brockmann, G., et al. (2005). Identification of quantitative trait loci controlling cortical motor evoked potentials in experimental autoimmune encephalomyelitis: correlation with incidence, onset and severity of disease. *Hum. Mol. Genet.* 14, 1977–1989. doi: 10.1093/hmg/ddi203
- Murphy, M., Drago, J., and Bartlett, P. F. (1990). Fibroblast growth factor stimulates the proliferation and differentiation of neural precursor cells in vitro. *J. Neurosci. Res.* 25, 463–475. doi: 10.1002/jnr.490250404
- Murthy, S. R., Thouennon, E., Li, W. S., Cheng, Y., Bhupatkar, J., Cawley, N. X., et al. (2013). Carboxypeptidase E protects hippocampal neurons during stress in male mice by up-regulating prosurvival BCL2 protein expression. *Endocrinology* 154, 3284–3293. doi: 10.1210/en.2013-1118
- Park, J. J., Cawley, N. X., and Loh, Y. P. (2008). A bi-directional carboxypeptidase E-driven transport mechanism controls BDNF vesicle homeostasis in hippocampal neurons. *Mol. Cell. Neurosci.* 39, 63–73. doi: 10.1016/j.mcn.2008. 05.016
- Pla, V., Paco, S., Ghezali, G., Ciria, V., Pozas, E., Ferrer, I., et al. (2013). Secretory sorting receptors carboxypeptidase E and secretogranin III in amyloid beta-associated neural degeneration in Alzheimer's disease. *Brain Pathol.* 23, 274–284. doi: 10.1111/j.1750-3639.2012.00644.x
- Qian, Y., Varlamov, O., and Fricker, L. D. (1999). Glu300 of rat carboxypeptidase E is essential for enzymatic activity but not substrate binding or routing to the regulated secretory pathway. J. Biol. Chem. 274, 11582–11586. doi: 10.1074/jbc. 274.17.11582
- Qin, X. Y., Cheng, Y., Murthy, S. R., Selvaraj, P., and Loh, Y. P. (2014). carboxypeptidase E-DeltaN, a neuroprotein transiently expressed during development protects embryonic neurons against glutamate neurotoxicity. *PLoS One* 9:e112996. doi: 10.1371/journal.pone.0112996
- Raballo, R., Rhee, J., Lyn-Cook, R., Leckman, J. F., Schwartz, M. L., and Vaccarino, F. M. (2000). Basic fibroblast growth factor (Fgf2) is necessary for cell proliferation and neurogenesis in the developing cerebral cortex. *J. Neurosci.* 20, 5012–5023. doi: 10.1523/JNEUROSCI.20-13-05012.2000
- Rodriguiz, R. M., Wilkins, J. J., Creson, T. K., Biswas, R., Berezniuk, I., Fricker, A. D., et al. (2013). Emergence of anxiety-like behaviours in depressive-like Cpe(fat/fat) mice. *Int. J. Neuropsychopharmacol.* 16, 1623–1634. doi: 10.1017/ S1461145713000059
- Russo, V. C., Gluckman, P. D., Feldman, E. L., and Werther, G. A. (2005). The insulin-like growth factor system and its pleiotropic functions in brain. *Endocr. Rev.* 26, 916–943. doi: 10.1210/er.2004-0024
- Selvaraj, P., Huang, J. S., Chen, A., Skalka, N., Rosin-Arbesfeld, R., and Loh, Y. P. (2015). Neurotrophic factor-alpha1 modulates NGF-induced neurite outgrowth through interaction with Wnt-3a and Wnt-5a in PC12 cells and cortical neurons. *Mol. Cell. Neurosci.* 68, 222–233. doi: 10.1016/j.mcn.2015.08.005
- Selvaraj, P., Xiao, L., Lee, C., Murthy, S. R., Cawley, N. X., Lane, M., et al. (2017). Neurotrophic Factor-alpha1: a Key Wnt-beta-Catenin Dependent Anti-Proliferation Factor and ERK-Sox9 activated inducer of embryonic neural stem

cell differentiation to astrocytes in neurodevelopment. Stem Cells 35, 557-571. doi: 10.1002/stem.2511

- Skalka, N., Caspi, M., Caspi, E., Loh, Y. P., and Rosin-Arbesfeld, R. (2013). Carboxypeptidase E: a negative regulator of the canonical Wnt signaling pathway. Oncogene 32, 2836-2847. doi: 10.1038/onc.2012.308
- Song, L., and Fricker, L. (1995). Processing of procarboxypeptidase E into carboxypeptidase E occurs in secretory vesicles. J. Neurochem. 65, 444-453. doi: 10.1046/j.1471-4159.1995.65010444.x
- Stahl, S., Reinders, Y., Asan, E., Mothes, W., Conzelmann, E., Sickmann, A., et al. (2007). Proteomic analysis of cathepsin B- and L-deficient mouse brain lysosomes. Biochim. Biophys. Acta 1774, 1237-1246. doi: 10.1016/j.bbapap. 2007.07.004
- Strittmatter, S. M., Lynch, D. R., and Snyder, S. H. (1984). [3H]guanidinoethylmercaptosuccinic acid binding to tissue homogenates. Selective labeling of enkephalin convertase. J. Biol. Chem. 259, 11812-11817.
- Weber, S. J., Louis, R. B., Trombley, L., Bissette, G., Davies, P., and Davis, T. P. (1992). Metabolic half-life of somatostatin and peptidase activities are altered in Alzheimer's disease. J. Gerontol. 47, B18-B25.
- Woronowicz, A., Cawley, N. X., Chang, S. Y., Koshimizu, H., Phillips, A. W., Xiong, Z. G., et al. (2010). Carboxypeptidase E knockout mice exhibit abnormal dendritic arborization and spine morphology in central nervous system neurons. J. Neurosci. Res. 88, 64-72. doi: 10.1002/jnr.22174
- Woronowicz, A., Cawley, N. X., and Peng Loh, Y. (2012). Carbamazepine prevents hippocampal neurodegeneration in mice lacking the neuroprotective protein, Carboxypetidase E. Clin. Pharmacol. Biopharm. Suppl. 1:2.
- Woronowicz, A., Koshimizu, H., Chang, S. Y., Cawley, N. X., Hill, J. M., Rodriguiz, R. M., et al. (2008). Absence of carboxypeptidase E leads to adult hippocampal neuronal degeneration and memory deficits. Hippocampus 18, 1051-1063. doi: 10.1002/hipo.20462
- Xiao, L., Chang, S. Y., Xiong, Z. G., Selveraj, P., and Peng Loh, Y. (2017). Absence of Carboxypeptidase E/Neurotrophic Factor-Alpha1 in Knock-Out Mice Leads to Dysfunction of BDNF-TRKB Signaling in Hippocampus. J. Mol. Neurosci. 62, 79-87. doi: 10.1007/s12031-017-0914-0

- Xiao, L., Yang, X., Sharma, V. K., and Loh, Y. P. (2018). Cloning, gene regulation, and neuronal proliferation functions of novel N-terminal-truncated carboxypeptidase E/neurotrophic factor-alphal variants in embryonic mouse brain. FASEB J. 33, 808-820. doi: 10.1096/fj.201800359R
- Yang, X., Cong, L., Lou, H., and Loh, Y. P. (2017). A novel 40kDa CPE-∆N isoform promotes proliferation and invasion in pancreatic cancer cells. Cancer Res. 77, 1967-1967. doi: 10.1158/1538-7445.AM2017-1967
- Zechel, S., Werner, S., Unsicker, K., and von Bohlen und Halbach, O. (2010). Expression and functions of fibroblast growth factor 2 (FGF-2) in hippocampal formation. Neuroscientist 16, 357-373. doi: 10.1177/1073858410371513
- Zheng, M., Streck, R. D., Scott, R. E., Seidah, N. G., and Pintar, J. E. (1994). The developmental expression in rat of proteases furin, PC1, PC2, and carboxypeptidase E: implications for early maturation of proteolytic processing capacity. J. Neurosci. 14, 4656-4673. doi: 10.1523/JNEUROSCI.14-08-04656. 1994
- Zhou, A., Minami, M., Zhu, X., Bae, S., Minthorne, J., Lan, J., et al. (2004). Altered biosynthesis of neuropeptide processing enzyme carboxypeptidase E after brain ischemia: molecular mechanism and implication. J. Cereb. Blood Flow Metab. 24, 612-622. doi: 10.1097/01.WCB.0000118959.03453.17
- Zhu, X., Wu, K., Rife, L., Cawley, N. X., Brown, B., Adams, T., et al. (2005). Carboxypeptidase E is required for normal synaptic transmission from photoreceptors to the inner retina. J. Neurochem. 95, 1351-1362. doi: 10.1111/ j.1471-4159.2005.03460.x

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Xiao, Yang and Loh. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Xiao et al